

Structural base for the transfer of GPI-anchored proteins into fungal cell walls

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Zusammenfassung

Pilze, wie der einzellige Modellorganismus *Saccharomyces cerevisiae*, besitzen eine Zellwand aus Polysacchariden und Proteinen, die für lebensfähige und gesunde Zellen essentiell ist. Während die Synthese ihrer Komponenten entlang des sekretorischen Weges und an der Plasmamembran erfolgt, wird die korrekte Verarbeitung auf der Außenseite der Plasmamembran bewerkstelligt. Dies wird durch sogenannte Glykosidhydrolasen (GH), realisiert, welche die Spaltung und Neuverknüpfung von glykosidischen Bindungen katalysieren. In *S. cerevisiae* und dem Humanpathogen *Candida albicans* wurde gezeigt, dass Mitglieder der GH76-Familie (*Dfg5*-Subfamilie) für den Einbau von GPI-verankerten Proteinen in die Zellwand verantwortlich sind, ein Vorgang, der für Hefen überlebenswichtig ist. Obwohl bakterielle Homologe dieser Klasse bereits beschrieben wurden, sind die zugrunde liegenden Mechanismen der pilzlichen Vertreter, trotz ihres außergewöhnlichen Potenzials als Wirkstoff Zielstruktur, noch immer unbekannt.

Um diese Lücke zu schließen, wurde die GH76-Familie zunächst einer phylogenetischen Analyse unterzogen, die Einblicke in ihren multifunktionalen Charakter gewährte. Die genaue Rolle der *Dfg5*-Proteine konnte durch Struktur- und Funktionsanalysen des Orthologs *CtDfg5* aus dem thermophilen Schimmelpilz *Chaetomium thermophilum* erklärt werden. Seine mit atomarer Auflösung bestimmte Kristallstruktur zeigte, dass die Faltung und das aktive Zentrum zwischen Pilz- und Bakterienhomologen konserviert ist, jedoch war es nicht möglich die bekannte α 1,6-Mannanaseaktivität *in vitro* zu zeigen. Stattdessen konnte die GPI-Glykanstruktur (Man α 1,2-Man α 1,6-Man α 1,4-GlcN) innerhalb der Bindetasche von *CtDfg5* durch hochmolekulares Zuckerfragmentscreening assembliert werden. Dies ermöglichte nicht nur einen detaillierten Blick auf das tatsächliche Substrat der *Dfg5*-Proteine, sondern auch erste experimentell abgeleitete Erkenntnisse über die dreidimensionale Architektur des GPI-Ankerglykans. Zusammen mit der Struktur eines potentiellen Akzeptormoleküls konnte der von *Dfg5*-Proteinen katalysierte Lipid-zur-Zellwand-Transfermechanismus abgeleitet werden. Darüber hinaus ermöglichten die strukturellen Einblicke in die Substratkoordinierung eine mögliche Lenkungsfunktion verschiedener GPI-Modifikationen, mit deren Hilfe die endgültige Lokalisation von GPI-verankerten Proteinen kodiert wird. Außerdem konnte durch Docking eine Leitstruktur (FP-1) identifiziert werden, das im aktiven Zentrum von *CtDfg5* bindet. FP-1 zeigt spezifische Effekte bei millimolaren Konzentrationen in Bezug auf die Lebensfähigkeit von *S. cerevisiae*. Schließlich wurde, durch die strukturelle Charakterisierung eines pilzlichen Homologs aus einer weiteren GH76-Unterfamilie, α 1,6-Mannobiose als zentrales Element von GH76-Substraten identifiziert.

Summary

Fungi, such as the unicellular model organism *Saccharomyces cerevisiae*, possess a thick cell wall composed of polysaccharides and proteins, which is essential for viable and healthy cells. While the mere synthesis of its components happens along the secretory pathway and at the plasma membrane, the correct processing is established on the exterior side of the plasma membrane. This is realized by a set of enzymes, called glycoside hydrolases (GH), which act on the hydrolysis and rearrangement of glycosidic bonds. In *S. cerevisiae* and the human pathogen *Candida albicans*, it has been shown that members of the GH76 family (*Dfg5*-subfamily) carry out the incorporation of GPI-anchored proteins into the cell wall, which is essential for these organisms. Although bacterial homologs of that class were already described, our understanding of the fungal counterparts and their underlying mechanism with its exceptional potential as a drug target still lack behind.

In order to fill this gap, the GH76 family has been subjected initially to phylogenetic analysis providing insights into its multifunctional character with up to ten different subfamilies. The exact role of *Dfg5*-proteins could be explained by an in-depth structural and functional analysis of one of its members, *CtDfg5*, from the thermophilic mold *Chaetomium thermophilum*. Its crystal structure determined at atomic resolution showed that the overall fold and the active site motif is shared between fungal and bacterial homologs, however an annotated function as α 1,6-mannanases could not be shown *in vitro*. Instead it was possible to reassemble the GPI-core glycan structure (Man α 1,2-Man α 1,6-Man α 1,4-GlcN) within the substrate-binding pocket of *CtDfg5* by screening crystals with high molar sugar-fragments. This did not only provide a detailed view on the true substrate of *Dfg5*-proteins, but also first experimentally derived insights into the three-dimensional architecture of the GPI-anchor glycan. Together with the complex structure of a putative acceptor molecule, a lipid-to-wall transfer mechanism catalyzed by *Dfg5*-proteins could be derived. Moreover, the structural insights suggested a possible way of using different GPI-modifications as a coding system to determine the final localization of GPI-anchored proteins. Furthermore, structure-based docking of a commercially available lead library helped to identify a small molecule (FP-1) that binds to the active site of *CtDfg5*. FP-1 shows specific effects at milli molar concentrations in terms of the viability of the model organism *S. cerevisiae*, assuming a high potential for further drug development. Finally, another fungal homolog from a so far uncharacterized subfamily showed that all GH76 family members recognize α 1,6-mannobiose as a central element of cognate substrates.

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1 Introduction

1.1 The fungal cell wall

The cell wall describes an envelope that sits just behind the plasma membrane, surrounding the whole cell like a protective armor. It is essential for the cell's morphology, its osmotic integrity, and it facilitates resistance to external harmful aggressors. Further roles of cell walls are sensing of the cell's environment, adhesion to surfaces, biofilm formation, and cell-to-cell communication, thus making it critical for pathogenicity. While absent in animals, cell walls appear in many different forms, as well as in many different classes of organisms throughout nature. For example, microalgae known as diatoms have developed a cell wall that basically consists of silicate, while bacteria have a cell wall composed of linear glycans connected by oligo-peptides, thus called peptidoglycan, whereas plants possess a cell wall made of cross-linked cellulose and other glycans (Drum and Gordon, 2003; Lampugnani et al., 2018; Vollmer et al., 2008).

Fungi possess a cell wall composed of different types of glucans, chitin and proteins (Free, 2013). The wall's outstanding importance for the organism becomes obvious when looking at the sheer mass of ~1200 genes in the genome of *Saccharomyces cerevisiae* (i.e. one fifth of all genes), which were found to be involved in its synthesis, maturation, and general maintenance (De Groot et al., 2001). A complex interplay of all the different factors enables a highly dynamic adaptation to the environment and the growing cell, while its osmotic integrity must be maintained at all time. Errors in wall assembly have fatal consequences for the fungal organism due to the high internal turgor pressure of up to 10 MPa, which would lead to spontaneous cell lysis (Money, 2001). This, together with the absence of the involved constituents in humans, is the reason, why factors involved in the biosynthesis of cell walls provide excellent targets for antifungal drugs.

In principal, fungal cell walls (FCW) from all described ascomycota are built in a similar fashion with species-specific adaptations. For the fundamental understanding, the different aspects of the FCW will be described for the model organism *S. cerevisiae* in the following. However, important differences especially to filamentous fungi will be pointed out.

1.1.1 The structural organization of the fungal cell wall

The CW of the yeast *S. cerevisiae* is responsible for up to 30% of the cell's dry weight and distributed over a 100-200 nm thick structure forming the outermost layer of the organism

(Aguilar-Uscanga and François, 2003; Dupres et al., 2010; Yamaguchi et al., 2011; Yin et al., 2007). Electron micrographs revealed a bipartite architecture with an inner layer comprised of glucans and chitin, which is covered by an outer layer composed of heavily glycosylated proteins (Figure 1) (Baba et al., 1989; Hagen et al., 2004; Kapteyn et al., 1999; Kopecka et al., 1974; Osumi, 1998; Zlotnik et al., 1984). This composition is species-specific and can change upon developmental stage changes, within different phases of the cell cycle, and in response to extracellular stimuli such as nutrients, osmotic stressors or drugs (Hopke et al., 2018).

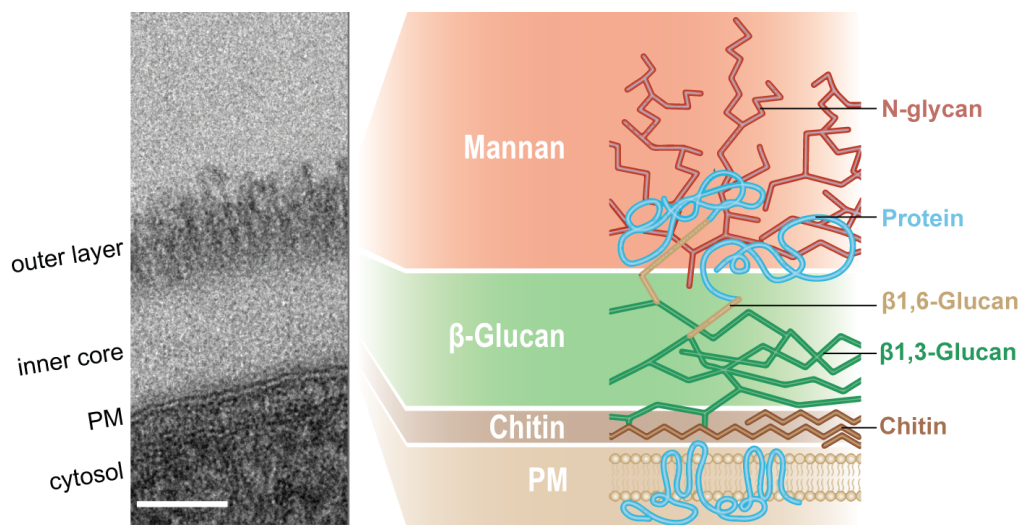


Figure 1: Architecture of the yeast cell wall. An electron microscopic picture of the fungal cell wall from *S. cerevisiae* is shown on the left. The respective structures found in the cell wall are schematically shown on the right (modified from Erwig and Gow, 2016). Scale bar=100 nm.

In yeast, the polysaccharides of the inner core consist of three different types (Kollár et al., 1995; Kopecka et al., 1974). As shown in the schematic representations linear chitin is the inner layer of the yeast cell wall (Figure 1). It is assembled of β 1,4-linked N-acetylglucosamine (GlcNAc) and accounts for only 1-2% of the CW's dry weight, while filamentous fungi such as *Aspergillus fumigatus* possess up to 15% (Gastebois et al., 2009; Hu et al., 2007; Lesage and Bussey, 2006). The degree of polymerization ranges from 110 to 190 residues. In unstressed cells most chitin is found in the bud tip and the scar (Holan et al., 2004). Chitin at the bud neck comes as a chitin ring that stops cell wall growth and a chitin-based septum, which finally leads to the separation of mother and daughter cells (Lippincott and Li, 1998; Molano et al., 1980; Schmidt et al., 2002; Shaw et al., 1991). It exists in three forms within the CW: (i.) as a free form, (ii.) bound to β 1,3-glucan and (iii.) linked to β 1,6-glucan (Cabib, 2009; Cabib and Durán, 2005). Chitin is a stiff molecule that facilitates

rigidity to the cell wall (Kang et al., 1984). This explains elevated levels in the lateral cell wall upon cell wall related stress to compensate for loss of integrity, which can be visualized by the chitin-binding fluorophore Calcofluor white (Kapteyn et al., 1997; Ram and Klis, 2006; Ram et al., 1994).

The most abundant molecule in the CW of *S. cerevisiae* is β 1,3-glucan, which accounts for half of the CW dry mass and therefore makes up most of the inner electron transparent layer (Figure 1) (Fleet, 1991). The mature molecule consists of up to 1500 linear β 1,3-linked glucose units, which is generally found as a major constituent of the fungal cell wall in all ascomycota (Free, 2013; Manners et al., 1973). β 1,3-glucan is an acid and alkali insoluble molecule with β 1,6-side branches that interconnect the linear molecules. Due to the high abundance and the extensive cross-linking, β 1,3-glucan is not only the main component responsible for the resilience of the cell wall, but also the basis for its flexibility during remodeling. Another type of glucan from the inner core of the yeast cell wall is β 1,6-glucan, which accounts for 5% of the dry weight and is composed of about 140 residues. While β 1,6-glucan is responsible for the attachment of cell wall proteins (CWP) in *S. cerevisiae* and *Candida albicans*, it is supposed to be absent in filamentous fungi like *A. fumigatus* and *Neurospora crassa*. Such fungi contain other types of polysaccharides described as β 1,3/1,4-mixed glucan, α 1,3-glucan, and galactomannans (Fontaine et al., 2000; Gastebois et al., 2009; Maddi and Free, 2010).

The outer, brush-like layer of the cell wall from *S. cerevisiae* can be digested by proteases and was found to consist of proteins highly decorated with N- and O-glycosylations (Kollár et al., 1997; Zlotnik et al., 1984). These posttranslational modifications are mannose-oligosaccharides and the eponymous element of mannoproteins forming the fibrillar structures (Figure 1). While polysaccharides are the main players for the structural framework of the cell wall, mannoproteins facilitate its functional features. There are three types of CWP: The first group contains proteins with enzymatic activities contributing to the synthesis and maintenance of the cell wall by their ability to hydrolyse and reconnect glycans. The second group has no enzymatic functions. They facilitate the capability of adhesion and flocculation in addition to further cross-linking the cell wall glucans (Dranginis et al., 2007; Goossens and Willaert, 2010; Klis et al., 2006, 2010; Kraushaar et al., 2015; Veelders and Essen, 2012; Yin et al., 2005). The third group is composed of mechanosensors that are able to detect osmotic stress, which is transmitted via a membrane helix to the cytosol to activate the cell wall integrity pathway (Rodicio and Heinisch, 2010).

Crucial for a functional cell wall is the covalent attachment of CWPs to the glycan matrix. One class is attached via an alkali labile ester bond of amino acid side chains to β 1,3-glucan. These proteins contain internal repeats and are therefore referred to as PIR-CWPs (Ecker et al., 2006). Due to their attachment, PIR-proteins can be released from the CW upon β 1,3-glucanase treatment (De Groot et al., 2005). Another way of CWP-attachment is based on the C-terminal posttranslational modification called the *glycosylphosphatidylinositol* (GPI)-anchor, which will be explained later in detail. A part of the GPI-glycan is linked to the β 1,6-glucan in *S. cerevisiae* cell walls (Gonzalez et al., 2009). The linkage is sensitive to hydrogen fluoride (HF)/pyridine-treatment due to the hydrolysis of the phosphodiester between the ethanolamine phosphate (EtN-P) of the GPI-remnant and the peptide-bond linked C-terminal amino acid of the GPI-CWP (Yin et al., 2005). Another, indirect linkage to the CW is established by disulfide bonds, as some proteins were found to be extractable from the glycan-meshwork by reducing sulfhydryl reagents (Cappellaro et al., 1998; Orlean et al., 1986).

1.1.2 Synthesis and maturation of the fungal cell wall

The synthesis of the fungal cell wall is a multi-layered process. It includes the production of core polysaccharides at the plasma membrane, extracellular glycan remodeling, and the synthesis of mannoproteins along the secretory pathway.

The biosynthesis of β 1,3-glucan and chitin is a well characterized process, although structural studies and a precise mechanistical understanding are still lacking. Both polysaccharides are synthesized by transmembrane proteins, which utilize nucleotide diphosphate sugars as a substrate for the vectorial synthesis of the polysaccharide precursor chains through the membrane into the extracellular space (Cabib et al., 1983; Duran et al., 1975; Shematek et al., 1980). In yeast, three proteins of the Fks family from the glycosyl transferase (GT) 48 class are involved in the β 1,3-glucan synthesis (Bowman and Free, 2006). The functional complex requires the binding of a regulatory subunit, the lipid-bound small G-protein Rho1 that is activated upon GTP-binding (Drgonová et al., 1996; Inoue et al., 1999; Mazur and Baginsky, 1996; Mol et al., 1994). The activated complex utilizes UDP-Glc as sugar donor, which is transferred to the growing glucan chain to result in precursor chains of 60-80mers (Douglas et al., 1994; Shematek et al., 1980). Although having been described to possess redundant functions with a lethal double-mutant phenotype, Fks1 is described as the major synthase during vegetative growth, whose deletion results in a 75% reduction of synthesized β 1,3-

glucan, while Fks2 is implicated in glucan-synthesis mainly under starvation, during mating and sporulation (Inoue et al., 1995; Mazur et al., 1995). The role of Fks3 is not entirely clear. It seems to be involved in ascospore wall formation, as its deletion results in abnormal spore morphology (Ishihara et al., 2007). In the filamentous pathogen *A. fumigatus* only one essential *FKS* gene is present (Mouyna et al., 2004). In contrast to the glucan synthases, the nature of chitin synthases (Chs) is more diverse. Two families subdivided into seven subclasses are described in literature, of which some fungi possess more than 20 *CHS* genes, while others contain just one (Lenardon et al., 2010; Morozov and Likhoshway, 2016). *S. cerevisiae* possesses three genes for Chs-proteins that belong to the GT2 family. Chitin synthases use UDP-GlcNAc as a substrate to produce up to 170mers *in vitro* (Kang et al., 1984; Orlean, 1987). The major synthase in yeast is Chs3, which produces more than 90% of chitin in vegetative cells with Chs2 accounting for ~5%, while no measurable contribution of chitin production could be determined for Chs1 *in vivo* (Shaw et al., 1991; Valdivieso et al., 1991).

In contrast to chitin and β 1,3-glucan, the synthesis of β 1,6-glucan in yeasts remains elusive. So far, no synthase could be identified to be responsible for the production of β 1,6-glucan. The synthesis might be associated with the membrane as well, as crude membrane extracts from *S. cerevisiae* are described to show β 1,6-glucan production *in vitro* (Vink et al., 2004). Mutants that possess a defective β 1,6-glucan synthesis can be identified by their increasing K1 killer toxin resistance (Kre). The toxin is a secreted protein that uses β 1,6-glucan as an acceptor and activates potassium efflux causing cell death (Sesti et al., 2001). Accordingly, a reduction of the acceptor is accompanied by an increase in resistance (Bussey, 1991; Hutchins and Bussey, 1983). Most of the discovered Kre-proteins, such as Kre5, Kre6, and Kre9, are proteins localized in the secretory pathway and possess β 1,6-synthesis rates that are reduced up to 95% compared to wild type. However, a direct catalytic role is unlikely, as no intracellular β 1,6-glucan could be observed so far and their role is more likely related to protein quality control (Meaden et al., 2015; Montijn et al., 1999; Shahinian and Bussey, 2000). Extracellular Kre-proteins do not seem to be directly involved in synthesis as well. Kre1 is a GPI-anchored protein (GPI-AP) with unknown function, which makes about 40% of β 1,6-glucan compared to wild type upon deletion (Boone et al., 1990; Breinig et al., 2004). Mutants in the β 1,3-glucan synthase Fks1 possess a K1 killer toxin phenotype accompanied by a modest reduction in β 1,6-glucan. This phenomenon is explained by a reduced availability of its acceptor β 1,3-glucan, although a dual function of Fks1 and Fks2 in the biosynthesis of both types of glucans is discussed (Dijkgraaf et al., 2002). Evidence for a membrane

associated synthesis is given by a recently characterized protein from the GT2 family, which was described to be a membrane embedded β 1,6-glucan synthase found in the bacterium *Mycoplasma agalactiae* (Gaurivaud et al., 2016). Cell walls from filamentous fungi such as *A. fumigatus* lack β 1,6-glucan, although this was recently questioned (Kang et al., 2018).

In order to generate the extensively cross-linked meshwork of polysaccharides and proteins of the fungal cell wall described above, the elongation and processing of the polysaccharide precursors, as well as the incorporation of proteins into the FCW is carried out extracellularly by carbohydrate active enzymes with hydrolyzing (and associated transglycosylation) activities. The precise temporal sequence is not entirely clear. Attempts of elucidating the sequence of events of the cell wall construction have been performed on yeast spheroplasts, which possess the ability to regenerate the removed cell wall. The regeneration process suggested β 1,3-glucan as the starting component, followed by the incorporation of β 1,6-glucan, mannoproteins and lastly chitin (Figure 2) (Kapteyn et al., 1997; Kreger and Kopecka, 1976; Lu et al., 1995; Roh et al., 2002; Shaw et al., 1991).

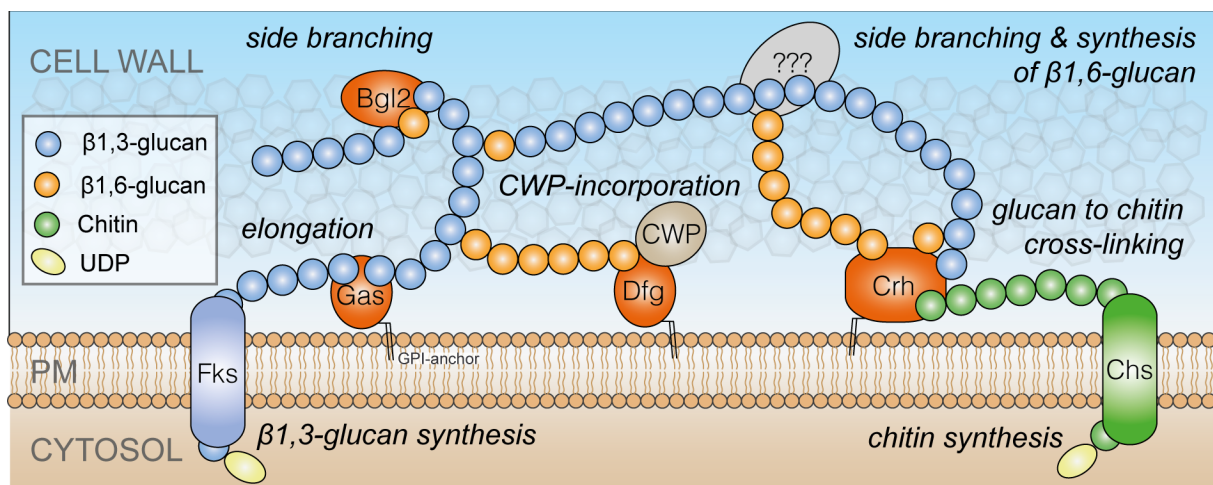


Figure 2: Schematic overview of the fungal cell wall biogenesis. Polysaccharides are produced at the plasma membrane (PM) by vectorial synthesis in the extracellular space. β 1,3-glucan precursors are elongated by proteins from the Gas-family (GH72) and side branched by Bgl2-proteins (GH17). How the synthesis of β 1,6-glucan happens is still unknown. In *S. cerevisiae*, CWPs are attached to the non-reducing end of β 1,6-glucan by proteins from the *Dfg5*-family (GH76). Chitin cross-linking is established with variable cross-link types by Crh-proteins (GH16) (figure adapted from Cabib and Arroyo, 2013 and Teparić and Mrša, 2013).

All modifications require the cleavage of glycosidic bonds, a reaction catalyzed by glycoside hydrolases (GHs). To date, 165 different families are listed in the carbohydrate active enzyme database, of which 115 families are curator approved (www.cazy.org, June 2019) (Lombard et al., 2014). Despite this diversity, there are merely two mechanistic ways GHs follow to

hydrolyze glycosidic bonds (Figure 3). One directly displaces the leaving group by water, which leads to an inversion of the anomeric configuration, thus called *inverting* glycosidases (Figure 3A). The other type follows a *retaining* mechanism utilizing a double-displacement reaction including a covalent glycosyl-enzyme intermediate (Figure 3B) (McCarter and Withers, 1994). Both classes possess a catalytic pair of carboxylate side chains, which are further apart in inverting (~ 9 Å) than in retaining glycosidases (~ 5 Å) to allow the direct intervention of a water molecule (Wang et al., 1994). While one of the residues acts as a general acid and the other one as a general base in inverting enzymes, retaining glycosidases utilize one of the carboxylates as the nucleophile and leaving group and the other residue as a general acid/base. However, both utilize an oxocarbenium ion-like transition state (McCarter and Withers, 1994). In the transglycosylase class of retaining GH enzymes, the intercepting molecule is not water, but an acceptor molecule (usually another carbohydrate alcohol), resulting in a new glycosidic bond (Grout and Vici, 1998).

Although sugars can adapt a broad spectrum of different conformations classified as chair (C), half-chair (H), boat (B), skew-boat (S), and envelope (E), just a couple of them favor the oxocarbenium ion transition state and are therefore utilized during the conformational distortion required for enzymatic catalysis (highlighted states in the Stoddart diagram shown in Figure 3C) (Raich et al., 2016). The Stoddart diagram is used to visualize the possible conformations of a pyranose ring, from which the distortions during catalysis (called the *catalytic itinerary*) at the -1 subsite moiety can be presented (Stoddart, 1971). The conventionally used sugar-binding subsite nomenclature labels the subsites from $-n$ to $+n$ (Davies et al., 1997). While $-n$ labels the non-reducing end with increasingly negative values towards the non-reducing end and away from the cleavage site, $+n$ accounts for moieties at the reducing end of the sugars, with increasing values away from the cleavage site towards the reducing terminus. The cleavage occurs between subsites -1 and +1.

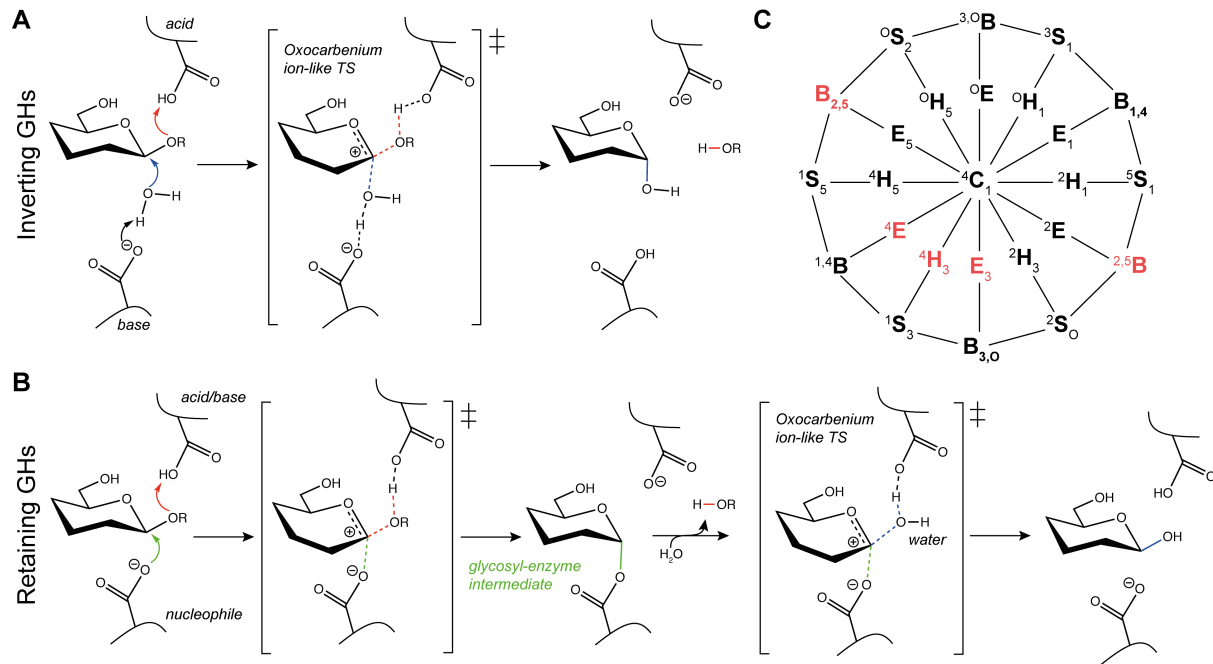


Figure 3: General mechanisms of glycoside hydrolases. The mechanisms of *inverting* (A) and *retaining* (B) GHs are shown with the characteristic oxocarbenium ion-like TS. **C** The Stoddart diagram centered at the 4C_1 conformation is shown. Conformations highlighted in red show the preferred distortions of the pyranoses to stabilize the oxocarbenium ion-like TS during catalysis (adapted from Raich et al., 2016).

While genetic analyses with functional predictions are numerous, biochemical *in vitro* analysis combined with structural characterizations of cell wall remodeling enzymes have been performed just for a couple of proteins (see Figure 2 and Figure 4). In yeasts, GPI-APs of the GH72 family, called Gas-proteins, have been described as β 1,3-glycosyltransferases and are thought to be responsible for the β 1,3-glucan elongation, as they are capable to cleave a donor β 1,3-glucan chain and subsequently attach it to the non-reducing end of an β 1,3-acceptor (Hurtado-Guerrero et al., 2009; Maz  n et al., 2011). Recently, a dual function of GH72 proteins has been proposed, contributing elongation and cross-linking activities to homologs from yeast and *A. fumigatus* (Aimanianda et al., 2017). There, it is suggested that GH72 and GH17 members act together on the side-branching of β 1,3-glucans. GH17 proteins, such as the Bgl2 homolog from *Rhizomucor miehei*, are described as β 1,3-glucan branching enzymes by exhibiting an endo-glycosidase activity coupled with the subsequent transfer of the reducing end to the 6-hydroxy group of an acceptor β 1,3-glucan chain, thereby forming a β 1,6-side branch (Goldman et al., 1995; Qin et al., 2015). Controversially, genetic studies in *N. crassa* suggested the GH72 family not to be involved in the β 1,3-glucan processing, but to act on the incorporation of CWPs into the cell wall by cross-linking the N-linked galactomannan to lichenin (Ao and Free, 2017; Kar et al., 2019). Crh-proteins (GH16-family

members) are responsible for the chitin capping of glucan-chains by connecting the reducing end of chitin with the non-reducing end of a glucan, a notion that is strongly supported by a recently deposited crystal structure of Crh5 (PDB: 6IBW) from *A. fumigatus* in its glycosyl-enzyme intermediate state with bound chitin (Cabib, 2009).

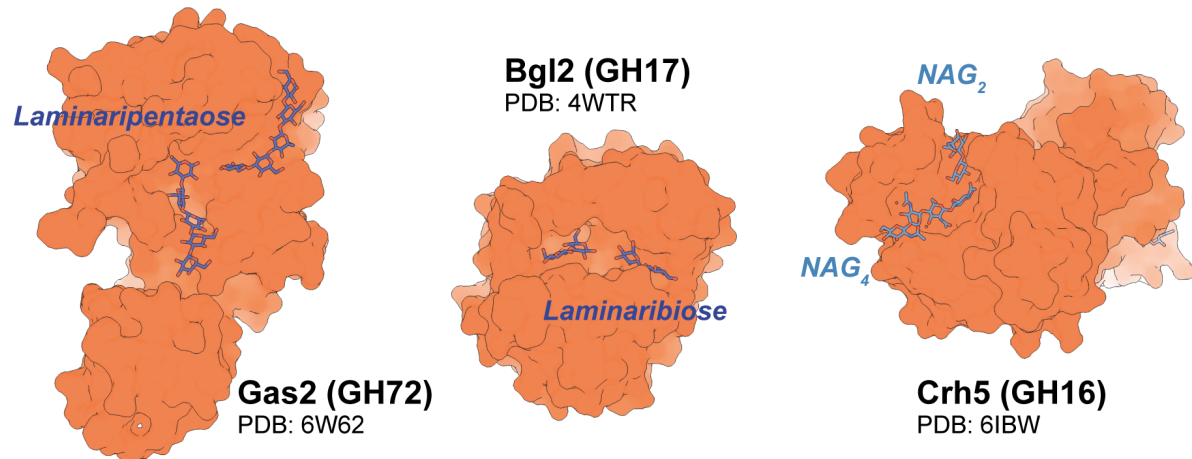


Figure 4: Overview of fungal GH-proteins involved in cell wall biosynthesis. Surface representation is shown for the structurally described GH-proteins in complex with their ligands, which are displayed as sticks. Laminaribiose/pentaose= β 1,3-linked glucose (di-,pentamer), NAG_{2/4}= β 1,4-linked N-acetyl glucosamine (di-/tetramer).

1.1.4 Incorporation of proteins into the cell wall

A further essential step in the synthesis of a functional fungal cell wall is the incorporation of cell wall proteins (CWP). As described above, PIR-CWP are connected via an alkali-sensitive linkage to the cell wall. It was shown for Pir4 from *S. cerevisiae* that a specific glutamine (Q74) of the repetitive sequence Q₆₉IGD₇₂GQ₇₄VQ₇₆ is identified as a glutamic acid upon hydrolytic release from the cell wall, while Pir4 purified from the medium still contained the glutamine (Ecker et al., 2006). It is further suggested that this conversion and the resulting ester linkage formation between the γ -carboxyl group of the glutamic acid and the hydroxyl group of a β 1,3-glucan glucose happens in an autocatalytic manner assisted by the surrounding glutamines and the aspartate, as their exchange led to a loss of cell wall attachment.

The second type of CWPs attached to the cell wall via the GPI-remnant requires the action of two homologous proteins called Dfg5 (*defective for filamentous growth*) and Dcw1 (*defective cell wall*) from the GH76 family in *S. cerevisiae* (Kitagaki et al., 2002). Both proteins are predicted as GPI-APs and are predominantly found at the plasma membrane and to a minor extent in the cell wall (Kitagaki et al., 2002; Spreghini et al., 2003). Single deletion mutants

are viable, but possess hypersensitivity against zymolyase in case of $\Delta dcw1$ and are not able to invade agar via filament formation in pseudohyphal cells upon nitrogen starvation in the $\Delta dfg5$ background (Kitagaki et al., 2002; Mösch and Fink, 1997). In contrast to that, the double mutant is lethal and promotor shut-off cells deficient in both proteins exhibit rounded and large morphologies with an increased chitin content and released the GPI-CWP Cwp1 into the medium (Kitagaki et al., 2002). Similarly, Dfg5 and Dcw1 were identified in a screen of mutants deficient in GPI-CWP anchorage (Gonzalez et al., 2010). Conditional mutants of $dcw1$ showed a cell cycle arrest in the early G2 phase and an aberrant cell wall upon elevated temperatures (Kitagaki et al., 2004). Studies in the pathogenic yeast *C. albicans* could strengthen the findings of a redundant and essential function of Dfg5 and Dcw1, as they were also found to be synthetically lethal (Spreghini et al., 2003). Here, $\Delta dfg5$ mutants were unable to form pH-induced hyphae, as the hyphae-specific gene (*HWT1*) was not expressed, while $\Delta dcw1$ did not show such a defect. In a very recent study it is shown that among seven *DFG* genes found in the genome of *A. fumigatus*, six of them are expressed under lab conditions and their complete deletion results in the absence of cell wall bound galactomannan (GM) accompanied with severe growth defects, but not lethality (Muszkieta et al., 2019). Interestingly, GM in *A. fumigatus* is a special type of polysaccharide, as it is attached to a GPI-anchor (Costachel et al., 2005).

It is suggested that Dfg5 and Dcw1 are responsible for the incorporation of GPI-CWP by cleavage of the Man α 1,4-GlcN linkage within the GPI-anchor core and the subsequent transfer of the terminal mannose to β 1,6-glucan, as such linkages exist in the cell wall of *S. cerevisiae* (Figure 5) (Fujii et al., 1999; Kitagaki et al., 2002; Kollár et al., 1997).

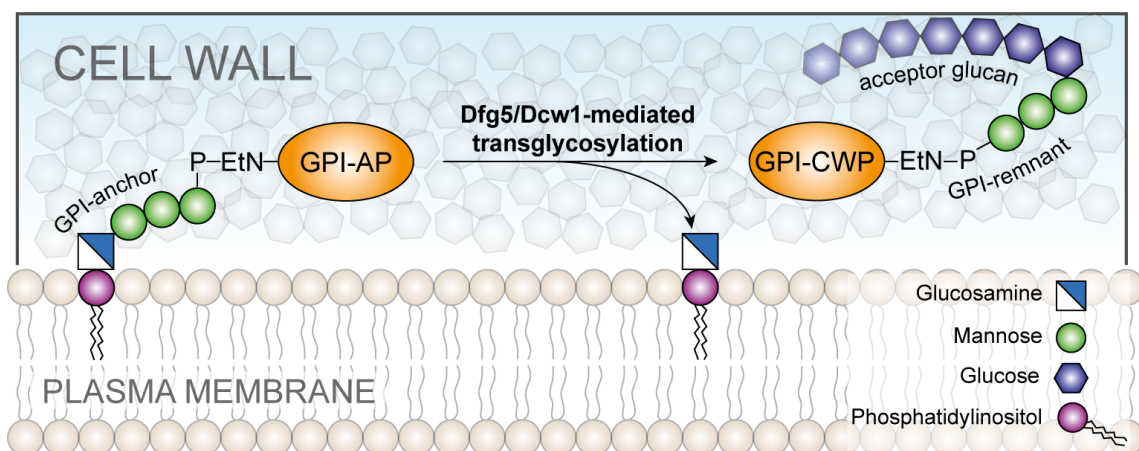


Figure 5: Principal model for CWP-incorporation by fungal GH76-proteins. GPI-APs are transported to the plasma membrane, where the recognition of the GH76-members Dfg5 and Dcw1 occurs. By cleaving the Man α 1,4-GlcN linkage within the GPI-anchor, the transglycosylation is facilitated into the cell wall glycan matrix.

In contrast to this pathway, an alternative route of CWP-attachment catalyzed by Dfg5/Dcw1 is suggested for orthologs from the filamentous fungi *N. crassa*. This is based on the annotated α 1,6-mannanase activity of GH76-proteins in the CAZy-database and the importance of the α 1,6-mannose core structure of N-linked GM observed in $\Delta och1$ mutants from *N. crassa* (Maddi and Free, 2010; Maddi et al., 2012). Note that GM from *N. crassa* is the glycoprotein decorating polysaccharide, while the above-mentioned GM from *A. fumigatus* is a lipopolysaccharide. Initially it was described that mutants lacking the α 1,6-mannosyltransferase Och1 show a severe defect in the incorporation of GPI-CWP (Maddi and Free, 2010). Och1 catalyzes the addition of the terminal α 1,6-mannose of N-glycans on which the α 1,6-mannan backbone is built on, thus leading to an immature form of the N-linked oligosaccharides (Nakanishi-Shindo et al., 1993). While *och1* mutants from a number of fungi possess a cell wall-related phenotype, it is not a general phenomenon found throughout ascomycota, as no cell wall defect was observed in *A. fumigatus* (Lambou et al., 2010; Moo et al., 2006; Schekman, 2010; Uccelletti et al., 2006; Yoko-o et al., 2001). Just two of the nine analyzed GH76-proteins from *N. crassa* were shown to possess morphological impact on the cells, of which $\Delta dfg5$ showed a spreading colonial phenotype, while $\Delta dcw1$ built thinner hyphae than the wild type (Maddi et al., 2012). The double mutant was not lethal like it is known from *S. cerevisiae*, which is explained with the remaining seven homologs that may function as cross-linking enzymes as well. Still, such cells released large amounts of cell wall proteins into the medium, a phenomenon that very well resembles the promotor shut-off cells from yeast (Kitagaki et al., 2002; Maddi et al., 2012). Further evidence for the N-glycan cross-linking hypothesis was later provided by studies of α -mannanases found in polysaccharide utilization loci (PULs) from the human gut bacterium *Bacteroides thetaiotaomicron* (Cuskin et al., 2015; Thompson et al., 2015a). They describe a sophisticated system established by this Gram-negative to degrade the yeast's outer mannan by a set of secreted mannanases including those of the GH76 family, which were shown to specifically hydrolyze the α 1,6-mannose backbone (Cuskin et al., 2015). Further studies successfully cocrystallized a GH76 protein from *Bacillus circulans* in complex with α 1,6-mannopentaose unveiling its substrate bound state (Thompson et al., 2015b).

1.2 GPI-anchored proteins

Aside from classical transmembrane helices, integral membrane proteins can also be produced by the attachment to lipids during their synthesis in the endoplasmic reticulum (ER). In eukaryotes, proteins can be transferred onto a glycolipid called *glycosylphosphatidylinositol anchor* (GPI-anchor) (Ikezawa, 2002). GPI-anchored proteins (GPI-AP) were discovered in the 70's upon mammalian tissue treatment with a phosphatidylinositol phospholipase C (PI-PLC), which released proteins from the plasma membrane and therefore was suggested to be covalently bound to a PI-anchor (Ikezawa et al., 1976; Low and Finean, 2015). They appear to preferentially localize at plasma membrane domains called lipid rafts, which consist of sphingolipids, sterols and certain proteins (Surma et al., 2012).

All characterized GPI-anchors from protozoa, fungi, plants, and mammals revealed a common core structure composed of an ethanolamine phosphate (EtN-P) attached to the C-terminus of the GPI-anchored protein, followed by three mannoses (Man1, Man2, Man3), glucosamine (GlcN), and an inositol-phospholipid (PI) (Figure 6) (Fankhauser et al., 1993; Ferguson et al., 1988; Fontaine et al., 2003; Homans et al., 1988; Mukasa et al., 1995; Nakano et al., 1994; Oxley and Bacic, 1999). The glycan-core side chains as well as the lipid composition not only vary between different species but can also be different within a single organism and even the same protein, as shown for a 5'-nucleotidase from bovine liver and a surface glycoprotein from *Trypanosoma brucei* (Ferguson et al., 1988; Ikezawa, 2002; Paulick and Bertozzi, 2008; Taguchi et al., 1994).

GPI-APs can be predicted *in silico* due to their terminal amino acid composition. The number of predicted GPI-APs vary between different fungal species, ranging from just 33 in *Schizosaccharomyces pombe*, ~60 putative GPI-APs in *S. cerevisiae*, to ~100 in *N. crassa* and more than 160 putative GPI-APs in *C. albicans*. The numbers can differ significantly depending on the applied search strategies (Caro et al., 1997; Eisenhaber et al., 2004; de Groot et al., 2003). The attachment to a GPI-anchor is predicted if a protein contains (i.) a hydrophobic signal peptide at its N-terminus, which is responsible for the cotranslational translocation of the protein into the ER-lumen and (ii.) a C-terminally located GPI-attachment signal (Caro et al., 1997; Pierleoni et al., 2008). The amino acid, which is attached by a GPI-transamidase with an amide linkage between its carboxyl group and the ethanolamine of the GPI, is called the ω -residue. Preceding residues belong to the ω -minus region and residues following the ω -site are designated as ω -plus (Figure 6). All characteristic features of such a GPI-attachment signal are found in these ω -regions. While the ω -site residue itself can be an

alanine, aspartate, asparagine, cysteine, glycine, or serine, the ω -minus region from ω -10 to ω -1 consists of polar amino acids. ω +2 is mainly an alanine, glycine or serine, followed by a moderately polar spacer sequence from ω +3 to ω +9. Finally, a hydrophobic patch with a length that is able to pass the membrane completes the GPI signal sequence (Eisenhaber et al., 1998, 2004; de Groot et al., 2003).

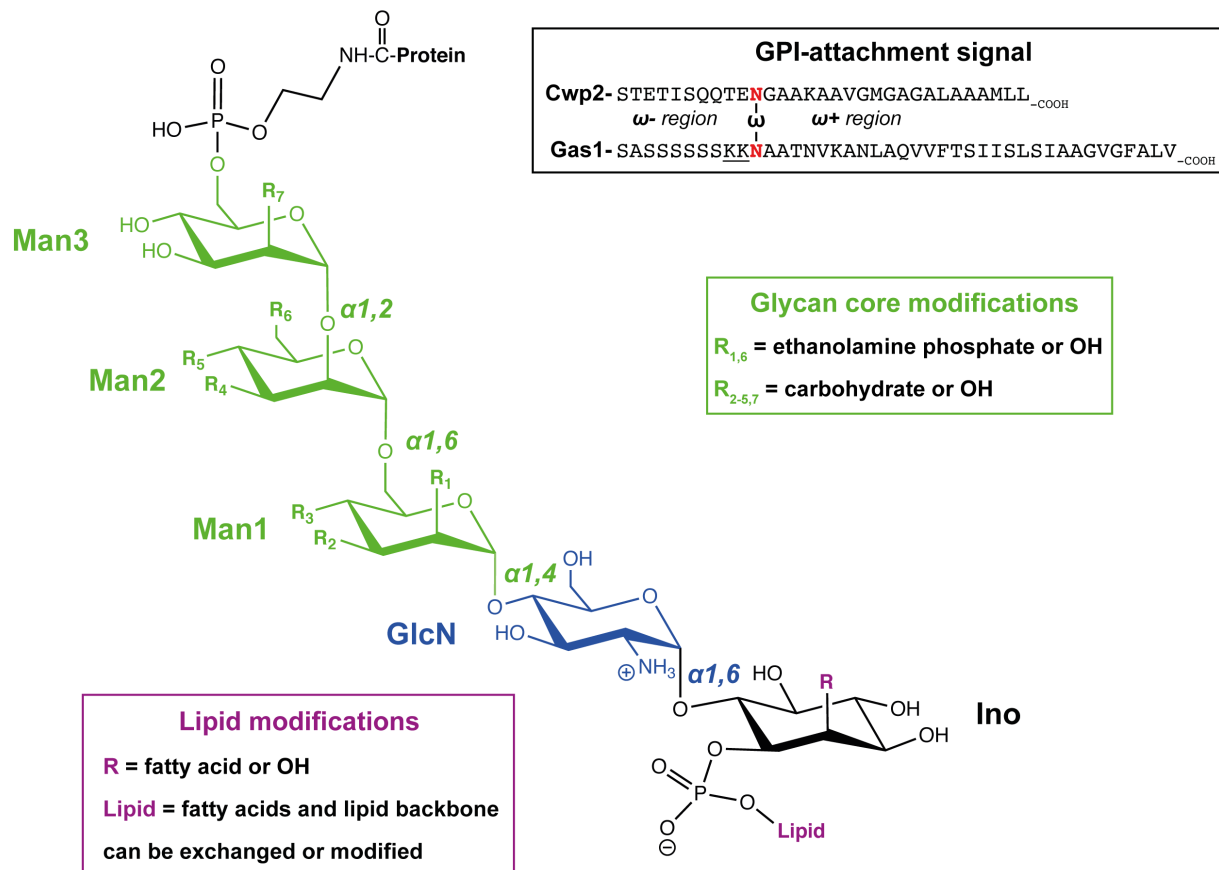


Figure 6: General GPI-anchor structure attached to a protein. The schematic structure of a GPI-anchor is shown, starting at the C-terminus of the attached protein, the bridging ethanolamine phosphate, three mannoses (Man1-3 starting from the moiety at the reducing end), a glucosamine (GlcN), and the phosphatidylinositol (Ino=Inositol). Characteristics of the different regions are highlighted in the boxes. The type of linkages between the GPI-core sugars is indicated (adapted and modified from Caro et al., 1997; Ferguson et al., 2009; Mayor and Riezman, 2004; Orlean and Menon, 2007).

The synthesis of the GPI-anchor and its transfer to the C-terminus of GPI-AP requires more than 20 genes in mammals and yeasts, which are associated with the ER-membrane (Orlean and Menon, 2007). In principle, three major tasks must be completed. Firstly the GPI precursor assembly occurs, which is a topologically separated process including a flip of the GlcN-PI from the cytosolic to the luminal side of the ER-membrane. Secondly, the recognition, cleavage, and attachment of the C-terminal GPI-anchoring propeptide from a freshly synthesized GPI-AP onto the GPI-anchor must happen. In a third step, modifications

on the GPI-lipid and the glycan side-chains in the ER and the Golgi take place (Orlean, 2012). Due to the architecture of the GPI-anchor it is intuitive that the major steps in GPI precursor synthesis and protein attachment are highly conserved throughout eukaryotes, while the subsequent processing to the mature GPI-anchor is more diverse (Fujita and Kinoshita, 2012).

In *S. cerevisiae*, the GPI-anchor harbors four mannoses, of which three possess an EtN-P-modification attached by Mcd4, Gpi7, and Gpi13 to the 2-hydroxy group of Man1 and the 6-hydroxy-group of Man2 and Man3, respectively, and an acylated inositol at its 2-hydroxy group, which has been added in the first step of the luminal GPI assembly by the acyltransferase Gwt1 (Benachour et al., 1999; Galperin and Jedrzejewski, 2001; Gaynor et al., 1999; Orlean, 2009; Tsukahara et al., 2003; Umemura et al., 2003). Upon protein attachment, the first steps in GPI-maturation consist of the removal of the inositol-linked acyl-chain by Bst1 and the remodeling of the diacylglycerol by substitution of the *sn*-2 chain to a longer C26:0-fatty acid catalyzed by Per1 and Gup1. These can subsequently be replaced with a ceramide by Cwh43 (Bosson et al., 2006; Fujita et al., 2006b, 2006a; Ghugtyal et al., 2007; Martin-Yken et al., 2001; Tanaka et al., 2004; Umemura et al., 2007). After the removal of the EtN-Ps from Man1 by Cdc1 and Man2 by Ted1, the GPI-AP is transported to the Golgi where 20-30% of the GPI-glycan is extended by α 1,2- or α 1,3-linked mannoses to Man4, before it is finally delivered to the plasma membrane (Fankhauser et al., 1993; Haass et al., 2007; Vazquez et al., 2014). Here, a final GPI-processing step is attributed to two GPI-anchored proteins, Dfg5 and Dcw1, which were described to be responsible for GPI-CWP attachment to the CW by cleaving the GPI-glycan and its subsequent transfer onto the β 1,6-glucan (Fujii et al., 1999; Kitagaki et al., 2002; Kollár et al., 1997).

Defects of GPI-anchor maturation result in delayed or false delivery of GPI-APs. Mutants deficient in *bst1*, *per1* and *gup1* that target the lipid moieties are not able to cluster into ER exit site (ERES) domains anymore, which are necessary for efficient ER to Golgi transport (Castillon et al., 2009). In *cwh43* mutants, the β 1,3-glucanase Gas1, usually residing in the plasma membrane, tends to be transferred to the cell wall and the genetically related Ted1 is crucial for p24-mediated COPII vesicular from ERESs (Manzano-Lopez et al., 2015; Yoko-o et al., 2018). While not having a compromised ER-exit, conditional mutants of *cdc1^{ts}*, still containing the EtN-P at Man1, show a partial secretion block in the late secretory pathway and a fragile cell wall due to a compromised CWP-attachment to β 1,6-glucans (Vazquez et al., 2014). Interestingly, the disruption of GPI-anchor synthesis in *S. cerevisiae* is lethal, while *A. fumigatus* survives with a defective cell wall, abnormal hyphae formation, rapid conidial germination, and aberrant conidiation (Leidich et al., 1995; Li et al., 2007).

1.3 Objectives of the thesis

Fungal infections are an increasing threat for human health, especially to immuno-compromised patients in clinical environments, with an estimated number of over two million infections per year and mortality rates up to 95% (Brown et al., 2012a). Due to their high similarity to humans on a cellular level, many antimycotica possess negative side effects on patients. A striking difference between mammalian and fungal cells is the fungal cell wall, which is essential for the survival of fungi and necessary for successful pathogenicity, yet absent in humans and therefore provides an excellent target for low side-effect drug development as shown for echinocandins (Perfect, 2017). An essential step in cell wall biosynthesis of yeasts such as *Saccharomyces cerevisiae* and the human pathogen *Candida albicans* is the incorporation of cell wall proteins, which is carried out by two homologous proteins called Dfg5 and Dcw1 belonging to the GH76 class (Kitagaki et al., 2002; Spreghini et al., 2003). However, a precise biochemical and structural understanding is still lacking behind. While GH76 proteins are essential in yeasts, they seem not to be essential for filamentous fungi like *Aspergillus fumigatus* and *Neurospora crassa* and may even act on other substrates (Maddi et al., 2012; Muszkieta et al., 2019).

In order to clarify the apparent contradictions and to shed light on this important step during fungal cell wall synthesis, this work will firstly address the evolutionary features of the GH76 family by utilizing *in silico* analysis to obtain a better understanding of its isofunctional subfamilies. Then, an in depth functional and structural characterization of a fungal *Dfg5*-member from the thermophilic mold *Chaetomium thermophilum* will enable us to not only allow structure-function based *in vivo* experiments, but shall also clarify the functional paradox of fungal *Dfg5*-proteins. This will be tackled by assembling its actual substrate into the binding pocket by high molecular sugar soaking. The determination of the crystal structure shall also set the basis for structure-based drug discovery. To do so, likely targets will be identified by docking and then used for soaking experiments, thereby identifying a true candidate for further drug development by combined structural and *in vivo* analysis. By expanding the structural knowledge to a further fungal GH76-homolog, the common feature of GH76-proteins shall finally be deduced.

2 Materials

2.1 Chemicals

(3-{4-imino-5,6-dimethyl-3H,4H-furo[2,3-d]pyrimidin-3-yl}propyl)dimethylamine (FP-1)	<i>Vitas-M Laboratory</i>
1H -Imidazole-2- methanamine, N ,1-dimethyl (FRAG37)	<i>Jena Bioscience</i>
2-(N-morpholino)ethanesulfonic acid (MES)	<i>Th. Geyer</i>
3-(N-morpholino)propanesulfonic acid (MOPS)	<i>Roth</i>
3-[3-(1H-imidazol-1-yl)propyl]-5,6-dimethyl-3H,4H-furo[2,3-d]pyrimidin-4-imine (FP-2)	<i>Vitas-M Laboratory</i>
4-(2-hydroxyethyl)-1-piperazineethanesulfonic acid (HEPES)	<i>VWR</i>
4-Nitrophenyl α -D-mannopyranoside	<i>Merck</i>
Acetate	<i>VWR</i>
Acetonitrile	<i>VWR</i>
Agar-agar	<i>Roth</i>
Agarose	<i>Invitrogen</i>
Ammonium iodide (NH ₄ I)	<i>Fluka</i>
Ammonium persulfate (APS)	<i>Merck</i>
Borate	<i>Grüssing</i>
Bromphenolblue	
Calcium chloride (CaCl ₂)	<i>Fluka</i>
cOmplete™ Protease Inhibitor Cocktail	<i>Merck</i>
Coomassie brilliant blue R-250	<i>Serva</i>
Dithiothreitol (DTT)	<i>Merck</i>
Ethanol	<i>VWR</i>
Ethylenediaminetetraacetic acid (EDTA)	<i>Merck</i>
Gadolinium(III)acetate (Gd(OAc) ₃)	<i>Alfa Aesar</i>
Glucosamine	<i>TCl chemicals</i>
Glucose	<i>Roth</i>
Glycerol	<i>Roth</i>
Glycin	<i>Sigma</i>
Hydrochloric acid (HCl)	<i>VWR</i>
Imidazole	<i>Merck</i>
Isopropanol	<i>VWR</i>
Isopropyl β -D-1-thiogalactopyranoside (IPTG)	<i>Gerbu</i>

Kanamycinsulfate	VWR
Manganese(II) chloride (Mn ₂ Cl)	
Mannose	Merck
Midori Green	Biozym
<i>p</i> -Anisaldehyde	
Peptone	Difco
Phenylmethylsulfonyl fluoride (PMSF)	Fluka
Polyethylene glycol 3350 (PEG3350)	Fluka
Polyethylene glycol 8000 (PEG8000)	Fluka
Potassium acetate	
Rotiphorese® Gel 30 (37,5:1)	Roth
Rubidium chloride (RbCl)	
Saccharose	VWR
Sodium chloride (NaCl)	VWR
Sodium dodecyl sulfate (SDS)	AppliChem
Sodiumhydroxide (NaOH)	AppliChem
Sulfuric acid (97%)	VWR
Tetramethylethylenediamine (TEMED)	Roth
Tris(hydroxymethyl)aminomethane (Tris)	Roth
Tryptone	Th. Geyer
Virkon	VWR
Yeast extract	Th. Geyer
α1,2-mannobiose	Dextra
α1,6-mannobiose	Dextra
β-mercaptoethanol	Roth
β1,3-laminaribiose	MEGAZYMES

Chemicals mentioned in this thesis, which are not listed here, were purchased from either *Fluka*, *VWR*, *Roth*, *Merck*, *AppliChem* or *TCI chemicals* in the highest best purity.

2.2 Equipment

Devise	Model	Manufacturer
AEC column	Bio-Scale Mini UNOsphere High Q	Bio-Rad
Agarose gelelectrophorese	Chamber	Feinmechanik, Chemistry department, PUM

Amylose resin HF	15 ml	NEB
Balance	PC2200	Mettler
Centrifuges	Centrifuge 5810 R	Eppendorf
	Fresco21	Heraeus
	Lynx 6000	Sorvall
	J2-HS / J2-21 M/E	Beckman
	L7-65	Beckman
Centrifuge Bottles	1l Superspeed CB with sealing	Nalgene
	JA-20	Beckman
	Polycarbonate tubes and bottles for 55.2 Ti	Beckman
Centrifuge Rotors	F6S 6x1000Y	Thermo Fisher
	JA-20 Fixed Angle Rotor	Beckman
	55.2 Ti	Beckman
Crystallization robot	Oryx8	Douglas Instruments
	Honeybee 963™	Digilab
Documentation of agarose gels	Computer E.A.S.Y.	UVP
	UV-transilluminator	Herolab
	Thermal printer UP-D 895	Sony
Documentation System Crystal plates	Rock Imager	Formulatrix
French Press	French Press	Aminco
	High pressure cell	Feinmechanik, Chemistry department, PUM
Heating block	BT3	Grant Instruments
IMAC column	Protino Ni-NTA Column 5mL	Macherey-Nagel
Incubators	BFED-53	Binder
	Multitron	InforsHT
	Innova S44i	Eppendorf
LC-System	NGC Chromatography System	Bio-Rad
Microfluidizer	Emulsiflex	
Microwave		LG
MilliQ water dispenser	Seralpur Pro90CN	Seralpur
NanoDrop	NanoDrop 200 Spectrophotometer	Thermo Scientific
OD-spectrometer	OD 600	Implen
Peristaltic pump	Pumpdrive 5201	Heidolph
pH meter	HI2020 edge®	Hanna Instruments
Pipets	Research	Eppendorf
Potter Homogenizer	15 ml glass cylinder and plunger	Sartorius
Power Boxes	EPS 301	Amersham Biosciences

Real-Time PCR-Cycler	Rotor-Gene Q	<i>Qiagen</i>
SDS-PAGE chamber	Mini-PROTEAN® Tetra Vertical Electrophoresis Cell	<i>Bio-Rad</i>
SEC column	HiLoad 26/600 Superdex 200pg	<i>GE Healthcare</i>
Spin concentrator	Amicon Concentrators (3-30 kDa MWCO)	<i>Millipore</i>
Thermocycler	GeneAmp PCR System 2400	<i>Perkin Elmer</i>
Thermomixer	Comfort	<i>Eppendorf</i>
Waterbath	NK22	<i>Haake</i>
X-ray sources/beamlines	Synchrotron Beamlines ID23-1/2, ID29	<i>ESRF, Grenoble</i>
	Synchrotron Beamline PXIII	<i>SLS, Villigen</i>
	In house source	<i>AG Klebe, Pharmacy department, PUM</i>

2.3 Commercial kits, enzymes, and consumables

Crystallization Screen	NeXtal Tubes JCSG Core Suite I	<i>Qiagen</i>
DNA Ladder	1 kb DNA Ladder	<i>NEB</i>
DNA-Ligase	T4-Ligase	<i>NEB</i>
	Ligase buffer 10x	<i>NEB</i>
DNA-Polymerase	Phusion Polymerase (2U/μL)	<i>NEB</i>
	Phusion HF-Buffer (5x)	<i>NEB</i>
GelEx Kit	QIAquick Gel Extraction Kit	<i>Qiagen</i>
Mini prep kit	QIAprep Spin Miniprep Kit	<i>Qiagen</i>
PCR purification	QIAquick PCR Purification Kit	<i>Qiagen</i>
Phospholipase C	PI-PLC from <i>Bacillus cereus</i>	<i>Merck</i>
Pipet tips		<i>Sarstedt</i>
Protein Ladder	Pierce Unstained Protein MW Marker	<i>Fermentas</i>
Reaction tubes	PCR-cups, 1.5 ml, 2.0 ml	<i>Sarstedt</i>
Restriction Enzymes	<i>NheI</i>	<i>NEB</i>
	<i>XhoI</i>	<i>NEB</i>
	<i>HindIII</i>	<i>NEB</i>
	<i>BamHI</i>	<i>NEB</i>
	Cutsmart 10x	<i>NEB</i>
Sypro Orange	SYPRO™ Orange Protein Gel Stain	<i>Thermo Fisher</i>
TLC plates	TLC Silica Gel 60 F254 (5x10cm)	<i>Merck</i>
Zymolyase	100T	<i>Roth</i>

Consumables, enzymes, and kits mentioned in this thesis, which are not listed here, were purchased from either *NEB*, *Merck*, *Roth*, *Sarstedt* or *Qiagen*.

2.4 Oligonucleotides, vectors, and DNA

2.4.1 Oligonucleotides for gene amplification

Name	Sequence (5'-3')
CtDfg5_fwd	ATG CGCTAG CCAGCAGCAGTATTACAAGATCG
CtDfg5_rev	ATG CCTCGAG TTACCGGTCACCGGCATTAATCTCC
ScDfg5_fwd	GCATG CGCTAG CATGGATTTGGATACTACTAGCAAAACG
ScDfg5_rev	ATG CCTCGAG TTAGCGATCACCTTTCTTAATG
ScDcw1_fwd	GCATG CGCTAG CGTCGAATTGGATCTAGACAACACTACG
ScDcw1_rev	ATG CCTCGAG TTACTTTGAACCCCTTAGTGATATT
CgDfg5_fwd	GCATG CGCTAG CATCAATTTGCAAGACAATAAGG
CgDfg5_rev	ATG CCTCGAG TTAACGGTCACCACCAGTGATTTT
CgDcw1_fwd	GCATG CGCTAG CCTGGAGCTTGATCTGGATGATTACG
CgDcw1_rev	ATG CCTCGAG TTATCTAGAACCACCAGTAATATG
MBP-ScDcw1_fwd	ATC GCGATCC GTGAATTGGATCTAGACAAC
MBP-ScDcw1_rev	AGT CAAGCTT CTTTGAACCCCTTAGTGATATT

The **bold** characters indicate the introduced restriction sites: *NheI* (GCTAGC), *XhoI* (CTCGAG), *BamHI* (GGATCC), and *HindIII* (AAGCTT).

The primers were used to amplify the defined regions from genomic DNA of *Saccharomyces cerevisiae* (Sc) and *Candida glabrata* (Cg), which we had available in our lab-stocks, and from cDNA of *Chaetomium thermophilum* (Ct), a kind gift of Dr. Patrick Pausch.

2.4.2 pET-28a(+)-Vector

The pET-system is a commonly used vector system for recombinant protein production in *E. coli*. Although the used pET-28a(+)-vector is a low copy plasmid, genes can directly be cloned in its *multiple cloning site* yielding enough plasmid for the different working steps. The plasmid provides a kanamycin resistance for selection purposes and stable maintenance within the bacterium in the presence of the antibiotic. The expression of the target gene is under the control of a T7-promotor (and terminator) as well as the lac-operator. Together with a suitable expression host (see below) an efficient protein production can be induced by lactose or its structural analog IPTG, reprogramming almost the complete metabolism of the cell towards gene expression. The vector furthermore encodes a hexa-histidine tag for efficient protein purification. The fully assembled pET-28a(+)-CtDfg5 vector used in this study is shown in Figure 7.

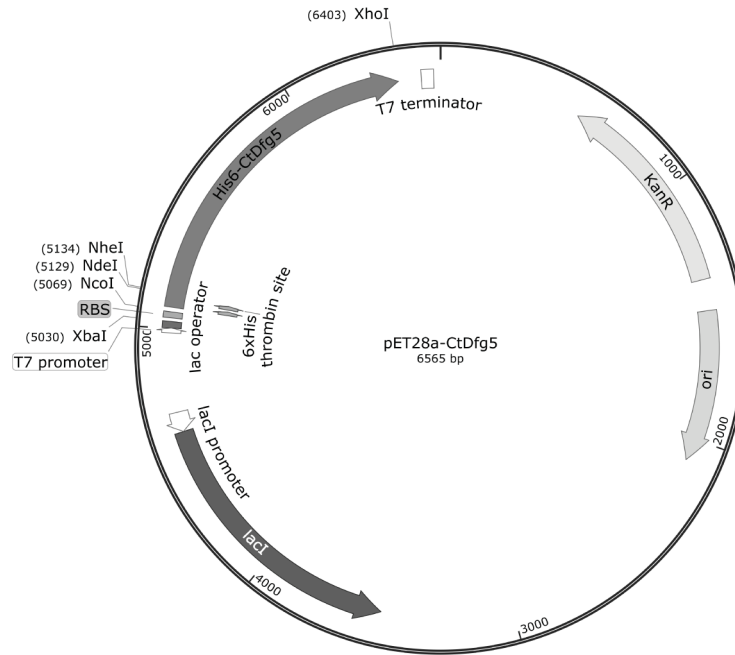


Figure 7: Vector map of pET28a-CtDfg5 used for heterologous protein production.

2.4.3 pMAL-cRI-Vector

pMAL-cRI is an old vector from *NEB* used for N-terminal MBP-fusion to the protein of interest. The plasmid provides an ampicillin resistance to the host organism for selection and maintenance. The expression is under the control of a tac-promoter and can be induced by lactose or its structural analog IPTG.

2.5 Organisms

2.5.1 *Escherichia coli* DH5 α Competent Cells

The *E. coli* strain DH5 α purchased from Invitrogen has been used for the efficient production and isolation of recombinant plasmid-DNA. Due to its mutations in endonucleases and recombination systems it is especially suited for that purpose, yielding high plasmid concentrations.

Genotype: F- ϕ 80*lacZ* Δ M15 Δ (*lacZYA-argF*)U169 *recA1 endA1 hsdR17*(rk-, mk+) *phoA supE44 thi-1 gyrA96 relA1* λ -

2.5.2 *Escherichia coli* SHuffle T7 Express Competent

E. coli SHuffle T7 cells from *NEB* were used for heterologous protein production of CtDfg5 and CtGH76. The deletions of *gor* and *trxB* promote the formation of disulfide bonds, so does a constitutively expressed disulfide isomerase called DsbC. The chromosomal copy of T7 RNA-polymerase under the control of the wild type *lacZ*-promoter enables the compatibility with the T7-expression system together with the pET-vector system used in this study. Deficiencies in proteases furthermore facilitate higher yields of recombinant protein.

Genotype: *fhuA2 lacZ::T7 gene1 [lon] ompT ahpC gal λ att::pNEB3-r1-cDsbC (Spec^R, lacI^q) Δ trxB sulA11 R(mcr-73::miniTn10--Tet^S)2 [dcm] R(zgb-210::Tn10 --Tet^S) endA1 Δ gor Δ (mcrC-mrr)114::IS10*

2.5.3 *Escherichia coli* BL21-Gold(DE3)

The *E. coli* BL21-Gold(DE3) strain from *NEB* is a protease deficient B strain derivative with high transformation efficiency optimized for the heterologous production of recombinant proteins using the T7 expression system.

Genotype: *F⁻ ompT gal dcm lon hsdS_B(r_B⁻m_B⁻) λ (DE3 [lacI lacUV5-T7p07 ind1 sam7 nin5]) [malB⁺]_{K-12}(λ ^S)*

2.5.4 *Saccharomyces cerevisiae* BY4741

Haploid cells of *S. cerevisiae* have been used for the preparation of soluble GPI-anchored proteins (see below). Beside common auxotrophic selection markers, they possess a deletion of the *ykl046c* locus, which encodes for the protein Dcw1. The strain was purchased from *Euroscarf* and is a S288c derivative, where many selection markers have been deleted

Genotype: BY4741; MATa; ura3 Δ 0; leu2 Δ 0; his3 Δ 1; met15 Δ 0; YKL046c::kanMX4

2.6 Software and Algorithms

<i>EFI-EST</i>	(Gerlt et al., 2015)
<i>CCP4i2</i>	(Nicholls, 2017)
<i>Clustal Omega</i>	(Sievers et al., 2011)
<i>ConSurf</i>	(Ashkenazy et al., 2016)

<i>Coot v0.8.3</i>	(Emsley et al., 2010)
<i>CRANK2</i>	(Skubák and Pannu, 2013)
<i>Cytoscape</i>	(Cline et al., 2007)
<i>GraphPad Prism</i>	(GraphPad Software)
<i>Phaser</i>	(McCoy et al., 2007)
<i>PHENIX suite</i>	(Adams et al., 2010)
<i>SeeSAR</i>	(BioSolveIT GmbH)
<i>UCSF Chimera</i>	(Pettersen et al., 2004)
<i>WEBLOGO3</i>	(Crooks et al., 2004)
<i>XDS</i>	(Kabsch, 2010)

3 Methods

3.1 Bioinformatics for biochemists

A major goal of biochemists is the comprehension of proteins and their underlying processes and mechanisms with the final understanding of the organisms' physiology. With the increasing number of genomes it is possible to combine already known principles and transfer them on so far uncharacterized proteins. While a comparison on DNA level is not practical, because of uncertainties concerning codon usage and silent mutations, it is much handier to use the amino acid sequence for global functional analysis. By comparing sequences it is possible to attribute uncharacterized proteins the functions of those already described. The whole procedure is based on the fact that proteins with similar sequences also share similar functions. Enzyme functions are classified in seven main classes characterized by the enzyme commission (EC) number classification consisting of four digits. With every digit the classification becomes more accurate. Using the famous enzyme lysozyme (EC 3.2.1.17), EC 3.-.-.- is attributed to the class of hydrolases, EC 3.2.1.- stands for glycoside hydrolases that hydrolyze O- and S-glycosyl compounds and 3.2.1.17 describes the class of lysozyme, which is responsible for the hydrolysis of β 1,4-linkages between N-acetylmuramic acid and N-acetyl-D-glucosamine in bacterial peptidoglycan. A threshold of 40% sequence identity can be used with a good confidence to transfer protein functions to a homolog concerning the first three digits of the EC number, while 60% and above are necessary for a 90% accuracy for all four digits (Tian and Skolnick, 2003). Although the actual function can vary from already characterized proteins of that class, such classifications are helpful to get a rough idea about protein properties to plan promising experiments. A good way of presenting such analyses is introduced by so called *sequence similarity networks*.

3.1.1 Protein Sequence Similarity Network (SSN)

The increasing number of available sequences requires reasonable approaches for their analysis. The *Enzyme Function Initiative* (EFI) developed a web tool called EFI-EST (*Enzyme Function Initiative-Enzyme Similarity Tool*), which allows users to analyze protein sequences according to their similarity (Gerlt et al., 2015). The aim of such sequence similarity networks (SSN) is to intuitively visualize whole protein families segregated into isofunctional subclasses by applying user-specified expectations values (E-values) on the collection of independent pairwise alignments (Atkinson et al., 2009). In such networks, *nodes* (representing one or more sequences with a certain sequence identity) are connected by *edges*,

if the applied stringency threshold is fulfilled. An example is shown in Figure 8, where it can easily be seen that moderate thresholds (or low *alignment scores*) result in large “hairball” clusters. By using more stringent cutoffs, edges between the nodes are removed and the hairball falls apart into distinct subclasses. The alignment score is increased until the family is split into isofunctional subfamilies.

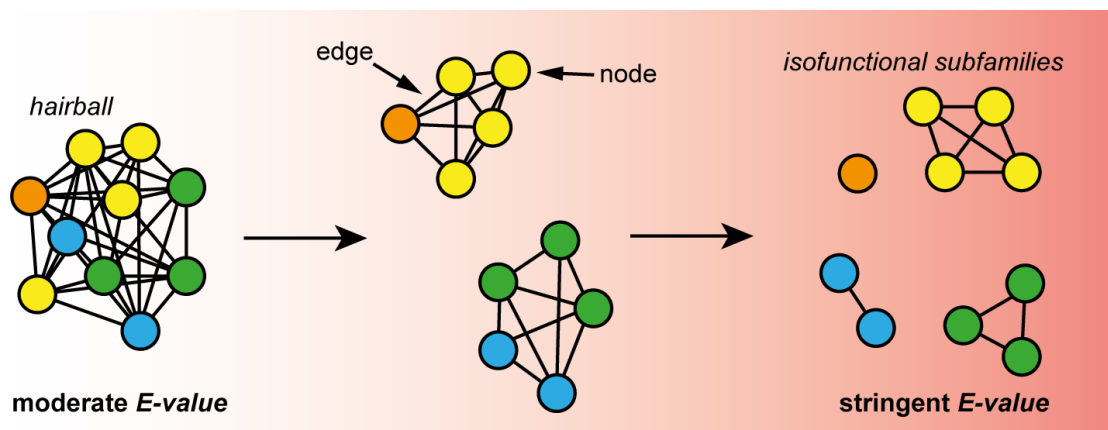


Figure 8: Effect of increasing the *E-value* stringency on the results of *sequence similarity networks*. The circles are described as *nodes*, which harbor sequences with a sequence identity of a user-defined threshold. The nodes are connected by *edges*, if the applied *E-value* cutoff is fulfilled.

For the SSN analysis, the protein family PF03663 (Glycosyl Hydrolase 76) was used to generate an EFI-EST dataset with a BLAST-derived E-value of 10^{-5} without further modifications. In a second step, different alignment scores were applied to the initial data and thereby generating the final SSN, which is produced as an .XGML file composed of 7485 sequences. It can be visualized and analyzed with Cytoscape (v3.5.1) (Cline et al., 2007). For further analysis of the resulting subfamilies, multiple sequence alignments and weblogs are generated by ClustalΩ and WEBLOGO3, respectively (Crooks et al., 2004; Sievers et al., 2011).

3.2 Molecular Biology

3.2.1 Polymerase Chain Reaction (PCR)

Starting in 2015, the DNA-template based polymerase chain reaction (PCR), which has been described 30 years before, was still the method of choice to get your desired DNA in hand (Mullis et al., 1986). Within repetitive rounds of DNA denaturation, annealing, and elongation, complementary primers, dNTPs and a heat stable polymerase are utilized to

amplify a specific region from a given template. The region is defined by gene flanking primers, which also introduce restriction sites for later processing. Site-directed mutagenesis was used to generate the wanted mutant in a two-step *fusion PCR* approach with two complementary primers of ~30 bp and the gene-flanking primers. A typical PCR reaction and the respective parameters used for the experiments are described below.

	Volume or weight	
Template DNA	20-30 ng	
Forward Primer (10 μ M)	2.0 μ l	
Reverse Primer (10 μ M)	2.0 μ l	
Phusion HF-Buffer (5x)	10.0 μ l	
Phusion Polymerase (2U/ μ l)	0.5 μ l	
dNTP (10 mM each)	1.0 μ l	
MilliQ water	ad 50 μ l	

	Temperature	Duration	
Initial Denaturation	98 °C	5 min	30x
Denaturation	98 °C	20 s	
Annealing	55-58 °C	20 s	
Elongation	72 °C	15-30 s/kbp	
Final Elongation	72 °C	5 min	
Cool down	4 °C	∞	

In the meantime genes are usually ordered at companies (here BioCat), which synthesize the genes template independently. As gene synthesis became cheap in the last few years, it allows an economic access to target genes, without the need of a template DNA, that already come in the vector of choice and are codon-optimized for the respective expression host.

3.2.2 Agarose gelelectrophoresis

To analyze amplified DNA or to check the success of restriction digestions, agarose gel electrophoresis was performed. This key technique in molecular biology is used to separate DNA in an electric field according to its size following the principle: The smaller the negatively charged DNA, the higher the ability to migrate through a matrix of an agarose gel towards the anode. The samples are visualized by DNA-intercalating dyes, which can be detected under UV-light. For a typical 1% agarose gel, 0.65 g agarose were added to 70 ml TBE-buffer (0.1 M Tris, 0.1 M borate, 2 mM EDTA) and boiled in a microwave until the agarose was dissolved completely. 2.5 μ l MIDORI Green were added to the solution before pouring the gel. Depending on the type of gel, 5 μ l samples for an analytical run or 45 μ l for a preparative gel were used.

3.2.3 PCR purification and Gel Extraction

Depending on the result of the analytical gel, a PCR product could directly be purified from the reaction by the QIAquick PCR Purification Kit from QIAGEN. Here, an isopropanol and guanidium chloride-containing buffer is added to the PCR reaction that facilitates binding of DNA to a silica matrix. While remaining nucleotides, primers, enzymes, and salts are removed with an ethanol-containing wash buffer, DNA molecules with more than 100 bp stick to the column and can finally eluted with a low salt buffer or water. To extract specific bands from a preparative gel, the QIAquick Gel Extraction Kit from QIAGEN was used. As an additional step, the DNA band must be dissolved before it is applied to the column. The exact procedures were performed according to the manufacture's guides.

3.2.4 DNA-modification: digest and ligate

In order to combine the target gene in the correct manner with the expression plasmid, compatible restriction sites are used. Usually, such sites consist of 4, 6, or 8 bp with a palindromic nature, which are recognized by endonucleases that cut the DNA in a *blunt* or *sticky end* manner. The digest is incubated at 37 °C for 1-2 h. A typical restriction digest consists of the following:

	Volume or weight
DNA	1 µg
Restriction Enzyme I	1 µl
Restriction Enzyme II	1 µl
Cutsmart (10x)	2 µl
MilliQ	ad 20 µl

When two different *sticky ends* present on the vector and the insert are used, the two DNA molecules can be combined in a specific orientation by the T4 DNA ligase. It catalyzes the formation of a phosphodiester bond between the 5'-phosphate and the 3'-OH groups of duplex DNA in an ATP-dependent manner. The ligation is carried out at RT for 30 min with a subsequent inactivation at 65 °C for 10 min using the following composition:

	Volume or weight
Vector	50 ng
Insert	*
T4-Ligase	0.5 μ l
Ligase buffer (10x)	1.0 μ l
MilliQ	ad 10 μ l

*A molar ratio of 1:3 vector to insert is used

3.2.5 Plasmid preparation and determination of DNA concentration

When transformed into a bacterium, the respective plasmid is maintained unmodified in the organism as long as the selective pressure is present. This feature is used to multiply the number of plasmids by growing the cells in LB-medium supplemented with 35 μ g/ml kanamycin until stationary phase over night at 37 °C shaking at 225 rpm. The plasmid is isolated by using the QIAprep Spin Miniprep Kit, which is based on the alkaline lysis protocol from Birnboim and Doly (Birnboim and Doly, 1979). The plasmid extraction follows a three-step protocol. The first step contains a buffer including RNase A (digestion of RNA) and EDTA (destabilizing membranes and inactivating DNases). The second buffer leads to the alkaline cell lysis and denaturation of DNA and proteins by NaOH supported by SDS, and, finally, the mixture is neutralized with potassium acetate buffer, which results in renaturation of plasmid DNA. Here lipids, proteins, and most of the genomic DNA remain insoluble and can be removed by centrifugation. The exact procedure was performed according to the manufactures guide.

LB-medium	
Tryptone	10 g/l
Yeast extract	5 g/l
NaCl	10 g/l
NaOH (10 M)	400 μ l/l

3.2.6 Preparation of competent *E. coli* and plasmid transformation

The native ability of *E. coli* to take up naked DNA from its environment is limited. In 1970 it was demonstrated that the treatment with calcium chloride enhances that ability and confers the bacterium an artificial (or chemical) competence (Mandel and Higa, 1970). Later, this principle was improved by the addition of further metals to the buffer (Hanahan, 1983; Kushner, 1988). For one batch of chemical competent *E. coli* cells, 50 ml LB-medium is inoculated with 1 ml of a preculture grown over night at 37 °C. After incubation at 37 °C and 225 rpm until an OD₆₀₀ of 0.5-0.6, the exponentially grown cells are harvested for 15 min and

3200g at 4 °C. The pellet is resuspended in 15 ml TBF-I buffer and incubated on ice for 2 h and subsequently pelleted again. After resuspension in 2 ml TBF-II buffer, the suspension is split up in 50 µl aliquots, flash frozen in liquid nitrogen and stored at -80 °C.

TBF-I		TBF-II	
RbCl	100 mM	RbCl	10 mM
CaCl ₂ x2H ₂ O	10 mM	CaCl ₂ x2H ₂ O	75 mM
Potassium acetate	30 mM	Glycerol	15% (v/v)
MnCl ₂	50 mM	MOPS	10 mM
Glycerol	15% (v/v)		

For transformation, 10-100 ng plasmid-DNA is added to the cells after they thawed on ice. In case of ligations, the complete reaction was used for the transformation. After a heat shock of 45 s at 42 °C, 300 µl LB-medium are added and subsequently the cells are incubated at 37 °C shaking with 225 rpm for 50 min. The recovered cells are then pelleted at 14000 rpm, resuspended in 50 µl of the medium, and then plated on LB-agar plates supplemented with 35 µg/ml kanamycin. Colonies are grown over night at 37 °C and the plates can be stored at 4 °C for 2-3 weeks.

LB-agar	
Tryptone	10 g/l
Yeast extract	5 g/l
NaCl	10 g/l
NaOH (10 M)	400 µl/l
Agar-agar	15 g/l

3.3 Protein biochemistry

There is one major bottleneck in protein biochemistry: No protein, no experiments.

This often leads to rampant trials of test expressions to find a condition that results in soluble protein. Due to the fact that there was an established protocol for the production of other extracellular proteins from different fungi in the lab, I will directly proceed with the condition that worked for the proteins subjected in this thesis.

3.3.1 Heterologous overproduction in *E. coli* and cell lysis

The protocol for producing extracellular fungal proteins combines the ability of the *E. coli* strain SHuffle T7 Express, which is optimized for the production of proteins with multiple disulfide bonds within its cytosol, and a low temperature. For heterologous overproduction of

proteins without a solubility tag, 2 l of sterile DYT medium supplemented with 35 µg/ml kanamycin in a 5 l chicane flask are inoculated with 40 ml of an LB-grown overnight culture and incubated at 37 °C and 150 rpm until an OD₆₀₀ of 0.2-0.3. The culture is then shifted to 12 °C and after cooling down for approx. 60 min, the production is induced with 0.1 mM IPTG. The cells are harvested at 4000 rpm and 4 °C for 20 min after an incubation of 72 h. The pellet from 2 l culture is resuspended in 25 ml lysis buffer, flash frozen in liquid nitrogen and stored at -80 °C. For cell lysis, the pellet is thawed in a water bath at RT until all ice disappeared. Steps from now on are kept on ice or at 4 °C.

For cell lysis, the suspension is poured into a pre-chilled French pressure cell press (short: French press, Aminco Inc.). A high pressure of 1000 psi is generated in the pressure cell, which is manually controlled by the outlet valve. When passing through the valve, the cells are decompressed and exposed to shear forces that finally disrupt the bacteria. The process is repeated two times. Afterwards, the lysate is centrifuged for 20 min at 4 °C and 18000 rpm, resulting in a cell-debris cleared lysate that is used for subsequent protein purification.

DYT-medium		Lysis buffer	
Tryptone	16 g/l	NaCl	200 mM
Yeast extract	10 g/l	HEPES	20 mM
NaCl	5 g/l		pH 8.0
NaOH (10 M)	400 µl/l		

3.3.1.1 Production of maltose-binding-ScDcw1 fusion protein

To produce MBP-ScDcw1 from *Saccharomyces cerevisiae* in *E. coli* BL21-Gold(DE3), 2 l LB-medium is inoculated with 40 ml over night culture supplemented with 50 µg/ml ampicillin and 12.5 g/l lactose for autoinduction (Studier, 2005). After 40 h at 20 °C shaking at 150 rpm in 5 l chicane flasks, cells are harvested and processed as described before.

3.3.2 Protein Purification

3.3.2.1 Immobilized Metal Affinity Chromatography (IMAC)

After cell lysis, the cleared lysate contains the overproduced protein and all soluble proteins from *E. coli*. To make life easy, the desired protein is produced with a His-tag at its N-terminus. The His-tag is utilized in the affinity chromatography technique called Ni-NTA, which is based on nitrilotriacetic acid (NTA) chelating nickel (Ni) ions. Polyhistidines are able to bind to nickel ions, while non-tagged proteins do not bind or can be removed

quantitatively during wash steps with moderate concentrations of the histidine analogon imidazole. To elute the protein competitively from the column, the imidazole concentration is simply increased. Alternatively, the pH can be shifted below 7, as the histidines become protonated and therefore loose their affinity for nickel ions.

The cleared supernatant including the His-tagged protein is loaded with a peristaltic pump using 3 ml/min flow rate onto a 5 ml Protino Ni-NTA column, which was equilibrated with 10 column volumes (CV) lysis buffer before. The column is then washed with 8 CV wash buffer and subsequently the recombinant protein is eluted by 4 CV elution buffer. Afterwards the protein is spin-concentrated with filter membranes of appropriate cutoff sizes to half of the injection loop volume used for size exclusion chromatography.

Wash buffer		Elution buffer	
NaCl	200 mM	NaCl	200 mM
HEPES	20 mM	HEPES	20 mM
Imidazole	25 mM	Imidazole	500 mM
	pH 8.0		pH 8.0

3.3.2.2 Size Exclusion Chromatography (SEC)

The principle of size exclusion chromatography (SEC) is based on the migration of macromolecules in a size-dependent manner through a chromatography column that consists of porous beads. Depending on the type of column, such beads can be made of dextran (Sephadex), polyacrylamide (Sephacryl) or agarose (Sephacrose). While small molecules are occasionally trapped in the pores of the stationary phase, larger molecules pass directly through the column. As a consequence, bigger proteins of similar shape elute with a smaller retention volume from the column compared to smaller proteins. This is used to separate proteins of different size from each other to get the best possible monodisperse and pure protein solution in a defined buffer, which is an important aspect in protein crystallization. With the help of size standards, good analyts, i.e. proteins that do not interact with the matrix and have a similar shape as the proteins used for calibration, can be attributed an apparent size in solution, which gives information about the oligomeric state of the protein.

The concentrated elution from the IMAC is injected onto a HiLoad 26/600 Superdex 200 pg (GE Healthcare) that has been equilibrated with a sterile-filtered and degassed SEC-buffer until the conductivity reached a stable niveau prior to use. The protein containing fractions are detected at 280 nm and collected in 1.5 ml fractions. After SEC, the protein is either flash

frozen in liquid nitrogen and stored at -80°C , directly used for crystallization or further purified by anion exchange chromatography, if necessary.

SEC-buffer	
NaCl	200 mM
HEPES	20 mM
	pH 7.5

3.3.2.3 Anion Exchange Chromatography

Proteins produced with the low-temperature protocol described above showed minor, but specific impurities, which could be removed for initial crystallization screening by a further purification method called *anion exchange chromatography* (AEC). The stationary phase of ion chromatography columns binds anions or cations depending on its matrix. The selection depends on the isoelectric point (pI, i.e. the pH value without a net charge) of the protein and the buffer system that is used. In buffers with a pH lower than the respective pI, the protein will be positively charged and *vice versa*. The working pH should be 1-2 units above or below the protein's pI. In low salt buffer systems, all proteins with the same charge will bind to the chosen column. By using increasing concentrations of salt or a change in pH, the bound proteins can be eluted from the column.

Here, the protein is loaded in AEC-1 buffer on a 1 ml Bio-Scale Mini UNOsphere High Q column from Bio-Rad. With a continuous gradient using AEC-1 and AEC-2, the protein of interest eluted at 12% of AEC-2.

AEC-1		AEC-2	
NaCl	10 mM	NaCl	1 M
HEPES	20 mM	HEPES	20 mM
	pH 8.0		pH 8.0

3.3.2.4 Amylose affinity chromatography

MBP can not only be used as a carrier protein to drive difficult, recombinant proteins in the soluble state, but it can also be used as an affinity tag for initial purification. MBP has a natural affinity for α 1,4-maltodextrins, which is used in amylose resins as an affinity matrix for capturing the fusion protein. The elution occurs competitively by the addition of maltose to the buffer.

MBP-ScDcw1 is loaded after removing the cell debris on a 15 ml amylose column equilibrated with 10 CV MBP-1 buffer. The column is washed with the buffer until A_{280}

reached baseline. Subsequently, the protein is eluted with MBP-2 buffer containing 10 mM maltose.

MBP-1		MBP-2	
NaCl	300 mM	NaCl	300 M
HEPES	20 mM	HEPES	20 mM
	pH 7.5	Maltose	10 mM
			pH 7.5

3.3.3 Protein Analytics

3.3.3.1 Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE)

Sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) describes a technique, which allows the separation of biological macromolecules according to their molecular weight under denaturing conditions (Laemmli, 1970). To ensure that, proteins are treated with SDS, which denatures proteins and simultaneously facilitates a negative charge that is proportional to the size of the protein. Furthermore, β -mercaptoethanol in the SDS-buffer reduces possible disulfide bonds. The generally described heat step can be skipped for most proteins in solutions. SDS-samples are loaded on a discontinuous SDS-PAGE. This type of SDS-PAGE allows nice results by sample focusing in a stacking gel that is based on isotachopheresis established by chloride and glycine at pH 6.8 and a size-dependent migration of the denaturated proteins in the separation gel with a pH of 8.8. By using a size standard it is possible to attribute apparent molecular weights to the protein bands and herewith control the progress of protein purification upon visualization with Coomassie brilliant blue R250, which binds to basic amino acids, especially arginine side-chains.

3-12 μ l SDS-sample and 5 μ l Pierce Unstained Protein MW Marker are pipetted in the pockets of an SDS-PAGE with a 4.5% (v/v) stacking gel and a 12% (v/v) separation gel. After the run, the gel is stained in hot Coomassie and subsequently destained in hot destain solution until the bands are well visible.

2x SDS-buffer		12% SDS-PAGE	Stacking Gel	Separation Gel
Tris/HCl, pH 6.8	62.5 mM	Acrylamide (30%)	6.67 ml	32 ml
Glycerol	15%	SDS (10% w/v)	500 μ l	800 μ l
SDS	4% (w/v)	dH ₂ O	29.8 ml	32 ml
Bromophenolblue	<i>a pinch</i>	Buffer*	12.5 ml	20 ml
β -mercaptoethanol	4% (v/v)	APS (10% w/v)	500 μ l	800 μ l
		TEMED	50 μ l	80 μ l

Stacking Gel Buffer*		Separation Gel Buffer*	
Tris/HCl, pH 6.8	625 mM	Tris/HCl, pH 8.8	1.125 M
Ethanol	400 ml	Saccharose	30% (w/v)

Coomassie		Destain	
Coomassie brilliant Blue R250	3.2 g	Ethanol	400 ml
Ethanol	400 ml	Acetic Acid	80 ml
Acetic Acid	80 ml	dH ₂ O	400 ml
dH ₂ O	400 ml		

3.3.3.2 Determination of Protein Concentration

For the sake of reproduction and reliability in protein biochemistry, it is necessary to know the concentration of the components including that of the protein used. To determine the concentration of the purified protein the extinction E of the respective protein at a wavelength of 280 nm was measured with a NanoDrop. It applies the law of Lambert-Beer

$$E = \varepsilon \cdot c \cdot d \quad \text{eq. 1}$$

E : Extinction, ε : molar extinction coefficient, c : concentration, d : thickness of irradiated object

that can also be given as

$$c_m = \frac{E \cdot MW}{\varepsilon \cdot d} \quad \text{eq. 2}$$

c_m : mass concentration, MW : molecular mass

The theoretical molecular mass and the extinction coefficient is calculated based on the amino acid sequence of the protein using the online tool Protparam (Gasteiger et al., 2005).

3.3.3.3 Thermal Shift Assay

The *Thermal Shift Assay* (TSA) is a biochemical method to easily determine the thermal stability of a protein due to sensitive fluorescence detection of an externally added dye, which binds to the protein upon denaturation. Interestingly, the thermal stability of proteins highly depends on the composition of the buffer cocktail. Usually, standard buffer systems are used for protein purification that can still be optimized for individual proteins in terms of pH, salts and further additives. Especially in X-ray crystallography, the optimal buffer composition can be crucial for successful protein crystallization as the protein is more stable and conformationally homogeneous (Huynh and Partch, 2015; Reinhard et al., 2013). As the

binding of small molecules can also have an impact on thermal stability, this method can also be used to screen for potential ligands.

To do so, the protein is diluted in the TSA-mixture containing the fluorescence dye Sypro Orange. The fluorescence of the dye is detected depending on the applied temperature gradient. As long as the protein is natively folded, the dye cannot bind and remains in solution. Here, the fluorescence is quenched. As the temperature increases constantly, the protein starts to denature at some point. The unfolding of the protein unmasks the unpolar core-residues to which the dye can bind and develop its full fluorescence. This increase of fluorescence is detected and the resulting curve is used to determine the melting point of the protein, which equals the inflection point of the melting peak (i.e. the maximum of the first derivation).

The buffers tested differed in their buffer substances used and the respective pH-value. All tested buffers contained 100 mM NaCl. The 5000x stock solution of Sypro Orange is 1:62.5 diluted with distilled water before usage.

The TSA-mixture and the respectively used buffers are listed below:

TSA-mixture					
Sypro Orange dilution		4 μ l			
CtDfg5 (100 μ M)		2 μ l			
Buffer		ad 40 μ l			
Acetate buffer		MES buffer		HEPES buffer	
NaCl	100 mM	NaCl	100 mM	NaCl	100 mM
Acetate	50 mM	MES	50 mM	HEPES	20 mM
adjust to pH 4.5 or 5.5		adjust to pH 6.0 or 6.5		(DTT 2 mM)	
				adjust to pH 7.5	

3.3.3.4 α 1,6-Mannanase Assay

In order to test fungal GH76-proteins concerning their ability to hydrolyse α 1,6-mannooligosaccharides, 10 μ M CtDfg5 and 5 mM α 1,6-linked mannotriose or mannohexaose buffered in acetate buffer (100 mM NaCl, 50 mM acetate, pH 5.5) or HEPES buffer (100 mM NaCl, 20 mM HEPES, pH 7.5) are prepared. The reactions and the negative control without protein are incubated at 30 °C for 22 h and subsequently analyzed by thin layer chromatography in comparison with standard solutions. The standards are composed of 5 mM sugar solutions from mannose, α 1,6-mannobiose, α 1,6-mannotriose, and α 1,6-mannohexaose.

3.3.3.5 Thin layer chromatography

The biochemical properties can be determined by enzyme activity assays and the analysis of the resulting products. Unfortunately, carbohydrates lack chromophoric groups making them invisible for commonly used UV/Vis-detectors in liquid chromatography systems. However it is easy to analyze mono-, di-, and shorter oligosaccharides by thin layer chromatography (TLC). To do so, 5 μ l of the sugar-containing solutions (i.e. reaction mixtures or standards) are spotted on the stationary phase (i.e. precasted silica gel 60 F₂₅₄ plates (5 cm x 10 cm), Merck). The development of the TLC occurs by using a liquid phase consisting of 70:30 acetonitrile/water in a TLC chamber that was saturated with the mobile phase over night prior to use. The run starts from the bottom line (ca. 1 cm distance to the lower edge) sitting in <0.5 cm of the mobile phase and is terminated when the solvent front reaches a virtual line, which is about 2 cm from the top end of the plate. Then, the plate is dried with hot air and subsequently dipped in the *p*-anisaldehyde containing visualization reagent, which is subsequently developed at 110 °C for 10 min. Depending on the type of sugar, the sugar-derivates can produce blue, green and violet spots.

TLC visualization reagent

<i>p</i> -Anisaldehyde	1 ml
Sulfuric acid (97%)	1 ml
Ethanol	18 ml

3.3.3.6 Preparation of soluble GPI-APs from *S. cerevisiae* and the GPI-glycan hydrolysis assay

As a suitable GPI-anchored substrate is not commercially available and the chemical synthesis of complex sugars is extremely difficult, a promising approach could be the isolation of the substrate from the fungus directly. In order to do so, *S. cerevisiae* cells with a BY4741 background possessing a $\Delta dcw1$ knockout are taken from cell patches grown on YPD agar plates to inoculate 40 ml YPD medium, which is incubated over night at 30 °C shaking with 250 rpm.

YPD medium

Yeast extract	10 g/l
Peptone	20 g/l
Glucose	20 g/l
Agar-agar (for plates only)	20 g/l
dH ₂ O	ad 1 l

The media are autoclaved before use at 121 °C for 10 min. A preculture is taken for starting 2 l main culture in YPD medium, which grows another 24 h at 30 °C in baffled flasks at 150 rpm. Upon harvesting with 3000g for 20 min at room temperature, the cell pellet is resuspended in the lysis buffer (10 mM NaCl, 20 mM HEPES (pH 8.0), 2 mM DTT (added freshly prior to use), 10 mM EDTA, 2 mM PMSF (added freshly prior to use), and one tablet of cOmplete™ Protease Inhibitor Cocktail to a final volume of 50 ml. Subsequently, 10 mg Zymolyase 100T is added to the suspension and incubated for 90 min at 30 °C. Then, the cells are lysed by cycling the suspension in a microfluidizer at 200000 psi for 20 min. From now on, the steps are prepared on ice or at 4 °C. Unbroken cells and insoluble debris is removed from the cell lysate by two low speed centrifugation steps at 3000g for 5 min each. Subsequently, membranes are pelleted with 100000g for 1 h, the supernatant is discarded and the pellet is homogenized with a Potter homogenizer in the lysis buffer without DTT and the protease inhibitor cocktail prior to another centrifugation step at 100000g for 1 h. This wash step is repeated a second time, upon which 1 U of PI-PLC from *Bacillus cereus* is added to the homogenized and washed membrane to release GPI-anchored proteins from the membrane. The reaction is incubated over night on a roller wheel and the membrane is pelleted again at 100000g for 1 h. The resulting supernatant is analyzed by loading on an SDS-PAGE (see above), however the sample is just allowed to enter the separation gel. Upon Coomassie-staining, the protein bands are cut out and the proteins are identified by mass spectrometry. The presence of GPI-anchored proteins suggested the successful release and the conversion of GPI-APs to soluble GPI-APs (sGPI-AP). The supernatant, which is stored upon flash freezing in liquid nitrogen and kept at -80 °C, is then used for the final treatment with CtDfg5. 2 mg of CtDfg5 are added to 2 ml of the supernatant. The reaction is incubated at 30 °C for 6 h, the negative control is incubated without the added protein. In order to remove the proteins and other high molecular substances, the reaction is then filtered through a 3 kDa cutoff membrane and subsequently analyzed by the mass spectrometry facility and the NMR facility concerning the expected product mass and specific chemical shifts.

3.4 Protein crystallography

How proteins can be attributed certain functions according to their primary amino acid sequence was subject of the bioinformatics part above. However, such *in silico* analysis can only help to make hypotheses. They are the basis to plan experiments, which support the theories or disprove them. One major aspect in the comprehension of protein function is the

knowledge of their three-dimensional structure. The three major techniques to get atomic insights (i.e. yielding resolutions corresponding to $<3 \text{ \AA}$) of biological macromolecules are nuclear magnetic resonance spectroscopy (NMR), electron microscopy (EM), and X-ray crystallography. While NMR is limited in the size of the sample (routinely studied are proteins up to $\sim 30 \text{ kDa}$), the great potential of structure determination at high resolution by cryo-EM arose in the last years and is optimal for huge complexes such as ribosomes, polymerases, and proteins that do not crystallize. However, today (April 2019) approx. 10.5% of the structures deposited in the protein data bank (PDB) were solved by NMR ($\sim 8.5\%$) and cryo EM ($\sim 2\%$), while more than 89% have been determined by X-ray crystallography. Basically, the three-dimensional structure of biological macromolecules of any size and kind (DNA, RNA, proteins, and their complexes) can be determined by X-ray crystallography. As the name indicates, the *only* requirement is their crystalline state. The steps that are necessary to determine the crystal structure of a protein will be described in the following with reference to the beautiful monographies of Jan Drenth, Bernard Rupp, and Gale Rhodes (Drenth, 2010; Rhodes, 2006; Rupp, 2010)

3.4.1 Protein crystallization

Although proteins have been crystallized on purpose since the middle of the 20th century, their individual properties (e.g. sequence, weight, and pI) still do not allow the a priori prediction of working crystallization conditions. While data collection and structure refinement is becoming more and more time efficient due to technical and computational improvements, the crystallization of proteins is still the bottleneck in X-ray crystallography. Nonetheless, a large number of commercially available screens and pipetting robots allow the screening of thousands of conditions in a straight-forward way.

The actual process of protein crystallization requires a pure and monodisperse protein solution with a sufficient concentration. This means that the solution contains the protein of interest in a distinct oligomeric state (almost) without other protein impurities. Said state is realized by consecutive steps of different purification methods up to the experimenter's satisfaction. Typical concentrations are between 5-50 mg/ml that require a reasonable yield of the protein. It can also be helpful to choose an optimal buffer system for the protein, which is determined by a thermal shift assay. The crystallization of the protein is achieved by reaching a supersaturated solution in which the protein starts to precipitate in a highly ordered manner, i.e. spontaneous formation of critical nuclei that can grow to well ordered macroscopic

crystals. This uncertain process depends not only on the protein concentration, but also on factors like pH, temperature, salt, precipitant, and other additives.

Using high-throughput screening and a pinch of luck the correct conditions can be identified. A common method to achieve supersaturation is vapor diffusion, which was also used in this work in its two variants *sitting* and *hanging drop*. Here, the undersaturated protein solution is mixed with the mother liquor in defined ratios. Within a closed system the water net diffusion is via the vapor phase from the drop into the mother liquor, as the substances from the reservoir are diluted within the drop. This results in a gradually decreasing volume within the drop and thereby an increase of the concentration (Figure 9). In an optimal experiment, this process leads from an undersaturated condition (yellow circle) to a concentration within the nucleation zone (red circle), where critical nuclei can form spontaneously by overcoming the energy barrier. Once happened, the state drops into the metastable zone, as the protein concentration is decreasing (blue circle). In that zone, crystals can grow without further nucleation. Finally, the condition reaches equilibrium of the undersaturated and saturated phase, where no further growth can happen. Within the precipitation zone the protein precipitates in an amorphous state and is therefore undesirable. However, in practice the general absence of amorphous precipitate is a good indicator that the initial protein concentration is not high enough.

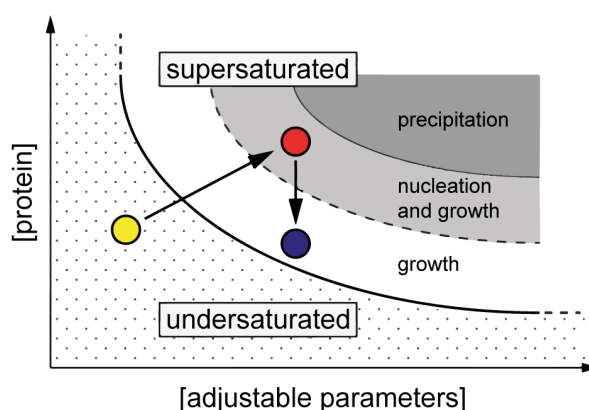


Figure 9: Phase diagram of an ideal crystallization experiment by vapor diffusion. The protein concentration is shown as a function of the precipitant concentration. Starting from the undersaturated situation (yellow circle), the concentration is shifted into supersaturation by vapor diffusion. Upon reaching the nucleation zone, critical nuclei can form and the crystal can start to grow (red circle). As the protein is removed from solution, the protein concentration decreases into the metastable growth zone, where only growth and no further nucleation can occur (blue circle) (adapted from Rhodes, 2006).

Commercially available screens were used for initial crystallization trials. Such *screening suites* contain 96 different conditions, which have been empirically shown to result in a good

number of crystal hits. The initial screens are pipetted in MRC 2 well crystallization sitting drop plates (*Swissci*) with the crystallization robot Honeybee 963TM (*Digilab*) by filling 80 µl mother liquor in the reservoir and subsequently using 300 nL to fill each well. After that 300 nL of protein solution is added to each well and finally sealed with sealing foil. The incubation is carried out at 4 °C and 18 °C in a Rock Imager (*Formulatrix*) documentation system that is taking pictures of each well according to a given schedule. Based on grown protein crystals, a second type of initial screens using microseeding has been used. The advantage here is that spontaneous nucleation is not necessary as micro crystals are added to the condition directly. Herewith it is possible to find conditions that are optimal for crystal growth, but not for nucleation. To prepare a seed stock, a seed bead is chilled on ice in a 1.5 ml reaction tube with 50 µl of the respective reservoir solution. A few nice looking crystals are then crushed in its well with a suitable glass probe until no bigger crystals are visible anymore. The drop is diluted with 6 µl of the reservoir solution and then transferred into the seed bead tube. This is repeated two more times and after that the tube is vortexed for two minutes with 30 s cool down phases after every 30 s. The resulting seed stock is stored at -20 °C. For pipetting with the Oryx8 (*Douglas Instruments*), a 1:10 dilution of the seed stock with the respective reservoir condition is prepared and directly used. For each condition, 300 nL protein are mixed with 200 nL reservoir solution and 100 nL seed stock.

For reproduction of *CtDfg5* crystals, the seeded JCSG core IV E7 condition has been used in a 24-well hanging drop approach. While the 3:2:1-ratio has been maintained, the final volume was increased by doubling the initial volumes. In fact, the crystallization of *CtDfg5* using this approach was successful for protein concentrations between 10-30 mg/ml.

3.4.2 Soaking of protein crystals

Before going to the synchrotron for collecting diffraction data, it may be necessary to prepare the crystals in certain cases. Soaking of protein crystals describes the exposure of grown crystals to small molecules in a solution that is as close as possible to the growth condition. Soaking can be necessary to protect the crystal from ice formation, to solve the structure or to get ligand-bound states.

Ice formation can occur while flash freezing the crystals in liquid nitrogen, as the crystals consist of a remarkable amount of water (i.e. ~30-70%) and they are harvested from an aqueous solution. Ice rings result in a loss of information during data collection, as they overlay with the actual diffraction data. By adding cryogenic protection substances such as

glycerol, glucose or high molecular polyethylene glycols to the growth condition, the formation of ice can be avoided by soaking the crystal in cryo solution prior to freezing. Cryo solutions used in this work contained 30% glycerol. As they can have negative impact on crystal quality, it is advisable to check the diffraction without cryo protection and, if necessary screen for better working compounds. When such chemicals are present in the mother liquor, cryo protection may not be required at all.

In order to solve the phase problem of protein crystallography (described below), it is possible to utilize heavy metal atoms to recalculate the phases lost during data imaging. One way to introduce heavy metals in the protein structure is by substituting the native methionine with selenomethionine (i.e. the sulphur is changed to a selenium). However, the whole process of protein production, purification and crystallization needs to be repeated without a guarantee of success. Another approach for introducing heavy metals is using existing crystals and soaking them in a solution containing the respective atoms. In this work, gadolinium acetate, $\text{Gd}(\text{OAc})_3$, was used as the acidic pI of 4.76 for *CtDfg5* suggested a reasonable amount of negatively charged amino acids, which are suitable for specific Gd^{3+} -binding. The lanthanide was used as previously described in our lab by soaking with a concentration of 50 mM $\text{Gd}(\text{OAc})_3$ with subsequent back soaking into the mother liquor containing 30% glycerol before flash freezing (Veelders and Essen, 2012).

The third purpose for the use of soaking was used in this study is to achieve substrate bound states. In principle, there are two ways. One is cocrystallization, which means that the ligand has been added to the protein prior to crystallization. This can happen by co-purification, if the ligand is present during protein production or purification and the affinity is high enough to keep it bound. Alternatively, ligands can be added prior to the crystallization screening. However, sometimes the actual ligand is not available, crystals do not grow in the presence of the ligand or the affinity is not high enough to force the protein into the bound state. Here, soaking can help. One main issue is that the addition of ligands can destroy the crystal lattice upon binding, either by interfering with the crystal contacts, or by inducing a conformational change that is also not compatible with the apo crystal form. On a macroscopic level, cracks can become visible within the crystal, the crystal can lose its sharp shape or they just dissolve. Furthermore, the crystal quality can suffer from the treatment. However, if the ligand is suitable for soaking, it enables the experimenter to utilize the apo crystals and convert them into ligand-bound states in a quite easy manner. For *CtDfg5*, GPI-anchor glycans were not commercially available. Therefore, fragments of the potential substrates have been used in

different concentrations and time points. Soaked crystals were directly flash frozen in liquid nitrogen from the soaking solution.

3.4.3 Principles of X-ray diffraction and data collection

The aim of the whole crystallographic procedure is to get the three-dimensional distribution of electrons from a molecule, which maps its appearance and therefore provides the structural biologist a precise picture (or model) of the protein. That the combination of crystals and X-rays is a good idea to study the atomic structure of molecules has been discovered 100 years ago, which has been awarded with two Nobel prizes in physics. The first one in 1914 honoring the work of Max von Laue for showing X-ray diffraction by crystals made of copper sulfate. One year later, in 1915, father and son William Henry Bragg and William Lawrence Bragg received the Nobel prize for extending that knowledge to analyze the actual crystal structure by means of X-ray diffraction. This is based on the periodic arrangement of the molecules within the three-dimensional crystal lattice. The repeating unit of the lattice is called *unit cell* of which the whole crystal lattice can be reconstructed by simple translation operations (Figure 10A). The unit cell itself consists of the *asymmetric unit* (AU), which is the smallest unit of a crystal. By applying rotational and translational symmetry operations on the AU, the unit cell can be generated. Basically, a diffraction pattern is the joined version of the objects that make the crystal. In the Bragg model, crystals are made up of a set of equally spaced planes that are parallel to each other and act as mirrors for the incident X-ray beam (Figure 10B).

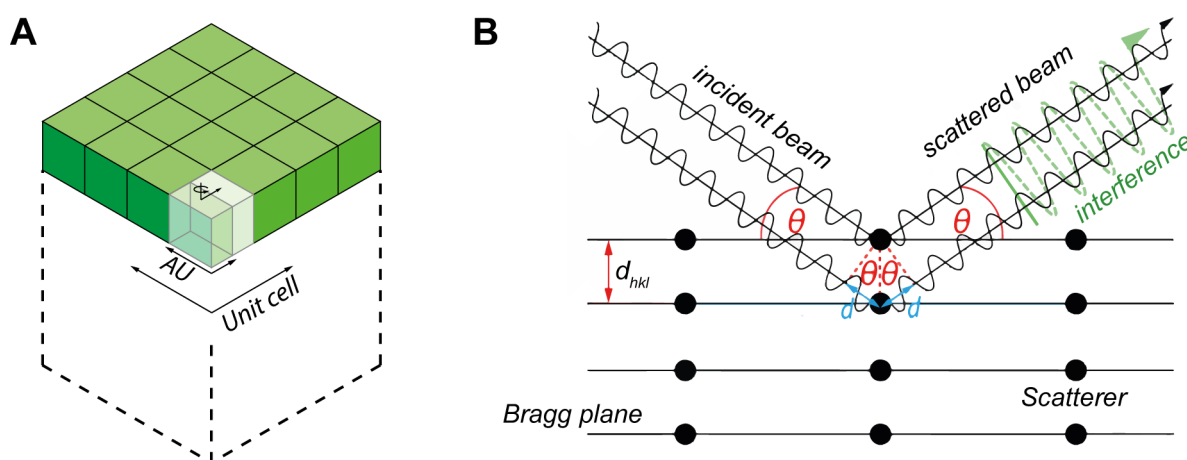


Figure 10: Schematic presentation of a three-dimensional crystal lattice and Bragg diffraction. **A** Crystals are made up of a periodic arrangement of its molecules. The *unit cell* is the repeating unit of the crystal, which reconstructs the crystal by translation operations in all three dimensions. The unit cell itself can consist of an *asymmetric unit* (AU). By rotational and translational operations, the unit cell can be built from the AU. **B** When applied on a crystal, the incident X-ray beam paths

on scatterers (black dots) within the crystal lattice, resulting in interaction with the scatterers. The horizontal line represents the Bragg planes, which are separated to each other by the interplanar distance called d_{hkl} . θ indicates the scattering angle, d is the path difference and the green dashed arrow indicates the resulting interference wave, when Bragg's law is fulfilled (see text for details).

When the beam interacts with the electrons of the atoms, it is converted into secondary spherical waves. This means that the scattered beam in principle goes into all directions, however only in one special case the resulting beam can interfere constructively and produce Bragg reflexes. This special case is mathematically described by *Bragg's law* and can be well explained by the illustration shown in Figure 10B. Here, the horizontal lines represent planes of one family of Bragg planes with incident and reflected beams that make an equal angle with the respective planes. To allow constructive interference of the two incident rays intersecting the plane, the scattered rays must be exactly in phase. This is only the case, if the lower scattered beam travels the additional distance depicted by d and if the traveled distance is equal to the wavelength λ , as the constructive interference requires the wave crests to be in phase. By applying simple trigonometric rules this results in *eq. 3*.

$$\lambda = d_{hkl} \sin \theta \quad \text{eq. 3}$$

That distance described by *eq. 3* must actually traveled twice by the beam as indicated in Figure 10B, it follows *eq. 4*.

$$\lambda = 2 \cdot d_{hkl} \sin \theta \quad \text{eq. 4}$$

Given the fact that for all other Bragg planes for that angle the distance to travel becomes a multiple of λ , the formula must be extended by the number of planes n , where n is an integer to allow constructive interference, resulting in *eq. 5* that is known as *Bragg's law*.

$$n\lambda = 2d_{hkl} \sin \theta \quad \text{eq. 5}$$

n : integer, λ : wavelength, d_{hkl} : distance between lattice planes, θ : diffraction angle

To get all the information of a crystal (i.e. expose all Bragg planes to the primary beam in the correct angle) many orientations must be analyzed during data collection.

For data collection, a protein crystal is picked with CryoLoopsTM directly from the growth condition and is flash frozen in liquid nitrogen. Using the rotation method, crystals were

measured at synchrotron beamlines (ID23-1 and ID-29 at the ESRF; PXIII at the SLS) with the exception of the initial dataset needed for phase determination, which has been measured at the inhouse source of AG Klebe (Pharmacy, Philipps-Universität Marburg) using a I μ S microfocus tube (*Incoatec*) and a MAR345 image plate detector (*Incoatec*) (Arndt, 1968; Nurizzo et al., 2006; De Sanctis et al., 2012). At the synchrotron, the loop is mounted automatically to a goniometer head with the crystal being protected by a cryostream running at 100 K. The goniometer enables the experimenter to rotate the crystal in any possible orientation through the primary beam, which is necessary to center the crystal in the beam in any orientation. Before collecting datasets, the crystal is characterized by two single measurements differing by 90° to assess an optimal collection strategy in terms of completeness and resolution by adjusting parameters such as exposure time, oscillation range, number of images and detector distance.

3.4.4 Processing of diffraction data

To proceed from data collection to structure determination, two major tasks must be fulfilled. The first task includes the assessment of crystal symmetry and the integration of the observed reflexes, which was done with the X-ray Detector Software (*XDS*) (Kabsch, 2010). After that data are reduced and judged by several parameters using the data reduction tool from the CCP4i2 software package (Potterton et al., 2018).

The input file *xds.INP* is automatically created at the ESRF and SLS and is executed by the *xds_par* command in the shell. The software follows the path of eight different program steps that must be fulfilled in a fixed order, as the generated output files are used in the next steps. The software starts with the correction of geometrical distortions by using *XYCORR* (which are in fact usually already corrected and saved in the provided folder), followed by *INIT* that classifies pixels to discriminate between background and diffraction. Then diffraction spots are identified and located by *COLSPOT*, which are used in the subsequent indexing step, where *IDXREF* determines the orientation and symmetry of the crystal lattice, which explains the found spots best. As a result, every reflection of every diffraction image is assigned to their respective position in the reciprocal space using Miller indices (*h,k,l*). After that *DEFPIX* defines a trusted region and defines pixels as untrusted, if they are obscured by hardware or excluded by a defined resolution range. The next step *XPLAN* is a tool for planning data collection, for which it estimates the completeness of new data for each starting angle and the crystals' total rotation. *INTEGRATE* then uses a two-step procedure to integrate

the reflexes and generates the `INTEGRATE.HKL`, which is used by *CORRECT*. It corrects intensities and standard deviations and writes out the finally integrated reflexes in `XDS_ASCII.HKL` including space group determination, refined unit cell constants and overall quality tables of the dataset.

The resulting `XDS_ASCII.HKL` file is directly imported into the X-ray data reduction and analysis module of the CCP4i2 software package, which comprises four programs. A few of these steps are redundant to some options that are possible within *XDS*, however rerunning data reduction here provides a good graphical output and careful data analysis can be done. After space group determination by *POINTLESS*, *AIMLESS* scales the data to a user defined cutoff value and *CTRUNCATE* generates amplitudes from the given intensities. Where to set the resolution cutoff to assess significant diffraction data is still a matter of debate. Current literature suggests to use the internal consistency determined by $CC_{1/2}$ as the major quality parameter (Karplus and Diederichs, 2012, 2015). This value randomly splits the data into half and gives values from 1 (for perfect correlation) to 0 (for no correlation) (see eq. 6). Depending on the crystallographers philosophy, data are suggested to be insignificant if the coefficient is below 0.15, while a more conservative value sets the cut-off at 0.5.

$$CC = \frac{\sum (x - \langle x \rangle)(y - \langle y \rangle)}{\sqrt{\sum (x - \langle x \rangle)^2 \sum (y - \langle y \rangle)^2}} \quad \text{eq. 6}$$

Historically spoken, crystallographic data were typically truncated at a resolution, where a signal-to-noise ratio reached ~ 2 and merging R -factors exceeded $\sim 80\%$. R_{merge} indicates how well symmetry-related reflex intensities merge, which is defined by eq. 7.

$$R_{\text{merge}} = \frac{\sum_{hkl} \sum_i |I_i - \langle I \rangle|}{\sum_{hkl} \sum_i \langle I \rangle} \quad \text{eq. 7}$$

I_i = intensity for a reflex at position hkl in measurement i , $\langle I \rangle$ = average intensity for reflex hkl

Typically, R -values are given as percentage and in principle it says that an ideal dataset would have an R -factor of zero. However, in reality this is of course not the case and indeed this quality parameter is vulnerable in terms of data redundancy, which results in increasing R -values for data with higher multiplicity (Weiss and Hilgenfeld, 1997). Unlike R_{merge} , the redundancy independent merging R -factor R_{rim} or R_{meas} is a quality factor, which is robust to

multiplicity, as it adds a factor for data redundancy (*eq. 8*) (Diederichs and Karplus, 1997). These values are suggested to be useful additional parameters also in terms of the correct space group assessment, however they should not play a role in determining the resolution cutoff (Karplus and Diederichs, 2015)

$$R_{meas} = \frac{\sum_{hkl} \sqrt{\frac{N}{N-1}} \times \sum_i |I_i - \langle I \rangle|}{\sum_{hkl} \sum_i |\langle I \rangle|} \quad eq. 8$$

$N = \text{redundancy of data}$

Beside the merged set of observed intensities, a second output is created by *FREERFLAG* that uses 5% of the data to generate a free-*R* set. Herewith a set of reflexes is excluded from the structure determination, which is later used for cross-validation of the refinement (see below).

3.4.5 Structure determination by solving the phase problem

The overall goal in protein structure determination is to obtain a three-dimensional map of the electron density ($\rho(x, y, z)$) in the volume of the unit cell (V), which is defined as a sum of a Fourier series of all structure factors measured in the diffraction experiment (*eq. 9*).

$$\rho(x, y, z) = \frac{1}{V} \sum_h \sum_k \sum_l F_{hkl} e^{-2\pi i(hx + ky + lz)} \quad eq. 9$$

The structure factor F_{hkl} is the Fourier transform of the electron density within a unit cell and is described as an electromagnetic wave. Therefore it is necessary to know the three terms that describe a wave to get the information of the electron distribution: the wavelength, amplitude and phase (Figure 11).

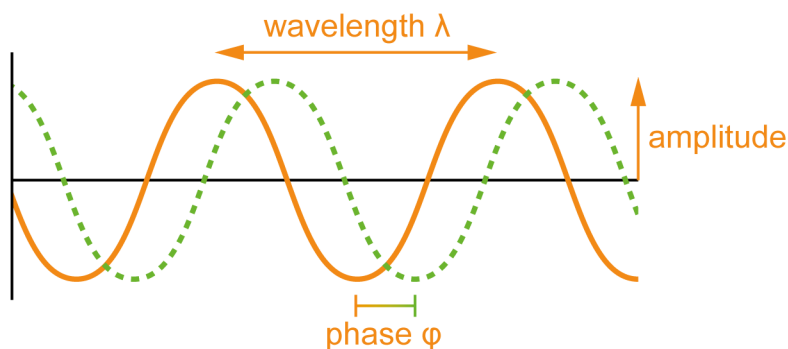


Figure 11: Characteristics of an electromagnetic wave. Two different waves are shown, which can be described by their amplitudes, wavelength λ and their phase ϕ .

While the wavelength is known from the experiment parameters used for data collection (no change of the incident beam upon elastic scattering) and the amplitude $|F_{hkl}|$ can be calculated as it is proportional to the measured spot intensity $I_{hkl}^{1/2}$, the phase can not be measured with the detection system used and is therefore lost by the experiment. This issue is called the *phase problem of protein crystallography*. A number of different methods are described to solve the phase problem (Taylor, 2010). Two of them have been used in this study: *single-wavelength anomalous diffraction* (SAD) and *molecular replacement* (MR).

The general principle of the SAD approach is based on the breakdown of *Friedel's law*, which says that the intensities of a pair of centrosymmetric reflections (the *Friedel pair*) at positions h,k,l and $-h,-k,-l$ are equal for normal scattering. This is true for crystals, which only possess light atoms such as C, O, N, H and S, as they do not exhibit strong anomalous scattering effects (although *native SAD is maturing* (Rose et al., 2015)). However, this effect can be very strong for heavy atoms such as selenium or heavy metals, which can be incorporated into the protein crystal by amino acid derivatives or crystal soaking. When the incident X-ray beam is close to the absorption edge of such atoms, the strength (f') and the phase (f'') of the scattering changes. As the differences of changed scattering factors are not equally, Friedel's law becomes violated: i.e. that the intensities of h,k,l and $-h,-k,-l$ are not the same anymore. This behavior is called *anomalous scattering* and depends on the type of heavy atom and its characteristic absorption maximum, which can be experimentally determined at tunable synchrotron beamlines. By using the *Patterson* method it is possible to determine the position of the heavy atom within the unit cell (Patterson, 1934). Here, the electron density function (eq. 9) is used, however the phases are set to zero and just the experimentally given intensity data are required, which is the square of the structure factors.

$$P(x, y, z) = \frac{1}{V} \sum_h \sum_k \sum_l |F_{hkl}|^2 e^{-2\pi i(hx+ky+lz)} \quad eq. 10$$

As a result, the Patterson function peaks do not correspond for the exact atom positions, but to the vectors between the atoms. This works well for substructure determination when only a few sites are present and improves the efficiency of substructure solution with *SHELXD* when Patterson seeding is used instead of random starting atoms (Schneider and Sheldrick, 2002).

For solving the structure of *CtDfg5*, the *CRANK2* pipeline of the ccp4i2 suite was used for gadolinium-soaked crystals from *CtDfg5* including the program suite *SHELXC/D/E* (Sheldrick, 2010; Skubák and Pannu, 2013). With the obtained structural model for *CtDfg5*, all other data sets were solved by *molecular replacement* (MR).

In this approach, an already known structure, which has a similar primary sequence to the target (40% is high, 30% identity is usually successful, 20% can be successful) is used as a search model to determine initial phases (Dimaio et al., 2011). Basically, similar structures possess the same overall fold and by placing the search model in the correct orientation (α, β, γ) and position (x, y, z) of the unknown unit cell it is possible to obtain the correct phase. As a prerequisite, the Patterson maps from the measured intensities of the diffraction experiment (the observed structure factor amplitudes F_{obs}) and from the search model (the structure factor amplitudes F_{calc} derived from the model) must be calculated. Then, the six-dimensional problem is split in two three-dimensional problems. In the first step, the resulting maps are rotated against each other to get a best possible correlation between both maps and in the second step a translation function is used to position the search model to the correct coordinates within the asymmetric unit (Figure 12). If the positioning occurred correctly, initial phases can be calculated deriving from the information given by the input model. The software used in this study is *Phaser* implemented in the CCP4i2 suite (McCoy et al., 2007).

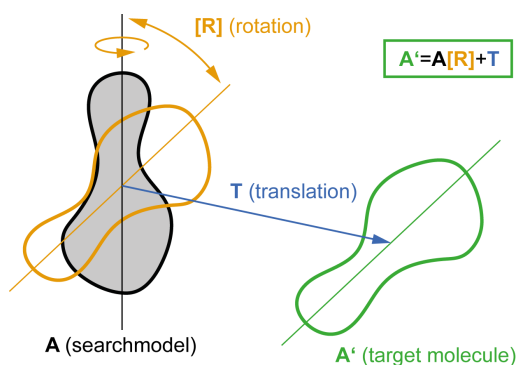


Figure 12: Visual explanation of molecular replacement. By applying rotational and translational variables on the search model A, its phases can be *borrowed* to calculate the phases of the target molecule A' without obtaining its phases experimentally.

In the optimal case, *Phaser* finds a unique solution at the end of the search. Three different parameters are considered to judge on the given solution. The *Log Likelihood Gain* (LLG) indicates how much better the model explains the diffraction pattern than a random distribution of atoms. As it is defined as the difference of the model likelihood and the Wilson distribution likelihood, the value must be positive and as high as possible. The second score is the *Translation Function Z-score* (TFZ), which is generally above 5 if the solution is likely to be correct and can clearly be distinguished from further solutions. As shown in Table 1, a definite solution can be assumed from *Z-scores* above 8.

Table 1: Probability table for a correct solution from *Phaser* after a successful molecular replacement job. The table is modified from https://www.phaser.cimr.cam.ac.uk/index.php/Molecular_Replacement (July 2019).

TFZ	Solved?
<5	no
5-6	unlikely
6-7	possibly
7-8	probably
>8	definitely

A further indication for a correct solution is the *clash score*, which should be below 10. However, looking at the clashes is in particular interesting, when a correct solution could not be obtained with the given search model. When using the default options of *Phaser*, 5% clashes are allowed for C α -atoms in the packing step. Solutions with higher clashes are rejected from further computing, although the *Z-scores* may indicate a reasonable solution. For such cases, either the threshold could be adjusted or the input model must be modified in terms of peripheral loop regions.

3.4.6 Structure refinement

Upon structure solution, first estimated phases with an initial model are obtained, but the job is not done yet, as the model still contains errors or is simply incomplete. The actual goal is to get a final model, which explains the crystallographic data as good as possible. The process from the initial model to the final structure is called *refinement* and consists of an iterative process of (1) manual model building by visual evaluation and modification of the model and its correlation with the electron density map in *Coot* and (2) the actual computational refinement using *Phenix.refine* (Afonine et al., 2012; Emsley et al., 2010). *Phenix.refine* works automatically by introducing small changes to the model that optimize the stereochemical properties and thereby generating better model amplitudes (F_{calc}). These are compared to the observed amplitudes (F_{obs}) upon which a new cycle is started. It therefore is intuitive that a correlation factor between F_{calc} and F_{obs} is used to evaluate the obtained structures and the refinement strategies (eq. 11).

$$R = \frac{\sum ||F_{obs}| - |F_{calc}||}{\sum |F_{obs}|} \quad eq. 11$$

Accordingly, a model that perfectly explains the data would result in an *R*-factor of zero, however due to imperfections of the crystal lattice these values usually range within 0.15-0.25. Since datasets with higher resolutions contain more detailed observations, it is possible to refine such models to better *R*-values than model from low-resolution data. In fact, two *R*-values are commonly used for structure evaluation. The above described factor is calculated from the *working* model, thus called R_{work} . As mentioned earlier, 5% of the merged data are used to generate a free-*R* flag, which is now used as a cross-validation method to further evaluate the refinement quality. R_{free} is calculated with the same equation as applied for R_{work} , however using the omitted diffraction data instead of F_{obs} (Brünger, 1992). As the model amplitudes are not fitted to the excluded data, this value is always greater than the R_{work} . Significant differences of the two parameters indicate an over-fitting of the model.

3.4.7 Structural analysis and visualization

The obtained structural model coordinates are written out as *pdb*-files, which can be visualized by UCSF Chimera (Pettersen et al., 2004). All figures containing protein structures were generated with this tool. The *ConSurf*-server has been used to plot evolutionary information on the single amino acids written in a *pdb*-file, resulting in nine differently

scored conservation categories that can be visualized by different colors (Ashkenazy et al., 2016)

3.5 Structure-based drug design using SeeSAR

The underlying rationale of finding potent antibiotics for a specific protein is to identify small molecules that are able to bind the target in such a manner that its actual biological task cannot be fulfilled anymore. In the case of essential proteins, such an interaction causes the death of the organism, which basically is the name-giving determinant of *antibiotics*. A plausible region, where such molecules should bind to enzymes is their substrate-binding region including the active site. A schematic workflow for structure-based drug development is shown in Figure 13.

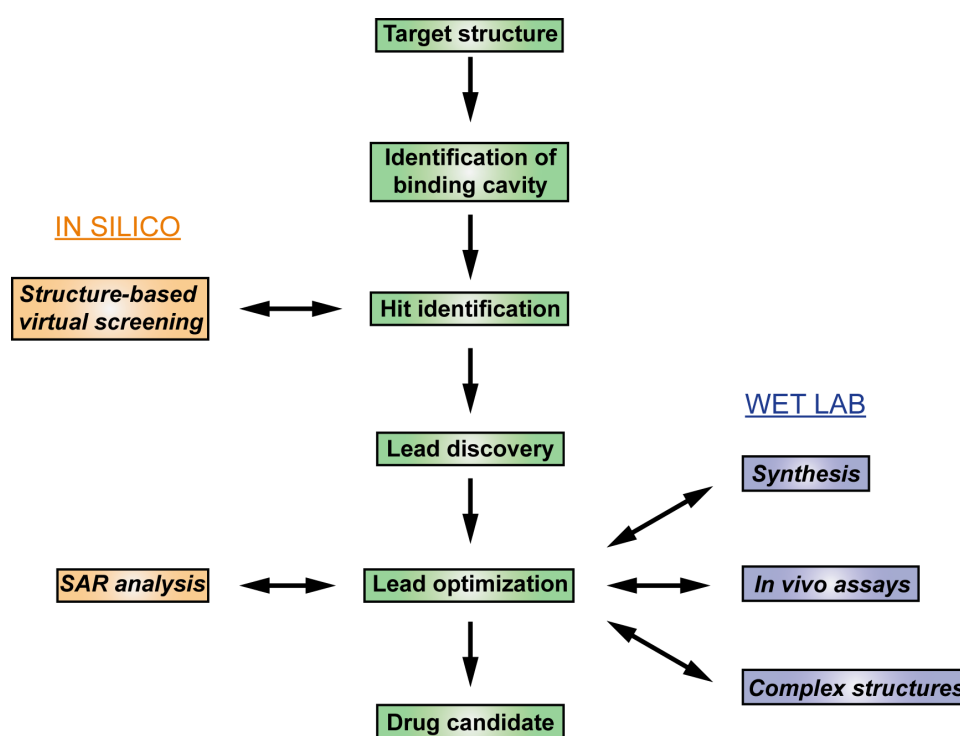


Figure 13: General workflow for structure-based drug discovery. The central green flow shows the general steps along which some tasks must be performed either *in silico* or in the wet lab (adapted from Andricopulo et al., 2009). SAR=structure-activity relationship

The absolute requirement of structure-based drug discovery is the three-dimensional protein structure, which is obtained in this work by X-ray crystallography. By comparing the protein to similar, already described structures, the binding cavity can be derived by structural alignments. Without prior knowledge, plausible regions can be assumed by visual inspection

of the structure concerning prominent cavities or pockets. Such a function is implemented in the software *SeeSAR*, which can automatically detect parts of the protein that are likely to bind molecules (BioSolveIT GmbH). The selected clouds are used to specify the potentially druggable binding sites, which are subsequently used for *Hit identification*. To do so, a molecule is loaded in the software, which then generates different poses with estimated binding affinities. The underlying algorithm scores the hydrogen bond and dehydration (HYDE) energies in the posed protein-ligand complex and thereby estimates the binding affinity (Reulecke et al., 2008; Schneider et al., 2013). The name-giving element of *SeeSAR* is that it allows to visually (so to say: *to see*) inspect the structure-activity relationships (SAR) of the leads bound to the protein. Atoms contributing to the interaction are highlighted in green, non-favoring atoms are labeled in red. Together with statistically derived torsion-angles from the CCDC small molecule crystal database, the user can modify and optimize the bound ligand with instant feedback of the applied structural changes (Guba et al., 2016; Schärfer et al., 2013).

In this work, a test license of *SeeSAR* v.8.1 was used to screen for the best potential binders from the commercially available *Frag Xtal Screen* from Jena Bioscience. Ten poses for the 96 molecules were generated in the substrate-binding pocket of *CtDfg5*. The generated poses were then prioritized by the estimated affinities and the best twelve binders were used for lead discovery by soaking experiments as described above.

4 Results

4.1 Phylogenetic analysis of GH76 proteins

The power of bioinformatic tools to predict protein functions based on sequence similarities is commonly used in protein biochemistry to assume probable functions and to plan promising experiments for the protein in hand. To get an overview about the current knowledge of the glycoside hydrolase 76 family (GH76), the respective PFAM entry PF03663 has been subjected to an in-depth sequence similarity network (SSN) analysis with the goal to divide the family in isofunctional subfamilies (see Figure 14).

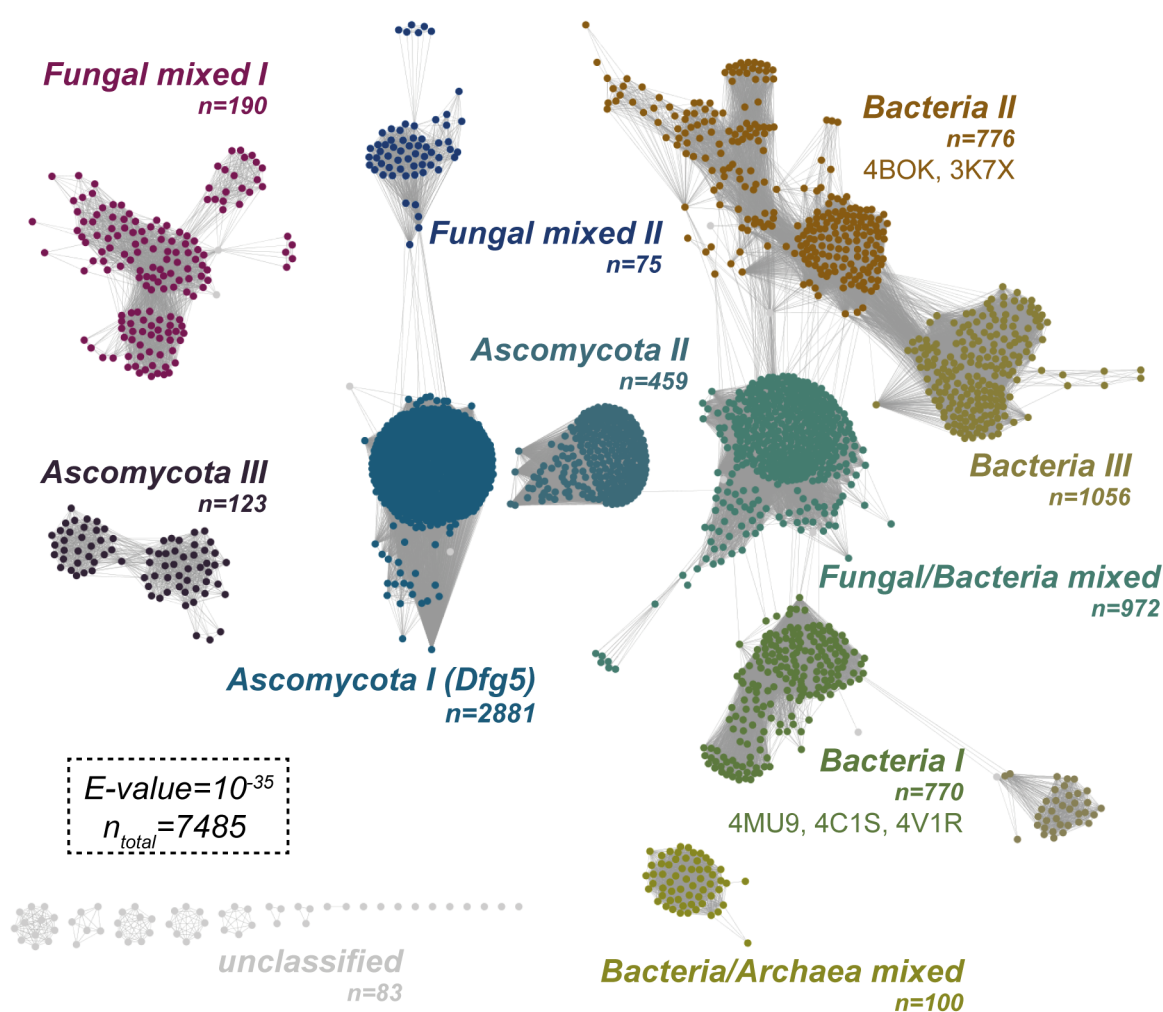


Figure 14: Sequence similarity network of GH76 proteins. The EFI EST-SSN of PF03663 shows the relationship of GH76 proteins using an E-value of 10^{-35} for all protein sequences with 300-700 amino acids. Every node harbors sequences of 40% identity. The applied parameters resulted in 7485 sequences in 2474 nodes and clustered into ten major subfamilies. The number of sequences *n* per subfamily is given. 83 sequences remained unclassified. If already deposited, PDB codes belonging to respective subfamilies are noted.

Using an E-value of 10^{-35} , the GH76 family falls apart in ten major subclasses. Five of them contain mostly bacterial proteins (*Bacteria I-III* and *mixed Bacteria/Archaea mixed*) and one subclass contains proteins of bacterial and fungal origins (*Fungal/Bacteria mixed*). Furthermore, three different subclasses were found to harbor proteins exclusively from Ascomycota (*Ascomycota I-III*) and another two subclasses contain mixed populations of fungal species (*Fungal mixed I/II*).

Two of the bacterial subclasses (*Bacteria I* and *II*) have already been characterized on a structural and biochemical level and both subclasses were attributed an α 1,6-mannanase function (Cuskin et al., 2015; Thompson et al., 2015b). In the literature, this activity has been generalized to the whole family. The largest subfamily in the SSN is the *Ascomycota I* class. Its first described member from *Saccharomyces cerevisiae* was found to have *defects in filamentous growth* and was called Dfg5 (Mösch and Fink, 1997). For clarity, the *Ascomycota I* subfamily will therefore be called *Dfg5* from here on. A few years later, Dfg5 and its homolog Dcw1 (*defective cell wall*) were described as essential factors for yeasts that are responsible for the incorporation of proteins into the fungal cell wall and it was suggested that the crosslinking happens via GPI-anchors (Kitagaki et al., 2002; Spreghini et al., 2003). Although being essential in *S. cerevisiae* and *Candida albicans*, a $\Delta dfg5/\Delta dcw1$ double-mutant in *Neurospora crassa* has been shown to be viable (Maddi et al., 2012). The present SSN provides a suitable explanation for this phenomenon. Unlike yeasts, the filamentous ascomycete *N. crassa* possesses seven different GH76 proteins, which are members of the *Dfg5*-subfamily (see Table 2). In the context of described functional redundancy, it is plausible that a double disruptant is not lethal in organisms with more than two of such paralogs.

Interestingly, the presence of just two copies of *Dfg5*-proteins seems to be rather the exception than the rule. Other ascomycetes, such as the thermophilic fungus *Chaetomium thermophilum* or the human pathogen *Aspergillus fumigatus* each possess six *Dfg5*-orthologs. Furthermore, *N. crassa*, *C. thermophilum*, and *A. fumigatus* harbor more GH76 proteins in the *Ascomycota II* and *mixed Fungal/Bacteria mixed* subfamilies. Apart from that, *C. albicans* possesses another five GH76 proteins in an additional *Ascomycota III* class, while just two homologs of *Dfg5*-proteins are present.

Table 2: Distribution of GH76 proteins in selected ascomycetes. Uniprot-IDs are shown for the respective organism and subclass. The proteins marked with an asterisk were used for calculating the identities shown in **Table 3**.

	<i>Saccharomyces cerevisiae</i>	<i>Candida albicans</i>	<i>Neurospora crassa</i>	<i>Chaetomium thermophilum</i>	<i>Aspergillus fumigatus</i>
Dfg5	Q05031*	Q5ACZ2*	Q1K7A8*	G0S3F2*	Q4WKP7*
	P36091*	Q5AD78*	Q7S4K4*	G0SFA3*	Q4WG09*
			Q1K7I4	G0SE99	Q4WFX5
			Q7SAB2	G0RXS3	Q4W985
			Q7RUC1	G0S8L9	Q4WFJ1
			Q7S3X3	G0S159	Q4WA43
			Q1K918		
			Q7SGV4*	G0SHT7*	Q4WI31
Ascomycota II					
Ascomycota III		Q59XS8*			
		A0A1D8PIN7*			
		A0A1D8PIJ7			
		A0A1D8PJF0			
		A0A1D8PFZ7			
Fungal/Bacteria mixed			Q7RYP1*	G0S5Y9*	Q4WIT8
			Q7S863	G0S8A0	

On a sequential level, selected *Dfg5*-proteins from Table 2 share sequence identities between 37-72% (see Table 3). It is suggested that proteins with a sequence identity of ~40% are similar enough that the general function can be transferred from characterized to uncharacterized proteins with a good degree of certainty (Tian and Skolnick, 2003). However, when looking at proteins from different subfamilies, the identities go down to ~20% and below. Such values have been described as the *Twilight zone of protein sequence alignments* and at this point reliable predictions of protein functions are not possible anymore (Rost, 1999). This supports the parameters applied to generate a SSN of GH76 proteins with segregated, isofunctional subclasses.

Table 3: Percent identities of selected proteins from the *Dfg5*-subfamily and the whole GH76 family

Dfg5		GH76	
Q4WKP7	44%	Q59XS8 (<i>Ascomycota III</i>)	14%
Q4WG09	39%	A0A1D8PIN7 (<i>Ascomycota III</i>)	13%
Q1K7A8	44%	Q9Z4P9 (<i>Bacteria II</i>)	18%
G0SFA3	48%	Q8A184 (<i>Bacteria I</i>)	17%
Q7S4K4	72%	G0S3F2 (<i>Dfg5</i>)	100%
G0S3F2	100%	Q1K7A8 (<i>Dfg5</i>)	44%
Q5ACZ2	37%	G0S5Y9 (<i>Fungal/Bac. mixed</i>)	20%
Q05031	40%	Q7RYP1 (<i>Fungal/Bac. mixed</i>)	21%
P36091	41%	Q7SGV4 (<i>Ascomycota II</i>)	20%
Q5AD78	37%	G0SHT7 (<i>Ascomycota II</i>)	23%

Taken together, the exact set of GH76 proteins in fungi is species-specific and the actual functions of the different subclasses are elusive, as for most classes *in vivo* data are not yet published. Despite the proposed contribution in cell wall protein incorporation, the mode of action is still a matter of debate and *in vitro* data still lack behind. While one side assumes that *Dfg5*-proteins hydrolyze the Man α 1,4-GlcN-linkage within the GPI-anchor, the other suggests the α 1,6-mannose backbone of N-linked outer chain mannan to be hydrolyzed and transferred to the CW glucan (Ao et al., 2015; Kitagaki et al., 2002; Maddi et al., 2012). The following chapter will solve the mechanistic paradoxon present in *Dfg5*-proteins.

4.2 The *Dfg5*-subfamily: Responsible for the incorporation of GPI-CWPs

As shown above, *S. cerevisiae* possesses only two paralogs of *Dfg5*-proteins and would be hence a good choice for *in vitro* studies, as a sophisticated model organism is available. Unfortunately, *Dfg5*-proteins from *S. cerevisiae* and from the closely related *Candida glabrata* could not be heterologously produced in the *Escherichia coli* expression system solely (SFigure 1 on p.134). A common approach to overcome this bottleneck in protein science is to use solubility tags, which can drive the fused protein of interest in a functional and soluble state. Dcw1 from *S. cerevisiae* could be produced with an N-terminally fused maltose-binding protein (MBP) using an amylose column for initial purification and subsequent SEC (SFigure 2 on p.134). MBP-ScDcw1 has been used for activity assays by Thierry Fontaine, where it was found to be inactive for all tested substrates (Muszkieta et al., 2019). As removal of the MBP-tag was not successful, it was not used for further studies.

Another approach to obtain soluble protein is to screen orthologs from thermophilic organisms, which have been described to have improved properties concerning their behavior in heterologous production and the following structural and biochemical studies due to their enhanced thermal stability (Cava et al., 2009). Unlike in bacteria and archaea, thermophilicity is less common in eukaryotes, but some fungi adopted an enhanced heat tolerance. Originally isolated from dung, the filamentous ascomycete *Chaetomium thermophilum* tolerates temperatures up to 60 °C and grows in standard media with a temperature optimum of 50-55 °C (Bock et al., 2014). The genome of *C. thermophilum* was sequenced in 2011 in order to provide a platform to study the nuclear pore complex (Amlacher et al., 2011). Therefore proteins from this fungus were chosen to be the subjects of an in-depth analysis.

4.2.1 Cloning, production and purification of recombinant CtDfg5

Suggesting an isofunctional subpopulation within the GH76-family, G0S3F2 from *Chaetomium thermophilum* (now termed as CtDfg5) found in the Dfg5-cluster is subjected to a detailed analysis in order to characterize the functional aspects of this subfamily. According to the annotations in the Uniprot entry, CtDfg5 is synthesized as type-I membrane protein containing an N-terminal signal peptide, which guides the nascent chain into the ER-lumen, and a C-terminal transmembrane helix (TMH). The presence of a TMH differs from the architecture of its yeast homologs, as the latter were described to harbor an ω -signal at the C-terminus that is attached to a GPI-anchor during protein maturation. In the final expression plasmid used for recombinant protein production in *Escherichia coli*, the terminal pro-peptide and TMH have been removed to obtain the protein in a soluble and matured state. CtDfg5 was amplified from cDNA of *Chaetomium thermophilum* and cloned into pET28a cut with *NheI/XhoI*. Production of soluble, recombinant protein could be achieved with the disulfide bond-promoting strain *E. coli* Shuffle T7 Express for three days at 12 °C induced with 0.1 mM IPTG (see Table 4 for protein chemical parameters).

Table 4: Theoretical properties of CtDfg5 calculated with ProtParam (Gasteiger et al., 2005).

Name	Uniprot-ID	Native amino acid range	Properties	Length	pI	MW	Ext. coefficient
CtDfg5	G0S3F2	30-449	His ₆ , Thrombin	443 aa	5.9	49.5 kDa	110.2 mM ⁻¹ cm ⁻¹

The protein was purified from pellets of 2 l liquid cultures that were lysed either with a microfluidizer or with a French press. The initial purification started with a Ni-NTA affinity chromatography of CtDfg5. To achieve a best pure and homogenous protein solution for further experiments, the elution of the IMAC was concentrated and further purified with size exclusion chromatography (SEC) (see Figure 15). About 8 mg per liter cell culture could finally be obtained after SEC. Although a slight delay could be observed compared to standards, the elution volume of CtDfg5 suggests a monomeric state of the protein in solution.

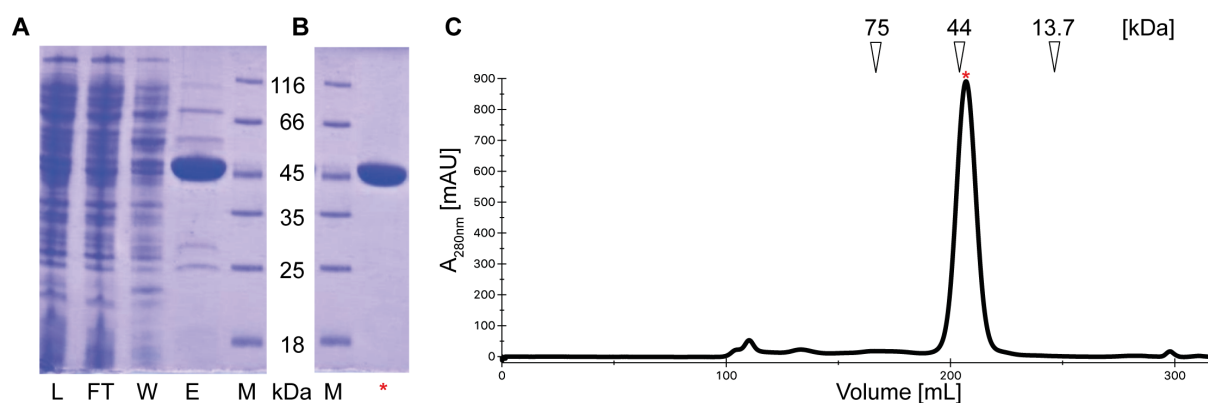


Figure 15: Purification of *CtDfg5*. **A** shows the IMAC purification with L=Load, FT=Flowthrough, W=Wash, E=Elution and M=Marker. **B** shows the peak of the size exclusion indicated by a red asterisk. **C** The chromatogram of the SEC is shown with the peak fraction from the SDS-PAGE indicated by the red asterisk and the arrows marking the elution volume of standard proteins with the respective molecular weight.

For the first crystallization screening that yielded the crystals shown in Figure 17, a further purification step was performed after the SEC step, as minor contaminations were still present. For that purpose, an anion exchange chromatography (AEC) has been performed, upon which the remaining contaminations were removed (SFigure 3 on p.135). The protein fractions indicated were subsequently concentrated to 3 ml and dialyzed over night with a 3 kDa cutoff dialysis membrane against 1 l AEC-1 buffer and then directly used for crystallization (see below).

As literature describes enzymatic assays for extracellular fungal proteins in acidic conditions to around pH 5, a buffer optimization screening was performed (Hurtado-Guerrero et al., 2009; Qin et al., 2015). Interestingly, the highest melting temperature was observed in the presence of 50 mM acetate buffered at pH 5.5, which is close to the pI of *CtDfg5* (see Figure 16). Furthermore the addition of 2 mM DTT leads to a decrease in stability of 5 °C. This supports the suggestion that disulfide bonds are present in the protein, as seven cysteines are present in the amino acid sequence.

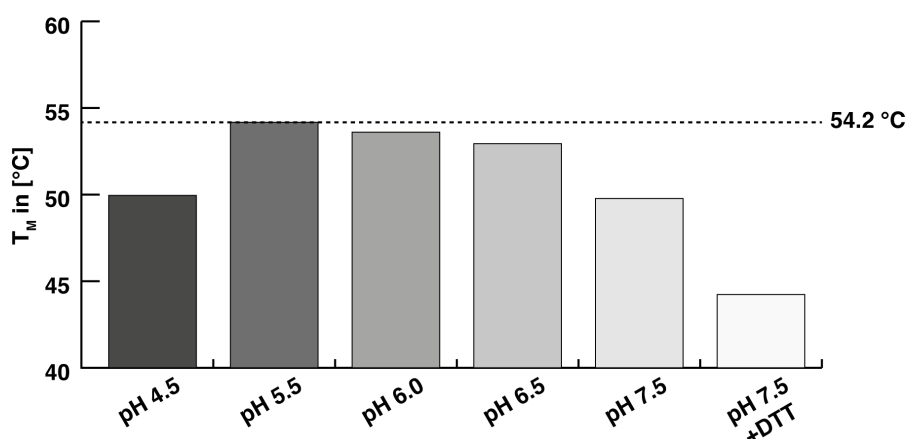


Figure 16: Thermal shift assay of CtDfg5. Different buffers with indicated pH-values have been used to determine the optimal conditions for CtDfg5. The highest melting temperature is indicated with the dashed line at 54.2 °C in the presence of pH 5.5.

4.2.2 Crystallization and structure determination of CtDfg5

In order to determine the first crystal structure of a fungal GH76 ortholog, *CtDfg5* was concentrated to 30 mg/ml in 10 mM NaCl, 20 mM HEPES at pH 7.5 (AEC-1 buffer) and subsequently used for initial crystallization screening. Macroscopic crystals appeared after two days at 18 °C in the presence of 20 mM NH₄I and 20% (w/v) PEG3350 (JCSG Core IE4). The crystals appeared with a monoclinic shape and were sufficient after two weeks of growth for diffraction studies. The course of growth is shown in Figure 17.

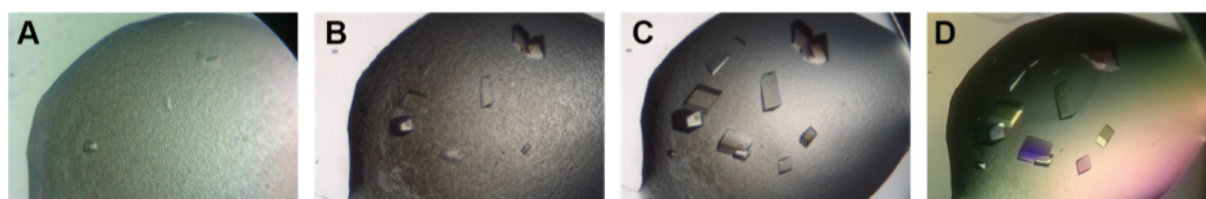


Figure 17: Crystallization of CtDfg5 in JCSG IE4. First crystals appeared after two days within amorphous precipitate (A). With advancing crystallization (B after four days, C after six days), the precipitate disappeared. D shows the crystals directly before freezing after two weeks of growth.

First diffraction studies resulted in a dataset with a ranking resolution of 1.2 Å, however with a rather low completeness (~84%). The space group as suggested from these data was the monoclinic C2 (complete: C 1 2 1). As the available homologs from the *bacterial* subfamilies in the PDB have low identities (3K7X 22%; 4BOK 20%; 4MU9 18%; 4C1S 17%; 4V1R ~16%), molecular replacement (MR) was not successful at this point. For phase

determination, the apo crystals were soaked in mother liquor with 50 mM Gd(OAc)₃ for 60 minutes, before they were shortly back soaked in mother liquor supplemented with 30% (v/v) glycerol as cryo protectant and flash frozen in liquid nitrogen. The diffraction studies were performed close to the L-II absorption edge of gadolinium (i.e. 1.56 Å) at a rotating CuK α -anode with a wavelength of 1.5418 Å, resulting in a dataset with significant anomalous signal to 1.96 Å and an overall resolution of 1.7 Å (see Table 5). A first model could be solved by SAD-phasing using the CRANK2 pipeline of the CCP4i2 suite (Potterton et al., 2018). A total number of 17 gadolinium atoms with an occupancy of at least 25% were found during substructure determination, which led to 404 amino acids that could automatically be build.

Table 5: Crystallographic data collection statistics of gadolinium soaked CtDfg5 crystals. Values in parentheses are for the highest resolution shell.

CtDfg5 _{Gd}	
Data collection	
X-ray source	I μ TM Mikrofocus source (Bruker)
Space group	C 1 2 1
Unit-cell parameters (Å, °)	$a=83.65$, $b=54.82$, $c=80.51$, $\alpha=90.00$, $\beta=91.14$, $\gamma=90.00$
Wavelength (Å)	1.54
Resolution range (Å)	40.03-1.8 (1.84-1.8)
Completeness (%)	99.9 (99.9)
Observed reflections	207832 (12113)
Unique reflections	33956 (2030)
Multiplicity	6.1
Wilson B factor (Å ²)	19.15
R_{merge} (%)	7.9 (38.4)
Mean $I/\sigma(I)$	12.8 (3.3)
CC($\frac{1}{2}$)	99.5 (89.9)
Anomalous completeness	99.8 (99.9)
Anomalous multiplicity	3.1 (3.0)
Anomalous CC($\frac{1}{2}$)	53.7 (4.7)

As the pH has a huge influence on the stability of CtDfg5, further crystallization trials have been performed in the presence of 50 mM acetate, 100 mM NaCl at pH 4.5 to bias the final pH-values towards the protein's optimum. The initial crystals were used as a seed stock to increase the hit rate. CtDfg5 crystallized at RT in JCSG Core IE7 (0.2 M Ca-acetate, 0.1 M MES, pH 6.0, and 20% (w/v) PEG 8000) and IF8 (0.1 M Na-citrate, pH 5.5, 20% (w/v) PEG 3000) by mixing 300 nl of 30 mg/ml protein solution with 200 nl mother liquor and 100 nl of the seed stock (1:10 dilution) (Figure 18). The crystals could be reproduced by using the same seed stock for the entire duration of this work in hanging and sitting drop conditions with variable final volumes (0.6-2 μ l). Like it was observed for the initial condition JCSG Core IE4, the protein crystallized in three-dimensional, monoclinically shaped crystals with very

good diffraction properties. Both conditions crystallized in space group $C 1 2 1$ with one monomer per asymmetric unit.

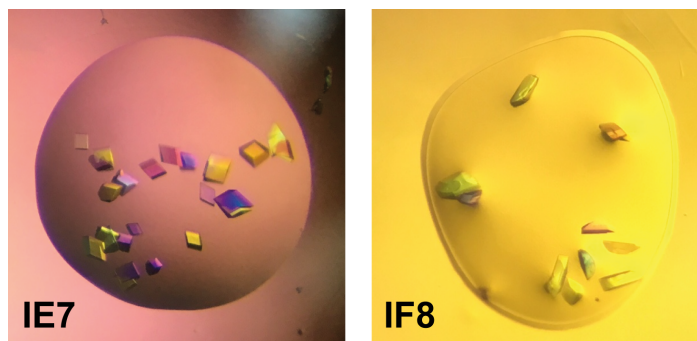


Figure 18: Crystallization of CtDfg5 in the presence of acetate buffer in JCSG Core IE7 and IF8.

The final dataset that resulted to the apo structure discussed in the following was obtained from soaking experiments of IE7-crystals with Frag37 from the *Frag Xtal Screen* (Jena Bioscience) at the European Synchrotron Radiation Facility (ESRF, Grenoble) (see Table 6). The structure has been solved by using the initial model from above for direct molecular substitution in *Phenix.refine*. Manual model building and refinement were performed with *Coot* and *Phenix.refine*, respectively (Afonine et al., 2012; Emsley et al., 2010).

Table 6: Crystallographic data collection and refinement statistics of the apo structure from CtDfg5. Values in parentheses are for the highest resolution shell.

CtDfg5 _{apo}			
PDB code	6RYO	Refinement	
Data collection			
X-ray source	ID23-1, ESRF	Resolution range (Å)	30.28-1.05
Space group	$C 1 2 1$	R_{work}/R_{free} (%)	10.6/12.3
Unit-cell parameters (Å, °)	$a=83.32$, $b=54.99$, $c=80.41$, $\alpha=90.00$, $\beta=90.27$, $\gamma=90.00$	Average B factor (Å ²)	13.4
Wavelength (Å)	0.972	No. of atoms	
Resolution range (Å)	30.28-1.05 (1.09-1.05)	Total	3935
Completeness (%)	97.20 (97.36)	Protein	3348
Observed reflections	297179 (28157)	Ligand	15
Unique reflections	164339 (16424)	Solvent	572
Multiplicity	1.8 (1.7)	R.m.s.d., bond lengths (Å)	0.013
Wilson B factor (Å ²)	9.66	R.m.s.d., bond angles (°)	1.62
R_{merge} (%)	1.9 (22.3)	Ramachandran plot	
R_{meas} (%)	2.7	Favored (%)	99
Mean $I/\sigma(I)$	17.06 (3.29)	Allowed (%)	1
$CC_{1/2}$ (%)	99.9 (88.4)	Disallowed (%)	0

4.2.3 The first crystal structure of a fungal GH76 protein

While bacterial GH76 proteins were shown to share their overall fold, general substrate specificity, and the catalytic DD-motif, the fungal *Dfg5*-subfamily remained uncharacterized (Cuskin et al., 2015; Thompson et al., 2015b). To gain detailed insights into that subclass, the crystal structure from the thermophilic fungus *Chaetomium thermophilum* has been determined at 1.05 Å resolution in space group C 1 2 1 with a continuous density map from residue Q32 till F441. The solvent content of the crystal is estimated to a low value of ~33%. Therefore, the tight packing of protein molecules within the crystal as shown in Figure 19B is not surprising. Ca^{2+} from the condition is traceable in electron density and responsible for crystal contacts between symmetry mates (Figure 19B). The ion is well coordinated by D79 from one molecule and E285 from one molecule and E285 from its neighbor.

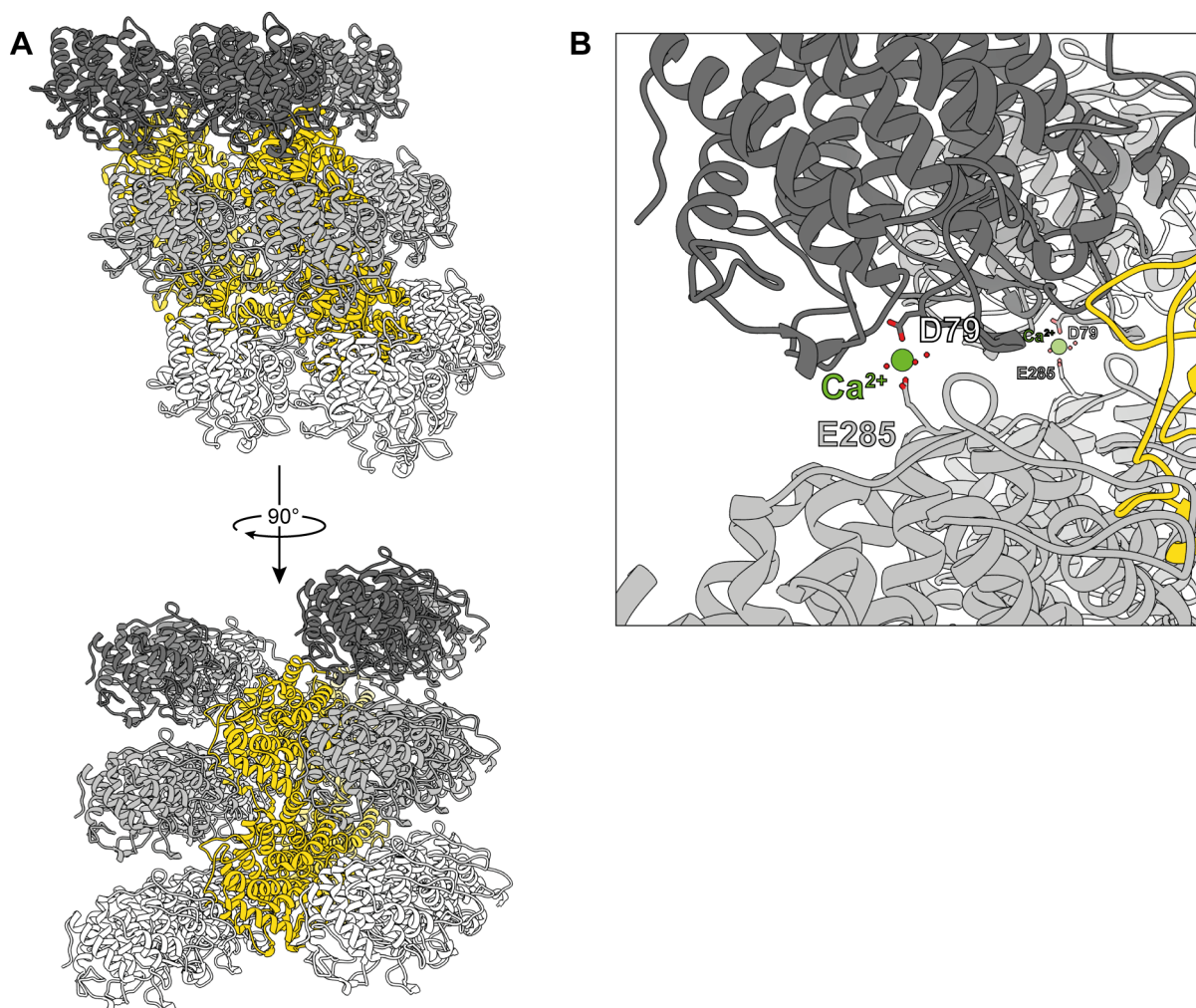


Figure 19: Crystal lattice of CtDfg5. **A** The arrangement of 16 molecules from CtDfg5 in the crystal lattice is shown from two sides as indicated by the arrow. The respective molecules are shown as ribbons. **B** Ca^{2+} (green sphere) establishes crystal

contacts between two molecules of CtDfg5 by being coordinated from D79 from one molecule and E285 from the neighboring lattice mate. Waters are shown as red dots.

The overall structure of CtDfg5 revealed a monomeric protein comprised of twelve GH76-characteristic α -helices arranged in an $(\alpha/\alpha)_6$ -helical barrel fold (Figure 20).

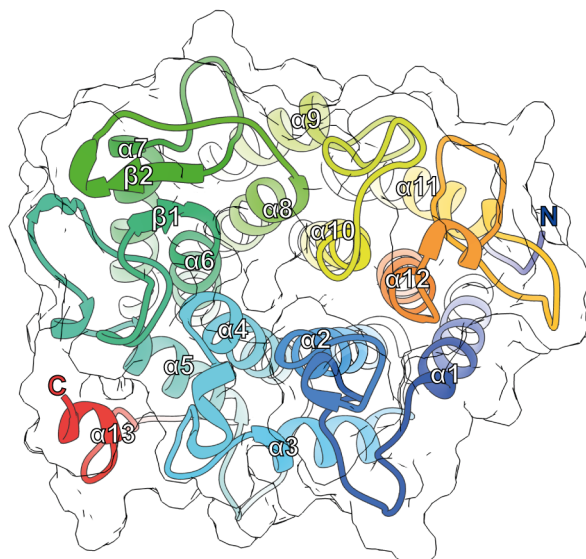


Figure 20: Overall structure of CtDfg5. The crystal structure of CtDfg5 is represented as a ribbon, which is colored from N- to C-terminus in rainbow colors from blue to red surrounded by white, transparent surface. The secondary structure elements are indicated.

A short alpha turn ($\alpha 13$) is formed at the C-terminus from A437 to the terminal F441. Beside the helical part, two β -strands are found between $\alpha 5/6$ and $\alpha 7/8$. As suggested from the thermal shift experiment (Figure 16), three disulfide bonds formed by cysteine pairs C182:C258, C315:C322, and C366:C375 stabilize the tertiary structure of CtDfg5.

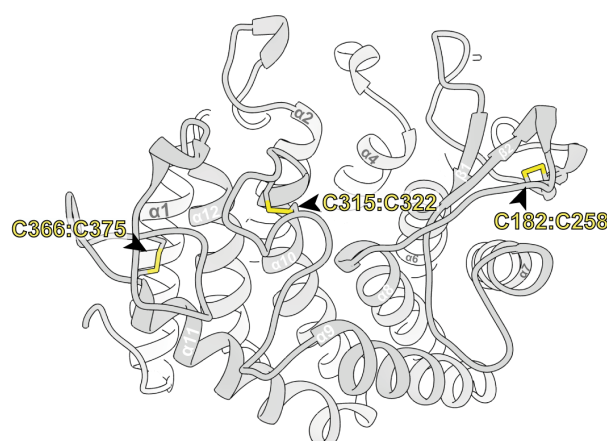


Figure 21: Disulfide bridges in CtDfg5. The three stabilizing disulfide bonds indicated by black arrows are shown as sticks in the context of the proteins secondary structure. Sulfur atoms are colored in yellow.

As mentioned above, the identities of already described bacterial homologs and CtDfg5 from the *Dfg5*-subfamily are at low values between ~15-20%. However, the superposition of CtDfg5 and BcGH76 (PDB: 4BOK) as the representative for bacterial GH76 proteins shows a well conserved overall fold with an r.m.s.d. of 3.2 Å for 400 aligned C α -atoms (see Figure 22).

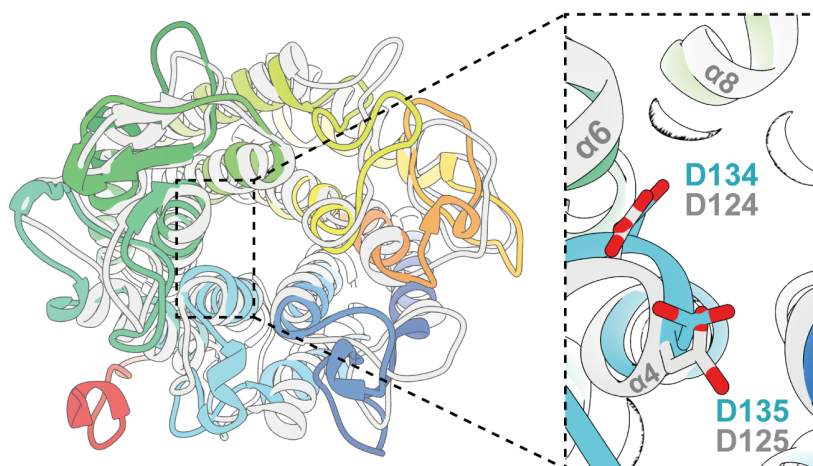


Figure 22: Comparison of bacterial and fungal GH76 proteins. A The conserved overall fold of GH76 proteins is represented by the superposition of CtDfg5 (rainbow) and BcGH76 (grey, PDB: 4BOK) shown as ribbons. The dashed box shows the conserved position of the putative active site DD-motif represented as sticks and colored according to the secondary structure. The oxygen's are red.

The major difference of the overall structure lays in the extended C-terminus of *CtDfg5*. While the structure of the bacterial protein terminates at $\alpha 12$, *CtDfg5* spans A413 to G436 over its back before it continues with $\alpha 13$. The active site of *BcGH76* was identified as a DD-motif consisting of the catalytic nucleophile D124 and the general acid/base residue D125 (Thompson et al., 2015b). In the structural alignment shown above, the active site of the bacterial protein and D134/D135 of *CtDfg5* superimpose very well (box in Figure 22). Finding such significant overlapping features of *bacterial* and *Dfg5*-proteins, it raises the question, how the identified putative active site mediates the function of *Dfg5*-proteins, and how the conserved active site is consistent with different substrate specificity.

4.2.4 Bacterial and Fungal GH76 proteins act on different substrates

One hypothesis of how fungal GH76 members are involved in the incorporation of cell wall-resident proteins is based on the annotated $\alpha 1,6$ -mannanase activity of the described bacterial homologs (Cuskin et al., 2015; Maddi et al., 2012; Thompson et al., 2015b). This model combines similar defects of the cell wall in *Δoch1* mutants, which is responsible for the transfer of the initial mannose of the N-glycan $\alpha 1,6$ -mannose backbone, and *Δdcw1/Δdfg5* deletion mutants in *Neurospora crassa* and *Candida albicans* (Ao et al., 2015; Maddi and Free, 2010; Maddi et al., 2012). The authors suggested that *Dfg5*-proteins manage the cell wall incorporation of GPI-APs by cleaving the $\alpha 1,6$ -mannose backbone of N-linked outer chain mannan, as it is known from the bacterial homologs, and subsequently transfer them onto the cell wall glycan by a so far unknown mechanism. Accordingly, *CtDfg5* should be able to catalyze the first reaction step, i.e. the cleavage of an $\alpha 1,6$ -mannooligomer with a minimum length of a triose, as found for the bacterial member from *Bacillus circulans*, *BcGH76* (Thompson et al., 2015b).

In order to test that hypothesis *in vitro*, *CtDfg5* reaction mixtures were incubated with $\alpha 1,6$ -linked manno-oligosaccharides (*kind gift of Dr. Daniel Varón Silva, MPI Potsdam*) and analyzed by thin layer chromatography (TLC) (Figure 23A). However, aside from a mannose contamination within the mannotriose sample that is present in both, the negative control and the respective reaction, no migration shift or a formed product could be observed.

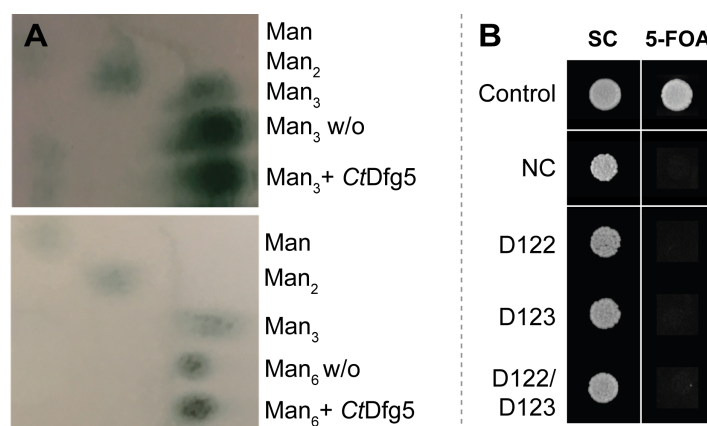


Figure 23: TLC of manno-oligosaccharides and *in vivo* test of the putative active site. **A** Upper panel: Mannose (Man), α 1,6-linked mannobiose (Man_2), and α 1,6-linked mannotriose (Man_3) are shown as standards. The incubated samples without (w/o) and with (+CtDfg5) protein are shown below. Lower panel: Size standards are shown as above. Here, an α 1,6-linked mannohexaose (Man_6) has been analyzed. **B** A plasmid-loss assay shows the essential role of the putative active site residues D122 and D123 from *S. cerevisiae*. Gesa Schmitz (AG Mösch) performed the *in vivo* work. SC: minimal medium for *S. cerevisiae*. 5-FOA: SC-medium supplemented with 5-Fluoroortic acid. NC: negative control.

The second route using the GPI-anchor as the unit to transfer proteins into the cell wall is based on the findings of GPI-remnants found in HF-threatened cell walls and the simultaneous release of proteins, which are predicted to possess an ω -signal at their C-terminus that is necessary for posttranslational modification of a GPI-anchor (Fujii et al., 1999; Kollár et al., 1997). Together with the findings of GPI-anchored proteins released into the medium in double knockouts of $\Delta dcw1/\Delta dfg5$ in *Saccharomyces cerevisiae*, it is proposed that the transfer is catalyzed by Dfg5-proteins cleaving the $\text{Man}\alpha 1,4\text{-GlcN}$ linkage within the GPI-anchor (see Figure 24) (Kitagaki et al., 2002). Unfortunately, such a substrate is not commercially available and the synthesis is difficult to realize, even for experts. Consequently, it was not possible to test hydrolysis or defective mutants *in vitro* due to the lack of a suitable substrate. However, it has been suggested by the SSN analysis (Figure 14 and Table 2) that the proteins involved in the lipid-to-wall transfer are all resident in the Dfg5-subfamily, including Dcw1 and Dfg5 from *S. cerevisiae*. Working in the living cell would not only enable us to analyze structure-function relationships in the context of the living cell and thereby bypass the lack of substrate, but also make a more general statement for the whole Dfg5-subfamily, as the predictions made on the basis of a protein from *Chaetomium thermophilum* are examined in another fungal species. Therefore, a collaboration with the group of Prof. Hans-Ulrich Mösch has been started. Gesa Schmitz established the double knockout strains of $\Delta dcw1/\Delta dfg5$ in *S. cerevisiae* and performed the *in vivo* work shown in this thesis.

As shown in Figure 22, the DD-motif identified as the active site in bacterial GH76 proteins is also highly conserved in *CtDfg5*. Thus, the residues in *Dfg5* from *S. cerevisiae* (*ScDfg5*) have been specified as residues D122/D123 (see Figure 28A on p.74) and the respective active-site mutants D122N, D123N and D122N/D123N were analyzed concerning their ability to complement the wildtype protein in a plasmid-loss assay (Figure 23B). As expected, this is not possible as their function should be absolutely essential for the organism in a double-knockout background.

Resulting from the first functional results from *Dfg5*-proteins, the active site seems to be highly conserved throughout the whole GH76 family, however at least the so far characterized bacterial homologs and those from the *Dfg5*-subfamily do not share the same substrate specificity. Following this, the next part of this work will deal with the identification of the real substrate to shed light on the last step of cell wall protein maturation.

4.2.5 Mapping the GPI-core glycan by sugar-fragment screening

To learn more from a structural perspective, substrate bound states are sought to be determined. The true substrate of *Dfg5*-proteins might be the GPI-anchor, as its mannose core structure is found covalently attached to the glucan matrix in the fungal cell wall and the delivery of GPI-AP into the CW is compromised in $\Delta dcw1/\Delta dfg5$ promoter shutoff cells (Kitagaki et al., 2002; Kollár et al., 1997). As GPI-anchors or derived carbohydrates are not commercially available, the primary goal was to utilize sugar fragments for cocrystal structures with *CtDfg5* (Figure 24).

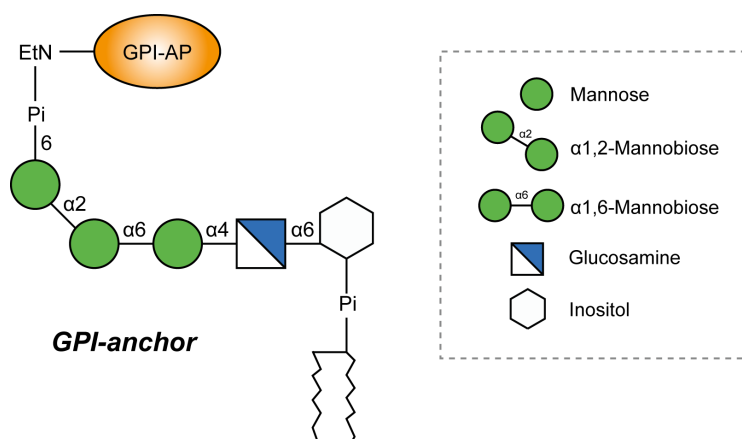


Figure 24: Schematic representation of the GPI-anchor and its tested fragments. The core structure of the GPI-anchor is shown with the conventionally used symbols for glycan representation with the respective glycosidic bonds being indicated. The edged line shows the lipid moieties. The box on the right shows the respectively used fragments in the crystallographic experiment. *GPI-AP*: glycosylphosphatidylinositol-anchored protein; *EtN*: Ethanolamine phosphate; *Pi*: inorganic phosphate.

To elucidate the structure bound state of *CtDfg5*, the protein was crystallized like described for the apo structure and used after one week of growth for further soaking experiments with different mono-, di-, and higher oligosaccharides. The sugars were solved in a 1:1 mixture composed of JCSG Core IE7 and 100 mM NaCl buffered with 50 mM acetic acid at pH 4.5 to final concentrations of 0.5-1 M. The apo crystals were transferred in 1 μ l of the respective solution and incubated for several hours. The exact concentrations and durations for the individual structures are shown in Table 7.

Table 7: Sugar concentrations and soaking times used for successful cocrystal structures of *CtDfg5*.

	Man	α 1,2-Man ₂	α 1,6-Man ₂	GlcN
Concentration	0.5 M	0.5 M	1 M	0.5 M
Duration	4 h	4 h	8 h	4 h

A critical point for successful soaking was to omit backsoaking after the incubation. Due to the similar crystal lattice parameters, the structures were also solved by molecular substitution using the apo structure as template. The collection and refinement statistics are shown in Table 8. While the soaking was successful for the GPI-glycan fragments listed above, soaking with inositol under the similar conditions did not result in additional electron density. The data collection statistics are given in Table S 1 on p.144.

Table 8: Crystallographic data collection and refinement statistics of the fragment bound states from *CtDfg5*. Values in parentheses are for the highest resolution shell.

	<i>CtDfg5</i> _{Man}	<i>CtDfg5</i> _{α1,2M}	<i>CtDfg5</i> _{α1,6M}	<i>CtDfg5</i> _{GlcN}
PDB code	6RY1	6RY2	6RY5	6RY6
Data collection				
X-ray source	ID23-1, ESRF	ID23-1, ESRF	ID29, ESRF	ID23-2, ESRF
Space group	C 1 2 1	C 1 2 1	C 1 2 1	C 1 2 1
Unit-cell parameters (\AA , $^\circ$)	$a=83.86$,	$a=83.72$,	$a=83.65$,	$a=83.42$,
	$b=55.02$,	$b=54.86$,	$b=54.82$,	$b=55.02$,
	$c=80.49$,	$c=80.13$,	$c=80.51$,	$c=80.12$,
	$\alpha=90.00$,	$\alpha=90.00$,	$\alpha=90.00$,	$\alpha=90.00$,
	$\beta=90.45$,	$\beta=90.45$,	$\beta=91.14$,	$\beta=90.37$,
	$\gamma=90.00$	$\gamma=90.00$	$\gamma=90.00$	$\gamma=90.00$
Wavelength (\AA)	0.972	0.972	1.54	0.873
Resolution range (\AA)	41.93-1.3	41.86-1.3	45.93-1.3	40.25-1.3
	(1.35-1.3)	(1.35-1.3)	(1.35-1.3)	(1.35-1.3)
Completeness (%)	99.76 (99.89)	94.32 (91.56)	96.40 (95.07)	96.27 (94.52)
Observed reflections	175788	161441	159977	166168
	(17387)	(15866)	(15490)	(15984)
Unique reflections	89845 (8959)	84221 (8154)	85994 (8464)	86433 (8481)
Multiplicity	2.0 (1.9)	1.9 (1.9)	1.9 (1.8)	1.9 (1.9)
Wilson <i>B</i> factor (\AA^2)	10.47	10.59	13.03	10.66
R_{merge} (%)	2.7 (16.2)	6.0 (11.6)	2.0 (9.4)	2.5 (14.0)

R_{meas} (%)	3.8	8.4	2.8	3.5
Mean $I/\sigma(I)$	15.10 (4.25)	8.63 (6.01)	20.02 (6.93)	15.54 (4.51)
CC(½)	99.9 (92.8)	99.0 (94.7)	99.9 (97.5)	99.9 (94.6)
Refinement				
Resolution range (Å)	41.93-1.3	41.86-1.3	45.93-1.3	40.25-1.3
R_{work}/R_{free} (%)	9.6/12.5	10.5/12.9	10.2/12.3	11.3/12.6
Average B factor (Å ²)	16.0	15.8	17.5	15.1
No. of atoms				
Total	4070	4078	3811	3947
Protein	3335	3335	3410	3331
Ligand	28	39	35	23
Solvent	707	704	366	593
R.m.s.d., bond lengths (Å)	0.010	0.011	0.013	0.011
R.m.s.d., bond angles (°)	1.51	1.52	1.62	1.44
Ramachandran plot				
Favored (%)	99	99	99	99
Allowed (%)	1	1	1	1
Disallowed (%)	0	0	0	0

As shown in Figure 24, the core component of the GPI-anchor is composed of three mannoses, glucosamine and inositol. While the latter one could not be observed in the respective soaking experiment, mannose alone could be unambiguously identified at GPI-glycan binding site -2 in the substrate binding canyon of *CtDfg5* (see Figure 25A). It is well bound by electrostatic and hydrophobic interactions mainly from residues of helices $\alpha 6$, $\alpha 8$, and $\alpha 10$. Soaking with $\alpha 1,2$ -mannobiose led to a complex structure that also occupies subsite -2, which is now extended to subsite -3 (Figure 25B). The -3 mannose is bound by fewer interactions than the -2 sugar and it is more surface exposed, however kept in place by an H-bond between its OH3-group and N262. By soaking the crystals with $\alpha 1,6$ -mannobiose, the central mannose could further be expanded to subsite -1 (Figure 25C). The -1 sugar sits deeply in the binding pocket of *CtDfg5* above the active site residue D134 and is stacked by W83. Interestingly, its OH1 and OH2 groups replace waters that used to coordinate a Ca^{2+} ion, which is coordinated above D135 and surrounded by three waters roughly on the plane of the hydroxyl groups of the sugar and a water opposing to D135. To fit the stereochemical properties of the -1 moiety, a β -mannose had to be modeled into the electron density, while all other mannoses were found in the α -state. The rearrangement probably happens due to the coordinated calcium. In fact, this kind of coordination seems to provide no further insights into the true distortion of the -1 position during catalysis, as this would also require the presence of the +1-sugar. This position however could be attributed to glucosamine, which has been found at GPI-glycan subsite +1 above the general acid/base residue D135 (Figure 25D). The sugar is bound by H-bonds with Q189 from the front and stacking interactions of Y81 from the back. Further electrostatic interactions are established by D325 and S130.

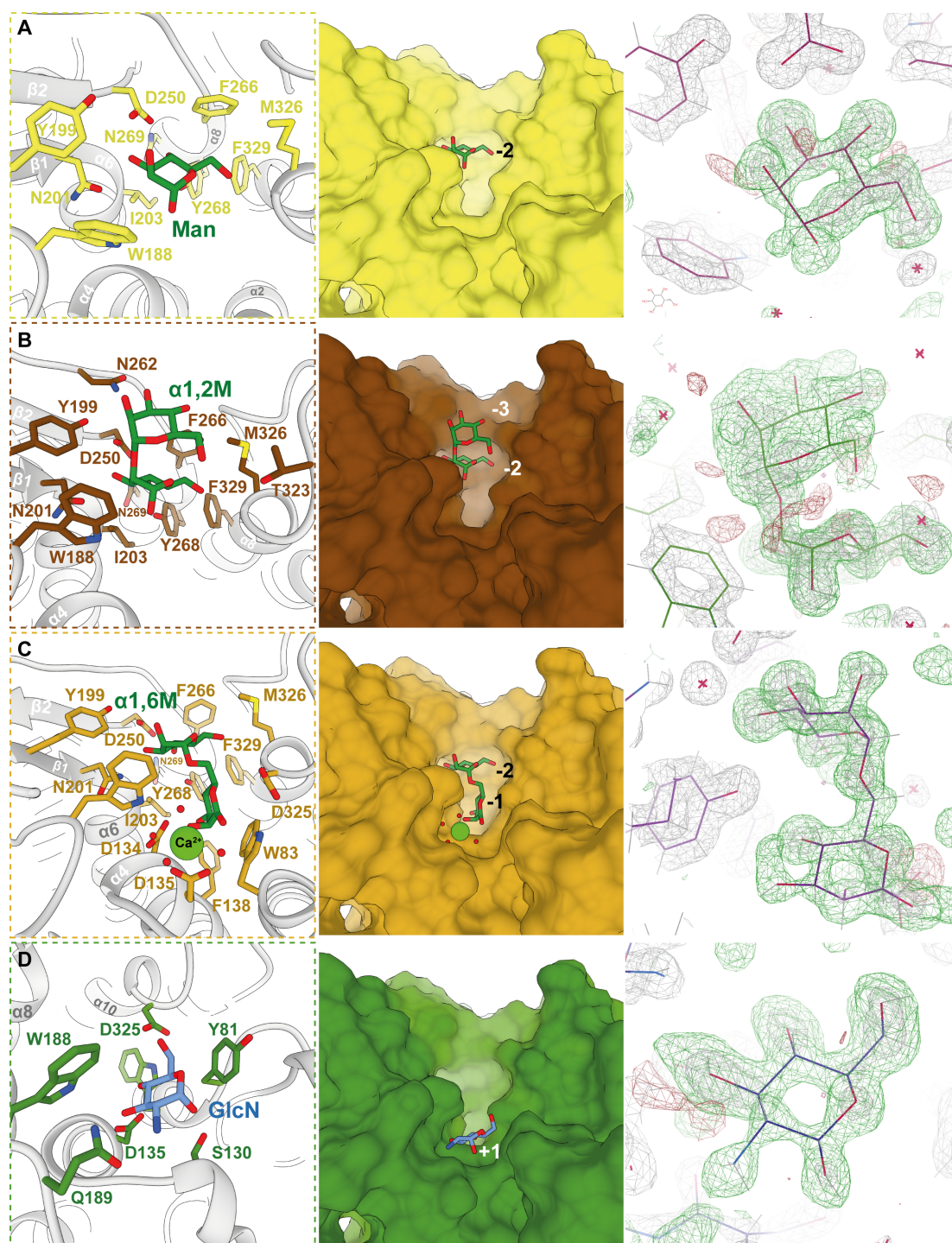


Figure 25: Sugar fragments resemble the GPI-anchor core in the binding pocket of CtDfg5. **A** The monosaccharide mannose (Man) is bound to the sugar-binding site -2 in the binding pocket of CtDfg5. **B** That central mannose could be extended by α 1,2-mannobiose (α 1,2M) to subsite -3 and with **C** α 1,6-mannobiose (α 1,6M) to the -1 site on top of the active site residues D134/D135 (**C**). **D** Glucosamine (GlcN) is bound to the +1 sugar-binding site. On the left, the sugar and its coordinating residues are presented as sticks colored with oxygens in red, nitrogens in blue, sulphur in yellow, and carbon atoms of Man in green, of GlcN in light blue and of the protein in the color given. The arrangement is shown in the context of the secondary structure elements, which are colored in grey. Ca^{2+} is shown as a light green sphere with its coordinating waters as red dots. In the middle panel, the sugar is shown as in the left panel in the context of the surface from CtDfg5 in the respective color scheme. The sugar-binding sites are indicated. In the right panel the OMIT-maps of the respective carbohydrate are shown with the difference map colored in green and contoured at σ -level 3.0.

By superimposing the four different states shown in Figure 25 it is possible to reconstruct the GPI-core structure from glucosamine to mannose 3 within the binding pocket of *CtDfg5* (Figure 26). The GPI-core occupies the GPI-glycan binding sites from -3 to +1 and is found in a *U-* or *C-shaped* conformation that very well fills out the complete binding pocket.

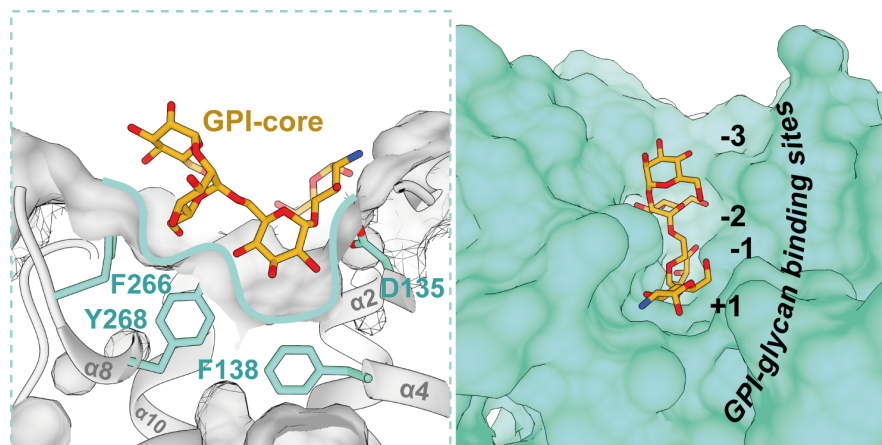


Figure 26: Assembled GPI-core bound to the binding pocket of *CtDfg5*. From the top view on the right with all four GPI-glycan binding subsites labeled, the GPI-core is shown on the left from the left site with the surface being capped in front of the binding pocket, which is highlighted by the turquoise line. Residues (turquoise sticks) and secondary structure elements (grey ribbon) are shown for orientation, the GPI-core is presented as orange sticks with red oxygens and blue nitrogen.

With this finding, the exact role of *Dfg5*-proteins seem to be clear, i.e. utilization of the GPI-anchor as a donor substrate to transfer CW-resident proteins from the plasma membrane into the wall. The differences and similarities between GH76 proteins from the *Bacteria II* and *Dfg5*-subfamilies concerning their substrate binding mode are shown in the following and allow an explanation, why *CtDfg5* is incapable of α 1,6-mannan hydrolysis.

4.2.6 Structural differences between bacterial and fungal GH76 proteins

In 2015, the first crystal structure of a GH76 protein in complex with its substrate α 1,6-mannopentaose spanning over the whole binding canyon from subsites -4 to +1 has been published (PDB: 5AGD) (Thompson et al., 2015b). This structure can now be used to compare the substrate bound states of two members of different GH76 subfamilies. In Figure 27A, *CtDfg5* is shown in a similar manner as in Figure 26, but superimposed with the complex structure of *BcGH76*. It clearly shows a quite similar binding mode of the α 1,6-linked mannoses at subsites -2 and -1 with a slightly tilted mannose 2, however it reveals huge differences in the binding modes of the peripheral glycan-binding sites. This becomes

already obvious looking at the predicted clash of the mannopentaose with the surface of *CtDfg5* at subsites -4 and -3 (Figure 27A).

A closer look on that region is given in Figure 27B. The upper panel shows the binding mode of the GPI-core in *CtDfg5*, the lower panel that of *BcGH76*. While the interactions at subsite -2 are conserved by the tyrosine interaction from below (Y268 and Y243) and from the front by an aspartate (D250 and D228), they differ in the interactions involved in the binding site close to C6 of mannose 2. One striking example is the already mentioned clash of the mannopentaose with the surface of *CtDfg5*. Responsible for this clash is F266, which stacks the C6 atom of mannose 2. This leads to the OH6-group swiveling away from F266. Consequently, an α 1,2-linkage to the mannose 3 of the GPI-core must be formed to circumvent the hydrophobic block that is different in *BcGH76*, where a threonine (T241) is present instead of the phenylalanine. In the latter enough space for the α 1,6-linkage is provided to continue the mannopentaose in a linear manner. Furthermore, residues from α -helix 10 also play an important role. The arrangement in *CtDfg5* shows a highly conserved F329, which is guiding M326 around the C6-OH6 configuration as described above. In *BcGH76*, this phenylalanine is substituted by a glycine and the methionine position shows a leucine (L295). The free space given by the glycine is used by L295 to stack the C6 atom of the -2 mannose, while it clashes with OH6 when superimposing the GPI-core glycan.

The other site of the binding pocket above the active site is compared in Figure 27C. Again, *CtDfg5* is shown in the upper and *BcGH76* in the lower panel. In both structures, an H-bond is formed by an aspartate (D325 and D294) sitting above α 10 with the sugar moiety at subsite +1. In *CtDfg5*, the α 1,4-linked glucosamine is bound from the front by electrostatic interactions of Q189, whereas from the back the C6 region is stacked by Y81. This binding mode is reversed in *BcGH76*, as the +1 mannose is α 1,6-linked with the -1 mannose. Accordingly, the stacking interaction is established by F122 from the front, while an electrostatic interaction is realized from the back by D71.

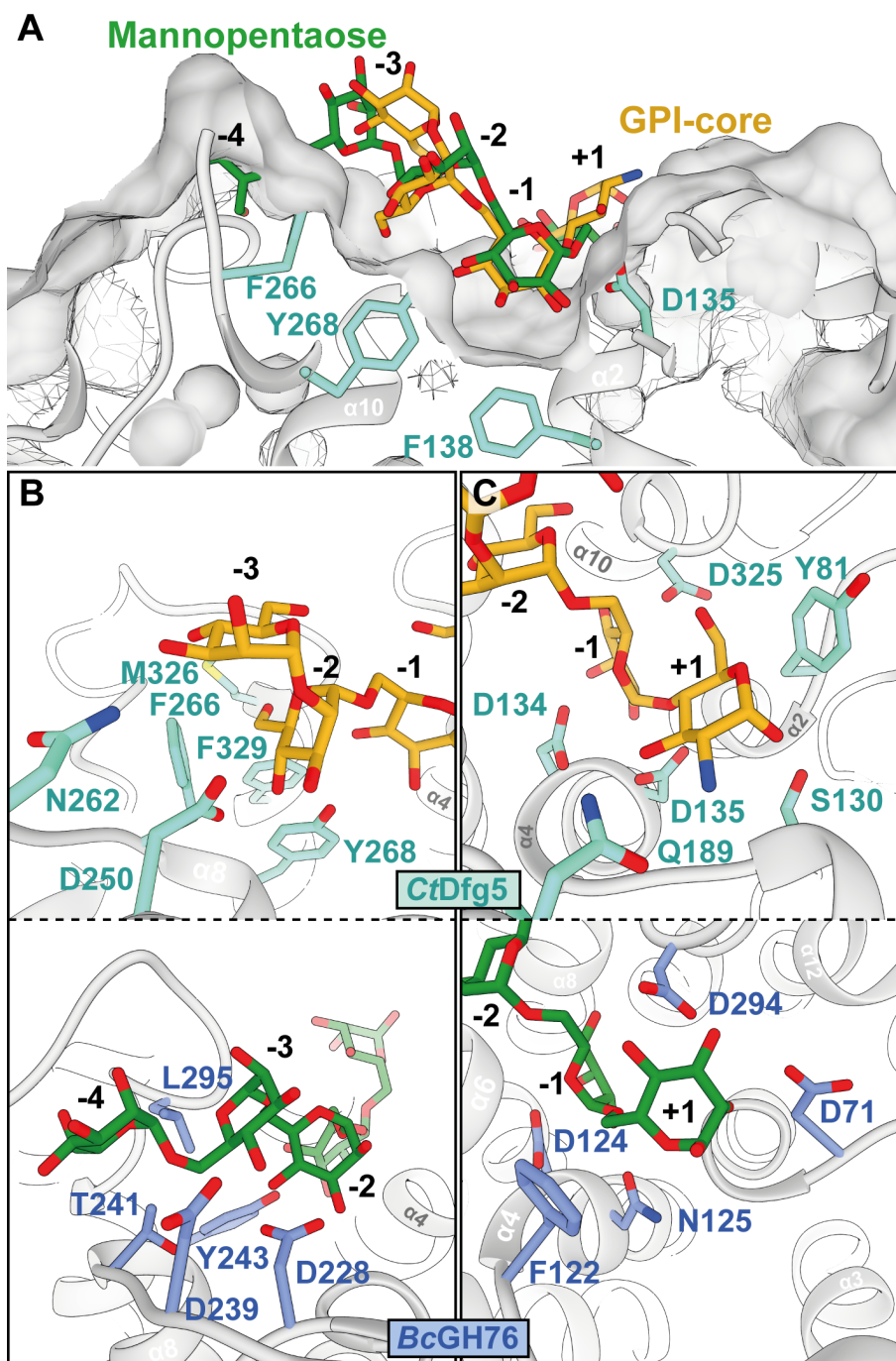


Figure 27: Comparison of CtDfg5 and BcGH76 in their substrate bound states. **A** The structure of BcGH76 in complex with α 1,6-mannopentaose (PDB: 5AGD) is superimposed with CtDfg5 and just the substrate (green sticks) is shown in the context of CtDfg5. The capped surface and secondary structure (both in grey), turquoise sticks for the residues and the orange GPI-core shown as sticks are presented of CtDfg5. **B** The upper panel shows the -1 to -3 sugar binding site of CtDfg5, the lower panel the -2 to -4 region in BcGH76. **C** The same arrangement as in **B** shows the substrate coordination at subsite +1. The shown secondary structure elements, the residues, and the sugar binding sites are indicated. The coloring in **B** and **C** is like in **A** with bluish sticks for the residues of BcGH76.

4.2.7 The conserved substrate binding pocket of CtDfg5

The analysis of evolutionary conserved and variable regions of proteins is a handy way to predict important residues and regions of a protein. On the left side of Figure 28A, a *ConSurf*-analysis of the *Dfg5*-family is plotted on the surface of CtDfg5 (Ashkenazy et al., 2016). It clearly shows the importance of the substrate-binding pocket, as almost all residues are attributed the highest conservational score possible (SFigure 4 on p.136). Further mutational studies based on this should strengthen the role of the substrate-coordinating residues, however as already explained above *in vitro* studies with defined substrates could not be performed due to a lack of suitable substrate. Like it was performed for the active site motif, structure-based mutants were planned according to the homology model of ScDfg5 (right side of Figure 28A) and analyzed *in vivo* by Gesa Schmitz (Figure 28B).

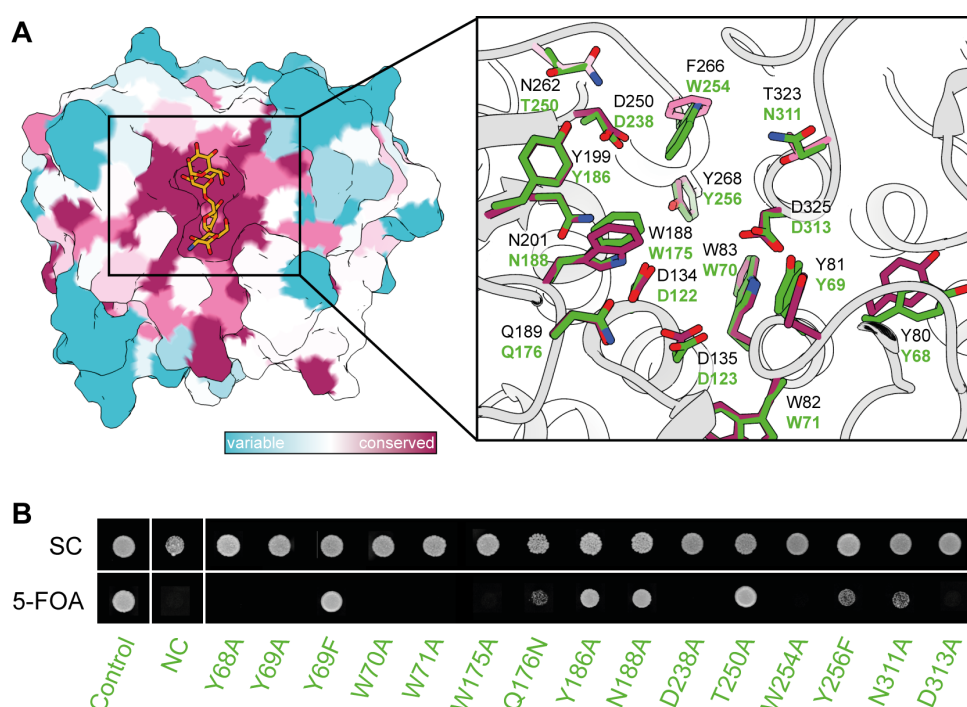


Figure 28: Structure-based mutants show the important role of residues in the binding pocket of *Dfg5*-proteins. **A** The surface of CtDfg5 is shown and colored according to the legend from variable (cyan) to conserved (dark magenta) residues. The assembled GPI-core is shown as orange sticks. The box on the right shows the binding pocket residues of CtDfg5 (*ConSurf* colors, indicated in black letters) superimposed with a structural model of ScDfg5 (green, indicated in green letters) in the context of the secondary structure of CtDfg5 (grey). **B** The structure-based mutants are analyzed in a plasmid-loss assay in *S. cerevisiae* on SC-medium with and without 5-FOA. The type of mutant is indicated. *In vivo* work has been performed by Gesa Schmitz (AG Mösch).

The plasmid-loss assay identified eight of 15 tested mutants as essential (Y68A, Y69A, W70A, W71A, W175A, D238A, W254A, D313A) for the viability of *Saccharomyces*

cerevisiae. Interestingly, the essential phenotype of Y69A is not given for the Y69F mutant, strengthening the importance of the glucosamine interaction described above by *CtDfg5*'s Y81. Apparently, the additional OH-group does not seem to be necessary, although tyrosine is fully conserved in the *Dfg5*-subfamily. On the other side of the glucosamine interaction, the rather conservative mutation of Q176N (*CtDfg5*: Q189) leads to a reduced growth rate, pointing out the importance of correct placements of the functional group of glutamine. In contrast to Y69, the OH group of Y256 (*CtDfg5*: Y268) seems to play a more important role, as the Y256F mutant exhibits a reduced growth. Interestingly, the hydrophobic block at mannose 2, here established by W254 instead of F266 in *CtDfg5*, is essential for correct protein function, while the N188A mutant, which is also involved in the binding of mannose 2, lacks any effect. Two residues coordinating at subsite -3 have been mutated as well. While T250A shows unaltered growth behavior in the spot assay, the growth rate of N311A is reduced.

4.2.8 *CtDfg5* hydrolyze GPI-core of soluble GPI-APs from *S. cerevisiae*

Taken the structural findings of *CtDfg5* and its structure-based analyses together, it becomes obvious why the hydrolysis of α 1,6-linked oligomannoses failed. Unfortunately, in terms of GPI-core glycan cleavage, any type of potential donor substrate is commercially not available. To show GPI-hydrolysis anyway, an *in vitro* assay with the following rational has been elaborated (see Figure 29):

As the GPI-core state provides no hint for the requirement of species-specific GPI-anchor modifications due to a lack of further space in the binding pocket of *CtDfg5*, it can be assumed that any GPI-anchor with that core structure should be suitable as substrate. Such GPI-anchors are synthesized by *S. cerevisiae*, where 80% contain an additional α 1,2-linked mannose at Man3 and some anchors possess a fifth mannose (Fankhauser et al., 1993). Nevertheless, these additional mannoses should not affect the binding within the pocket. Therefore, crude membrane extracts from *S. cerevisiae* were prepared, followed by a *Bacillus cereus*-PI-PLC treatment, which is known to release GPI-APs from membranes in a Ca^{2+} -independent manner and thereby producing a cyclic phosphate (see Figure 29, step 1) (Heinz et al., 1996). Upon centrifugation the supernatant contained soluble GPI-APs (sGPI-AP) such as Gas1 (Uniprot: P22146), Gas3 (Uniprot: Q03655) Gas5 (Uniprot: Q08193), Ecm33 (Uniprot: P38248) and YJL171C (Uniprot: P46992), as identified by tryptic digests using ESI-MS (Table S 2 on p.144). In step 2, *CtDfg5* is incubated with the supernatant,

which is subsequently (step 3) filtered through a 3 kDa MWCO cutoff filter membrane. In this step, the hydrolysis of the Man α 1,4-GlcN linkage within the GPI-core should occur. The flow through with molecules smaller than 3 kDa was again analyzed by ESI-MS that should find the resulting GlcN-Ino-Pi with a theoretical mass of 402.08 Da. Indeed, such a molecular weight could be identified (step 4, SFigure 5 on p.137). Furthermore, the flow through has been analyzed by Dr. Xiulan Xie from the NMR-facility concerning characteristic chemical shifts of GlcN-Ino-Pi atoms (step 5). One is a single anomeric carbon, which is known to show up in a certain region in 2-D ^1H ^{13}C spectra, while cyclic phosphates are identified by a chemical shift around 16 ppm with ^{31}P -NMR in the literature (Bubb, 2003; Chakraborty and d'Alarcao, 2005; Hecht et al., 2010; Schofield et al., 2002). Both, the specific region within the 2-D spectrum and the cyclic phosphate could be observed in this analysis (SFigure 6 and SFigure 7 on pp.138-139). This strongly suggests that *CtDfg5* is able to hydrolyze the Man α 1,4-GlcN linkage in the GPI-core for the subsequent transfer onto an acceptor sugar in the fungal cell wall.

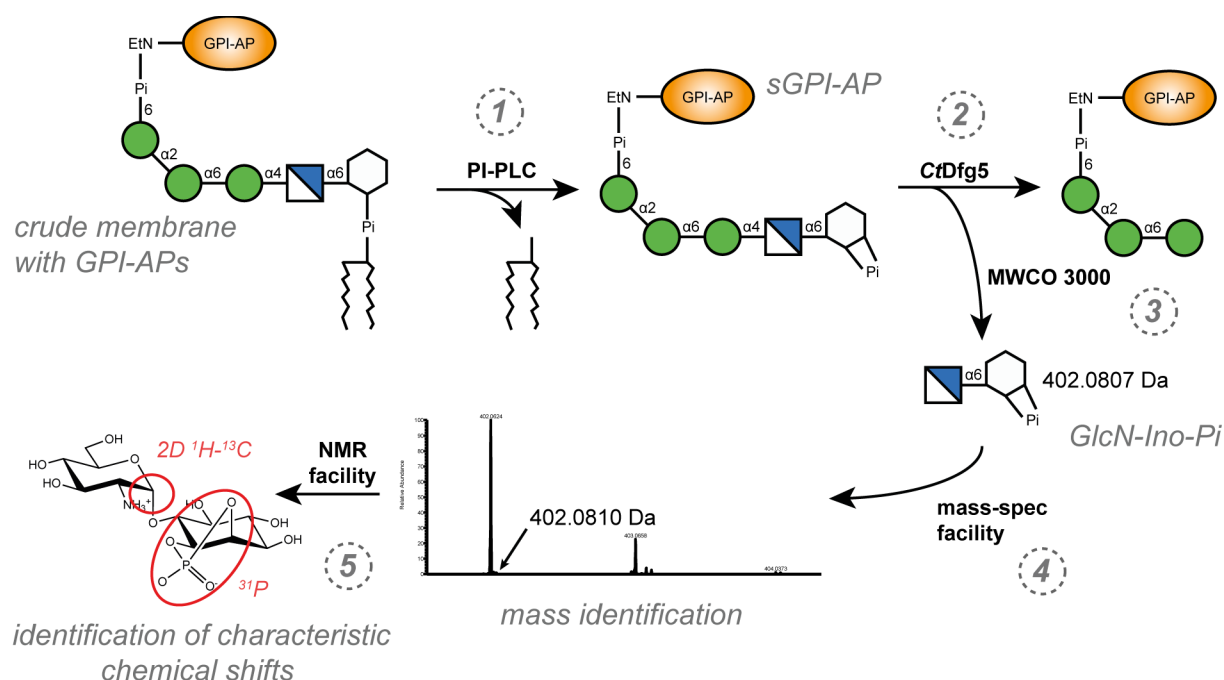


Figure 29: The rational principle of the *in vitro* assay. The workflow to obtain the product is shown. After membrane purification, PI-PLC (1) is used to release GPI-AP from the lipid moiety and thereby producing a cyclic phosphate. Upon ultracentrifugation, the supernatant undergoes a *CtDfg5* treatment (2) that releases a 402.08 Da large disaccharide-phosphate. Subsequently the reaction is filtered with a 3 kDa cut off filter membrane (3) and the flow through is analyzed by mass spectrometry (4) and NMR-facilities (5).

4.2.9 Structural base of the transfer mechanism catalyzed by *Dfg5*-proteins

From *Saccharomyces cerevisiae* it is known that GPI-CWPs are attached to the wall via β 1,6-glucan (Gonzalez et al., 2009). However, the situation in other fungi is less clear, as the CW architecture differs from species to species. While *Candida* spp. seem to have a similar ensemble, *Aspergillus fumigatus* apparently lacks the presence of β 1,6-glucans, but contains differently linked glycans. Therefore it is hard to predict, what the actual attachment-glycan of CWPs in the respective species is.

In order to answer the question of the acceptor identity for *CtDfg5*, possible acceptor-molecules were used again for soaking experiments that were performed as described before. While soaking with 0.5 M gentiobiose (Glc β 1,3-Glc) did not result in additional density, soaking of 1 M laminaribiose (Glc β 1,3-Glc) for 8 h resulted in another complex structure of *CtDfg5* (see Table 9 and Figure 30A).

Table 9: Crystallographic data collection and refinement statistics of *CtDfg5* soaked with laminaribiose. Values in parentheses are for the highest resolution shell.

<i>CtDfg5</i> _{β1,3G}			
PDB code	6RY7		
Data collection		Refinement	
X-ray source	ID23-1, ESRF	Resolution range (Å)	40.05-1.3
Space group	C 1 2 1	R_{work}/R_{free} (%)	11.9/9.4
Unit-cell parameters (Å, °)	$a=83.36$, $b=54.88$, $c=80.10$, $\alpha=90.00$, $\beta=90.45$, $\gamma=90.00$	Average B factor (Å ²)	16.1
Wavelength (Å)	0.972	No. of atoms	
Resolution range (Å)	40.05-1.3 (1.35-1.3)	Total	3866
Completeness (%)	97.41 (97.52)	Protein	3332
Observed reflections	169436 (16736)	Ligand	34
Unique reflections	86591 (8648)	Solvent	500
Multiplicity	2.0 (1.9)	R.m.s.d., bond lengths (Å)	0.009
Wilson B factor (Å ²)	11.99	R.m.s.d., bond angles (°)	1.41
R_{merge} (%)	2.6 (9.9)	Ramachandran plot	
R_{meas} (%)	3.7	Favored (%)	99
Mean $I/\sigma(I)$	16.58 (6.88)	Allowed (%)	1
CC _{1/2} (%)	99.8 (97.4)	Disallowed (%)	0

Laminaribiose could unambiguously be identified in the electron density, although the +1 subsite is much clearer than the +2 acceptor-subsite (Figure 30A+B). However, the β -linkage as well as C1, C2 and its hydroxy-group can be well seen and lowering the σ -level leads to the appearance of the full pyranose ring. This can be explained by partial occupation of the +2 subsite, which is only loosely bound by two loop regions termed as *acceptor binding loops* (ABL) 1 and 2 (Figure 30B+C). These regions are not conserved in *Dfg5*-proteins, but shape the conserved *canyon* that is suggested to harbor the PI-binding site. Strikingly, the non-

reducing sugar moiety of laminaribiose adapted a conformation quite similar to the observed +1-subsite conformation of glucosamine (Figure 30C).

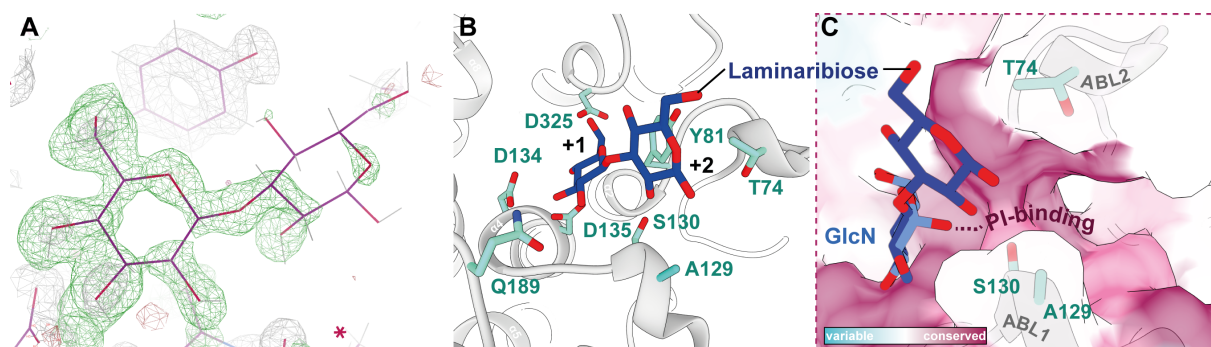


Figure 30: Laminaribiose bound state of CtDfg5. **A** OMIT-map of laminaribiose with the Fo-Fc map shown in green and contoured at σ -level 3.0. Laminaribiose is shown as blue sticks bound to +1 GPI-glycan binding site with its non-reducing end in the context of secondary structure elements of CtDfg5. The second moiety defines an acceptor-specific subsite +2. The coordinating residues are shown as turquoise sticks. **B** A superposition with the GlcN-bound state shows the highly similar coordination at subsite +1. The coloring of the surface is according to the ConSurf analysis, highlighting the conserved site where PI-binding is suggested and the ABL-regions, which are neither conserved nor variable. Oxygens are colored in red, nitrogens in blue.

4.3 The Dfg5-subfamily: A potential drug target against fungal infections

Most diseases, which are in the focus of the public are associated with multi-resistant bacterial or epidemic viral infections, however the influence of fungal-based infections in humans cannot be underestimated. The *Global Action Fund for Fungal Infections* (<https://www.gaffi.org>) suggests that fungal infections are the 4th most common illness on earth. As a functional human immune system is usually able to cope with fungal challenges, primary infections are rare. Due to the increasing number of modern medication especially in clinical environments, invasive systemic mycoses are an increasing problem that require effective and safe drugs (Brown et al., 2012b). While antibiotics against bacteria can easily target structures that are absent in humans and therefore work quite specifically, eukaryotic targets usually come with serious side effects. It is therefore desirable to develop antifungal drugs, which target components of the fungal cell that are absent in humans. Accordingly, the fungal cell wall is an excellent target, since the structural components as well as the synthesizing factors are unique to fungi. It is not surprisingly that current antimycotica that are well tolerated by the patient belong to the class of echinocandins, which target the β 1,3-glucan synthase (Perfect, 2017). However, due to the frequent use in hospitals the number of resistant fungi, in particular *Candida* spp. is an increasing phenomenon resulting in

therapeutic failure (Perlin, 2015). As *Dfg5*-proteins are essential for these organisms and they do not share homology with human proteins, it might be a good starting point for treating fungal infections. Therefore, a structure-based search for potential inhibitors of *Dfg5*-proteins has been started.

4.3.1 Structure-based prioritization of a commercial fragment screen library

If a known protein structure is available, virtual screening is the method of choice to identify potential inhibitors. Usually this requires screening of large libraries with millions of possible lead structures. In order to find a suitable molecule for *CtDfg5*, a commercially available fragment library was chosen for initial studies. The molecules from the Frag Xtal Screen (*Jena Bioscience*) were further prioritized by docking analysis. For that, its 96 fragments were docked with *SeeSAR* to the GPI-glycan binding pocket of *CtDfg5* (BioSolveIT GmbH). Ten poses of each fragment were calculated using the default parameter of the software. The twelve best fragments were chosen regarding their estimated binding affinities to *CtDfg5*. Three of the fragments (34, 35, and 37) were estimated to bind in the low to mid μM -range, for six fragments (2, 24, 32, 48, 51, and 85) a mid μM to low mM-range have been calculated and another three molecules were attributed binding affinities in the mM region (Figure 31).

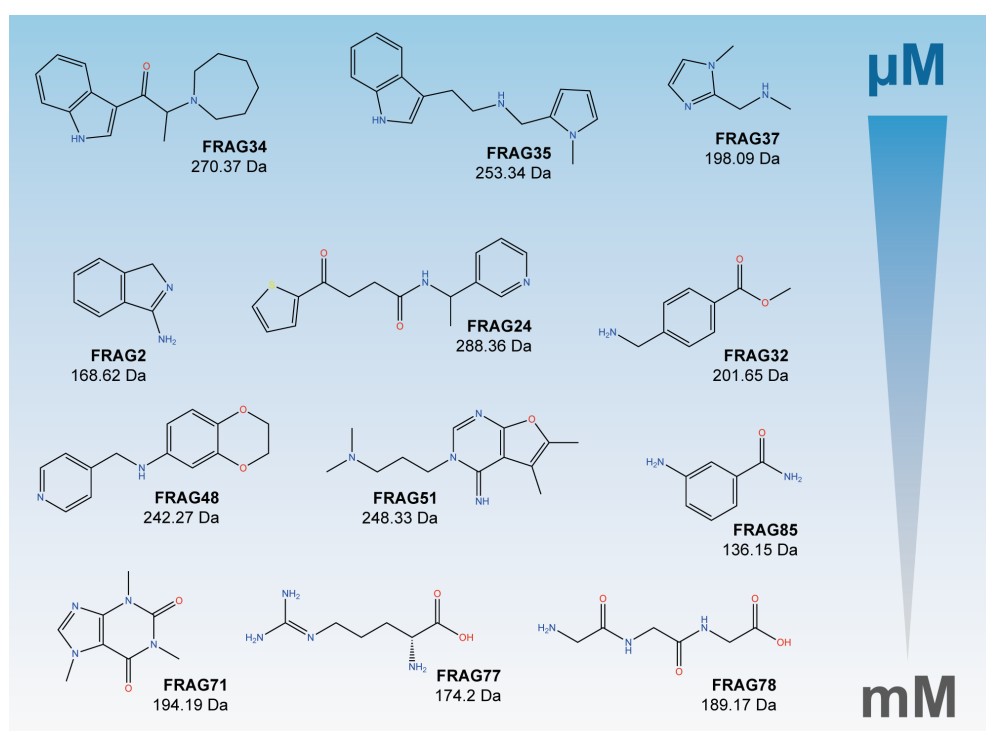


Figure 31: Twelve most promising hits from virtual ligand screening using *SeeSAR*. The chosen fragments are sorted according to their estimated binding affinities (see bar on the right). The fragments are presented as structural formulas and named according to their position in the Frag Xtal Screen. The respective molecular weight is given.

4.3.2 Crystal structure of CtDfg5 in complex with a potential inhibitor lead structure

The fragments identified above were diluted from 1 M DMSO stocks (*kind gift from AG Klebe*) to 100 mM concentrations in the already outlined buffer used for previous soaks. Interestingly, the condition with FRAG37, which could not be found in the crystal structure after 3 h of soaking, enhanced the diffraction quality of the crystal up to 1.05 Å resolution as mentioned above. Luckily, a positive hit for fragment binding could be found for FRAG51 (from now on referred to as FP-1 due to the furo[2,3-*d*]pyrimidin substructure that constitutes the majority of the molecule) after 3 h soaking time. The additional electron density from a high resolution data set (Table 10) could be unambiguously assigned to the fragment (see Figure 32A).

The binding within the GPI-glycan pocket of CtDfg5 is established by the furo[2,3-*d*]pyrimidin ring of FP-1, which is stacked by Y81. Furthermore, it forms a salt bridge between D325 and its propane-amine tail, which extends to subsite -1 and makes an electrostatic interaction with D134 (see Figure 32B). A superimposition with the GPI-core clearly reveals the excellent position at the active site of CtDfg5 giving a good initial hit for a competitive inhibitor lead structure (Figure 32C). Noteworthy, the observed orientation is contrary to the orientation that was predicted by *SeeSAR* (SFigure 8 on p.140).

Table 10 Crystallographic data collection and refinement statistics of CtDfg5 soaked with FP-1. Values in parentheses are for the highest resolution shell.

CtDfg5 _{FP-1}			
PDB code	6RY8		
Data collection		Refinement	
X-ray source	ID23-1, ESRF	Resolution range (Å)	30.27-1.3
Space group	C 1 2 1	R_{work}/R_{free} (%)	14.1/13.1
Unit-cell parameters (Å, °)	$a=83.31$, $b=55.04$, $c=80.35$, $\alpha=90.00$, $\beta=90.23$, $\gamma=90.00$	Average B factor (Å ²)	15.5
Wavelength (Å)	0.972	No. of atoms	
Resolution range (Å)	30.27-1.3 (1.35-1.3)	Total	4057
Completeness (%)	95.47 (96.61)	Protein	3346
Observed reflections	151167 (15034)	Ligand	33
Unique reflections	85345 (8647)	Solvent	678
Multiplicity	1.8 (1.7)	R.m.s.d., bond lengths (Å)	0.007
Wilson B factor (Å ²)	10.36	R.m.s.d., bond angles (°)	1.31
R_{merge} (%)	3.6 (25.3)	Ramachandran plot	
R_{meas} (%)	5.1	Favored (%)	99
Mean $I/\sigma(I)$	11.16 (2.77)	Allowed (%)	1
CC(½)	99.8 (88.1)	Disallowed (%)	0

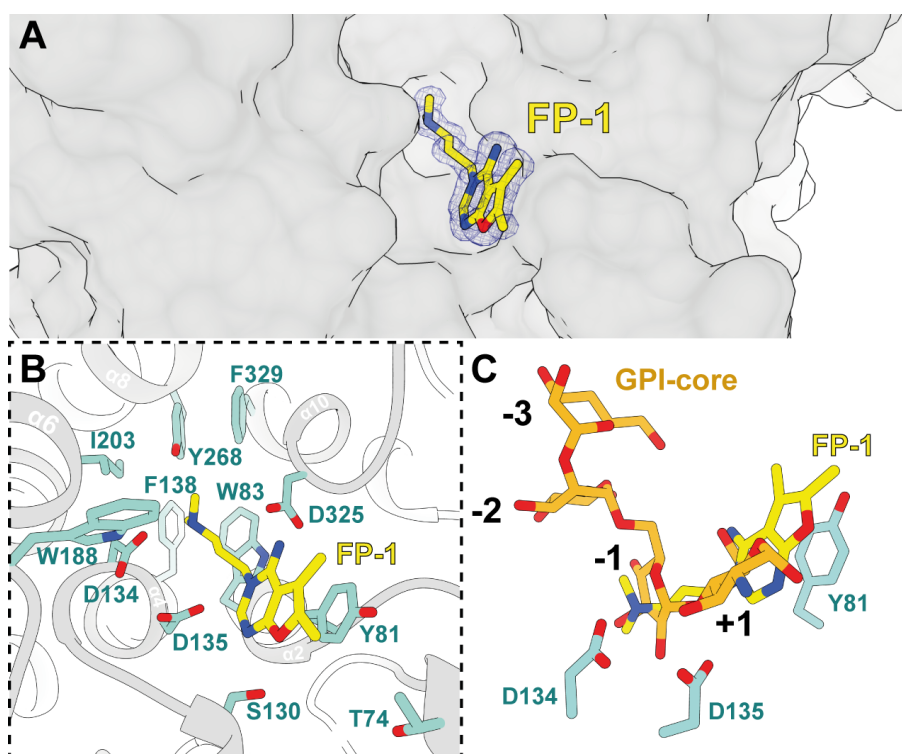


Figure 32: FP-1 bound above the active site of CtDfg5. **A** FP-1 is shown with its experimental electron density contoured at σ -level 1.0 within the binding pocket of CtDfg5. **B** FP-1 shown in the context of its interacting residues and the secondary structure of CtDfg5. **C** Superimposition of FP-1 and the GPI-core with Y81, D134, and D135 shown for orientation. The GPI-glycan binding sites are indicated. Residues and ligands are represented as sticks, all amino acids are colored in turquoise, FP-1 in yellow and GPI-core in orange, with the oxygens in red and nitrogens in blue.

As the binding mode of FP-1 suggests an inhibitory effect on the protein and non-functional *Dfg5*-proteins are lethal for *S. cerevisiae*, the effect of FP-1 supplementation in the yeast-growth medium will be analyzed in the following.

4.3.3 FP-1 impacts the viability of *S. cerevisiae*

In order to get an idea about a useful working concentration for *in vivo* tests, biolayer interferometry measurements were performed with the help of Dr. Sven-Andreas Freibert. The K_D of the FP-1:CtDfg5 interaction could be determined at 2.71 ± 0.96 mM (SFigure 9 on p.141). Based on that, Gesa Schmitz (AG Mösch) tested the effect of FP-1 on living cells. She tested three different strains: the wild type, and two single mutant strains of which one produced just Dfg5, while the other one produced only Dcw1. The growth behavior in YEPD medium alone of all three strains was almost identical (Figure 33A). Initially, 10 mM FP-1 was added to the YEPD medium, however no growth could be observed. Therefore, the concentration was reduced to 5 mM FP-1 (Figure 33B). As a result, wild type cells could not reach exponential growth phase within the observed time and finally reached a cell density

approx. half the cell density without FP-1. While this already shows an impact on the fitness of treated cells, the single mutants possessing only one *Dfg5*-protein were barely able to grow. Small differences can be observed between the *Dfg5* and *Dcw1* mutant, as a slight increase in optical density can be seen for *Dfg5*. In order to test, whether this effect indeed relies on the interaction between the supplemented FP-1 and the protein, a structural derivative called FP-2 has been analyzed as well. Instead of the two methyl groups, FP-2 possesses an imidazole ring at the furo[2,3-*d*]pyrimidin-distal end of the propyl-tail (Figure 33C). FP-2 failed to produce a complex structure upon soaking experiments and the K_D could not be determined by biolayer interferometry (SFigure 9 on p.141). Wild type cells could grow in the presence of 5 mM FP-2 until saturation, which is quite convincing in this context (Figure 33C). The *Dfg5* mutant also reached the saturation 2 h after the wild type. The *Dcw1* mutant clearly suffered from the FP-2 treatment, however it could also reach the exponential phase at the end of the observed period of time.

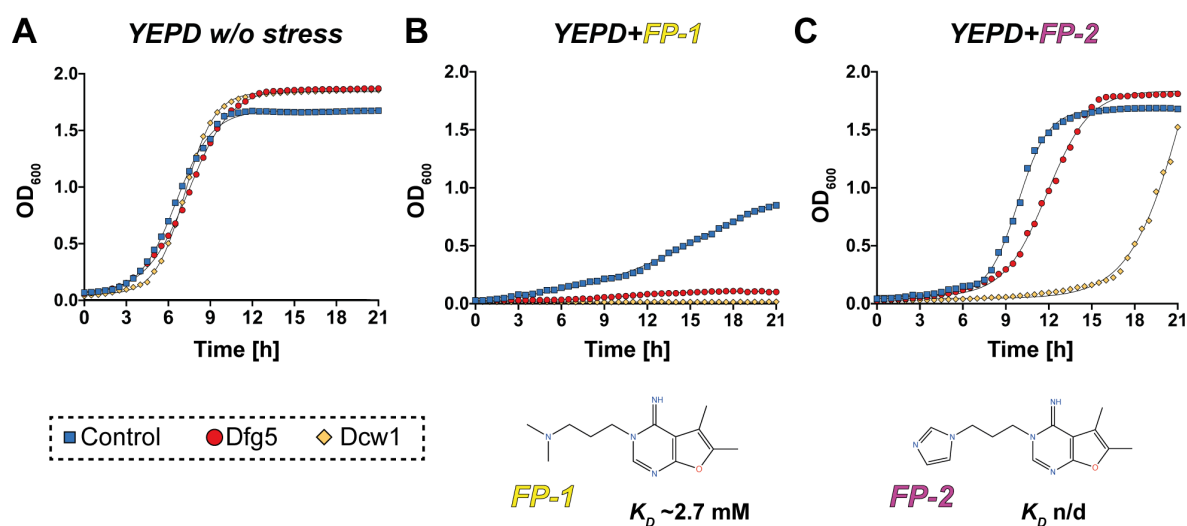


Figure 33: Effect of FP-1 and a derivate on the growth of *S. cerevisiae*. Growth curves of three different strains (indicated in panel A) are shown in YEPD (A), YEPD supplemented with 5 mM FP-1 (B), and YEPD supplemented with 5 mM FP-2 (C). The structural formulas of the respective supplement are given below the respective growth curve. The *in vivo* experiments were performed by Gesa Schmitz (AG Mösch). The binding studies were performed with the help of Dr. Sven-Andreas Freibert, who analyzed the data.

These findings indicate a promising starting point for an efficient drug against ascomycetes.

4.3.4 Combined fragment state as the structural basis for lead structure optimization

In order to optimize a lead structure in terms of its binding properties, one promising approach can be the combination of different binders. As we have seen in the sugar-fragment screen before, mannose is able to bind to GPI-glycan subsite -2 when *CtDfg5* crystals are soaked in 500 mM mannose containing mother liquor. This should allow a combination with FP-1, which was identified at subsites -1/+1.

To show simultaneous binding of mannose and FP-1, both substances were used for soaking experiments. The final dataset was collected from a *CtDfg5* crystal soaked in the mother liquor as described above supplemented with 1 M mannose and 100 mM FP-1 at the PXIII beamline at the Swiss Light Source (SLS) in Villigen, Switzerland (see Table 11).

Table 11: Crystallographic data collection and refinement statistics of *CtDfg5* soaked with FP-1 and mannose. Values in parentheses are for the highest resolution shell.

<i>CtDfg5</i> _{FP-1+Man}			
Data collection		Refinement	
X-ray source	PXIII, SLS	Resolution range (Å)	30.28-1.2
Space group	C 1 2 1	R_{work}/R_{free} (%)	12.4/13.8
Unit-cell parameters (Å, °)	$a=83.66$, $b=55.06$, $c=80.68$, $\alpha=90.00$, $\beta=90.60$, $\gamma=90.00$	Average B factor (Å ²)	18.73
Wavelength (Å)	0.999	No. of atoms	
Resolution range (Å)	30.28-1.05 (1.09-1.2)	Total	3929
Completeness (%)	96.56 (88.12)	Protein	3335
Observed reflections	207437 (16167)	Ligand	53
Unique reflections	110541 (10023)	Solvent	541
Multiplicity	1.9	R.m.s.d., bond lengths (Å)	0.015
Wilson B factor (Å ²)	14.34	R.m.s.d., bond angles (°)	1.68
R_{merge} (%)	2.2 (30.4)	Ramachandran plot	
R_{meas} (%)	3.1 (43.0)	Favored (%)	98.53
Mean $I/\sigma(I)$	14.56 (2.47)	Allowed (%)	1.47
CC(½)	99.9 (75.2)	Disallowed (%)	0

As it was expected from the individual structures, the simultaneous binding of both fragments together is not interfered (see Figure 34). Both molecules are found in the same conformation like observed before for the individual complexes. The shortest distance between them is 3.3 Å from the 1-hydroxy group of the mannose at subsite -1 (Man1) and the respective methyl group of FP-1.

Another interesting point of this complex structure is additional density of a mannose moiety (Man2) above Y199, which is located in the β 1-strand. The presence of that sugar can be explained due to the higher mannose concentration used for the soaking experiment compared to *CtDfg5*_{Man} (1 M instead of 500 mM). The closest contact is 3.8 Å established between

3-hydroxy group of Man1 and the 4-hydroxy group of Man2, which may provide another possible linker between to fragments to increase the binding properties of a *Dfg5*-subfamily specific inhibitor. Furthermore, it raises the possibility of another GPI-glycan binding site (i.e. subsite -4), as the common core structure of the GPI-anchor in fungi harbors another α 1,2-linked mannose moiety that branches off mannose 3.

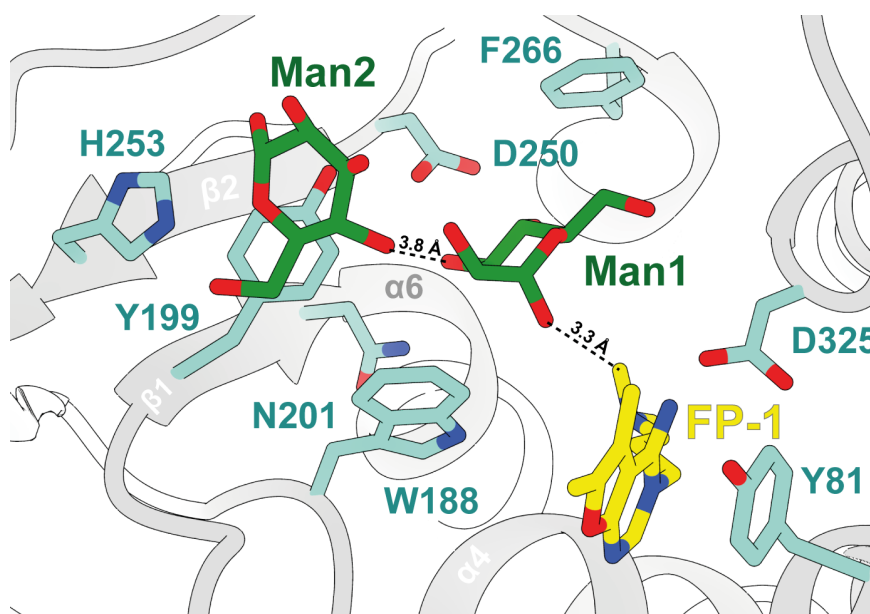


Figure 34: Combined soaking of mannose and FP-1. The bound ligands and their coordinating residues are shown as sticks in the context of the secondary structure elements from CtDfg5 colored in grey. Amino acid carbons are colored in turquoise, mannose carbons in green and FP-1 carbons in yellow. Oxygens are red and nitrogens blue. The closest distances between FP-1 and Man1, and Man1 and Man2 are indicated.

4.4 Expanding the knowledge to the *Fungal/Bacteria mixed* subfamily

To this point, bacterial and fungal GH76 proteins have been described for three different subfamilies and it was shown that their function differs drastically. After elucidating the functional background of *Dfg5*-proteins and their potential role as a drug target, the last part of this thesis will deal with another subfamily of GH76 proteins trying to emphasize the common features of the whole family.

So far, the knowledge about the *Fungal/Bacteria mixed* subfamily is rather limited. One of its members, which is in the following referred to as CtGH76 (Uniprot-ID G0S5Y9), is composed of 439 amino acids and in contrast to CtDfg5, no further annotations regarding its topology or posttranslational modifications are given. No signal peptide or ω -signal sequence are predicted by common *in silico* tools such as SignalP-5.0 or the *GPI Fungal Prediction*

Server. However, the transmembrane prediction tool TMHMM2.0 provides a hint about its topology (see Figure 35) (Eisenhaber et al., 2004; Sonnhammer et al., 1998).

Although the general output does not predict any transmembrane helix (TMH), the resulting expectation number is 17.6. This is close to the cutoff criterion 18, which is thought to predict a TMH to be very likely and, furthermore, a warning of a possible N-terminal signal sequence is produced. As a TMH at the N-terminal region cannot be excluded from this initial analysis, the first 50 amino acids were dismissed for soluble heterologous protein production in *E. coli* SHuffle, which was established by Markus Friedrich during his master thesis using the same expression condition as described in this thesis.

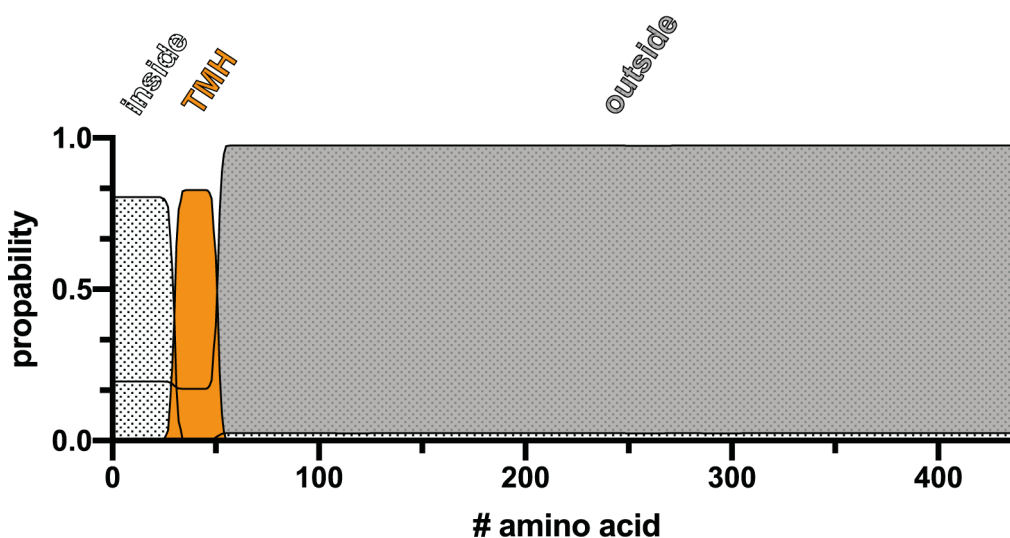


Figure 35: TMHMM probability plot of CtGH76. The topology of CtDfg5 is shown with a probability plotted against the amino acids. From the TMHMM prediction tool analysis a TMH can be predicted thus giving rise to a putative type-II membrane protein.

Although the phylogenetic analysis of the GH76 class suggested a quite close relationship of the *Fungal/Bacteria mixed* subfamily to the bacterial homologs, Markus Friedrich could not show hydrolysis of α 1,6-manno oligosaccharides by catalysis of CtGH76. This goes along with the previous findings for CtDfg5, though it was a little bit surprising due to the close relationship to the *bacterial* subfamilies (see Figure 14 on p.53).

Markus Friedrich was able to obtain the crystal structure of CtGH76. It crystallized in space group P 31 2 1 and diffracted to 2.0 Å resolution. As shown before for *bacterial* GH76 and Dfg5-proteins, this *mixed* subfamily has the GH76-typical $(\alpha/\alpha)_6$ -helical barrel fold as well (Figure 36). The most significant differences compared to the already described homologs are the regions indicated as β 1/2-wing and β 3/4-wing. In the superposition with CtDfg5 it is easy

to see that these regions are prolonged in *CtGH76*. This difference is unique for *CtGH76* and not found in other GH76 structures, which are excluded from superposition for clarity. These extended regions lead to a break of the antiparallel β -sheet between α -helices 5/6 and 7/8 described for *CtDfg5*, however forming two antiparallel sheets in the loop between α -helices 7/8 resulting in the β 1/2-wing. Noteworthy, the corresponding β -strands (i.e. β 1 of *CtGH76* and β 2 from *CtDfg5*) are tilted to each other. Furthermore, the loop extension between α -helices 11 and 12 results in the formation of two additional β -strands forming the antiparallel β 3/4-wing region in *CtGH76*, a structural element that is completely absent in the other GH76 proteins.

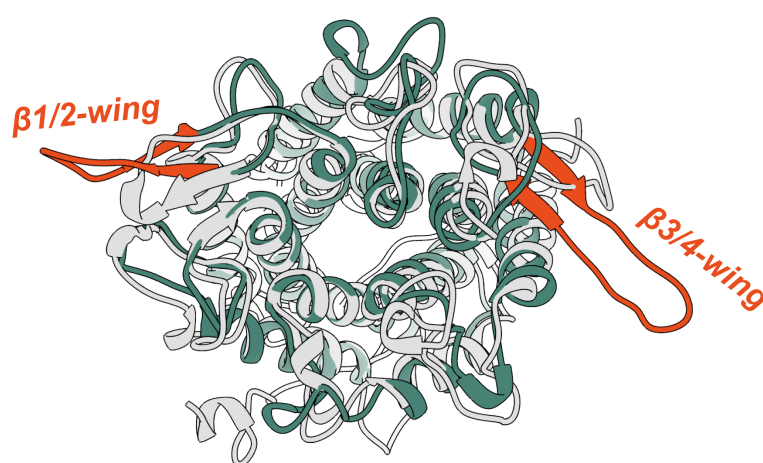


Figure 36: Superposition of *CtGH76* and *CtDfg5*. Both structures are shown with their secondary structure elements. *CtDfg5* is shown in grey, *CtGH76* is shown in blue-green and its indicated wing regions are colored in red.

The biological role of the *Fungal/Bacteria mixed* subfamily in fungi is still elusive. With the initial *in silico* findings it is even hard to predict the actual destination of such proteins making any functional prediction highly speculative. In the following, *CtGH76* is subjected to sugar-fragment soaking experiments as described for *CtDfg5* to get more insights into its functional properties.

Unlike it was found for *CtDfg5*, glucosamine could neither be identified at subsite +1, nor at any other possible glycan-binding site of *CtGH76*. An explanation might be that the glucosamine-coordinating residues Y81 and Q189 from *CtDfg5* are substituted by a proline and an aspartate in *CtGH76*. However, this binding site does not only seem to exclude the GPI-glycan binding ability described for *Dfg5*-proteins, but also does not support an α 1,6-linked mannose residue like it is known for the *bacterial* homologs.

As shown above the *bacterial* and *Dfg5*-subfamilies apparently share their binding mode at subsites -2 to -1, where an α 1,6-linked mannanose structure can be observed. However, the α 1,6-mannanose-bound states per se differ, as the biose is coordinated at subsites -3/-2 in the bacterial ortholog, while *CtDfg5* coordinates the disaccharide at -2/-1 (see Figure 48 on p.108). In order to achieve that state for *CtGH76*, α 1,6-mannanose is dissolved to a final concentration of 1 M in a solution consisting of equal volumes of the protein buffer (200 mM NaCl, 20 mM HEPES, pH 8.0) and the crystallization condition from the JCSG Core IV suite, E5 (3.6 M Sodium formate, 10% (v/v) glycerol). The soaking was performed at room temperature for three hours. The crystals were directly flash frozen in liquid nitrogen and diffraction studies resulted in a dataset that was solved at 2.1 Å resolution (Table 12).

Table 12: Crystallographic data collection and refinement statistics of *CtGH76* soaked with α 1,6-mannanose. Values in parentheses are for the highest resolution shell.

<i>CtGH76</i> _{α1,6M}			
Data collection		Refinement	
X-ray source	PXIII, SLS	Resolution range (Å)	49.24-2.25
Space group	P 31 2 1	R_{work}/R_{free} (%)	20.4/21.8
Unit-cell parameters (Å, °)	$a=106.84$, $b=106.84$, $c=127.0$, $\alpha=90.00$, $\beta=90.00$, $\gamma=120.00$	Average B factor (Å ²)	68.1
Wavelength (Å)	0.999	No. of atoms	
Resolution range (Å)	49.24-2.25 (2.33-2.25)	Total	2905
Completeness (%)	99.74 (99.57)	Protein	2806
Observed reflections	80464 (7980)	Ligand	50
Unique reflections	40269 (3990)	Solvent	49
Multiplicity	2.0	R.m.s.d., bond lengths (Å)	0.010
Wilson B factor (Å ²)	47.78	R.m.s.d., bond angles (°)	1.32
R_{merge} (%)	3.1 (24.5)	Ramachandran plot	
R_{meas} (%)	4.4 (34.6)	Favored (%)	98.33
Mean $I/\sigma(I)$	9.3 (2.0)	Allowed (%)	1.11
CC(½)	99.9 (94.0)	Disallowed (%)	0.56

Unlike in the bacterial homolog, the additional density could be observed at glycan-binding sites -2 and -1 corresponding to the two α 1,6-linked mannoses (Figure 37). The overall binding appears as it was observed for the respective state of *CtDfg5*, however a striking difference becomes obvious in that context. In all described GH76 structures so far, a tryptophan (W188 in *CtDfg5*, W172 in *BcGH76*) residue is directing the α 1,6-linkage and the -1 moiety in the correct position due to its steric presence. Conversely, *CtGH76* possess a valine (V210) at that position, which does not result in a narrow channel but a more open binding pocket. However, a functional relevance of that substitution is questionable, as the majority of *Fungal/Bacteria mixed* proteins possess a tryptophan at that position.

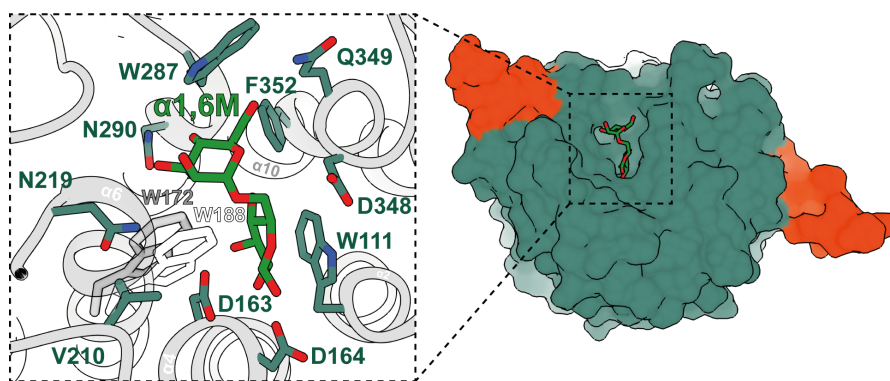


Figure 37: CtGH76 bound to α 1,6-mannobiose. On the left side, the coordination of α 1,6-mannobiose (α 1,6-M) at subsites -2 to -1 is shown in context of the secondary structure elements of CtGH76. The coordinating residues are shown as sticks as well as the residues W188 (white) and W172 (transparent grey) from CtDfg5 and BcGH76, respectively. On the right, the ligand is shown in the context of a surface representation of CtGH76, with the majority of the surface colored in blue-green and the above-explained wing-regions colored in red. Oxygens are shown in red, nitrogens in blue.

Due to the extended wing regions it seems to be possible that this region may be involved in carbohydrate coordination. Therefore a trisaccharide was tested in the soaking screen as described before, which is composed of three mannoses that resemble a core component of the high mannose N-glycan from fungi (Figure 38A). Here, two mannoses are α 1,3- and α 1,6-linked to a central mannose. The soaked crystals were treated as before with a 1 M soaking solution and a complex structure could be determined at 2.6 Å resolution (Table 13).

Table 13: Crystallographic data collection and refinement statistics of CtGH76 soaked with mannotriose. Values in parentheses are for the highest resolution shell.

CtGH76 _{Man3}			
Data collection		Refinement	
X-ray source	PXIII, SLS	Resolution range (Å)	49.1-2.6
Space group	P 31 2 1	R_{work}/R_{free} (%)	16.7/19.6
Unit-cell parameters (Å, °)	$a=106.59$, $b=106.59$, $c=126.24$, $\alpha=90.00$, $\beta=90.00$, $\gamma=120.00$	Average B factor (Å ²)	61.73
Wavelength (Å)	0.999	No. of atoms	
Resolution range (Å)	49.1-2.6 (2.69-2.6)	Total	2893
Completeness (%)	99.83 (99.92)	Protein	2806
Observed reflections	51928 (5077)	Ligand	46
Unique reflections	25983 (2539)	Solvent	41
Multiplicity	1.8	R.m.s.d., bond lengths (Å)	0.010
Wilson B factor (Å ²)	55.40	R.m.s.d., bond angles (°)	1.31
R_{merge} (%)	4.2 (30.6)	Ramachandran plot	
R_{meas} (%)	6.0 (43.2)	Favored (%)	98.05
Mean $I/\sigma(I)$	8.2 (2.0)	Allowed (%)	1.11
CC(½)	99.7 (83.0)	Disallowed (%)	0.84

Indeed, the additional density appeared close to the described β 1/2-wing region interacting with its proximal residues A271 and Y280 (Figure 38B). The C6-carbon involved in the α 1,6-linkage is stacked by Y217 and L284 guides the orientation of C2-C1-O1 from the α 1,3-linked mannose. Electrostatic interactions are facilitated by T216 with the 2-hydroxy group of the α 1,6-linked mannose and N282 to the reducing end of the central mannose. Additional density also appeared in the -2 subsite, which may be attributed to a single mannose that might be a contamination due to partial hydrolysis of the triose.

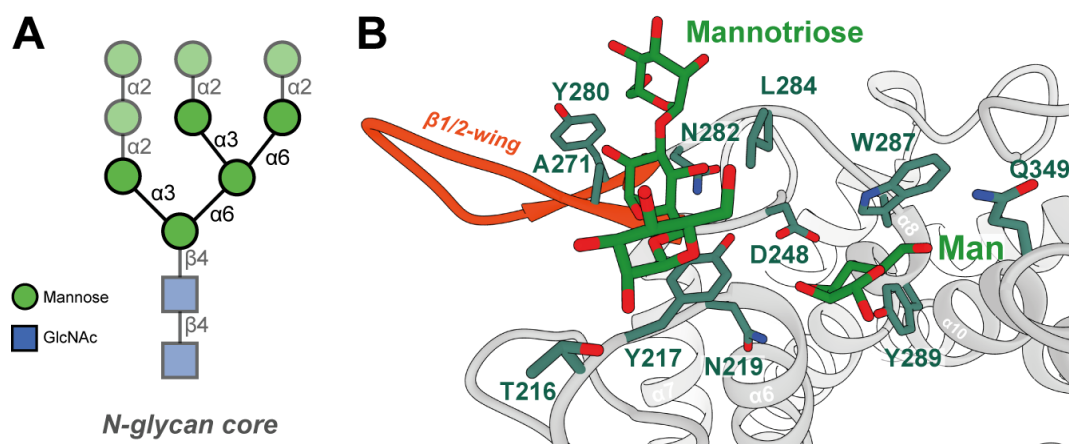


Figure 38: Mannotriose bound state of CtGH76. **A** The schematic representation of the high mannose core of fungal N-glycans is shown. The mannose is presented twice in that structure highlighted by its opacity. The symbols are explained at the bottom left and the respective linkages are indicated. **B** Mannotriose, mannose and their coordinating residues are shown as sticks in the context of the secondary structure elements from CtGH76. The β 1/2-wing is emphasized in red, while the remaining cartoon is in grey with labeled α -helices. Amino acid-carbons are colored in blue-green, carbohydrate-carbons in green, oxygens are red, and nitrogens are colored in blue. *GlcNAc*=N-acetylglucosamine; *Man*=mannose.

5 Discussion

The cell wall is a special part of the fungal cell. Although not being part of the interior, the outermost layer must be built and maintained in such a fine-tuned manner to simultaneously ensure the protective and rigid role, as well as its adaptable and highly dynamic character. While the mere synthesis of the polysaccharides and proteins of the fungal cell wall (FCW) happens along the secretory pathway either soluble or embedded in the plasma membrane, the further glycan-processing and the wall's maturation is established by a set of proteins that act extracellularly. An essential element of that system is the incorporation of GPI-anchored proteins (GPI-AP) into the FCW (Kitagaki et al., 2002). Glycoside hydrolases of the GH76 class were identified to be responsible for the lipid-to-wall transfer, however the exact mode of action was not clear, as already described bacterial homologs were attributed an enzymatic activity that contradicts the previously described way of GPI-CWP attachment (Ao et al., 2015; Cuskin et al., 2015; Fujii et al., 1999; Kitagaki et al., 2002; Kollár et al., 1997; Thompson et al., 2015b).

Initial sequence similarity analyses from this study gave a hint that GH76-proteins are not an isofunctional group of proteins, but possess a still elusive number of different functions in bacteria, fungi and archaea. One of the occurring subfamilies exclusively contained proteins from ascomycota, to which a cell wall defective phenotype had already been ascribed. A member of that *Dfg5*-subfamily from the thermophilic filamentous fungi *Chaetomium thermophilum*, *CtDfg5*, was then assigned to an in-depth analysis. This allowed insights into the substrate binding mode and the way of how CWP are transferred to the glucan meshwork. This work also showed the potential of *Dfg5*-proteins as a drug target and provides an idea about the general feature of GH76-proteins by expanding the structural repertoire of this family with complex structures of an additional member from the *Fungal/Bacteria mixed* subfamily, called *CtGH76*.

The different aspects of this thesis mentioned above will be discussed in the following.

5.1 A subfamily of GH76-proteins act on fungal cell wall biogenesis

Due to the technological advances in recent years, sequencing of entire genomes has become a routine, which provides us with millions of sequences. As plain sequences are useless for researchers, functional assessment is required. The first stage of protein domain identification is fulfilled by protein family databases such as InterPro (<http://www.ebi.ac.uk/interpro/>) or

Pfam (<https://pfam.xfam.org>). The developers are constantly challenged with the increasing number of sequences, trying to improve the classification and functional annotation based on existing experimentally characterized proteins (El-Gebali et al., 2019; Mitchell et al., 2019). A key problem of sequence-based protein function assignment to a complete protein family is the uncertainty in the low-identity region, which may still give an idea about superior features of the respective protein class, however the exact mechanistic function is often disguised in the *Twilight zone* and the general assignment of a few homologs to a complete family can result in misleading hypotheses (Rost, 1999).

In order to clarify the controversy in the current literature regarding the function of fungal GH76-proteins, the first aspect that was addressed was, whether this protein family is isofunctional per se, or not. This was tackled by generating a sequence similarity network (SSN) with the intention to produce isofunctional clusters. In the presented SSN (see Figure 14 on p.53), the family already falls apart at moderate cutoff criteria into ten major subfamilies. This behavior indicates that GH76 have diverse functional properties. It happens that all fungal proteins, which were previously described to be involved in cell wall biogenesis, are exclusively found in the *Ascomycota I* subfamily (Table 14). Therefore, this subfamily has been renamed to *Dfg5* with respect to the first ortholog that was described (Mösch and Fink, 1997). Consequently, it must be concluded that proteins from this cluster follow the same functional principles and thereby disagree with the current annotation as α 1,6-mannanase. Notably, the bacterial subfamilies well separate from the *Dfg5*-subfamily, strengthening the assumption that the latter's catalytic function is indeed different.

Table 14: Fungal *Dfg5*-proteins described in the literature regarding their role in the cell wall biogenesis. ^(#) In the cited publication, single disruptants were analyzed concerning CW-related phenotype for all nine GH76 proteins found in *N. crassa*, of which just the two shown in the table were further characterized. ^(*) Q4WIT8 (called Dfg2 in the cited publication) does not belong to the *Dfg5*-subfamily, but to the *Fungal/Bacteria mixed* cluster. A further GH76-protein belonging to the *Ascomycota II* subfamily (Q4WI31) was not analyzed in this study.

Organism	Uniprot	Publication
<i>S. cerevisiae</i>	Q05031, P36091	(Kitagaki et al., 2002, 2004)
<i>C. albicans</i>	Q5ACZ2, Q5AD78	(Ao and Free, 2017; Spreghini et al., 2003)
<i>N. crassa</i>	Q1K7A8, Q7S4K4	(Maddi et al., 2012) [#]
<i>A. fumigatus</i>	Q4WKP7, Q4WIT8, Q4WG09, Q4WFX5, Q4W985, Q4WFI1, Q4WA43	(Muszkieta et al., 2019) [*]

5.2 How GPI-AP's are attached to the cell wall glycan

In order to determine the catalytic mechanism within the *Dfg5*-subfamily, the crystal structure of a homolog from *Chaetomium thermophilum* was determined. The structure alone revealed the conserved (α/α)₆-helical barrel fold, which is known from the already described bacterial GH76-proteins, including the conserved position of the active site DD-motif (Cuskin et al., 2015; Thompson et al., 2015b). While the active site motif was found to be essential in structure-based mutants of *S. cerevisiae*, the α 1,6-mannanase activity on a mannotriose and a mannohexaose could not be observed for *CtDfg5*. Interestingly, this was suggested as a key feature regarding the hypothesis of CWP-attachment via N-linked outer chain mannans (Maddi et al., 2012).

For a better understanding of the functional properties of *CtDfg5*, complex structures were sought to be determined. As suitable substrates of complex carbohydrates are not commercially available, *CtDfg5* crystals were soaked with sugar fragments of the N-linked outer chain mannan and the GPI-anchor. Surprisingly and very convincingly at the same time, complex structures of *CtDfg5* with mannose, α 1,2-mannobiose, α 1,6-mannobiose, and glucosamine could be obtained, which together reassembled the core structure of the GPI-anchor within the binding pocket of the protein from GPI-glycan binding sites -3 to +1. This not only sets the basis for the true role of *Dfg5*-proteins in the lipid-to-wall transfer of GPI-CWPs, it also provides first insights into the conformational properties of GPI-anchors as discussed below. In addition to the assembled substrate-bound state of *CtDfg5*, the protein was also successfully soaked with laminaribiose, the disaccharide unit of the major cell wall polysaccharide β 1,3-glucan. This structure revealed the non-reducing end bound in the same manner coordinated to subsite +1, as it was observed for the glucosamine (Figure 39).

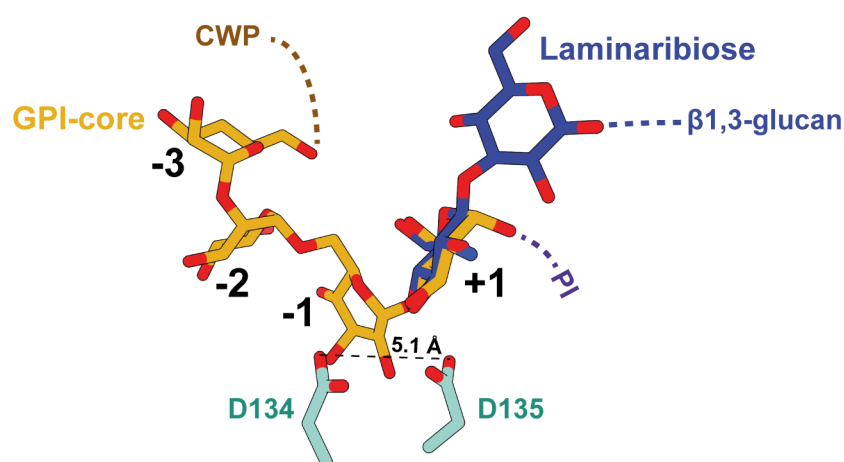


Figure 39: Superposition of the reassembled GPI-core and laminaribiose bound to *CtDfg5*. The active site DD-motif (turquoise), the GPI-core (orange) and laminarib

iose (blue) are shown as sticks. The *native* extensions are added to the respective termini by the colored dashed lines. The dashed line between the carboxy-groups of D134 and D135 shows their distance as indicated. The GPI-glycan binding subsites are indicated from -3 to +1. CWP=cell wall protein; PI=phosphatidylinositol.

With this state of an acceptor sugar ready to attack a covalent glycosyl-enzyme intermediate and the typical distance of the active site residues of ~ 5 Å for retaining glycosidases, it is possible to propose a structure-based mechanism for the two-step transglycosylation reaction of *Dfg5*-proteins exemplary for *CtDfg5* (see Figure 40). Upon binding of the GPI-core, the nucleophilic D134 attacks the anomeric center of Man1, while a proton donated by D135 supports the departure of the GlcN-PI leaving group. This results in a covalent glycosyl-enzyme intermediate, typical for retaining reaction mechanisms (Figure 40). After the release of the leaving group, the acceptor polysaccharide enters the scenery with its non-reducing end occupying subsite +1. In this particular case, the binding of a β 1,3-linked laminaribiose was observed, however from the coordination it seems to be important that the +1-penetrating sugar-moiety is the non-reducing end of a glucose-polymer. In the following step, the 4-hydroxy group is activated by the deprotonated D135, allowing a nucleophilic attack on the glycosyl-enzyme intermediate, which results in the formation of a new α 1,4-glycosidic bond established between mannose and glucose. Thompson and colleagues proposed a reaction coordination with a $B_{2,5}$ transition state following a ${}^0S_2 \leftrightarrow B_{2,5}^\ddagger \leftrightarrow {}^1S_5$ itinerary for GH76 retaining α -mannanases (Thompson et al., 2015b). To comment on the conformational distortion of the -1 subsite moiety of *CtDfg5* in order to support the glycosidic bond breakage, a -2 to +1 spanning molecule would probably be the minimum requirement to allow the necessary distortion. Unfortunately, such a state could not be achieved, moreover the -2/-1 subsite state mapped by a bound α 1,6-mannobiose presumably does not reflect the catalytically competent conformation of Man1, as the 1- and 2-hydroxy groups coordinate a Ca^{2+} together with D135. Anyway, the presence of calcium is an interesting fact regarding the preference of *Dfg5*-proteins to catalyze transglycosylation rather than hydrolysis observed for the bacterial homologs. The negatively charged plasma membrane is suggested to increase the proximal calcium concentrations up to 30 times compared to the bulk concentration and that the binding of calcium to the bilayer is an entropy-driven process due to the displacement of interfacial water (Binder and Zschörnig, 2002; Seelig, 1990; Sinn et al., 2006; Vernier et al., 2009). This seems to be a perfect environment for *Dfg5*-catalyzed transglycosylation due to proximity to the plasma membrane with an increased amount of calcium that inhibits

hydrolysis, a low water availability in general and the supply of acceptor molecules by their membrane embedded synthesis.

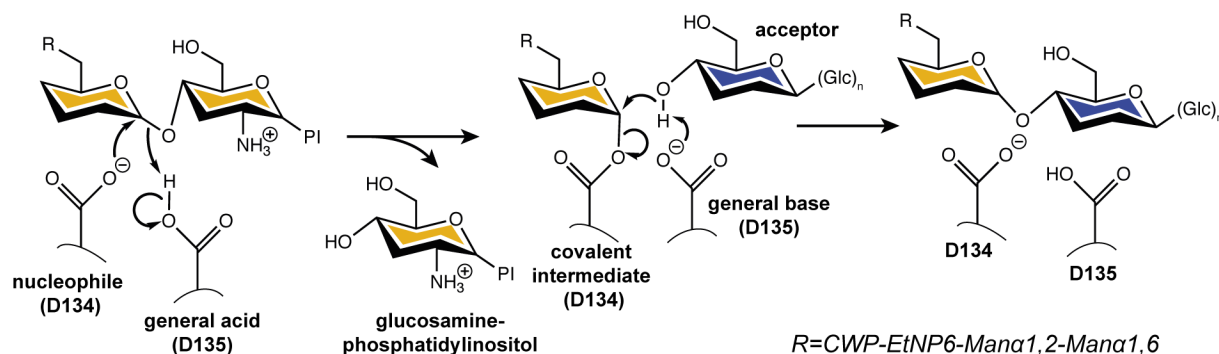


Figure 40: Mechanism of the transglycosylation mediated by *Dfg5*-proteins. A classical retaining mechanism can be suggested with D134 (CtDfg5-nomenclature) acting as the nucleophile and D135 as the general acid/base residue. Upon the first cleavage step of the glycosidic bond, the glycosyl-enzyme intermediate is formed. In the second step, the acceptor molecule (instead of the water from the general hydrolysis mechanism) intercepts the anomeric carbon and thereby forms a new glycosidic bond. PI=phosphatidylinositol; (Glc)_n=glucose polymer, here $\beta 1,3$ -glucan.

In the context of the incorporation of cell wall proteins, a model for the lipid-to-wall transfer can be suggested (Figure 41). The first step is the binding of the GPI-AP and the subsequent cleavage and release of the GPI-lipid, while the protein is covalently attached to Dfg5. This is a crucial step in the transglycosylation event, as it prevents the GPI-AP to be lost to bulk solvent. Dfg5 then presents the covalently bound GPI-AP to the cell wall glycan and allows the binding of a suitable acceptor molecule, which fits to the +1 GPI-glycan binding subsite. Subsequently, the new glycosidic bond is established and the GPI-AP becomes a GPI-CWP bound with its GPI-core to the non-reducing end of the cell wall glycan. The transfer essentially requires the GPI-core structure. Strikingly, the recent findings in *A. fumigatus* described that *Dfg5* proteins are responsible for the transfer of GPI-anchored galactomannans, which goes along with the findings within this work (Muszkieta et al., 2019).

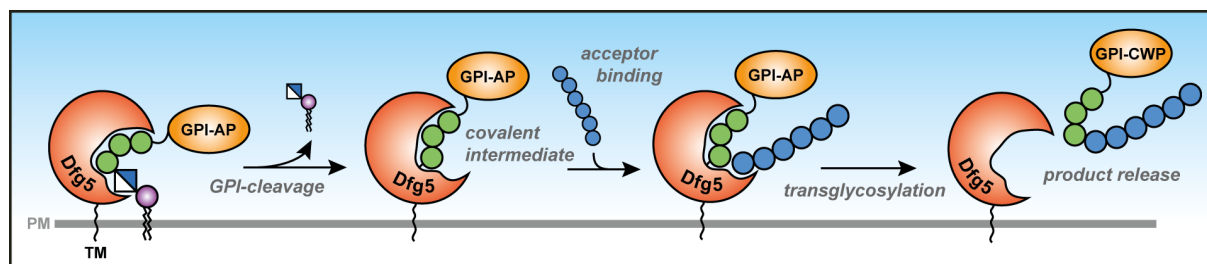


Figure 41: Model of how GPI-APs are attached to the cell wall glycan. Upon GPI-AP recognition, *Dfg5*-proteins cleave the GPI-anchor core by forming a covalent intermediate. After binding of the acceptor molecule, the GPI-remnant is transferred on the non-reducing end of the cell wall glycan. Finally, a GPI-CWP is released from the enzyme. TM=transmembrane helix; PM=plasma membrane.

5.3 To be or not to be sorted into the fungal cell wall

A major question in the context of GPI-CWPs is how the sorting into the cell wall occurs. In principle all GPI-APs end up at the plasma membrane. Some of these proteins can be classified as GPI-plasma membrane proteins (GPI-PMPs), others are transferred into the cell wall ending up as GPI-CWPs. However, this division seems to be not absolutely strict, as several GPI-APs end up in both locations (Pittet and Conzelmann, 2007). The transglycosidases Dfg5 and Dcw1 themselves are usually produced as GPI-APs in ascomycota, so it must be ensured that they remain at the plasma membrane to incorporate GPI-CWPs as described above.

In yeast, temperature-sensitive mutants of a gene involved in the cell-division cycle (*cdc1^{ts}*) were shown to stop growth in early G2 phase having small buds and replicated DNA, a phenomenon that was found in *dcw1^{ts}* mutants as well (Hartwell et al., 1970; Kitagaki et al., 2004). More recently it has been shown that Cdc1 removes the EtN-P added to Man1 by Mcd4 during GPI-anchor biosynthesis and that it can be deleted in a $\Delta mcd4$ background (Vazquez et al., 2014). It is not clear, what the consequences of EtN-P removal by Cdc1 exactly are regarding intracellular trafficking. Interestingly, Ted1, the phosphodiesterase that removes the EtN-P from Man2, is shown to act together with Emp24 and Erv25 at the ERES for facilitating efficient cargo exit from the ER (Haass et al., 2007). With the reassembled GPI-core structure of *CtDfg5*, however, it is possible to explain the consequences caused by the presence of an EtN-P at Man1. By adding an EtN-P group to the 2-hydroxy group of Man1 it becomes obvious that there is no space left at subsite -1 for suitable binding as it penetrates the proteins surface (Figure 42A). Such a modification most likely results in the non-binding of the GPI-anchor core, in any case there is no catalytically relevant conformation (and distortion) of Man1 anymore possible, thus preventing cleavage of the Man α 1,4-GlcN linkage and subsequent transfer onto the cell wall glycan. This explains the connected phenotypes of *cdc1* and *dfg5/dcw1* mutants, which basically show the same behavior for complete deletion (lethal) and the temperature-sensitive mutants (G2 arrest) as both result in an abolished or compromised lipid-to-wall transfer of GPI-CWPs, respectively.

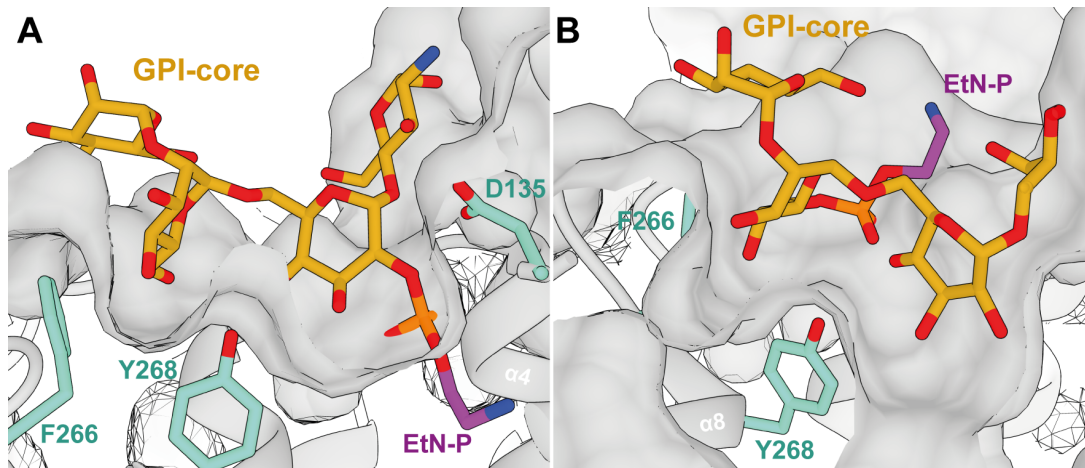


Figure 42: Hypothetical positions of EtN-P at Man1 and Man2. The additional, hypothetical EtN-P (purple) added to Mannose 1 (A) and Mannose 2 (B) of the GPI-core (orange) are shown in context of the clipped surface of CtDfg5 (grey) with marker residues (turquoise) shown for orientation.

In contrast to Cdc1, the removal of EtN-P from Man2 by Ted1 is not essential in *S. cerevisiae*, though it was shown that its deletion results in a delayed ER-export of GPI-APs, as it works in a concerted manner with the p24-complex that is responsible for efficient COPII traffic from ER to Golgi by interaction with the mature GPI-glycan (i.e. without the Man2-EtN-P) (Manzano-Lopez et al., 2015). The findings of $\Delta ted1$ mutants resemble the deletion mutants of the p24-complex components *emp24* and *erv25*, which show the same defect of ER-exit (Belden and Barlowe, 1996; Schimmöller et al., 1995). The consequences for the final cell wall delivery by Dfg5-proteins in $\Delta ted1$ -cells (i.e. the presence of EtN-P at the 6-hydroxy group of Man2) may not be as fatal as for the $\Delta cdc1$ mutant, as the 6-hydroxy group of Man2 is not buried in the binding pocket of the transglycosidase (Figure 42B). A slightly distorted coordination of Man2 is very likely, however it cannot be excluded that catalysis can still happen if the Man1-coordination is not substantially affected. This goes along with $\Delta ted1$ mutants that show a growth behavior in rich medium as observed for the wild type (Yoko-o et al., 2018). The latter study investigated the GPI-lipid remodeling mediated by Cwh43 from PI (phosphatidylinositol) to IPC (inositolphosphorylceramide), which is important for the final destination of GPI-APs. Interestingly, the EtN-Ps of Man1 and Man2 are required for ceramide-remodeling by Cwh43 (Benachour et al., 1999; Zhu et al., 2006). While the CWP Cwp2 possesses a PI in wild type cells, GPI-PMPs such as Gas1 harbor an IPC lipid. In $\Delta cwh43$ however, Gas1 with the PI-form lipid favored to be localized to the cell wall. Like observed for $\Delta ted1$, the $\Delta cwh43$ mutant possessed a doubling time like the wild type, while the double mutant $\Delta ted1/\Delta cwh43$ showed a reduced growth rate (Yoko-o et al., 2018). It is

not entirely clear, when and why Cwh43 acts on lipid remodeling. In yeast, two different models are proposed. A sequential model is suggested as the main pathway, where Cwh43 utilizes a PI containing diacylglycerol (pG2) after Gup1, while the divergent model suggests an alternative route utilizing lysoPI for the ceramide remodeling and thereby separating the reactions of Gip1 and Cwh43 (Ghugtyal et al., 2007; Umemura et al., 2007). A recent study suggests the absence of an alternative route in *A. fumigatus* (Li et al., 2018).

As a consequence, the fate of GPI-CWPs and GPI-PMPs is encoded during the maturation of the GPI-anchor in the secretory pathway, while *Dfg5*-proteins decode the made decision at the plasma membrane. Based on the structural model of the complex between the GPI-core glycan and *CtDfg5*, a sequence of events can be suggested in the ER of yeasts (and probably other fungi possessing two paralogs of the mammalian PGAP5, i.e. Cdc1 and Ted1), upon which the destination of GPI-APs is determined (Figure 43). As a start the remodeling of the lipid by Bst1, Per1, and Gup1 occurs in the first three steps of the sequential pathway directly after the transamidase added the GPI-AP to the GPI-precursor. The GPI-lipid remodeling (and not the GPI-glycan remodeling) is the crucial step for the concentration at ERESs, suggesting that this step is required for the presorting and concentration at the exit sites (Manzano-Lopez et al., 2015). The next step determines the destination. Future GPI-CWP must be recognized by Cdc1 at this point (upper route), which not only removes the EtN-P at Man1, but also prevents further lipid-modification by Cwh43, as this modification is required for ceramide-transfer (Vazquez et al., 2014; Zhu et al., 2006). Subsequently Ted1 removes the EtN-P from Man2, which results in the mature ER-form of the GPI-glycan and facilitates efficient COPII-vesicular traffic to the Golgi initiated by the p24-complex (Haass et al., 2007; Manzano-Lopez et al., 2015). Upon export to the plasma membrane, the glycan of such GPI-APs containing a PI-lipid are recognized by *Dfg5*-proteins and further transferred to the cell wall glycan. Note that further modifications such as the fourth and fifth glycan added during GPI-biosynthesis and remodeling in the Golgi, respectively, are omitted in this model for clarity. GPI-PMPs however are directly recognized by Cwh43 (lower route), which remodels the lipid from PI to IPC. As the preferred substrate of Cdc1 are GPI-anchors with a diacylglycerol lipid this step is omitted and the GPI-ceramide is directly handed over to Ted1 (Vazquez et al., 2014). Ted1 proceeds with the removal of the EtN-P from Man2 and thereby facilitates the efficient binding of the p24-complex for cargo export to the Golgi. This results in a secreted GPI-AP possessing an IPC and an EtN-P at Man1. This is the modification that is suggested to be incompatible with the efficient binding and the catalytic function of *Dfg5*-proteins, which finally leaves the GPI-PMP in the plasma membrane. As mammals do not have to

discriminate between GPI-PMPs and GPI-CWPs, this might also explain why they possess just one PGAP5-homolog for GPI-glycan processing, which ensures efficient ER to Golgi transport by removing EtN-P from Man2, while the EtN-P at Man1 is commonly found in those GPI-anchors (Fujita et al., 2009; Gonzalez et al., 2009).

It must be mentioned at this point that initial studies of the GPI-composition in *S. cerevisiae* proposed a structure totally lacking EtN-Ps moieties at the Man1 and Man2 positions using the GPI-anchors purified from crude membrane preparations standing in stark contrast to the proposed model, as the EtN-P at Man1 is an obligatory requirement for the glycan based discrimination by *Dfg5*-proteins (Fankhauser et al., 1993). Analyses in *A. fumigatus* used an approach including an HF-treatment that did not allow any statements concerning the EtN-P modifications as they were removed before analysis, while further studies in *S. cerevisiae* questioned the Fankhauser model by showing the presence of EtN-P at the α 1,4-linked mannose (i.e. Man1) (Canivenc-Gansel et al., 1998; Flury et al., 2000). To date it is clear that the addition of EtN-P to Man1-3 is essential for the synthesis and functionality of the GPI-anchor, suggesting a methodical problem in early fungal GPI-anchor studies due to harsh isolation conditions, however the EtN-P at Man3 linking the protein to the anchor remained untouched. In current literature reviewing the GPI-structure, no unambiguous model is proposed and recent attempts to explain the role of the GPI-anchor in intracellular trafficking indeed included the EtN-P, while claiming a lack of knowledge concerning the side-branches upon their cross-linking (Kinoshita and Fujita, 2016; Orlean and Menon, 2007; Pittet and Conzelmann, 2007).

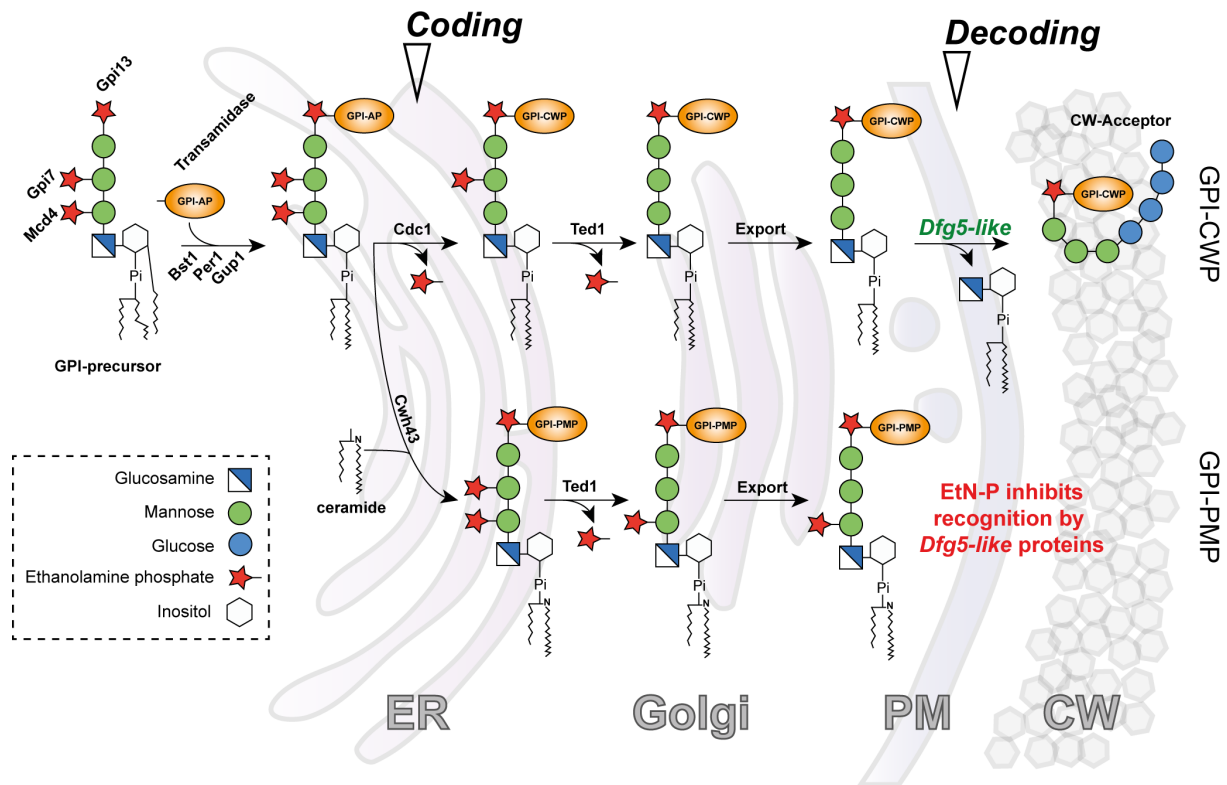


Figure 43: Presence of EtN-P at Man1 prevents GPI-based CW-incorporation by *Dfg5*-proteins. After GPI-precursor biosynthesis, the transamidase complex transfers the protein onto the EtN-P of Man3, upon which Bst1, Per1, and Gup1 remodel the GPI-lipids. Depending on the destination, either Cdc1 removes the EtN-P from Man1, or Cwh43 catalyzes the exchange from PI to IPC. Thereby, the EtN-P *code* for the final localization of the GPI-AP is written. Subsequently, Ted1 removes the EtN-P from Man2 to facilitate efficient ER-exit mediated by the p24 complex. Upon further processing in the Golgi, *Dfg5*-proteins are able to *decode* the presence or absence of an EtN-P within the GPI-core and thereby facilitating the transport of correctly processed GPI-CWPs into the CW, while GPI-PMPs remain bound to the membrane (adapted from Kinoshita and Fujita, 2016).

What remains is the question, how the discrimination between the Cdc1-route and the Cwh43-route works. Structural studies on the GPI-anchor and its interacting partners would be a way to shed light on this issue, but to my knowledge this thesis presents the first insights into experimentally supported GPI-glycan conformations, if only based on the assembly of its glycan-fragments. This is not really surprising, as the high degree of flexibility of glycosidic bonds suggest a highly dynamic behavior and simultaneously makes it hard to analyze it in solution (Vliegthart, 2011). However, a number of computational simulations exist, which try to suggest likely conformations and dynamic behaviors by searching for energetically favorable ensembles optima and highly occupied states. A tetrasaccharide consisting of Man α 1,2-Man α 1,6-Man α 1,4-GlcNAc- α OMe (simulating the GPI-core) was observed to adopt frequently a *C-like shaped* conformation with distances between C1 of GlcNAc (mimicking

GlcN) and C4 of Man3 (*r1'4-distance*) of about 12.7 Å (Figure 44A) (Wehle et al., 2012). Distances in other snapshots were further found to be as short as 5 Å up to 16 Å in a stretched conformation. Interestingly, the GPI-core as predicted to be bound to CtDfg5 adopts a conformation that is surprisingly similar to the described *C-like shape* with an *r1'4-distance* of ~11 Å (Figure 44B).

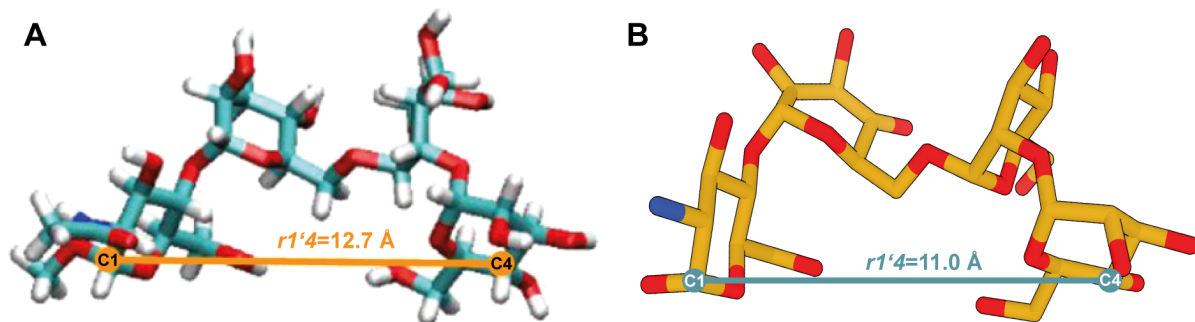


Figure 44: Comparison of simulated and fragment-derived GPI-shape. **A** *C-shaped* GPI-core state observed frequently in MD-simulations with an *r1'4-distance* of 12.7 Å (modified from Wehle et al., 2012). **B** The reassembled GPI-core from CtDfg5 is shown with an indicated *r1'4-distance* of 11.0 Å.

GPI-anchors are thought to exist in between two opposing states, a *lollipop* and a *flop-down* conformation, which have impact on the distance of the attached protein and its interaction with the bilayer as the *lollipop* stands straight up from the membrane while the *flop-down* remains in close contact with the lipid-head groups concerning all core-glycans (Lehto and Sharom, 2002). A recent study used the complete GPI-anchor inserted in a lipid bilayer with and without an attached GFP for their simulations (Banerjee et al., 2018). They describe the *lollipop* conformation as a rather unlikely configuration for the GPI-anchor alone and with the GFP attached to it, while their simulations clearly favor the *flop-down* state. However, due to its flexibility, additional modifications at the GPI-core itself and residues close to the C-terminal ω-site are good candidates to drive the GPI-anchor configuration in a way that favors certain shapes and thus the ability of proteins to bind more or less likely. As mentioned above, Cwh43 requires the presence of both charged EtN-Ps at Man1 and Man2, while the ER-exit by the p24-complex is highly enhanced upon removal of the Man2-EtN-P. While an explanation can be the direct steric hindrance of the modifications with the binding pockets of the respective proteins, another idea might be their influence on the GPI-shape. This may happen due to the negatively charged surface of the lipid bilayer and the positive amino group of the EtN-Ps. That positive charges indeed have an effect on the final localization has been shown for amino acids at the ω-minus region in *S. cerevisiae* and *A. fumigatus* (Frieman and

Cormack, 2003; Hamada et al., 1998, 1999; Ouyang et al., 2013). It was observed that basic amino acids at ω -1 or ω -2 lead to a residence in the plasma membrane, while tyrosine, valine, isoleucine, leucine or asparagine favor the attachment to the cell wall. The final destination could be modified by respective point mutations or chimeric proteins with exchanged ω -regions, thereby targeting plasma membrane proteins into the cell wall or *vice versa*. Hydrophobic residues may allow less stringent contact with the plasma membrane leading to a conformation that preferentially allows binding of the GPI-core glycan modifying enzyme Cdc1, while lysine at ω -1 or ω -2 helps to flatten the glycan on the lipid head groups, hiding it from Cdc1 and allowing the action of Cwh43, which anyway has to act on the lipid moiety, and not the glycan itself.

Clearly the GPI-AP/lipid interactions would not absolutely fix a certain conformation, but rather lead to a preferred state that still has intrinsic dynamics. Actually this kind of system would explain, why the analyses of GPI-PMPs and GPI-CWPs suggest a more relative than an absolute distribution, as several GPI-PMPs can be also found covalently attached to the cell wall.

5.4 Fungal cell wall biogenesis as a drug target

Secondary infections caused by fungal pathogens are an increasingly recognized threat for immunocompromised patients suffering for example from cancer, HIV or neutropenia. Ascomycota are commonly identified as the cause of such systemic fungal infections, among which *Candida* spp., *Aspergillus* spp., and *Cryptococcus* spp. belong to the most frequently found members (Enoch et al., 2006). Echinocandin-class drugs target the cell wall biosynthesis by inhibiting the β 1,3-glucan synthases. However, due to their good tolerance in patients, the high number of their usage led to an increased number of echinocandin-resistant strains, especially among *Candida* spp. (Perlin, 2015). Anyway, the cell wall of fungi seems to be a promising target for antimycotica, as its structures are absent in humans, thus implying a low degree of negative side effects. Recently, the development of potent inhibitors against the β 1,3-glycosyltransferases from the GH72 family has been published (Delso et al., 2018). The authors described an initially low-affinity laminaritrise-based binder, which was modified at its flexible linker at the reducing end to a final binding affinity in the low μ M-region (3.1 μ M for ScGas2, 1.5 μ M for AfGel4). However, the substances have not been tested *in vivo*. As the incorporation of proteins into the cell wall is an essential process for fungal life, the development of a potent Dfg5-inhibitor seems to be a promising approach. For

bacterial *endo*- α 1,6-mannanases, O- and S-linked isofagomine based inhibitors are known (ManIFG and ManSIFG), which bind to the -1/-2 subsites with affinities of about 1 μ M (Belz et al., 2017; Thompson et al., 2015b). The addition of a second mannoside to the non-hydrolyzable S-linked variant ((ManS)₂IFG) increased the affinity about 37-fold (Belz et al., 2017).

Compared to these molecules, the binding of FP-1 to *CtDfg5* ($K_D=2.71 \text{ mM} \pm 0.96$) is rather low, however a clear impact on the viability of *S. cerevisiae* could be observed, suggesting a good starting point for further drug development. An obvious step for optimizing the binding properties would be the rationalized binding optimization by the substitution of functional groups, which would contribute to an energetically favored ligand-protein interaction and thus lowered binding enthalpies. Another common approach during drug optimization is called *SAR by NMR* (i.e. structure-activity relationships obtained by nuclear magnetic resonance), where NMR-based screens identify two reasonable binders, which are then covalently linked and result in high affinity ligands (Shuker et al., 1996). For example, it was shown that the combination of a 17 mM and a 0.02 mM binder of stromelysin result in a potent inhibitor with a $K_D=15 \text{ nM}$ when linked together (Hajduk et al., 1997). This is achieved by a decrease of freedom in their free states and accordingly a lower entropy penalty upon binding, which results in better ΔG values. In fact, the simultaneous binding of FP-1 and mannose has been shown. The closest distance between FP-1 and the -2 subsite mannose is 3.3 Å, however when superimposed with the α 1,2-mannobiose state the actual distance is only 2.8 Å, which is quite close to the 2.77 Å needed for a thio ether linkage as used in the *endo*- α 1,6-mannanase inhibitors described above (Figure 45). The combination of the identified fragment together with a partial structure of the real substrate resulting in a ligand spanning the GPI-glycan binding site from subsite -3 to +1 is a promising candidate to inhibit the lipid-to-wall transfer already at low inhibitor concentrations.

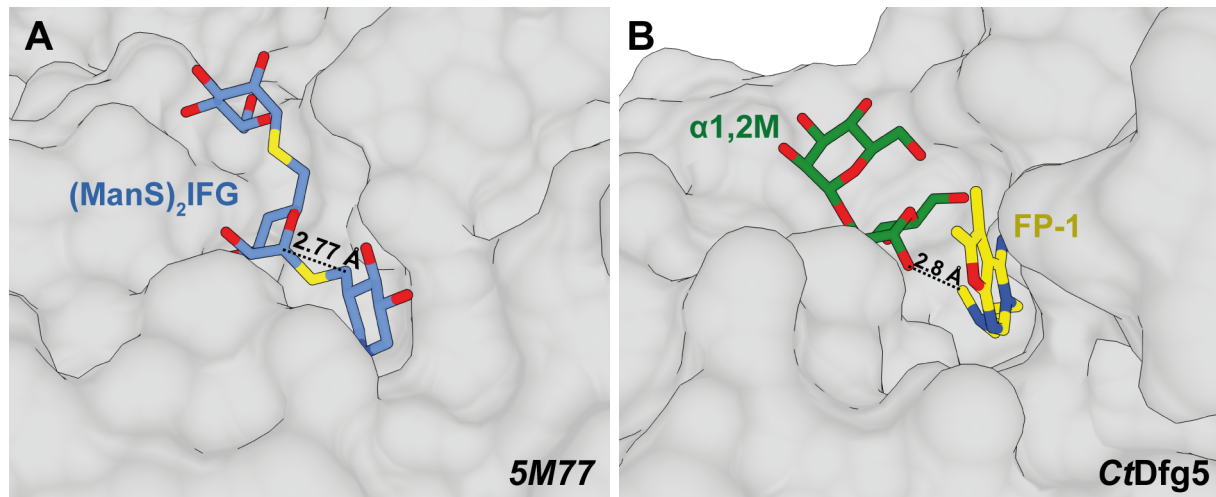


Figure 45: Comparison of the (ManS)₂IFG state of bacterial α 1,6-mannanases with a superposition of FP-1 and α 1,2-mannobiose states from CtDfg5. A The inhibitor is shown as blue sticks in the context of the protein surface. The distance between the adjacent carbons of the thio-ether linkage is indicated. The PDB code is given at the bottom right. **B** The CtDfg5-complex states of FP-1 (yellow sticks) and α 1,2-mannobiose (green sticks) are superimposed in the context of the protein surface and the resulting distance between the closest atoms is indicated. Surfaces are colored in grey.

Aside from the high potential as a specific antifungal drug, such a molecule can be an excellent tool to study the behavior of *Dfg5*-proteins *in vivo*. As discussed in the following, *Dfg5*-paralogs exhibit a number of individual preferences on their substrate. FP-1 may enable to shut off or reduce the action of *Dfg5*-single mutants to directly observe differences in the attachment of proteins onto the cell wall without changing the gene expression.

5.5 The functional diversity of *Dfg5*-paralogs

The SSN analysis not only revealed distinct clusters of bacterial and fungal homologs, but also provided insights into the diverse number of *Dfg5*-proteins within different organisms ranging from just two copies in *S. cerevisiae* up to seven paralogs in *Neurospora crassa* (see Table 2 on p.55). As only the deletion of all members is expected to cause a lethal phenotype in yeasts, the paralogs can complement the loss of the other protein at least to some extent, which implies a redundancy in the molecular function of *Dfg5*-proteins. This goes along with the proposed mechanism, as all *Dfg5*-proteins use the same GPI-structure as a substrate. Emphasizing this notion, the six *Dfg5*-members of *A. fumigatus* expressed under *in vitro* conditions were recently shown to incorporate the GPI-anchored galactomannan to β 1,3-glucan of the cell wall, expanding the range of substrates to lipid-anchored carbohydrates (Muszkieta et al., 2019). There it was necessary to delete all expressed *Dfg5*-paralogs to

abolish the presence of GM in the cell wall. Astonishingly, *AfDFG3* possessing the largest impact on cell morphology in *Aspergillus fumigatus*, could complement a temperature sensitive double mutant in *S. cerevisiae*, showing that although the GPI-GM is suggested as the substrate in the mold, it is able to transfer GPI-AP to the cell wall in the yeast, as the lipid-anchored galactomannan is a unique feature for *Aspergillus* spp. and the only plausible target in yeasts are GPI-APs (Muszkieta et al., 2019). However, unlike in yeasts, the function of *Dfg5*-proteins in *A. fumigatus* does not seem to be essential, which might be explained due to different cell wall architectures.

In yeasts more stringent culture conditions such as pH stress or nutritional starvation showed morphological differences between the two *Dfg5*-paralogs (Mösch and Fink, 1997; Spreghini et al., 2003). Both, *Candida albicans* and *S. cerevisiae*, were shown to have defects in their (pseudo)hyphae formation in the $\Delta dfg5$ background, but not in $\Delta dcw1$. A different effect was also observed regarding the adhesive ability of Flo11-expressing *S. cerevisiae* cells, where the $\Delta dfg5$ strain could be removed by a wash test indicating non-adhesive cells due to a compromised incorporation of Flo11 into the cell wall, while $\Delta dcw1$ cells are resistant to the wash test and stick to the solid medium (*personal communication with Gesa Schmitz, AG Mösch*). Furthermore, *N. crassa* exhibits a colonial phenotype with highly irregularly grown hyphae in the $\Delta dfg5$ background, while $\Delta dcw1$ has just a subtle phenotype (Maddi et al., 2012). Taken together the different paralogs possess specificity for certain targets, however the discrimination on the molecular level is still elusive, due to the apparent similarity of the used substrate. This is even true on an interspecies level as *Dfg5*-orthologs from filamentous fungi can complement in yeast (Muszkieta et al., 2019).

In this context, a sequence feature could be observed in the acceptor-binding loop (ABL1) described for the laminaribiose state. By looking at the overall conservation as it is displayed there (see Figure 30 on p.78), this loop region does not seem to have a certain function, as the level of preservations is *strikingly inconspicuous*. However, ABL1 was the only region of the protein showing an increased degree of flexibility. While this loop was missing in the electron density maps of the initial data used for structure determination, the loop became structured in the seeded screening upon lowering the overall pH. Surprisingly, this loop region was able to move up to 4 Å towards the active site when comparing different states, while the overall conformation of the protein remained unchanged (Figure 46).

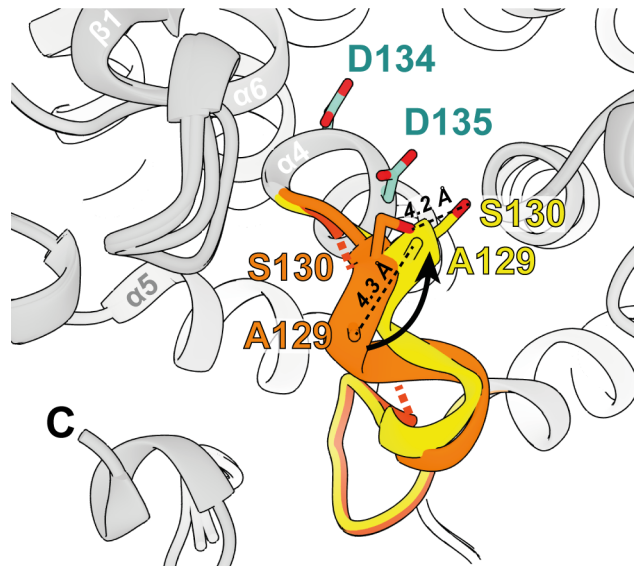


Figure 46: Dynamics within the ABL1-region. Three different ABL1-conformations were observed in the structures of CtDfg5, which are superimposed here. The first one lacks density for P125-S130 (colored in red, indicated by dotted line). The second conformation is shown in orange with density for the complete loop, and the third state colored in yellow is moved by approx. 4 Å indicated by the distances between the respective residues A129 and S130 and the arrow showing the *direction* towards the active site DD-motif. Secondary structure is indicated for orientation, residues are shown as sticks, oxygens are colored in red. The C-terminus is labeled.

When looking at the individual sequences of the ABL1 in the multiple sequence alignment, a surprising pattern can be observed, where several subpopulations emerge from the *Dfg5*-subfamily as shown in Figure 47 (SFigure 10 and SFigure 11 on pp.143-143 for the full ABL1-alignment). Intriguingly, some of these subpopulations exist only in filamentous fungi, others are exclusively found in Saccharomycetes, while being absent in the other subphylum. Surprisingly, it is possible to distinguish between the paralogs Dfg5 and Dcw1 in Saccharomycetes just by comparing their ABL1 motifs. All characterized and annotated Dfg5 homologs possess an invariable methionine together with valine or threonine (MV/T), while Dcw1 homologs usually possessed a TT-motif with few variations in the first threonine. Undeniably, the situation in Pezizomycotina is more complex, as the *Dfg5*-subfamily is more diverse. However, a pattern of conserved ABL1-motifs suggests a certain degree of substrate specialization within the different taxonomic classes, where some can be found in several classes (R/KT-motif), while others seem to be specific for a certain class (QW and N/SY for Eurotiomycetes, AS for Sordariomycetes). It is worth mentioning that by excluding apparent outliers of the shown ABL1-subpopulations, all protein sequences from Taphrinomycotina (e.g. the fission yeast *Schizosaccharomyces pombe*) disappeared from the final alignment. However, a similar pattern can be observed for these proteins as well, but the number of

sequences is rather limited (just 31 hits in the Uniprot database when searching *PF03663+Taphrinomycotina*) and therefore they are removed from this analysis.

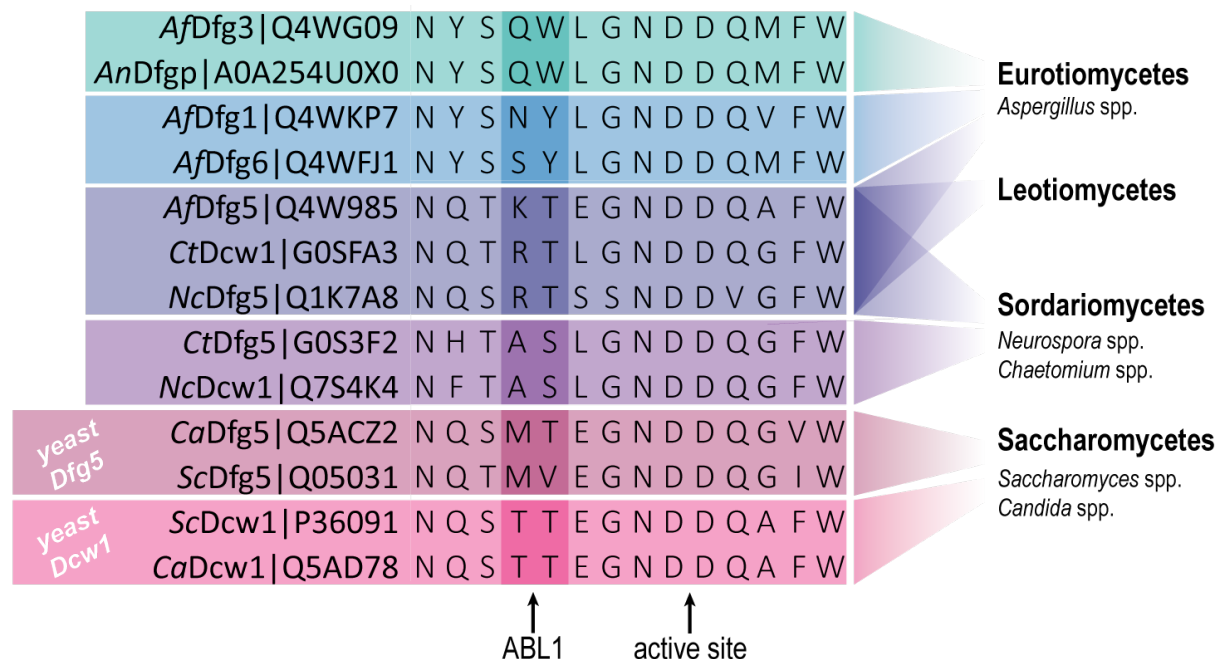


Figure 47: Multiple sequence alignment of *Dfg5*-proteins highlighting subpopulations in the ABL1-region. The ABL1 is highlighted by the boxes and the arrow, active site DD-residues are indicated by an arrow as well. The respective Uniprot-IDs and names according to the literature are given. The emerging subpopulations of the two homologs in yeast are indicated. *Af* *Aspergillus fumigatus*, *An* *Aspergillus niger*, *Ct* *Chaetomium thermophilum*, *Nc* *Neurospora crassa*, *Ca* *Candida albicans*, *Sc* *Saccharomyces cerevisiae*.

Since ABL1 is associated with acceptor binding (i.e. the cell wall polysaccharides), a correlation between the number of *Dfg5*-paralogs and the complexity of the cell wall could be assumed in order to optimize the lipid-to-wall transfer for certain proteins or in a special cell morphotype. However, whether other regions of *Dfg5*-proteins discriminate between different substrates cannot be concluded at this point. Anyway, as *AfDfg3* is able to complement the temperature-sensitive phenotype in yeast, the general function seems to be highly conserved throughout the *Dfg5*-subfamily, also supporting the notion that the principle type of the acceptor β -glucan is not depending on the protein, but on the supply at the plasma membrane (Muszkieta et al., 2019). It can also be speculated, whether the ABL1-loop may act like a flap for the efficient binding of the GPI-anchor, as this region clearly is in close contact to the membrane, if not even dipped into it. A dynamic, shovel-like behavior with different modifications at the shovel may contribute to a preferred or less favored binding of GPI-APs,

depending on their individual (most likely C-terminally influenced) shape of the GPI-core glycan.

5.6 The common feature of GH76-proteins

After describing the different aspects of the *Dfg5*-subfamily of GH76-proteins it becomes clear that this family separates from the already described bacterial homologs not only in the SSN analysis, but also on a functional level. Generally, different members of individual glycoside hydrolase families can have a broad spectrum of substrates (Naumoff, 2011). Apart *Dfg5*-proteins, another subfamily of GH76-proteins has been studied on a structural level in the scope of this thesis. This cluster found in the SSN is comprised of proteins from bacteria and fungi, therefore termed as the *Fungal/Bacteria mixed* subfamily. Only a fungal members have been assessed, however a further characterization concerning their cellular function is missing, as no phenotype could yet be observed (Maddi et al., 2012; Muszkieta et al., 2019). From the SSN analysis, this class is apparently closer related to the bacterial homologs, suggesting also a similar function. However, an α 1,6-mannanase activity could not be observed *in vitro* for *CtGH76* (Markus Friedrich, Master thesis). He also obtained the crystal structure of the apo-state, which showed differences in the two *wing* domains described above, while the overall $(\alpha/\alpha)_6$ -helical barrel fold is conserved (see Figure 36 on p.86).

To extend the functional knowledge on GH76-proteins, a similar glycan-fragment screening approach has been applied to the apo-crystals of *CtGH76* as done before for *CtDfg5*. The crystal structure in complex with an α 1,3- α 1,6-mannotriose showed that the extended wing region is involved in carbohydrate binding, however unlike for *Dfg5*-proteins no glucosamine could be observed at subsite +1 upon molar soaking. A striking difference between *BcGH76* and *CtDfg5* is the coordination of α 1,6-linked mannobiose. The bacterial protein utilizes α 1,6-oligomers as substrate, which enables it to bind the biose to its -3/-2 subsites (Figure 48B). A coordination at subsite -1 could only be achieved by a combination of a substrate-binding canyon spanning substrate and an active-site variant, as the protein provides a better binding site for the disaccharide at -3/-2, while at subsite -1 the sugar moiety is supposed to be distorted for catalysis (Thompson et al., 2015b). This is a further indication that the substrates differ between *bacterial* and *Dfg5*-subfamilies, as seen by the α 1,6-mannobiose state in *CtDfg5* (Figure 48A). Interestingly, the approach applied to *CtGH76* revealed the same occupied subsites as in *CtDfg5*, suggesting that linear α 1,6-mannooligomers are not the prime candidate for a substrate of the *Fungal/Bacteria mixed* subfamily (Figure 48C). This explains

the findings in Markus Friedrich's master thesis that *CtGH76* is incapable to hydrolyze α 1,6-mannooligomers.

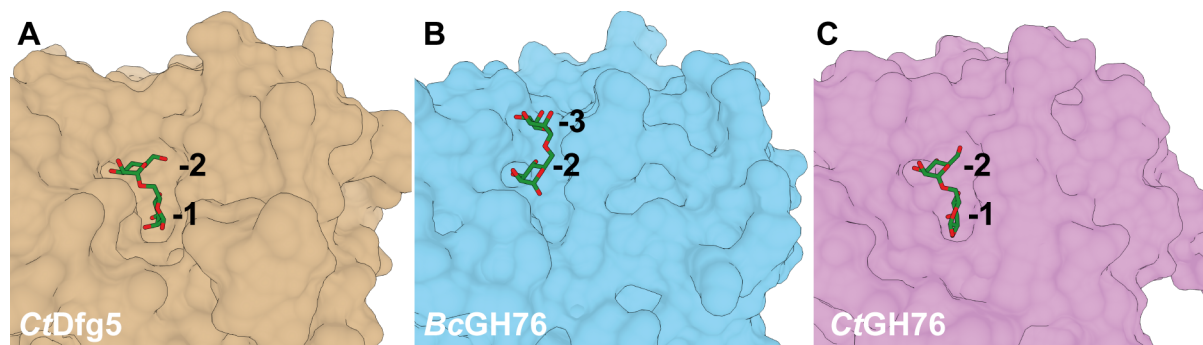


Figure 48: Known α 1,6-mannobiose states of GH76-proteins. GH76 family members from three different subfamilies are shown in complex with α 1,6-mannobiose with the respective protein surfaces and the ligand shown as green sticks with red oxygens bound to either subsites -2 to -1 in the fungal proteins (A+C) or -3 to -2 in the bacterial structure (B).

Due to the lack of *in vivo* work on this type of GH76-proteins it is currently hard to predict their function. The complex structures combined do not assemble a plausible glycan candidate on which *CtGH76* may act, although the α 1,3- α 1,6-Mannotriose exists within the high mannose N-glycan core structure known from yeast mannoproteins.

The α 1,6-mannobiose state helps to understand the common feature of that protein family. By comparing the complex states of *CtDfg5*, *CtGH76* and *BcGH76*, which occupy the -2/-1 subsites, the similar binding mode of that part of the binding site clearly stands out (Figure 49). It can therefore be hypothesized that GH76-proteins act on substrates that contain an α 1,6-mannobiose core unit, which fits to the -2/-1 subsites, while coordinating the neighboring glycan residues in a subfamily-specific manner.

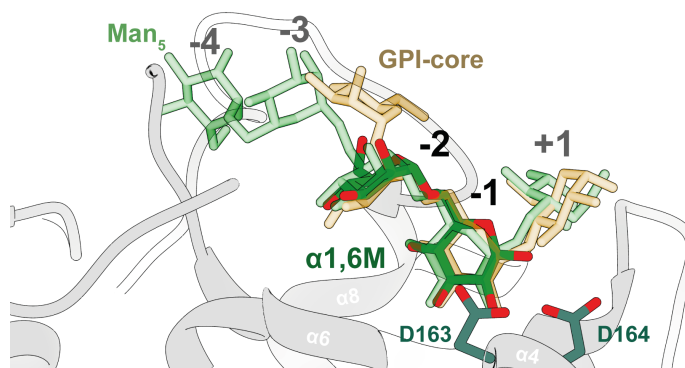


Figure 49: Comparison of substrate states from GH76-proteins in the context of *CtGH76*. A superposition of the mannopentose (Man_5) bound state of *BcGH76* (PDB 5AGD), the assembled GPI-state of *CtDfg5* and the α 1,6-mannobiose (α 1,6M) state of *CtGH76* is shown in the context of *CtGH76*. The active site residues (D163/D164) of *CtGH76* are shown as sticks for orientation to assign the substrate binding sites. The ligands are shown as sticks, Man_5 is colored in transparent

green, GPI-core in transparent orange and α 1,6M is shown as solid sticks to highlight the conserved substrate element of GH76-proteins.

5.7 Outlook

Within the scope of this work the role of *Dfg5*-proteins was analyzed and clarified by in depth structural analysis of a heterologously produced thermophilic homolog from *Chaetomium thermophilum* in terms of its underlying mechanism to transfer GPI-anchored substrates from the plasma membrane into the fungal cell wall. First experimentally derived insights into the spatial arrangement of the GPI-anchor core are provided. Furthermore, the suggested great potential as a drug target was pointed out by identifying a small molecule bound to the active site of the protein, which was shown to inhibit the growth of *Saccharomyces cerevisiae*. Finally, the common feature of GH76-proteins was suggested by combining the known insights of substrate coordination with a further member of another GH76-subfamily showing a conserved α 1,6-mannobiose coordination at the -2/-1 subsites.

These findings only opened the gate for further studies. Still a robust method for an *in vitro* assay showing the hydrolysis of the GPI-core and the transglycosylation onto the acceptor remains to be established. A major problem for a successful transglycosylation might be the absence of the membrane, where the transfer is supposed to happen. From a structural point of view, there is no reason why *Dfg5*-proteins should not act like their bacterial homologs. A functional assay may require a membrane-based system including embedded glucan synthases, correctly processed GPI-anchored substrates, membrane associated *Dfg5*-proteins in an aqueous solution containing native concentrations of calcium. An *in vivo* system with suitable reporters (such as described for *Aspergillus fumigatus* in Donat et al., 2012) and a sophisticated shutoff system in combination with a potent inhibitor such as an improved FP-1 might work as well. Such a system may also enable the analysis of the specificities of the two yeast-paralogs on different targets. Combined with MS-based cell wall compositional analyses it may show which GPI-APs are preferentially transferred by the respective paralogs. A further aspect in this context is the discrepancy of paralog numbers in individual organisms, which range from two in yeasts to nine in *Neurospora crassa*. This number may depend on different developmental stages with variable acceptors in the cell wall, and an adaptation to specific donor substrates.

The sorting of GPI-APs along the secretory pathway and the decision at the plasma membrane for the final destination by decoding the EtN-P modifications remains to be validated as well.

While it is clear that the cleavage of the GPI-core glycan by *Dfg5*-proteins with the Man1-EtN-P cannot occur, molecular dynamic (MD-) simulations may answer the question if binding in principle can happen with such modifications, or not. A full molecular understanding of the processing factors within the secretory pathway that are responsible for the coding (i.e. Cdc1, Cwh43, Ted1) still lacks behind. In order to understand the precise sequence of the GPI-sorting, this aspect must be addressed as well.

The initial hit of FP-1 by structure-based fragment screening shows principal biological activity against yeast and is a good starting point for further drug development. However, it requires optimization due to the low binding properties. The simultaneous binding of FP-1 and mannose has been shown experimentally and the combination of two fragments is discussed above. Also a combination with α 1,2-mannobiose seems attractive. This might be a promising approach to combine these two fragments in order to obtain an optimal inhibitor for *Dfg5*-proteins. Such inhibitors may be able to compete with native substrates already at low concentrations.

Finally, several subfamilies of the GH76-class remain to be characterized for a full understanding of these glycoside hydrolases. A start for the *Fungal/Bacteria mixed* class is already provided and a very likely common feature for GH76 proteins is discussed, however the actual function needs to be explained, also in terms of the function in archaea, where a few organisms contain such proteins.

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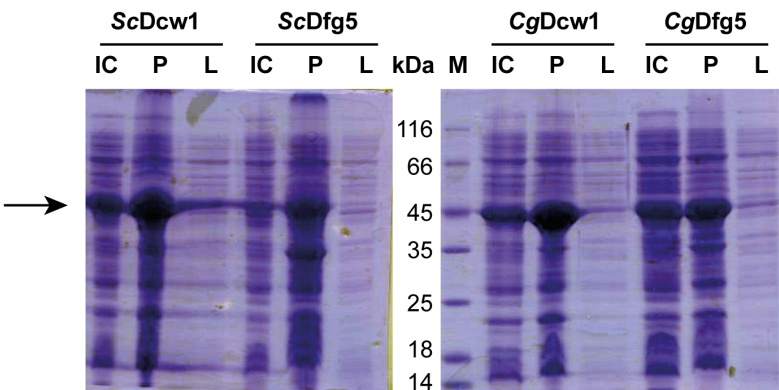
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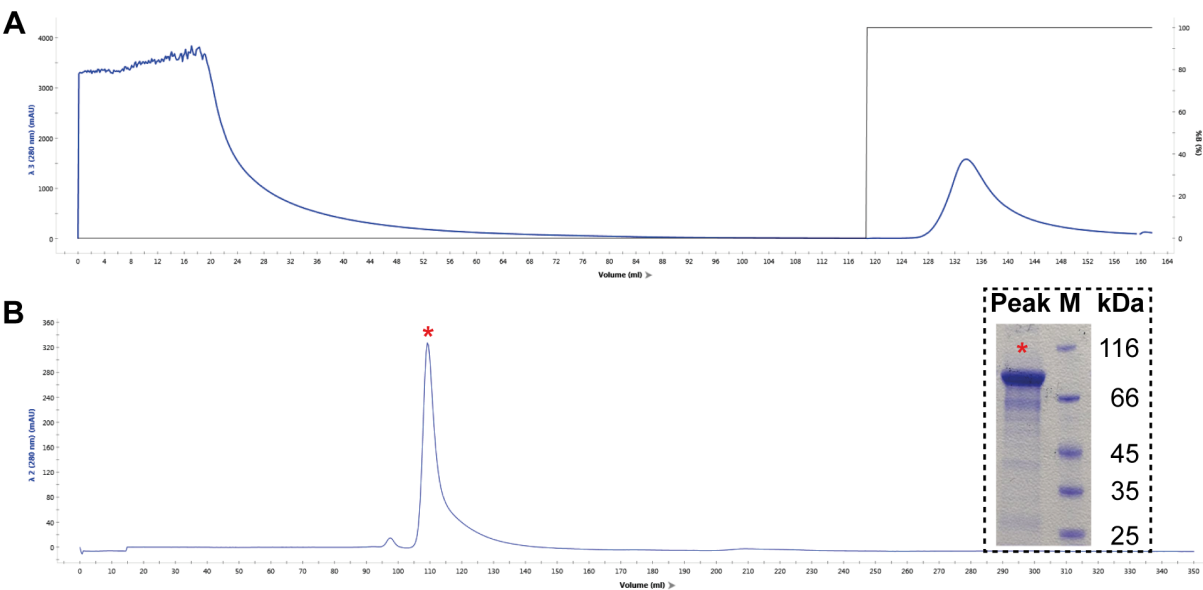
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7 Supplement

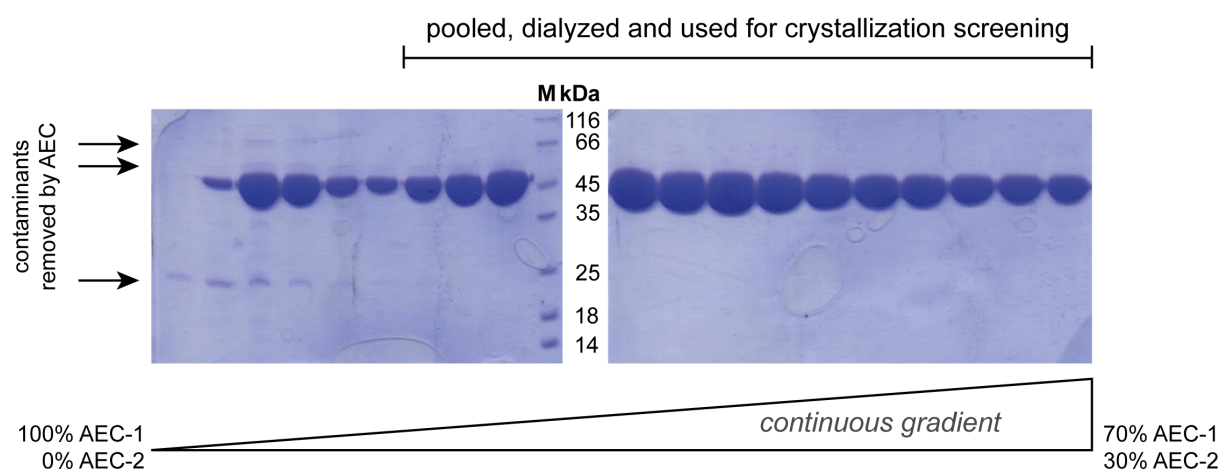
7.1 Figures



SFigure 1: Test expressions of Dfg5-homologs from mesophilic fungi. Heterologous production of Dfg5-orthologs from *Saccharomyces cerevisiae* (Sc) and *Candida glabrata* (Cg) were overexpressed, but only found insoluble. The arrow indicates the apparent size of over produced proteins, IC=induced cells, P=pellet, L=Load (for affinity chromatography, i.e. the supernatant of the lysis upon centrifugation), M=Molecular weight marker. The marker is labeled with the respective molecular weights in kDa.



SFigure 2: Purification of MBP-ScDcw1. **A** The amylose affinity purification with a single step gradient is shown. $A_{280\text{ nm}}$ is shown as a blue line and the percentage of MBP-2 buffer („B“) is shown as a black line. **B** The SEC run of MBP-ScDcw1 is shown. The peak fraction of the 87.3 kDa fusion protein is loaded on SDS-PAGE as indicated by the red asterisk. M=molecular weight marker



SFigure 3: Anion exchange chromatography of CtDfg5 for first crystal screen. *M*=Molecular weight marker.

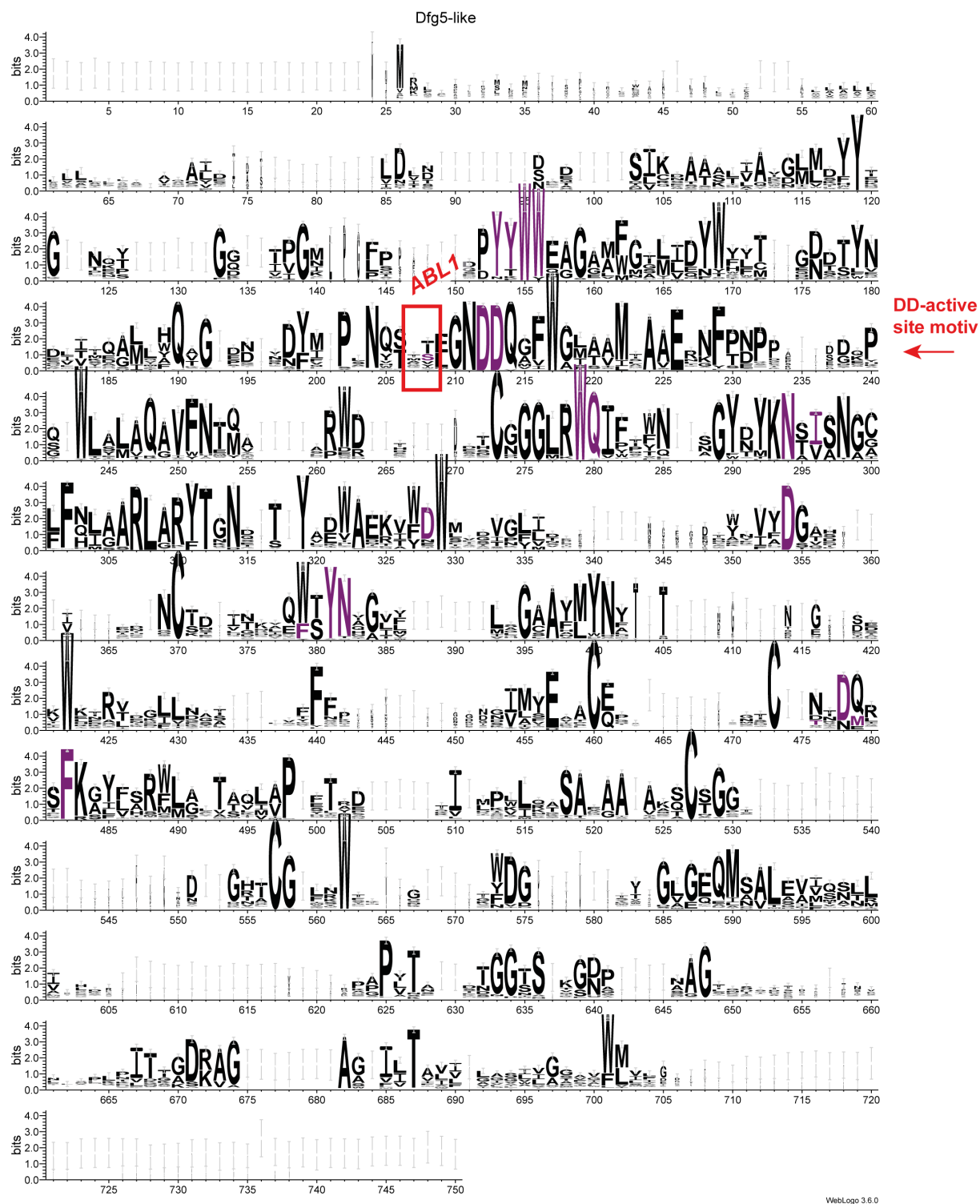
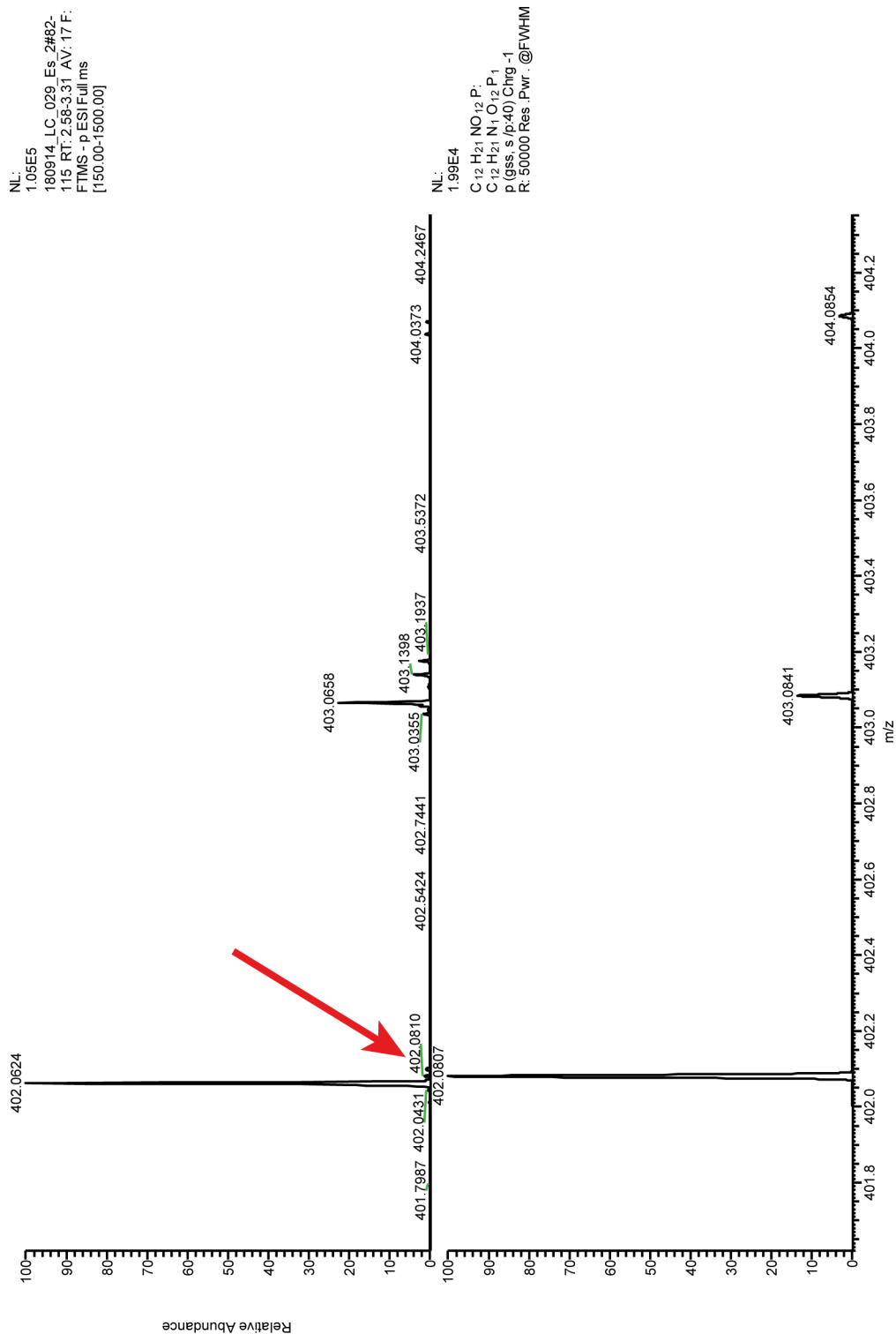


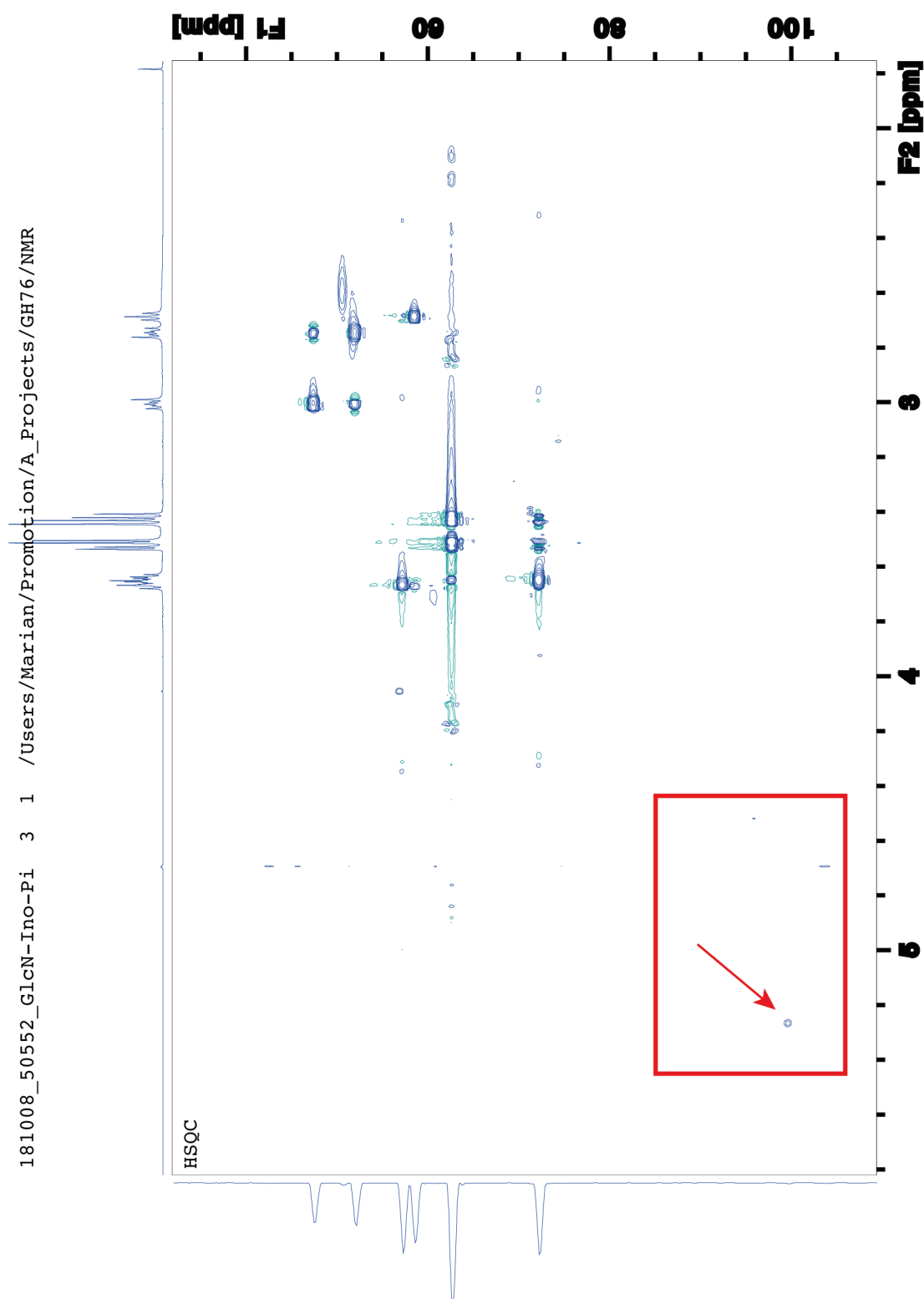
Figure 4: WEBLOGO of the *Dfg5*-subfamily calculated from the MSA that was used for the *ConSurf* analysis. The conserved residues are highlighted, as well as the ABL1 region. The full MSA comprising 250 *Dfg5*-sequences is only part of the electronic resource due to its length.

Experimental spectrum

Reference spectrum



SFigure 5: Identification of target mass calculated for GlcN-Ino-Pi (402.08 Da). MS analysis was performed by the MS-facility led by Dr. Uwe Linne.



SFigure 6: Identification of anomeric carbon by 2D- ^1H - ^{13}C NMR. The region marked in red indicates the typical chemical shift region for anomeric carbons in carbohydrates. A singlet peak corresponding to anomeric carbons was observed for the analyzed sample. NMR analysis was performed by Dr. Xiulan Xie.

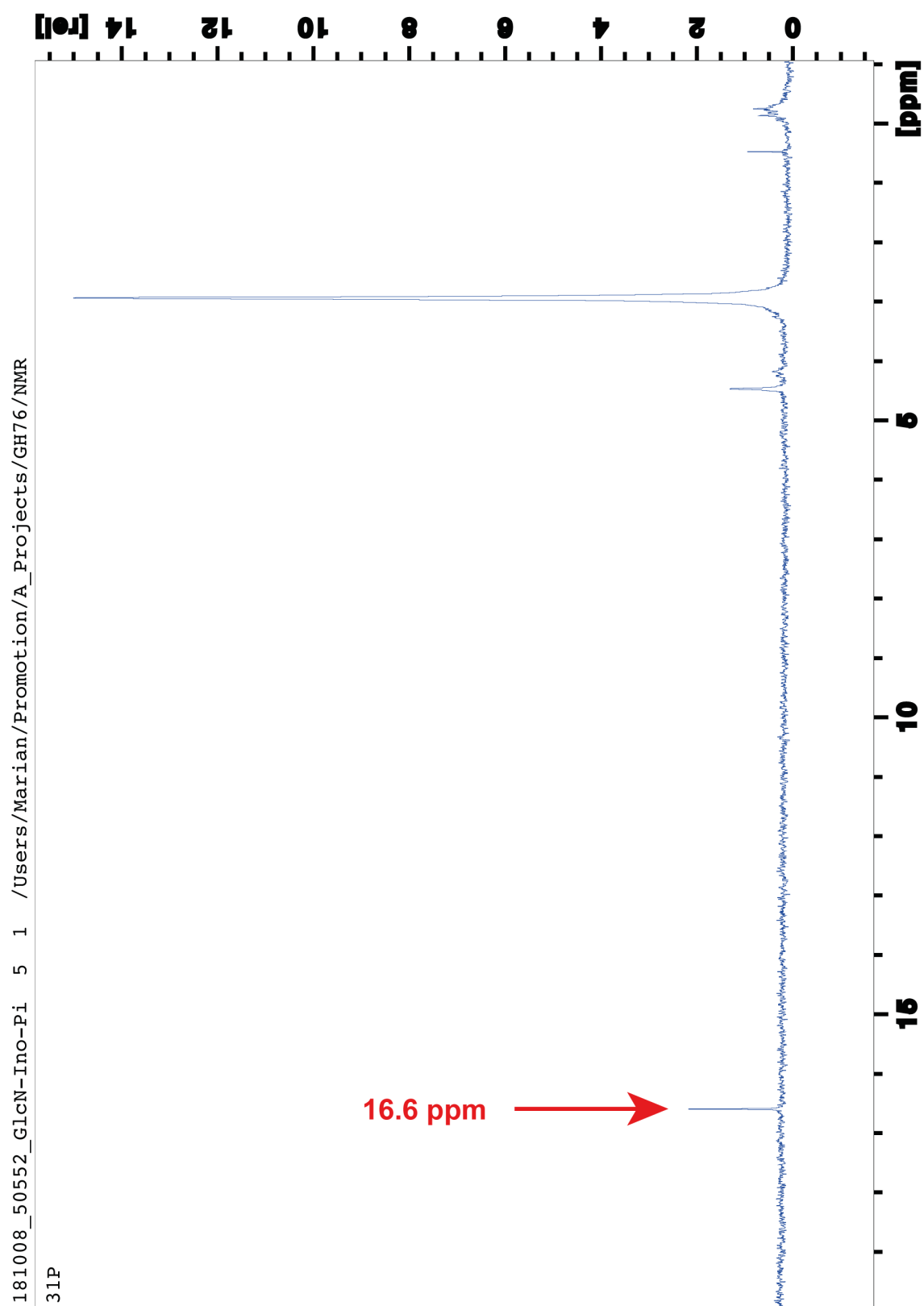


Figure 7: Identification of specific chemical shift of cyclic phosphate by ^{31}P NMR around 16 ppm in form of a singlet. NMR analysis was performed by Dr. Xiulan Xie.

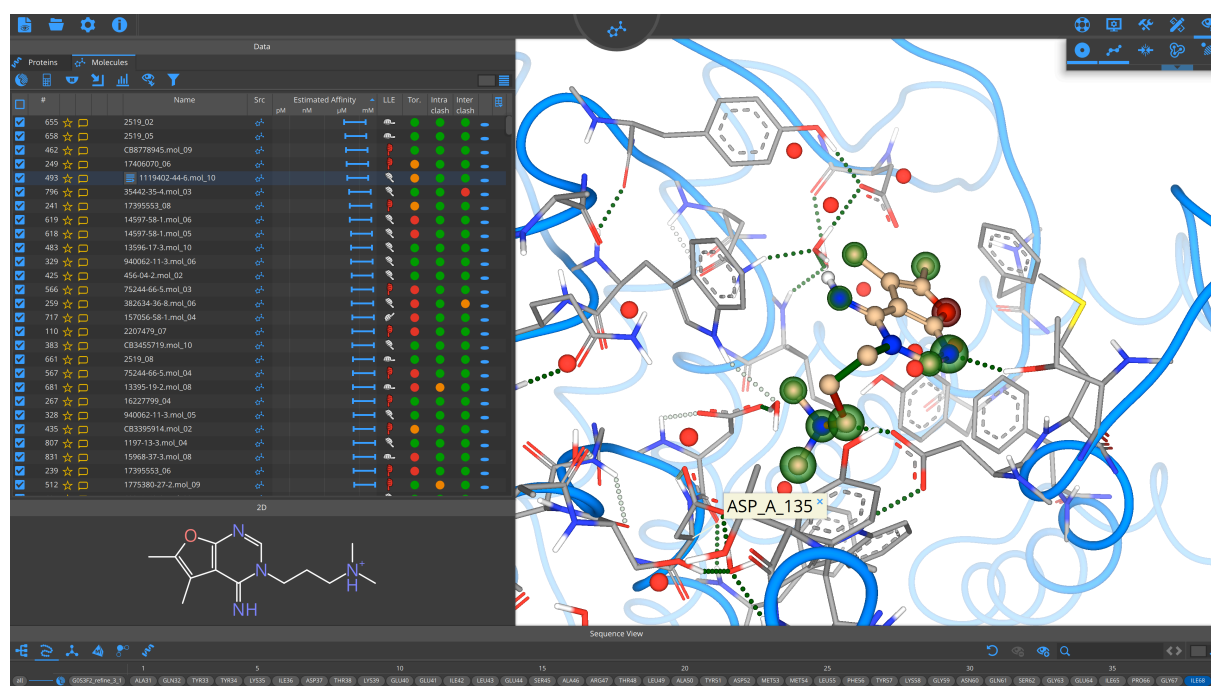
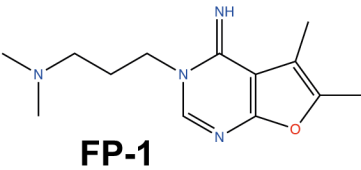
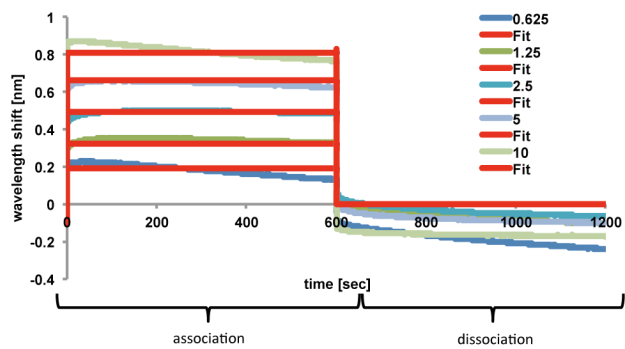


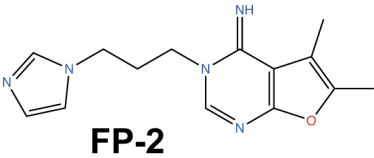
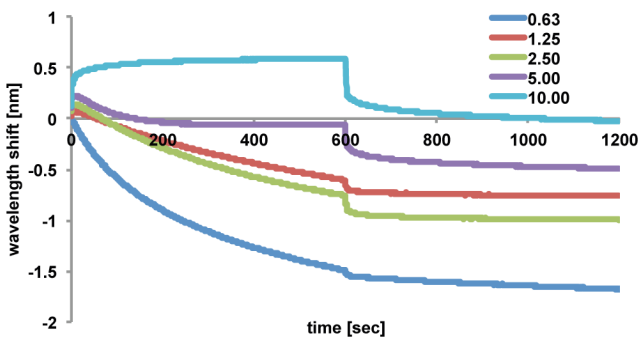
Figure 8: Docked pose of FP-1 generated by SeeSAR. The conformation from molecular docking is inverted to the real conformation found in the crystal structure.

FP-1 + CtDfg5



Loading Sample	Loading Conc. (µg/ml)	c(F51) (mM)	KD (M)	KD Error	ka (1/Ms)	ka Error	kdis (1/s)	kdis Error	Full R^2
Dfg5	49.5	0.625	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331
Dfg5	49.5	1.25	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331
Dfg5	49.5	2.5	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331
Dfg5	49.5	5	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331
Dfg5	49.5	10	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331

FP-2 + CtDfg5



Loading Sample	Loading Conc. (µg/ml)	c(F51a) (mM)	KD (M)	KD Error	ka (1/Ms)	ka Error	kdis (1/s)	kdis Error	Full R^2
Dfg5	49.5	0.625	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
Dfg5	49.5	1.25	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
Dfg5	49.5	2.5	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
Dfg5	49.5	5	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.
Dfg5	49.5	10	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

Summary Biolayer Inferometry

		cFrag (mM)	KD (M)	KD Error	ka (1/Ms)	ka Error	kdis (1/s)	kdis Error	Full R^2
Dfg5	FP-1	0.625	2.71E-03	9.56E-04	1.08E+04	2.70E+03	2.94E+01	7.32E+00	0.9331
	FP-2	10	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.	n.d.

Figure 9: Binding analyses of FP-1 and FP-2 with CtDfg5 using biolayer inferometry. The analysis was performed together with Dr. Sven-Andreas Freibert, who evaluated the data.

tr Q58GD7 Q58GD7_EMENI/112-125	NY\$QWLGNDDQMFV	tr G9MQD1 G9MQD1_HYPVG/113-126	NH\$ASLGNDDQGFV
tr B8NX26 B8NX26_ASPFN/112-125	NY\$QWLGNDDQMFV	tr I1RL09 I1RL09_GIBZE/120-133	NH\$ASLGNDDQGFV
tr A0A100I5A6 A0A100I5A6_ASPNG/108-121	NY\$QWLGNDDQMFV	tr C7Z068 C7Z068_NEC7/116-129	NH\$ASLGNDDQGFV
tr A0A254U0X0 A0A254U0X0_ASPNG/108-121	NY\$QWLGNDDQMFV	tr A0A2N6NY19 A0A2N6NY19_BEABA/113-126	NY\$ASLGNDDQGFV
tr B8NPZ1 B8NPZ1_ASPFN/111-124	NV\$DWLGNDDQMFV	tr J3NHD5 J3NHD5_GAGT3/113-126	NY\$ASLGNDDQGFV
tr Q2TWC1 Q2TWC1_ASPOR/111-124	NY\$QWLGNDDQMFV	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6/113-126	NY\$ASLGNDDQGFV
tr A1CD27 A1CD27_ASPCL/110-123	NY\$QWLGNDDQMFV	tr F7VZ72 F7VZ72_SORMK/113-126	NV\$ASLGNDDQGFV
tr Q4WG09 Q4WG09_ASPFU/111-124	NY\$QWLGNDDQMFV	tr Q96TX1 Q96TX1_NEUCS/113-126	NV\$ASLGNDDQGFV
tr A0A0J5PNX6 A0A0J5PNX6_ASPFM/111-124	NY\$QWLGNDDQMFV	tr Q1K7I4 Q1K7I4_NEUCR/113-126	NV\$ASLGNDDQGFV
tr A1DJ54 A1DJ54_NEOFI/111-124	NY\$QWLGNDDQMFV	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS/113-126	NV\$ASLGNDDQGFV
tr A0A254TXZ7 A0A254TXZ7_ASPNG/108-121	NV\$SYLGNDDQMAV	tr F8MXT4 F8MXT4_NEUT8/113-126	NV\$ASLGNDDQGFV
tr A0A124BYC5 A0A124BYC5_ASPNG/108-121	NV\$SYLGNDDQVAV	tr W3X8E3 W3X8E3_PESFW/113-126	NV\$ASLGNDDQGFV
tr B8MHG0 B8MHG0_TALSN/109-122	NY\$SYLGNDDQEFV	tr G0SE99 G0SE99_CHATD/117-130	NV\$ASLGNDDQGFV
tr Q5ATF9 Q5ATF9_EMENI/103-116	NY\$SYLGYDDQFFV	tr B2B747 B2B747_PODAN/114-127	NV\$ASLGNDDQGFV
tr A1CCM5 A1CCM5_ASPCL/111-124	NY\$SYLGNDDQMFV	tr G2Q8A7 G2Q8A7_MYCTT/111-124	NY\$ASLGNDDQAFV
tr A1DJ50 A1DJ50_NEOFI/111-124	NY\$SYLGNDDQMFV	tr G2QT05 G2QT05_THITE/111-124	NV\$ASLGNDDQGFV
tr A0A0J5PQ80 A0A0J5PQ80_ASPFM/136-149	NY\$SYLGNDDQMFV	tr E9DYA6 E9DYA6_METAQ/113-126	NV\$ASLGNDDQGFV
tr Q2TYU3 Q2TYU3_ASPOR/109-122	NY\$SYLGNDDQGFV	tr E9E4X8 E9E4X8_METAQ/131-144	NV\$ASLGNDDQGFV
tr Q0CB86 Q0CB86_ASPTN/117-130	NY\$SYLGNDDQGFV	tr Q4WFX5 Q4WFX5_ASPFU/117-130	NQ\$RTSLGNDDQAFV
tr A0A124BXU6 A0A124BXU6_ASPNG/110-123	NY\$SYLGNDDQMFV	tr A0A0J55N29 A0A0J55N29_ASPFM/117-130	NQ\$RTSLGNDDQAFV
tr A0A254U2J9 A0A254U2J9_ASPNG/110-123	NY\$SYLGNDDQMFV	tr A0A254UET1 A0A254UET1_ASPNG/116-129	NQ\$RTSLGNDDQAFV
tr Q2U557 Q2U557_ASPOR/111-124	NV\$SYLGNDDQMFV	tr A0A124BV85 A0A124BV85_ASPNG/115-128	NQ\$RTSLGNDDQAFV
tr A0A124BXU6 A0A124BXU6_ASPNG/107-120	NV\$SYLGNDDQMFV	tr B2W762 B2W762_PYRTR/110-123	NQ\$KTLGNDDQAFV
tr A2R8R5 A2R8R5_ASPNC/107-120	NV\$SYLGNDDQMFV	tr A1CMM3 A1CMM3_ASPCL/97-110	NQ\$RTSLGNDDQAFV
tr A0A254U3K7 A0A254U3K7_ASPNG/107-120	NV\$SYLGNDDQMFV	tr D4ATT7 D4ATT7_ARTBC/115-128	NQ\$RTSLGNDDQAFV
tr B6H7I2 B6H7I2_PENRW/111-124	NY\$SYLGNDDQMFV	tr C1H190 C1H190_PARBA/121-134	NQ\$RTSLGNDDQAFV
tr B6GZT8 B6GZT8_PENRW/111-124	NY\$SYLGNDDQMFV	tr K1XLH0 K1XLH0_MARBU/108-121	NQ\$RTSLGNDDQAFV
tr A1CSC2 A1CSC2_ASPCL/110-123	NY\$NYLGNDDQVFW	tr A0A0A2VM24 A0A0A2VM24_PARBA/120-133	NQ\$KTLGNDDQAFV
tr Q4WKP7 Q4WKP7_ASPFU/109-122	NY\$NYLGNDDQVFW	tr A0A1S9RH77 A0A1S9RH77_9EURO/116-129	NQ\$KTLGNDDQAFV
tr A0A0J5PM92 A0A0J5PM92_ASPFM/109-122	NY\$NYLGNDDQVFW	tr W6QCW3 W6QCW3_PENRF/116-129	NQ\$KTLGNDDQVFW
tr A1D587 A1D587_NEOFI/109-122	NY\$NYLGNDDQVFW	tr B6GZU2 B6GZU2_PENRW/119-132	NQ\$KTLGNDDQVFW
tr B8MYP3 B8MYP3_ASPFN/106-119	NY\$NYLGNDDQVFW	tr A1CVB0 A1CVB0_NEOFI/105-118	NQ\$KTLGNDDQAFV
tr Q2UR85 Q2UR85_ASPOR/106-119	ND\$NYLGNDDQVFW	tr Q4W985 Q4W985_ASPFU/105-118	NQ\$KTLGNDDQAFV
tr A0A117DW50 A0A117DW50_ASPNG/109-122	ND\$NYLGNDDQVFW	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM/105-118	NQ\$KTLGNDDQAFV
tr A0A254U5V1 A0A254U5V1_ASPNG/109-122	NW\$ASLGNDDQGFV	tr E2PT42 E2PT42_ASPNC/106-119	NQ\$RTSLGNDDQSFV
tr F7VVT4 F7VVT4_SORMK/125-138	NW\$ASLGNDDQGFV	tr A0A254TKQ9 A0A254TKQ9_ASPNG/106-119	NQ\$RTSLGNDDQSFV
tr G2QKJ0 G2QKJ0_MYCTT/119-132	NH\$ASLGNDDQGFV	tr A0A100ISD9 A0A100ISD9_ASPNG/106-119	NQ\$RTSLGNDDQSFV
tr J3NZQ6 J3NZQ6_GAGT3/124-137	AH\$ASLGNDDQGFV	tr Q0CG55 Q0CG55_ASPTN/105-118	NQ\$RTSLGNDDQAFV
tr J3P147 J3P147_GAGT3/136-149	KH\$ASLGNDDQGFV	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI/115-128	NQ\$KTLGNDDQGMV
tr J3PGN0 J3PGN0_GAGT3/120-133	KH\$ASLGNDDQGFV	tr A0A177AJ74 A0A177AJ74_9PEZI/115-128	NQ\$KTLGNDDQGFV
tr A0A0C4E991 A0A0C4E991_MAGP6/120-133	SK\$ASLGNDDQGFV	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI/111-124	NQ\$KTLGNDDQGFV
tr G4N3E1 G4N3E1_MAGO7/119-132	NF\$ASLGNDDQGFV	tr K1WJG5 K1WJG5_MARBU/111-124	NQ\$KTLGNDDQGFV
tr Q7S4K4 Q7S4K4_NEUCR/124-137	NF\$ASLGNDDQGFV	tr A7F3P0 A7F3P0_SCL51/113-126	NQ\$KTLGNDDQGFV
tr A0A0B0DFT3 A0A0B0DFT3_NEUCS/124-137	NF\$ASLGNDDQGFV	tr A7EIG7 A7EIG7_SCL51/111-124	NQ\$KTLGNDDQGFV
tr F8MRU1 F8MRU1_NEUT8/124-137	NY\$ASLGNDDQGFV	tr A0A1C1D213 A0A1C1D213_9EURO/113-126	NQ\$KTLGNDDQAFV
tr F7VVP8 F7VVP8_SORMK/124-137	NH\$ASLGNDDQGFV	tr F7VWZ9 F7VWZ9_SORMK/108-121	NQ\$RTSLNDDVGFV
tr G0S3F2 G0S3F2_CHATD/126-139	NH\$ASLGNDDQGFV	tr Q1K7A8 Q1K7A8_NEUCR/108-121	NQ\$RTSLNDDVGFV
tr B2AEF2 B2AEF2_PODAN/125-138	NH\$ASLGNDDQGFV	tr Q9C2J1 Q9C2J1_NEUCS/108-121	NQ\$RTSLNDDVGFV
tr G2QF51 G2QF51_MYCTT/125-138	NH\$ASLGNDDQGFV	tr F8MBP4 F8MBP4_NEUT8/108-121	NQ\$RTSLNDDVGFV
tr G2XFH9 G2XFH9_VERDV/119-132	NH\$ASLGNDDQGFV	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI/109-122	NQ\$RTSLGNDDQGFV
tr G3JGZ5 G3JGZ5_CORMM/118-131	NH\$ASLGNDDQGFV		

SFigure 10: Multiple sequence alignment of ABL2-region

tr W3WN68 W3WN68_PESFW/108-121	NQTRSLGNDDQGF	tr A3LN37 A3LN37_PICST/107-120	NQSTTBGNDDQGF
tr W3XAF6 W3XAF6_PESFW/107-120	NQTRSLGNDDQGF	tr G8BF46 G8BF46_CANPC/108-121	NQSTTBGNDDQGF
tr G2QB99 G2QB99_MYCTT/107-120	NQTRTLGNDDQGF	tr H8WVX8 H8WVX8_CANO9/108-121	NQSTTBGNDDQGF
tr G05FA3 G05FA3_CHATD/108-121	NQTRTLGNDDQGF	tr G3AMT4 G3AMT4_SPAPN/108-121	NQSTTBGNDDQGF
tr G2XGF6 G2XGF6_VERDV/107-120	NQTRTLGNDDQGF	tr A5DUW0 A5DUW0_LODEL/108-121	NQSTTBGNDDQGF
tr G4NGV8 G4NGV8_MAGO7/110-123	NQTRTLGNDDQGF	tr C5M5U7 C5M5U7_CANTT/110-123	NQSTTBGNDDQGF
tr A0A136J7D3 A0A136J7D3_9PEZI/107-120	NQTRTLGNDDQGF	sp Q5AD78 DCW1_CANAL/106-119	NQSTTBGNDDQGF
tr W3X554 W3X554_PESFW/108-121	NQTRSLGNDDQGF	tr B9WAI2 B9WAI2_CANDC/106-119	NQSTTBGNDDQGF
tr Q75CW7 Q75CW7_ASHGO/103-116	HQHMLVGNDDQGF	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO/106-119	NQSTTBGNDDQGF
tr Q75CW6 Q75CW6_ASHGO/113-131	HQHMLVGNDDQGF	tr Q6BZF0 Q6BZF0_DEBHA/107-120	NQSTTBGNDDQGF
tr G8JMI3 G8JMI3_ERECY/109-122	NQSTMVGNDDQGF	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO/107-120	NQSTTBGNDDQGF
tr C5DHG0 C5DHG0_LACTC/106-119	NQSTMVGNDDQGF	tr A0A1B2J789 A0A1B2J789_PICPA/108-121	NQSTTBGNDDQGF
tr A0A0C7N752 A0A0C7N752_9SACH/113-126	NQSTMVGNDDQGF	tr C4QXV4 C4QXV4_KOMPG/108-121	NQSTTBGNDDQGF
tr A0A1G4JM92 A0A1G4JM92_9SACH/113-126	NQSTMVGNDDQGF	tr G8J588 G8J588_ERECY/109-122	NQSTTBGNDDQGF
tr W0THH8 W0THH8_KLUMD/107-120	NQSTMVGNDDQGF	sp Q75D66 DCW1_ASHGO/109-122	NEITTBGNDDQGF
tr Q6CIP9 Q6CIP9_KLULA/106-119	NQSTMVGNDDQGF	tr R9XAT7 R9XAT7_ASHAC/109-122	NEITTBGNDDQGF
tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH/106-119	NQSTMVGNDDQGF	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA/108-121	NQSTTBGNDDQGF
tr J7RQR5 J7RQR5_KAZNA/118-131	NQSTMVGNDDQGF	tr A0A061B6H7 A0A061B6H7_CYBFA/106-119	NQSTTBGNDDQGF
tr G8ZPC1 G8ZPC1_TORDC/90-103	NQSTMVGNDDQGF	tr A0A157HQM3 A0A157HQM3_9SACH/109-122	NQSTTBGNDDQGF
tr C5DYD7 C5DYD7_ZYGRC/107-120	NQSTMVGNDDQGF	tr A0A157HZQ4 A0A157HZQ4_9SACH/109-122	NQSTTBGNDDQGF
tr S6E3R3 S6E3R3_ZYGB2/107-120	NQSTMVGNDDQGF	tr J75438 J75438_KAZNA/116-129	NQSTTBGNDDQGF
tr A0A157HIA4 A0A157HIA4_9SACH/109-122	NQSTMVGNDDQGF	tr W0TAH6 W0TAH6_KLUMD/108-121	NQSTTBGNDDQGF
tr J2GYH1 J2GYH1_TETBL/107-120	NQSTMVGNDDQGF	tr Q6CP42 Q6CP42_KLULA/108-121	NQSTTBGNDDQGF
tr H2AQJ6 H2AQJ6_KAZAF/104-117	NQSTMVGNDDQGF	tr A0A0A8L8U3 A0A0A8L8U3_9SACH/108-121	NQSTTBGNDDQGF
tr G0WHL4 G0WHL4_NAUDC/114-127	NQSTMVGNDDQGF	tr C5DGT1 C5DGT1_LACTC/108-121	NQSTTBGNDDQGF
tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU/114-127	NQSTMVGNDDQGF	tr A0A0C7N2N9 A0A0C7N2N9_9SACH/108-121	NQSTTBGNDDQGF
tr J8Q6F0 J8Q6F0_SACAR/114-127	NQSTMVGNDDQGF	tr A0A1G4JVK5 A0A1G4JVK5_9SACH/108-121	NQSTTBGNDDQGF
tr J6EGN8 J6EGN8_SACK1/114-127	NQSTMVGNDDQGF	tr A7TLL2 A7TLL2_VANPO/109-122	NQSTTBGNDDQGF
sp Q05031 DFG5_YEAST/114-127	NQSTMVGNDDQGF	tr J2H842 J2H842_TETBL/108-121	NQSTTBGNDDQGF
tr G2WKU6 G2WKU6_YEASK/114-127	NQSTMVGNDDQGF	tr G8ZQ93 G8ZQ93_TORDC/107-120	NQSTTBGNDDQGF
tr A6ZMV1 A6ZMV1_YEAS7/114-127	NQSTMVGNDDQGF	tr G0W5X4 G0W5X4_NAUDC/107-120	NQSTTBGNDDQGF
tr C7GRB3 C7GRB3_YEAS2/114-127	NQSTMVGNDDQGF	tr G0V8N4 G0V8N4_NAUCC/107-120	NQSTTBGNDDQGF
tr A0A0L8VJB2 A0A0L8VJB2_9SACH/114-127	NQSTMVGNDDQGF	tr H2AMQ5 H2AMQ5_KAZAF/109-122	NQSTTBGNDDQGF
tr G8YQC0 G8YQC0_PICSO/113-126	NQSTMVGNDDQGF	tr G8BXX2 G8BXX2_TETPH/112-125	NQSTTBGNDDQGF
tr G8YRT2 G8YRT2_PICSO/113-126	NQSTMVGNDDQGF	tr H2B005 H2B005_KAZAF/108-121	NQSTTBGNDDQGF
tr C4XWK1 C4XWK1_CLAL4/111-124	NQSTMVGNDDQGF	tr G0W7G8 G0W7G8_NAUDC/112-125	NQSTTBGNDDQGF
tr A3LMV8 A3LMV8_PICST/113-126	NQSTMVGNDDQGF	sp Q6FLP9 DCW1_CANGA/106-119	NQSTTBGNDDQGF
tr G3AKK4 G3AKK4_SPAPN/118-131	NQSTMVGNDDQGF	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB/106-119	NQSTTBGNDDQGF
tr A5DV30 A5DV30_LODEL/111-124	NQSTMVGNDDQGF	tr J8Q2K7 J8Q2K7_SACAR/109-122	NQSTTBGNDDQGF
tr C5M5J2 C5M5J2_CANTT/111-124	NQSTMVGNDDQGF	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU/109-122	NQSTTBGNDDQGF
sp Q5ACZ2 DFG5_CANAL/111-124	NQSTMVGNDDQGF	sp P36091 DCW1_YEAST/109-122	NQSTTBGNDDQGF
tr B9WAA8 B9WAA8_CANDC/111-124	NQSTMVGNDDQGF	tr A0A0L8VLX1 A0A0L8VLX1_9SACH/109-122	NQSTTBGNDDQGF
tr A0A0H5CF34 A0A0H5CF34_CYBJA/110-123	NQSTMVGNDDQGF	tr G2WHY6 G2WHY6_YEASK/109-122	NQSTTBGNDDQGF
tr A0A0H5CB47 A0A0H5CB47_CYBJA/105-118	NQSTMVGNDDQGF	tr A6ZZ50 A6ZZ50_YEAS7/109-122	NQSTTBGNDDQGF
tr A5DNV5 A5DNV5_PICGU/112-125	NQSTTBGNDDQGF	tr C7GP28 C7GP28_YEAS2/109-122	NQSTTBGNDDQGF
tr G8YB80 G8YB80_PICSO/107-120	NQSTTBGNDDQGF	tr J5RXY9 J5RXY9_SACK1/109-122	NQSTTBGNDDQGF
tr G8YM23 G8YM23_PICSO/107-120	NQSTTBGNDDQGF	tr Q6CAI2 Q6CAI2_YARLI/106-119	NQSTTBGNDDQGF
tr A0A1E45C85 A0A1E45C85_9ASCO/106-119	NQSTTBGNDDQGF	tr A0A167CDD3 A0A167CDD3_9ASCO/108-121	NQSTTBGNDDQGF
tr C4Y2X5 C4Y2X5_CLAL4/104-117	NQSTTBGNDDQGF	tr A0A167CDC5 A0A167CDC5_9ASCO/111-124	NQSTTBGNDDQGF
tr A0A0L0P6S8 A0A0L0P6S8_CANAR/107-120	NQSTTBGNDDQGF		

SFigure 11: Multiple sequence alignment of ABL2-region generated by Clustal Omega using 200 sequences.

7.2 Tables

Table S 1: Crystallographic data collection statistics of CtDfg5 crystals soaked with 500 mM inositol for 4 h. Values in parentheses are for the highest resolution shell.

CtDfg5 _{Ino}	
Data collection	
X-ray source	ID 23-2, ESRF
Space group	C 1 2 1
Unit-cell parameters (Å, °)	$a=83.41$, $b=55.07$, $c=80.34$, $\alpha=90.00$, $\beta=90.31$, $\gamma=90.00$
Wavelength (Å)	0.87
Resolution range (Å)	45.96-1.2 (1.22-1.2)
Completeness (%)	98.2 (84.1)
Observed reflections	479751 (14185)
Unique reflections	111409 (4656)
Multiplicity	4.3
Wilson B factor (Å ²)	10.01
R_{merge} (%)	7.6 (110.5)
Mean $I/\sigma(I)$	8.1 (1.0)
CC(½)	98.2 (84.1)

Table S 2: Identified GPI-APs upon PI-PLC treatment in the supernatant. The analytics were performed by the MS-facility led by Dr. Uwe Linne.

Uniprot-ID	Description	Coverage [%]	# Peptides	# PSMs	# Unique Peptides
P22146	1,3-beta-glucanosyltransferase GAS1 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=GAS1 PE=1 SV=2	14	16	105	16
Q03655	Probable 1,3-beta-glucanosyltransferase GAS3 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=GAS3 PE=1 SV=1	29	13	53	13
Q08193	1,3-beta-glucanosyltransferase GAS5 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=GAS5 PE=1 SV=1	14	5	26	5
P38248	Cell wall protein ECM33 OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=ECM33 PE=1 SV=3	39	25	145	25
P46992	Cell wall protein YJL171C OS=Saccharomyces cerevisiae (strain ATCC 204508 / S288c) GN=YJL171C PE=1 SV=2	28	7	11	7

Table S 3: List of abbreviations used in this thesis.

ABL	Acceptor binding loop
Af	<i>Aspergillus fumigatus</i>
An	<i>Aspergillus niger</i>
Bc	<i>Bacillus circulans</i>
C-terminus	Carboxyl-terminus
Ca	<i>Candida albicans</i>
Ct	<i>Chaetomium thermophilum</i>
CV	Column volume
CWP	Cell wall protein
Dcw1	Defective cell wall
DNA	Deoxyribonucleic acid
Dfg5	Defective in filamentous growth
e.g.	For example
et al.	Latin: <i>Et alii/aliae</i> – „and others“

EtN-P	Ethanolamine phosphate
ERES	Endoplasmatic reticulum exit site
FCW	Fungal cell wall
FP-1	(3-{4-imino-5,6-dimethyl-3H,4H-furo[2,3-d]pyrimidin-3-yl}propyl)dimethylamine
FP-2	3-[3-(1H-imidazol-1-yl)propyl]-5,6-dimethyl-3H,4H-furo[2,3-d]pyrimidin-4-imine
GH	Glycosyl hydrolase
Glc	Glucose
GlcN	Glucosamine
GlcNAc	N-acetylglucosamine
GPI-AP	Glycosylphosphatidylinositol-anchored protein
GPI-CWP	Glycosylphosphatidylinositol-cell wall protein
GPI-PMP	Glycosylphosphatidylinositol-plasma membrane protein
GT	Glycosyl transferase
GTP	Guanosine triphosphate
i.e.	<i>Latin: id est</i> – „that is to say“
IMAC	Immobilized metal ion affinity chromatography
Ino	Inositol
IPC	Inositolphosphorylceramide
JCSG	Joint Center for Structural Genomics
Man	Mannose
MBP	Maltose-binding protein
MR	Molecular replacement
MSA	Multiple sequence alignment
<i>n</i>	Integer
N-terminus	Amino-terminus
<i>Nc</i>	<i>Neurospora crassa</i>
Ni-NTA	Nickel-nitrilotriacetic acid
p.	Page
PDB	Protein data bank
PI	Phosphatidylinositol
PIR-CWP	Protein with internal repeats-cell wall protein
PM	Plasma membrane
SAD	Single-wavelength anomalous diffraction
SAR	Structure activity relationship
<i>Sc</i>	<i>Saccharomyces cerevisiae</i>
SEC	Size exclusion chromatography
SSN	Sequence similarity network
TM(H)	Transmembrane helix
UDP	Uridine diphosphate

Table S 4: List of amino acids and their one-letter codes.

Amino acid	One-letter symbol
Alanine	A
Arginine	R
Asparagine	N
Aspartic acid	D
Cysteine	C
Glutamic acid	E
Glutamine	Q
Glycine	G

Histidine	H
Isoleucine	I
Leucine	L
Lysine	K
Methionine	M
Phenylalanine	F
Proline	P
Serine	S
Threonine	T
Tryptophan	W
Tyrosine	Y
Valine	V

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Supplement of the electronic resource

Complete multiple sequence alignment used for *Consurf* analysis of CtDfg5

ConSurf Color-Coded MSA

001	tr A0A254U2J9 A0A254U2J9_ASPNG	-----	MRLV	RDC	L	----	YG	----
002	sp Q6FLP9 DCW1_CANGA	-----	-	-	-	M	----	RL
003	tr A1CCM5 A1CCM5_ASPCL	-----	MRLP	SLRV	V	----	GW	----
004	tr C1H190 C1H190_PARBA	-----	MRLFP	HSSSR	ST	SQ	SCT	A-ILLS
005	tr Q2UR85 Q2UR85_ASPOR	-----	-	MPPD	WL	-	-	-Y-
006	tr A5DV30 A5DV30_LODEL	-----	MEFV	-	-	F	-	-
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	-----	MKLS	-	-	S	-	-TY-
008	tr J3PGN0 J3PGN0_GAGT3	-----	MGRSR	-	LS	-	-	-QG-A-
009	tr A0A124BXU6 A0A124BXU6_ASPNG	-----	-	MAGNQ	WLR	-	-	-CV-
010	tr C5P4A1 C5P4A1_COCP7	-----	MRLF	-	SWS	-	SSS	-GWMA-QSL
011	tr Q0CG55 Q0CG55_ASPTN	-----	-	-	-	-	MRR	-PWH-LIL-
012	tr C4Y2X5 C4Y2X5_CLAL4	-----	-	-	-	M	-	-
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	-----	-	-	-	-	MLLR	-IWH-ILY-
014	tr C7Z068 C7Z068_NECH7	-----	MLSLK	-	S	-	-	-AAL-
015	tr G8BXX2 G8BXX2_TETPH	-----	MRFN	-	-	T	-	-
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	-----	MRL	-	-	L	-	-
017	tr A1CSC2 A1CSC2_ASPCL	-----	MNLL	GRSS	-	-	-	-S-
018	sp Q05031 DFG5_YEAST	-----	MIVN	ISA	-	KM	-	-ILS-ICFT
019	tr H2AQJ6 H2AQJ6_KAZAF	-----	-	-	-	-	M	-MIT-LLFP
020	tr G2XFH9 G2XFH9_VERDV	-----	-	MFN	-	S	-	-LQTVA-
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	-----	-	MQFR	-	-	K	-
022	sp Q5AD78 DCW1_CANAL	-----	-	MK	-	-	F	-
023	tr B6H7E8 B6H7E8_PENRW	-----	-	-	MPRW	L	-	-
024	tr B6GZU2 B6GZU2_PENRW	-----	-	MRVF	NYAS	QPSQ	P	-SWM-GLLA
025	tr E9E4W7 E9E4W7_METAQ	-----	-	MVA	-	P	-	-RLSPA-
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	-----	-	-	MRVAGH	STRP	-	-SWA-SLLT
027	tr G2R7G8 G2R7G8_THITE	-----	-	MGWVS	-	TSK	-	-ATALLSA-
028	tr G3JBY1 G3JBY1_CORMM	-----	-	MRG	-	-	-	-
029	tr A5DNV5 A5DNV5_PICGU	-----	-	MKFL	-	-	S	-CGS-
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	-----	-	-	-	MHL	-	G-IYAA-FVVG
031	tr Q2TWC1 Q2TWC1_ASPOR	-----	-	MRGL	SWQQ	L	-	-GG-
032	tr J5RXY9 J5RXY9_SACK1	-----	-	MLL	-	-	N	-
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	-----	-	MKAS	-	-	-	-
034	tr A0A254U5V1 A0A254U5V1_ASPNG	-----	-	MVV	PGWI	-	-	-W-
035	tr A0A061B6H7 A0A061B6H7_CYBFA	-----	-	-	-	-	-	-
036	tr G8ZQ93 G8ZQ93_TORDC	-----	-	MFV	-	-	M	-
037	tr Q9C2J1 Q9C2J1_NEUCS	-----	-	MRWN	-	-	-	-
038	tr C5NZK5 C5NZK5_COCP7	-----	-	MILPS	LFSNQP	GPPCS	-	A-LFLT
039	tr Q6C0T7 Q6C0T7_YARLI	-----	-	MRIQ	-	-	-	-
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	-----	-	-	-	-	M	-
041	tr F7VZ72 F7VZ72_SORMK	-----	-	-	-	MF	-	STLQRLA-
042	tr A3LN37 A3LN37_PICST	-----	-	MRFH	-	-	L	-
043	tr Q6CAI2 Q6CAI2_YARLI	-----	-	MKLT	-	-	-	-
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	-----	-	MKLT	-	-	I	-
045	tr G9MJS5 G9MJS5_HYPVG	-----	-	-	-	-	MK	-SFFRPGAM-
046	tr G9MHI4 G9MHI4_HYPVG	-----	-	MKSP	-	-	-	-
047	tr D4AP27 D4AP27_ARTBC	-----	-	MKLK	-	RLS	F	LSP--RWVV-L--
048	tr B6GZT8 B6GZT8_PENRW	-----	-	MHLL	GHCSS	-	-	-SR-
049	sp O74556 YCZ2_SCHPO	-----	-	MSLT	-	-	-	-
050	tr A0A0C4E991 A0A0C4E991_MAGP6	-----	-	MGRSR	-	LS	-	-QS-A-
051	tr B8MHG0 B8MHG0_TALSN	-----	-	MRSF	-	-	I	-
052	tr A3LMV8 A3LMV8_PICST	-----	-	MLNP	-	-	S	-
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	-----	-	MKLQ	-	-	H	-
054	sp Q9P6I3 YHG7_SCHPO	-----	-	MRYLS	-	F	-	-
055	tr Q2US57 Q2US57_ASPOR	-----	-	MVQG	VNWL	N	-	-GG-
056	tr W7N622 W7N622_GIBM7	-----	-	MRAS	-	-	-	-
057	sp Q9P6I4 YHG6_SCHPO	-----	-	MNIYS	V	G	-	-
058	tr C7GRB3 C7GRB3_YEAS2	-----	-	MIVN	ISA	-	KM	-
059	tr C7GP28 C7GP28_YEAS2	-----	-	MLV	-	-	N	-
060	tr G3JGZ5 G3JGZ5_CORMM	-----	-	MFS	-	-	L	-
061	tr C4QXV4 C4QXV4_KOMPG	-----	-	-	-	-	-	-
062	tr I1S0Z5 I1S0Z5_GIBZE	-----	-	-	-	-	MK	-SFYSAGA-
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	-----	-	-	-	-	MHCMQ	L--LWL-L--
064	tr G0W5X4 G0W5X4_NAUDC	-----	-	MK	-	-	-	-
065	tr G8ZPC1 G8ZPC1_TORDC	-----	-	-	-	-	-	-
066	tr A0A167CDC5 A0A167CDC5_9ASCO	-----	-	MKLP	-	-	-	-

[illegible]

136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH											MIM		LIMN
137	tr Q752P3 Q752P3_ASHGO													MT
138	tr B8MYP3 B8MYP3_AS PFN									MPPDWL		Y		
139	tr Q2TXL6 Q2TXL6_AS POR									MWSYF		W		
140	tr Q0CN17 Q0CN17_AS PTN									MQP NRL		W		
141	tr J3NHD5 J3NHD5_GAGT3									MF		APLV RVG		
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI										MKTG	SFFA		LAT
143	tr A1CVB0 A1CVB0_NEOFI									MHCMQL		LWL		L
144	tr B8NVI3 B8NVI3_AS PFN									MWSYF		W		
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH									M				RL
146	tr A1DJ54 A1DJ54_NEOFI									MRMLS LRQL		CG		
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS									MRTT	SSP	RGATWLTA		
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA													MKP
149	tr A0A177A618 A0A177A618_9PEZI									MRL		WSV	AR	
150	tr Q75CW6 Q75CW6_ASHGO								MTVA FVRVGAE					PS
151	tr Q7S4K4 Q7S4K4_NEUCR									MRTT	SSP	RGATWLTA		
152	tr G2QB99 G2QB99_MYCTT									MR I				SG
153	tr G0S3F2 G0S3F2_CHATD								MTTWS	SSK		RPLRWLVA		
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO									MK		L		
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB											MK		LLFS
156	tr A0A0J5PQ80 A0A0J5PQ80_AS PFM	MFTH PAPQPSNSDAL LNRLISENIAM								MRLQSLPRL		GW		
157	tr G2QKJ0 G2QKJ0_MYCTT									MAP		TTTVLLG		
158	tr Q4WFX5 Q4WFX5_AS PFU										MGRQCPC HWPL	SLRL		
159	tr Q5BGD7 Q5BGD7_EMENI									MRWGSLLSL		GG		
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH										M			KV
161	tr G8BYN9 G8BYN9_TETPH										M	FYL	TLL	
162	tr F7VVT4 F7VVT4_SORMK										ML	LTTKIAG		
163	tr G8YB80 G8YB80_PICSO									MKTL		P		
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA													
165	tr G2WRS7 G2WRS7_VERDV										MK	G SFVARA		
166	tr A0A124BVB5 A0A124BVB5_AS PNG											MHIP	PL RLWR	
167	tr A7EIG7 A7EIG7_SCLS1										MKLSTS	SSAL	AT	
168	tr C5M5U7 C5M5U7_CANTT								MKFSCS		F			
169	tr B9WAA8 B9WAA8_CANDC									MISL		Q		
170	tr F8MRU1 F8MRU1_NEUT8									MRTT	SSP	RGAKWLTA		
171	tr A1DJS0 A1DJS0_NEOFI									MRLQSLPRL		GW		
172	tr B6HLM8 B6HLM8_PENRW									MRFKGAGH		M		
173	tr A0A117DWS0 A0A117DWS0_AS PNG									MVVPGWI		W		
174	tr A0A124BXM1 A0A124BXM1_AS PNG									MRLVRDC	L	YG		
175	tr J3NZQ6 J3NZQ6_GAGT3											MARRWGATA LLL		
176	tr W0THH8 W0THH8_KLUMD													MR
177	tr B2AEF2 B2AEF2_PO DAN								MATTP	APK		SPSRWLTA		
178	tr Q96TX1 Q96TX1_NEUCS										ML	SPLPRLA		
179	tr E9DYA													

022	sp Q5AD78 DCW1_CANAL	----	SIYLIISLSSFSFSAIW	----	LDTN	----	NET
023	tr B6H7E8 B6H7E8_PENRW	----	TL-LTF-LLL--QNALAI	----	VNPE	----	DET
024	tr B6GZU2 B6GZU2_PENRW	A----	-----ALLGTSTVQAVE	----	LDVE	----	DGD
025	tr E9E4W7 E9E4W7_METAQ	----	LVAIASSLLFSGVAEAQYKVDT	----		----	RD
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	I----	-----GLAGLGYVSAFD	----	MDIE	----	DPD
027	tr G2R7G8 G2R7G8_THITE	----	--V-----LLAAT--	----	GAIAAGEAKLQVDLS	----	SPD
028	tr G3JBY1 G3JBY1_CORMM	A----	VSLVSLLLA--APSNAL	----	LS-SK	----	A
029	tr A5DNV5 A5DNV5_PICGU	----	SCGFLVVSALIWVVSALD	----	LQTD	----	DFD
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	A----	-----LLSICGISSAIH	----	VDLS	----	SSD
031	tr Q2TWC1 Q2TWC1_ASPOR	----	AVL--L-LLLQGQVALSALS	----	VQVN	----	SKD
032	tr J5RXY9 J5RXY9_SACK1	I----	GL--LSVLFFTKFSNAVE	----	LDLD	----	NYE
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	A----	QTLGLSAT--ASAAQLN	----	LDLT	----	SES
034	tr A0A254U5V1 A0A254U5V1_ASPNG	----	AV-FLTIFVSFQRTVTALQ	----	LDIN	----	DEQ
035	tr A0A061B6H7 A0A061B6H7_CYBFA	R----	LTSLLAVSVLNSVAQAIW	----	LTPD	----	DLD
036	tr G8ZQ93 G8ZQ93_TORDC	L----	A----ALASTFACANAVW	----	IDLN	----	NTE
037	tr Q9C2J1 Q9C2J1_NEUCS	----	VCGLMGLLA--QSATAIT	----	MDID	----	DTQ
038	tr C5NZK5 C5NZK5_COCP7	V----	-----LLGVQAVHAAIP	----	LDLG	----	SPD
039	tr Q6C0T7 Q6C0T7_YARLI	----	LVAALCGLITTVTC-ALN	----	LDLN	----	SPV
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	V----	TL--LSGLVSLVSVFGL	----	LDLD	----	DYA
041	tr F7VZ72 F7VZ72_SORMK	----	AVALV--GVDVA-NAAFSVAS	----		----	VA
042	tr A3LN37 A3LN37_PICST	----	--LTASVSLFMACASAMW	----	LDTN	----	NDT
043	tr Q6CAI2 Q6CAI2_YARLI	I----	-----SRLLFATTASALT	----	VDLD	----	DPE
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	----	--VLS-AVIGLVSTVQGIW	----	LDTN	----	NVT
045	tr G9MJS5 G9MJS5_HYPVG	----	AA-LLSVTDMAVAQQSPFSIAS	----		----	NS
046	tr G9MHI4 G9MHI4_HYPVG	A----	ATAALCLLG--QGANAI	----	LTVT	----	DSN
047	tr D4AP27 D4AP27_ARTBC	L----	-----LLLQSWLASAIK	----	LDLE	----	SDD
048	tr B6GZT8 B6GZT8_PENRW	----	AA-IFALLLSS--IARAI	----	LDIS	----	DEQ
049	sp O74556 YCZ2_SCHPO	L----	ATILFSFA-----EAI	----	VDLN	----	DTS
050	tr A0A0C4E991 A0A0C4E991_MAGP6	----	AAALLVAASLQTANAQYYKIDT	----		----	VD
051	tr B8MHG0 B8MHG0_TALSN	----	SI-LLGQLLLTMPFATALT	----	VNAN	----	DPN
052	tr A3LMV8 A3LMV8_PICST	----	--STFVLTLAISLSVSAVS	----	LDVD	----	SKE
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	----	--AALMVAGALASSATAIE	----	VELG	----	NEE
054	sp Q9P6I3 YHG7_SCHPO	----	----FFEFFFLFSFAFAFD	----	FDVT	----	SDD
055	tr Q2US57 Q2US57_ASPOR	----	ALVYLGVLRLR--TVKA	----	VDVS	----	SEQ
056	tr W7N622 W7N622_GIBM7	K----	TVAVTSALA--AVVHGID	----	VSWD	----	DDK
057	sp Q9P6I4 YHG6_SCHPO	----	----LFYFFLVFIGAQAMD	----	LDIT	----	DYQ
058	tr C7GRB3 C7GRB3_YEAS2	F----	L-----SFFKATHAMD	----	LDTT	----	SKT
059	tr C7GP28 C7GP28_YEAS2	I----	GL--LGVLFATRFTNAVE	----	LDLD	----	NYE
060	tr G3JGZ5 G3JGZ5_CORMM	----	AASLVLSGLYDSVQAQYYKVDT	----		----	KE
061	tr C4QXV4 C4QXV4_KOMPG	L----	--VLVLALLRVLSPAKGL	----	LELS	----	DLS
062	tr I1S0Z5 I1S0Z5_GIBZE	----	IALLASTGLVSAQVSPYSIS	----		----	TD
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	----	-----TLSPAYSIP	----	LDPN	----	DPT
064	tr G0W5X4 G0W5X4_NAUDC	R----	VFNILFLLTFTHRANALS	----	LDLN	----	NYQ
065	tr G8ZPC1 G8ZPC1_TORDC	----	-----MGLD	----	LDPS	----	SKD
066	tr A0A167CDC5 A0A167CDC5_9ASCO	V----	SLFALTLLQCVYNTVAIT	----	VDFQ	----	DDT
067	tr R9XAT7 R9XAT7_ASHAC	T----	FTVAATLSLLAGCGRTLQ	----	LDVD	----	DLQ
068	tr B2B747 B2B747_PODAN	----	AVLLS--GAAVANAATYSIAS	----		----	AA
069	tr G0SE99 G0SE99_CHATD	----	TAVLL--GARVT-YAAFKLDT	----		----	IE
070	tr J8Q6F0 J8Q6F0_SACAR	F----	L-----TFFKASHAMD	----	LDTT	----	SNT
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	I----	GLLSLFLAVT-SVLAI	----	LDTS	----	SKD
072	tr J3K6E2 J3K6E2_COCIM	A----	-----LVGMLSIVSAID	----	LNLD	----	DHE
073	tr G4N3E1 G4N3E1_MAGO7	----	T-ATLSLAAVPIANAQFYKISS	----		----	SQ
074	tr C5M5J2 C5M5J2_CANTT	----	SLIVYVLLLFTSVVYSVD	----	MDTS	----	SKE
075	tr A0A254U0X0 A0A254U0X0_ASPNG	----	FLL--L-LLLHSQRAYSIDL	----	IQVN	----	DAN
076	sp Q75DG6 DCW1_ASHGO	T----	FTAAAVLSLLAASGRTL	----	LDVD	----	DLQ
077	tr Q6BZF0 Q6BZF0_DEBHA	----	--VLS-AVIGLVSTVQGIW	----	LDTN	----	NVT
078	tr E9E3Q1 E9E3Q1_METAQ	----	WLAFA GTLLIGTTGAQYYKIDT	----		----	ED
079	sp P36091 DCW1_YEAST	I----	GL--LGVLFATRFTNAVE	----	LDLD	----	NYE
080	tr W0TAH6 W0TAH6_KLUMD	W----	KLMLTTTLTAFLTVA	----	LDVD	----	DLN
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	S----	LRALMVPMFLAQQALSID	----	LDVD	----	SET
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	T----	IAGMAGLLL--QPAAAI	----	VNFD	----	DTQ
083	tr Q2TYU3 Q2TYU3_ASPOR	----	AIVC-LAGIGR--VVEALS	----	IDIN	----	DAD
084	tr G2XGF6 G2XGF6_VERDV	----	LSAVCLM-G--NAAHAIQ	----	LDID	----	SEQ
085	tr G3AMT4 G3AMT4_SPAPN	----	--TLVLTTLCTITNVVHAIW	----	LDTN	----	NDT
086	tr A0A1C1D213 A0A1C1D213_9EURO	S----	-----ALLFTQNALAI	----	LDLS	----	SPD
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	----		----		----	
088	tr A0A100I5A6 A0A100I5A6_ASPNG	----	SLL--L-LLLHSQRAYSIDL	----	IQVN	----	DAD
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	G----	LLW--L-QLMVLPVRALD	----	LDTT	----	SKD
090	tr G2Q8A7 G2Q8A7_MYCTT	----	AAALA--GAGVA-SAYS	----	SIAT	----	TD
091	tr B8NPZ1 B8NPZ1_ASPFN	----	AVL--L-VFLSGQGALADLG	----	IDVN	----	NVD

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tr F7VVT4 F7VVT4_SORMK	M---	-VAP--L	LLAAASSVY-AATTTGQSTLQVNVN	---	DTA--
163 tr G8YB80 G8YB80_PICSO	----	--VFSALLGILVTVTGLE	-----LDVT	-----	SKS--
164 tr A0A0H5CB47 A0A0H5CB47_CYBJA	M---	-ILLPILTAFLVTRSGAID	-----LDLS	-----	SRD--
165 tr G2WRS7 G2WRS7_VERDV	----	-ATTLL--SAHTAHAAYYSIQS	-----	-----	DD--
166 tr A0A124BVB5 A0A124BVB5_ASPNG	S---	-LLVCVIWSLPCFQVLAI	-----LDLQ	-----	NAD--
167 tr A7EIG7 A7EIG7_SCLS1	A---	-----LIAGSHLVNAIE	-----VDLT	-----	SPD--
168 tr C5M5U7 C5M5U7_CANTT	----	--TAYVIIISIFSTLANAIW	-----LDTN	-----	NET--
169 tr B9WAA8 B9WAA8_CANDC	----	-QLIISILLFFTSSVQSVD	-----LTV	-----	DKD--
170 tr F8MRU1 F8MRU1_NEUT8	----	-LFAAAACLLP-AANAQGYAIDT	-----	-----	TD--
171 tr A1DJS0 A1DJS0_NEOFI	----	-NALWTTLIFSS--LAQAID	-----VNIN	-----	DDQ--
172 tr B6HLM8 B6HLM8_PENRW	----	-AI-VALRILPFIGHVYALD	-----LDVN	-----	SQE--
173 tr A0A117DWS0 A0A117DWS0_ASPNG	----	-AV-FLTIFVFFQRTVTALQ	-----LDIN	-----	DEQ--
174 tr A0A124BXM1 A0A124BXM1_ASPNG	----	-GLFLNTALFSQ--LSHAID	-----IDIS	-----	STS--
175 tr J3NZQ6 J3NZQ6_GAGT3	LH--	-ATASIQGVAAQANGPNFYKIST	-----	-----	KE--
176 tr W0THH8 W0THH8_KLUMD	L---	-GIWSYLILSISSLVSAID	-----LDTS	-----	SKD--
177 tr B2AEF2 B2AEF2_PODAN	----	-LLATTSCLLP-GARATVYKLGT	-----	-----	KA--
178 tr Q96TX1 Q96TX1_NEUCS	----	-AVALV--GVDVA-NAAYSIA	T-----	-----	VA--
179 tr E9DYA6 E9DYA6_METAQ	----	-SWTLLA-AGRLSAAASVFS	LDS-----	-----	DD--
180 tr B9WAI2 B9WAI2_CANDC	----	--STYIIISLFSLSHAIW	-----LDTN	-----	NET--
181 tr Q6C171 Q6C171_YARLI	A---	-ILSALAFLLGTARADLSF	-----MDLS	-----	KPD--
182 tr A6ZMV1 A6ZMV1_YEAS7	F---	-L-----SFFKATHAMD	-----LDTT	-----	SKT--
183 tr B6K7X7 B6K7X7_SCHJY	I---	-TTVLLGFLTTCIRLSFAFS	-----LDVT	-----	DEN--
184 tr I1RSA2 I1RSA2_GIBZE	T---	-TTAVLGWTA--VAANAYE	-----LT	-----	ITD--
185 tr F7VWZ9 F7VWZ9_SORMK	----	-AAGLMGLFA--QSATAIK	-----MDID	-----	NDQ--
186 tr A0A1B2J789 A0A1B2J789_PICPA	L---	--VLVLAFLRVLSPAKGLE	-----LELS	-----	DLS--
187 tr G0V8N4 G0V8N4_NAUCC	I---	-IYLLFTVSSFSFPIAAVD	-----LNLN	-----	SLQ--
188 tr H8WXN8 H8WXN8_CANO9	----	--SFAILLTSLLTFSQAIS	-----LDTN	-----	NVT--
189 tr A6ZZS0 A6ZZS0_YEAS7	I---	-GL--LGVLFATRFTNAVE	-----LDLD	-----	NYE--
190 tr G9MQD1 G9MQD1_HYPVG	----	-SFVIIATCFFYATVEAIYRIDN	-----	-----	AD--
191 tr A0A1G4JVK5 A0A1G4JVK5_9SACH	S---	-VAWILALLASFHNCA	GVY-----LDVN	-----	NTA--
192 tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	----	-ATT--L	LLSGQVALADLG-----IQAD	-----	NVD--
193 tr A0A254UET1 A0A254UET1_ASPNG	S---	-IILCVVWSLPCFHVLAID	-----LDLQ	-----	NPD--
194 tr A1D587 A1D587_NEOFI	----	-TS-LVTVVL	AGQGAVTAL-----LDIN	-----	DVQ--
195 tr Q1K7I4 Q1K7I4_NEUCR	----	-AVALV--GVDVA-NAAYSIA	T-----	-----	VA--
196 tr G8JS88 G8JS88_ERECY	W---	-VILGVFLNLLGTDVSALE	-----LDVD	-----	DFS--
197 tr A0A136J7D3 A0A136J7D3_9PEZI	----	--FAGLGLLA--GSVGAIQ	-----VNFD	-----	SDD--
198 tr E9E4X8 E9E4X8_METAQ	----	-GVMLLAAAFAQAQTS	PFSIDSTGKATPIPTPNT-----	-----	VRA--
199 tr Q6CP42 Q6CP42_KLULA	F---	-KLVL	SLVTLFSSVSFGLD-----LDVN	-----	DLN--
200 tr G0WHL4 G0WHL4_NAUDC	L---	-L-----LTVHNTQ	AID-----LDIT	-----	SKE--
201 tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	V---	-GL--LSVLFATRFTNAVE	-----LDLD	-----	NYQ--
202 tr F9WYX6 F9WYX6_ZYMTI	----	----LTSLALSAGLANAID	-----LDVN	-----	NRD--
203 tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	----	-ATALL--AGGLV-DAAYSIA	N-----	-----	TA--
204 tr J3P147 J3P147_GAGT3	VTAQ	APAAPAPAAPAGDPHPFYKIGT	-----	-----	RD--
205 tr I2H842 I2H842_TETBL	F---	-HSLLLKISVITTTITNAVS	-----LNLN	-----	DSS--
206 tr S6E3R3 S6E3R3_ZYGB2	I---	-IAW-LLH--AIIPVWALD	-----LEVE	-----	SKD--
207 tr E5AD94 E5AD94_LEPMJ	A---	-LSLLSS--HVVPQVQALVLD	--PTSPGKILTA-----	-----	TLE--
208 tr Q6FJM7 Q6FJM7_CANGA	L---	-QLW-VLIGTLFPDAGAIN	-----L-QD	-----	NKD--
209 tr A5DUW0 A5DUW0_LODEL	----	--VSCFITFCFLTTLTQAIW	-----LDTN	-----	NDT--
210 tr G8BF46 G8BF46_CANPC	----	--SLALLLANLLPFLQALP	-----LDTN	-----	NVT--
211 tr W3XAF6 W3XAF6_PESFW	----	-VFGLLQTLA--YQVSAVD	-----LDIT	-----	QPD--
212 tr B6H7I2 B6H7I2_PENRW	----	-GL-IATLLFST--LTPAIE	-----LDLN	-----	DEQ--
213 tr Q5ATF9 Q5ATF9_EMENI	----	-----ALLASLGRISALE	-----IQLN	-----	DPQ--
214 tr A0A167CDD3 A0A167CDD3_9ASCO	----	--ITASVALLVQQATS	SLT-----LDET	-----	SVE--
215 tr G0RXS3 G0RXS3_CHATD	----	--ALL--FLTAAC--	---VHAQDGRAKLQVDLD-----	-----	DYD--
216 tr B8NX26 B8NX26_ASPFN	----	-TLL--LLFLHAQTVWS	DLG-----IVAN	-----	NED--
217 tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	V---	-----LLGVQAVHAAIP	-----LDLG	-----	SPG--
218 tr A0A0J5SN29 A0A0J5SN29_ASPFM	S---	-LL--LSLSLLPLRATTLD	-----LNV	T-----	DPD--
219 tr A1CMM3 A1CMM3_ASPCL	A---	-----LLAGQMAQAID	-----LDLT	-----	S---
220 tr B6HU84 B6HU84_PENRW	----	-TL-LGGAFAV--QGALAIE	-----LNID	-----	DEMRL
221 tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	----	-AVALV--GVDVA-NAAYSIA	T-----	-----	VA--
222 tr G2WHY6 G2WHY6_YEASK	I---	-GL--LGVLFATRFTNAVE	-----LDLD	-----	NYE--
223 tr G0W7G8 G0W7G8_NAUDC	L---	-SIVTTTTTLLLSHSTKALE	-----IDL	D-----	DID--
224 tr G4NGV8 G4NGV8_MAGO7	----	-AAAACLLLQQLPGAVSLN	-----VNFN	-----	DPN--
225 tr W3X855 W3X855_PESFW	----	--YLSICVASVLPLVSAVD	-----IDWN	-----	DDS--
226 tr A0A2N6NY19 A0A2N6NY19_BEABA	----	-ATALL--S-LASGQNTYSIDS	-----	-----	DD--
227 tr J8Q2K7 J8Q2K7_SACAR	I---	-GV--LGVLFARSVA	AVE-----LDLD	-----	NYE--
228 tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	----	--ILS--LSTIISFTQAVF	-----LDTN	-----	NVT--
229 tr J6EGN8 J6EGN8_SACK1	F---	-L-----SFVGISHAMD	-----LDTT	-----	SKT--
230 tr A0A100ISD9 A0A100ISD9_ASPNG	----	-----TFIVVVQ	AID-----YDVD	-----	DPD--
231 tr G8YQC0 G8YQC0_PICSO	----	--VLLLLFLFNSLCFAVD	-----INPG	-----	DVD--

049	sp 074556 YCZ2_SCHPO	--	SVDL	ATSL	VADG	LLNY	YAG	--	QHK	----	GG	--	TIGM	----	FL	----
050	tr A0A0C4E991 A0A0C4E991_MAGP6	--	AIKQ	SAKT	LAYD	MMLM	YPG	--	NKT	----	GQ	--	EPGI	LPGP	PPTEN	-K
051	tr B8MHG0 B8MHG0_TALSN	--	SLKS	AAST	AAAS	AVNY	YNN	--	RES	----	KL	--	IPGK	----	FD	----
052	tr A3LMV8 A3LMV8_PICST	--	SVCD	AAKY	VLDG	TLNY	YEG	--	LKP	----	GG	--	TVGM	----	FA	----
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	--	SVCN	AATD	LDVG	IMDY	YLG	--	TRY	----	AG	--	TVGM	----	FQ	----
054	sp Q9P6I3 YHG7_SCHPO	--	SINS	ALT	TVTD	GMLN	YQS	--	TSH	----	----	--	TF	----	T	----
055	tr Q2US57 Q2US57_ASPOR	--	SIKDA	AAS	TAA	YGMM	HYHG	--	NES	----	GN	--	IPGK	----	LP	----
056	tr W7N622 W7N622_GIBM7	--	SIKQA	AAS	TVAY	GLVK	YTG	--	NNT	----	GD	--	TPGN	----	LP	----
057	sp Q9P6I4 YHG6_SCHPO	--	SIDN	TVN	IMMK	DLMN	YWN	A--	SSQ	----	----	--	AF	----	VA	----
058	tr C7GRB3 C7GRB3_YEAS2	--	SICD	ATAL	IQQG	MLDY	YEG	--	TRY	----	GG	--	TVGM	----	FQ	----
059	tr C7GP28 C7GP28_YEAS2	--	SLQN	ATSL	IAYG	LMDY	YTG	--	NQY	----	GK	--	TVGM	----	FS	----
060	tr G3JGZ5 G3JGZ5_CORMM	--	AIKTT	AST	LAYD	LMLL	YEG	--	NKT	----	GQ	--	IPGI	LPGP	PPSEN	-K
061	tr C4QXV4 C4QXV4_KOMPG	--	SLQH	ATSL	VADG	LMDY	YEG	--	FHL	----	GG	--	TIGM	----	FT	----
062	tr I1S0Z5 I1S0Z5_GIBZE	--	AIKKS	AKQL	AADL	IEY	YHG	--	NDP	----	GG	--	IPGI	LPGP	PPPN	----
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	--	SIKQA	AHH	VAA	NMLSH	YTG	--	MKP	----	GG	--	NPGN	----	LP	----
064	tr G0W5X4 G0W5X4_NAUDC	--	SLEND	TAL	VAYG	LMDY	YNG	--	DQY	----	GQ	--	TVGM	----	FS	----
065	tr G8ZPC1 G8ZPC1_TORDC	--	SICY	ATAL	IQKG	MLDY	YEG	--	TRI	----	GG	--	TVGM	----	FQ	----
066	tr A0A167CDC5 A0A167CDC5_9ASCO	--	SLNN	ALAL	VADG	LMDY	YNG	--	DQY	----	GG	--	TPGM	----	FV	----
067	tr R9XAT7 R9XAT7_ASHAC	--	SIRE	ATSL	VATG	LMDY	YHG	--	HDY	----	GQ	--	TVGK	----	FT	----
068	tr B2B747 B2B747_PODAN	--	DIKQT	SSLL	AWDL	LQY	YKG	--	NLT	----	GQ	--	TPGI	LPGP	PPPA	----
069	tr G0SE99 G0SE99_CHATD	--	DIKET	AAT	VAWD	LQY	YHG	--	NES	----	GQ	--	TPGI	LPGP	PPPA	----
070	tr J8Q6F0 J8Q6F0_SACAR	--	SICD	ATAL	IQDG	MLDY	YEG	--	TRY	----	GG	--	AVGM	----	FQ	----
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	--	SICD	ATSL	IQQG	ILDY	YAG	--	DDY	----	GG	--	AVGM	----	FV	----
072	tr J3K6E2 J3K6E2_COCIM	--	SIKLA	AKIA	AATG	MTSF	YTG	--	HHP	----	GG	--	IPGT	----	LP	----
073	tr G4N3E1 G4N3E1_MAGO7	--	EIKD	STTT	LAYD	LMTL	YPG	--	NQT	----	GK	--	EPGI	LPGP	PPSEN	-K
074	tr C5M5J2 C5M5J2_CANTT	--	SICE	AAK	VVAD	GMWN	YEG	--	FKY	----	GG	--	VVGM	----	FA	----
075	tr A0A254U0X0 A0A254U0X0_ASPNG	--	SLAN	AGST	IADP	LMFY	KQ	--	NQT	----	EG	--	IPGK	----	LT	----
076	sp Q75DG6 DCW1_ASHGO	--	SIRE	ATSL	LATG	LMDY	YHG	--	HDY	----	GE	--	TVGK	----	FS	----
077	tr Q6BZF0 Q6BZF0_DEBHA	--	NLKE	VSA	IFA	EGLD	YDG	--	NEY	----	GG	--	TIGM	----	FT	----
078	tr E9E3Q1 E9E3Q1_METAQ	--	AIKE	SART	LAYD	LMLL	YKG	--	NQS	----	GE	--	IPGI	LPGP	PPADG	-K
079	sp P36091 DCW1_YEAST	--	SLQN	ATSL	IAYG	LMDY	YTG	--	NQY	----	GK	--	TVGM	----	FS	----
080	tr W0TAH6 W0TAH6_KLUMD	--	SLQK	ATSL	VSSG	LMDY	YTG	--	LQK	----	GQ	--	TIGM	----	FA	----
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	--	SICD	ASAL	VIGG	VMDY	YQG	--	TRY	----	GG	--	TVGM	----	FQ	----
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	--	SVLD	AAST	ISYG	LLKF	YTG	--	NNT	----	GD	--	VDGN	----	LP	----
083	tr Q2TYU3 Q2TYU3_ASPOR	--	SIKSA	AAS	QTAY	GSML	WYSG	--	NET	----	GQ	--	IPGA	----	FP	----
084	tr G2XGF6 G2XGF6_VERDV	--	SVKD	AAS	TIAF	GLVS	FYTG	--	NNT	----	GD	--	VPGN	----	LP	----
085	tr G3AMT4 G3AMT4_SPAPN	--	TIRE	DCNL	IAKGL	LDY	YEG	--	TKY	----	GG	--	TIGM	----	FT	----
086	tr A0A1C1D213 A0A1C1D213_9EURO	--	SIKSA	AKIV	ADEM	VTY	YTG	--	YRP	----	GD	--	VPGN	----	LP	----
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	--	----	----	----	----	----	--	----	----	----	--	----	----	----	----
088	tr A0A100I5A6 A0A100I5A6_ASPNG	--	SLAN	AGST	IADP	LMVF	YKQ	--	NQT	----	EG	--	IPGK	----	LT	----
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	--	SICD	ATDL	IQQG	IMDY	YAG	--	SQY	----	GG	--	TVGT	----	FQ	----
090	tr G2Q8A7 G2Q8A7_MYCTT	--	GIKKT	AADV	AWDL	LQY	YHG	--	NES	----	GQ	--	TPGI	LPGP	PPPN	----
091	tr B8NPZ1 B8NPZ1_ASPFN	--	SLKQ	AGKA	VAA	PMMD	FYKK	--	NET	----	EG	--	IPGK	----	LT	----
092	tr Q7SAB2 Q7SAB2_NEUCR	--	----	----	----	----	----	--	----	----	----	--	----	----	----	----
093	tr A2R8R5 A2R8R5_ASPNC	--	SIKNA	AAS	IAAY	GMMKH	YKG	--	NES	----	GE	--	IPGK	----	IP	----
094	tr C5DHG0 C5DHG0_LACTC	--	SICD	ATSL	IQQG	IMDY	YAG	--	TRI	----	GG	--	TVGM	----	FQ	----
095	tr G0SFA3 G0SFA3_CHATD	--	SVKAA	AKT	IAFG	MTEY	YTG	--	DNP	----	GD	--	VPGN	----	LP	----
096	tr K1WJG5 K1WJG5_MARBU	--	SIKSA	AAS	TIA	YGMM	KYTG	--	NNT	----	GD	--	NPGN	----	LP	----
097	tr H2B005 H2B005_KAZAF	--	SLQN	ATAL	VAYG	LMDY	YTG	--	NQY	----	GK	--	TVGM	----	FS	----
098	tr C5DYD7 C5DYD7_ZYGRC	--	SVCS	ATAL	IQQG	MLDY	YAG	--	DKY	----	GG	--	AIGM	----	FQ	----
099	tr G8YM23 G8YM23_PICSO	--	SIQK	ATSL	IAQG	LLDY	YEG	--	TKY	----	GG	--	TIGM	----	FS	----
100	tr W3X8E3 W3X8E3_PESFW	--	DIKAT	TRTL	AADL	MTY	YKG	--	NQS	----	GQ	--	TPGI	LPGP	PPPA	----
101	tr A0A254U3K7 A0A254U3K7_ASPNG	--	SIKNA	AAS	IAAY	GMMKH	YKG	--	NES	----	GE	--	IPGK	----	IP	----
102	tr Q6CER8 Q6CER8_YARLI	--	TVDK	ALAL	IAEGL	LDY	YQG	--	FTY	----	GG	--	TIGM	----	FV	----
103	tr G2QT05 G2QT05_THITE	--	DIKRT	AADV	AWDL	LQY	YKG	--	NQT	----	GQ	--	TPGI	LPGP	PPPA	----
104	sp Q5ACZ2 DFG5_CANAL	--	SICSA	AKY	VVQG	IWNY	YEG	--	LKY	----	GG	--	TVGM	----	FA	----
105	tr G2WKU6 G2WKU6_YEASK	--	SICD	ATAL	IQQG	MLDY	YEG	--	TRY	----	GG	--	TVGM	----	FQ	----
106	tr A0A0C7N752 A0A0C7N752_9SACH	--	SICA	ATSL	IQQG	IMDY	YAG	--	SQY	----	GG	--	TVGM	----	FQ	----
107	tr W3WN68 W3WN68_PESFW	--	SIKE	GAA	TIAW	GLVK	YTG	--	NNS	----	GD	--	VAGN	----	LP	----
108	tr G2QFS1 G2QFS1_MYCTT	--	EIRE	SART	LAYD	LMLF	YKG	--	NQS	----	GE	--	IPGI	LPGP	PPTEH	-K
109	tr G3J9G4 G3J9G4_CORMM	--	SIKST	AST	LVY	GLMK	YNG	--	NVT	----	GG	--	IPGL	----	LP	----
110	tr Q2UJ03 Q2UJ03_ASPOR	--	SVKR	ASY	AVAN	NMLSH	YTG	--	MNP	----	GD	--	TPGV	----	LP	----
111	tr G8YRT2 G8YRT2_PICSO	--	SVCS	AAK	KVAD	GEFNY	YEG	--	LKK	----	GG	--	TVGM	----	FS	----
112	tr W3X554 W3X554_PESFW	--	SIKS	ASST	LA	FGLL	KYTG	--	NNT	----	GD	--	VPGN	----	LP	----
113	tr Q1K7A8 Q1K7A8_NEUCR	--	SVKD	AAAT	IAYG	MLKY	YTG	--	NNT	----	GD	--	TPGN	----	LP	----
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	--	SIKDA	AAAT	TAYN	MMSN	YTG	--	NQT	----	GQ	--	IPGK	----	LP	----
115	tr A7F3P0 A7F3P0_SCLS1	--	SIKTA	ASD	VAYD	MMKY	YTG	--	NRT	----	GD	--	VPGN	----	LP	----
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	--	SIKDA	AAS	TIA	YGLV	KYNG	--	NLT	----	GG	--	IPGL	----	LP	----
117	tr Q6CIP9 Q6CIP9_KLULA	--	SICT	ATAL	IQQG	ISDY	YEG	--	DVY	----	GG	--	TVGM	----	FV	----
118	tr H2AMQ5 H2AMQ5_KAZAF	--	SLQN	ATAL	VAWG	LMDY	YEG	--	TKY	----	GG	--	TVGM	----	FS	----

119	tr G8JMI3 G8JMI3_ERECY	--SICAATQKIQRGIMDY YWG--STP-----GG--IVGM--FI----
120	tr C5DGT1 C5DGT1_LACTC	--SIKNATALVAGGLLDY YTG--QQY-----GQ--TIGM--FS-----
121	tr K1XJR4 K1XJR4_MARBU	--SIKDGAAATLAYDMMSFYKG--NQS-----GG--IIGVLPGP PPNP-P
122	tr A0A124BYC5 A0A124BYC5_ASPNG	--SIKDAAATAAFNAMSYYTG--NQT-----GQ--IPGY--IN-----
123	tr A7TLL2 A7TLL2_VANPO	--SLRNATSLVAYGLMDY YNG--LQY-----GQ--TIGM--FS-----
124	tr Q0CB86 Q0CB86_ASPTN	--SVKSAASEAVYGAMKWYSG--NDT-----GG--IPGA--FP-----
125	tr K1XLH0 K1XLH0_MARBU	--SIIDAAGTVAYDMMTY YTG--NRT-----GD--TPGN--LP-----
126	tr J7S438 J7S438_KAZNA	--SIRNATSLIAYGLMDY YTG--EQY-----GK--TVGM--FA-----
127	tr Q4WG09 Q4WG09_ASPFU	--SLKSAAKTVAAPMMDFYDK--NQT-----EG--IPGK--LT-----
128	tr H8WZ09 H8WZ09_CANO9	--SICSAAKIVSDGMWNY YEG--FKH-----GG--TVGM--FA-----
129	tr I1RL09 I1RL09_GIBZE	--EIKQSARTLAYDLMLQYDG--NTT-----GM--IPGILPG PPTYEY-K
130	tr Q4W985 Q4W985_ASPFU	--SIKQAAHHVAANMLSHYTG--MKP-----GD--NPGN--LP-----
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	--SICDATA LIQDGM LDY YEG--TRY-----GG--AVGM--FQ-----
132	tr W6QCW3 W6QCW3_PENRF	--SIKKA AKQVANNMMSY YTG--MNP-----GD--NPGN--LP-----
133	tr W3WMD3 W3WMD3_PESFW	--SIKQAASQVAEDLLTFYRG--DEP-----GW--VPGILPG PPPD----
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	--TLQNATALVAKGLMDY YTG--NQY-----GQ--TIGM--FS-----
135	tr I2GYH1 I2GYH1_TETBL	--SICGATSLI IKGMLDY YWG--TQY-----GG--TVGM--FQ-----
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	--SVCAATALIQQGMLDY YAG--DKY-----GG--AVGM--FQ-----
137	tr Q752P3 Q752P3_ASHGO	--DVCHFTNEIQKGIMDY YWG--SKY-----GG--IVGM--FQ-----
138	tr B8MYP3 B8MYP3_ASPFN	--SIKSAAATAAYNMMSY YHG--NES-----GQ--TPGK--LP-----
139	tr Q2TXL6 Q2TXL6_ASPOR	--SIKDAAATAAYGMMTY YHG--NES-----GQ--IPGK--LP-----
140	tr Q0CN17 Q0CN17_ASPTN	--SIKTAAATTAFNMMSQYHG--NES-----GQ--IPGK--LP-----
141	tr J3NHD5 J3NHD5_GAGT3	--DIKSTASLVAADLMTY YKG--NLP-----GQ--VPGILPG PPPA----
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	--SVKSAASSIAFDLMSY YTG--NQT-----GQYNIPGLLEQPP-----
143	tr A1CVB0 A1CVB0_NEOFI	--SIKQAAHDVAANMLSHYTG--MKP-----GD--NPGN--LP-----
144	tr B8NVI3 B8NVI3_ASPFN	--SIKDAAATAAYGMMTY YHG--NES-----GQ--IPGK--LP-----
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	--ATYNATALVAQGLLDY YTG--LQY-----GK--PVG M--FS-----
146	tr A1DJ54 A1DJ54_NEOFI	--SLKSAAKTVAAPMMEFYDK--NQT-----EG--IPGK--LT-----
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	--NIRASAKTLAFDLMKFYNG--NQS-----GQ--IPGILPG PPSDG-K
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	--SLKNVTSLIADGLMDY YTG--DQY-----GQ--TIGM--FS-----
149	tr A0A177A618 A0A177A618_9PEZI	--SIKSATGEVAKKLMSFYTG--NKP-----GD--TPGY--LP-----
150	tr Q75CW6 Q75CW6_ASHGO	--SICDATKLIQVGILDY YEG--LRP-----GG--RVGF--FQ-----
151	tr Q7S4K4 Q7S4K4_NEUCR	--NIRASAKTLAFDLMKFYNG--NQS-----GQ--IPGILPG PPSDG-K
152	tr G2QB99 G2QB99_MYCTT	--SVKSAASTIAFGLVKY YTG--NNT-----GD--TPGN--LP-----
153	tr G0S3F2 G0S3F2_CHATD	--EILESARTLAYDMMLFYKG--NQS-----GE--IPGILPG PPTHEH-K
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	--NIKQNAALVADGLMNY YDG--YRR-----GG--TIGK--FI-----
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	--SICAATALIQQGMLDY YEG--TRY-----GG--TVGM--FQ-----
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	--SIKNAASTAAYGAMSYYHG--NES-----GQ--IPGA--FP-----
157	tr G2QKJ0 G2QKJ0_MYCTT	--SIKSAAKIVAKNLYSY YHG--NEP-----GQ--TPGILPG PPP-----G
158	tr Q4WFX5 Q4WFX5_ASPFU	--NIKEVASQLAWDLVSFYTG--NNT-----GD--VPGN--LP-----
159	tr Q5BGD7 Q5BGD7_EMENI	--SLKEAGKTITGPMWEY YLA--NQT-----EG--IPGK--LT-----
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	--SLQNATALVAYGLMDY YTG--LQY-----GE--AVGM--FA-----
161	tr G8BYN9 G8BYN9_TETPH	--SVCEATALIQKGMLDY YQG--TRY-----GG--TVGM--FQ-----
162	tr F7VVT4 F7VVT4_SORMK	--SIKTAAKTVAKNLISY YSG--DKP-----GQ--TIGILPG PPP-----A
163	tr G8YB80 G8YB80_PICSO	--SIQNATALIAKGLLDY YEG--TKY-----GG--TIGM--FT-----
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	--SICDAAALIQDGVLDY YEG--TRY-----GG--AVGM--FV-----
165	tr G2WRS7 G2WRS7_VERDV	--AIRSAAKTLAHDLVAY YHG--NET-----GQ--TPGILPG PPPA----
166	tr A0A124BVB5 A0A124BVB5_ASPNG	--SIKSVASDLAWDLVSFYTG--NNT-----GD--VPGN--LP-----
167	tr A7EIG7 A7EIG7_SCLS1	--SIKSAASTIAYDMMTY YTG--NQT-----GG--IPGN--LP-----
168	tr C5M5U7 C5M5U7_CANTT	--TIREDCNIIAKGLLDY YEG--TTY-----GG--TIGM--FT-----
169	tr B9WAA8 B9WAA8_CANDC	--SVCSAAKVVVQGVWNY YEG--LKN-----GG--TVGM--FA-----
170	tr F8MRU1 F8MRU1_NEUT8	--NIRASAKTLAFDLMKFYNG--NQS-----GQ--IPGILPG PPSDG-K
171	tr A1DJS0 A1DJS0_NEOFI	--SIKNAASTAAYGAMSYYHG--NES-----GQ--IPGA--FP-----
172	tr B6HLM8 B6HLM8_PENRW	--SIKDAGSTATYNMMTWYKQ--NGT-----D--NPGF--IS-----
173	tr A0A117DWS0 A0A117DWS0_ASPNG	--SIKDAARTTAFNMMSY YHG--NES-----GQ--TPGK--LP-----
174	tr A0A124BXM1 A0A124BXM1_ASPNG	--SIKDAASKTAYGSMTWYHG--NET-----GQ--IPGA--FP-----
175	tr J3NZQ6 J3NZQ6_GAGT3	--EVKQSAKDLAWHVM TLYPG--NQT-----GK--EVGIFPG PPSSEG-K
176	tr W0THH8 W0THH8_KLUMD	--SICSATSLIQQGIVDY YDG--YKY-----GG--SVGM--FT-----
177	tr B2AEF2 B2AEF2_PODAN	--EIKESAATLAYDLMLY YKG--NQS-----GE--IPGILPG PPTHEH-K
178	tr Q96TX1 Q96TX1_NEUCS	--DIKKT SADIAFDMMQY YKG--NLT-----GQ--TPGILPG PPPA----
179	tr E9DYA6 E9DYA6_METAQ	--AIKESASILAWDMLQHYHG--NES-----GG--TPGILPG PPPA----
180	tr B9WAI2 B9WAI2_CANDC	--TIREDCNIIAKGLLDY YDG--NKY-----GG--VIGM--FS-----
181	tr Q6C171 Q6C171_YARLI	--TVHEALALVAGGLMDY YDG--TRP-----GG--TVGM--FV-----
182	tr A6ZMV1 A6ZMV1_YEAS7	--SICDATA LIQGGMLDY YEG--TRY-----GG--TVGM--FQ-----
183	tr B6K7X7 B6K7X7_SCHJY	--SILDGLNIVKSGLMNY YDS--ST-------T--VF-----
184	tr I1RSA2 I1RSA2_GIBZE	--TIESAAAKAAKGLTSWYTG--MNP-----GD--TPGN--LP-----
185	tr F7VWZ9 F7VWZ9_SORMK	--SVKDAAATIAYGMMSY YTG--NQT-----GD--TPGN--LP-----
186	tr A0A1B2J789 A0A1B2J789_PICPA	--SLQHATSLVADGLMDY YEG--FHL-----GG--TIGM--FT-----
187	tr G0V8N4 G0V8N4_NAUCC	--SLQNATSLVAYGLMDY YTG--DQY-----GQ--TVGM--FS-----
188	tr H8WXN8 H8WXN8_CANO9	--SVREGCNLIAQGLLDY YEG--TKY-----GG--TIGE--FS-----

189	tr A6ZZS0 A6ZZS0_YEAS7	--SLQ	NATSLIAYGLMDY	YTG	--NQY	--GK	--TVGM	--FS	--	--
190	tr G9MQD1 G9MQD1_HYPVG	--NIRD	SARGLAYDLMLQ	YEG	--NKT	--GQ	--IPGK	--LP	GP	PPSEN-K
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	--SIYN	NATALVAEGLLDY	YTG	--LQY	--GQ	--PIGM	--FS	--	--
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	--SLKS	AAKTVAAPMMDF	YDK	--NQT	--EG	--IPGK	--LT	--	--
193	tr A0A254UET1 A0A254UET1_ASPNG	--SVKS	VASELAWDLVSFY	TG	--NNT	--GD	--VPGN	--LP	--	--
194	tr A1D587 A1D587_NEOFI	--SIKD	AAATTAYNMMSNY	TG	--NQT	--GQ	--IPGK	--LP	--	--
195	tr Q1K7I4 Q1K7I4_NEUCR	--DIKK	TSADIAFDMMQY	YKG	--NLT	--GQ	--TPGILPG	PPPA	--	--
196	tr G8JS88 G8JS88_ERECY	--SMQS	ATSLISTGVLDY	YTG	--LDP	--GN	--TIGM	--FS	--	--
197	tr A0A136J7D3 A0A136J7D3_9PEZI	--SIKAA	ASTVAFGLARFY	TG	--NNT	--GD	--VPGN	--LP	--	--
198	tr E9E4X8 E9E4X8_METAQ	--AIKQ	SSSTLAWDMLQY	YKG	--NLT	--GQ	--TPGILPG	PPPA	--	--
199	tr Q6CP42 Q6CP42_KLULA	--SLKN	ATSLVAYGLMDY	YTG	--LQY	--GK	--TIGM	--FA	--	--
200	tr G0WHL4 G0WHL4_NAUDC	--SICD	ATSLIQGGMDY	YWG	--TQY	--GG	--TVGM	--FQ	--	--
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	--SLQN	ATSLVAYGLMDY	YTG	--NQY	--GK	--TVGM	--FS	--	--
202	tr F9WYX6 F9WYX6_ZYMTI	--SVLK	ASKIVVDNILSV	YNNYTESP	--	--GG	--IPGL	--LP	--	--
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	--DIKT	TAASIAADLMAY	YKG	--NLP	--GQ	--VPGILPG	PPPA	--	--
204	tr J3P147 J3P147_GAGT3	--QIIQ	SSRDLAWEVMAL	YPG	--NQT	--GQ	--ETGILPG	PPDDG	-K	
205	tr I2H842 I2H842_TETBL	--SLDN	AALVAYGLMDY	YNG	--NQY	--GQ	--TIGM	--FS	--	--
206	tr S6E3R3 S6E3R3_ZYGB2	--SVCA	AATALIQQGMLDY	YAG	--DKY	--GG	--AVGM	--FQ	--	--
207	tr E5AD94 E5AD94_LEPMJ	--SIKD	VSSKVAKQLVAQ	YAK	--IDDKGVHVL	GG	--YPGI	--LY	--	--
208	tr Q6FJM7 Q6FJM7_CANGA	--SICA	AATALIQQGMLDY	YEG	--TRY	--GG	--TVGM	--FQ	--	--
209	tr A5DUW0 A5DUW0_LODEL	--TIRE	DICALIADGILDY	YDG	--YNY	--GG	--TIGE	--FV	--	--
210	tr G8BF46 G8BF46_CANPC	--SVRE	ACNLIAKGLLDY	YEG	--TKY	--GG	--TIGE	--FT	--	--
211	tr W3XAF6 W3XAF6_PESFW	--SVKE	AAATIAWGLVKY	YTG	--NNT	--GD	--VPGN	--LP	--	--
212	tr B6H7I2 B6H7I2_PENRW	--SIKD	AASTSTYSMMGW	YYG	--NET	--GQ	--IPGS	--FP	--	--
213	tr Q5ATF9 Q5ATF9_EMENI	--SIKD	AASKTAYGSLWY	YSG	--NET	--GG	--NPGA	--FP	--	--
214	tr A0A167CDD3 A0A167CDD3_9ASCO	--SIYG	AQALVAQGLMDY	YNG	--NQT	--GQ	--TPGM	--FS	--	--
215	tr G0RXS3 G0RXS3_CHATD	--SIRRA	AKIVAGNLWTY	YHG	--DKP	--GQ	--TPGILPG	PHPAVPS		
216	tr B8NX26 B8NX26_ASPFN	--SLKK	GAEIIPPMMEFY	KE	--NQT	--EG	--IPGK	--LT	--	--
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	--SIKS	AAKQIAGGMVKY	YTG	--YKP	--GD	--VPGN	--LP	--	--
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	--NIKE	VASQLAWDLVSFY	TG	--NNT	--GD	--VPGN	--LP	--	--
219	tr A1CMM3 A1CMM3_ASPCL	--		Q	--GRP	--GD	--VPGN	--LP	--	--
220	tr B6HU84 B6HU84_PENRW	SDSLKS	AAKTVAATTMMEY	YDA	--RES	--KD	--IPGK	--FD	--	--
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	--DIKK	TSADIAFDMMQY	YKG	--NLT	--GQ	--TPGILPG	PPPA	--	--
222	tr G2WHY6 G2WHY6_YEASK	--SLQN	NATSLIAYGLMDY	YTG	--NQY	--GK	--TVGM	--FS	--	--
223	tr G0W7G8 G0W7G8_NAUDC	--SLRN	NATSLVAYGLMDY	YTG	--NQY	--GK	--TIGM	--FS	--	--
224	tr G4NGV8 G4NGV8_MAGO7	--SVKS	AASTVAFGMMKY	YTG	--NNT	--GD	--TPGN	--LP	--	--
225	tr W3X855 W3X855_PESFW	--SIKD	GAAQIAYGLLQY	YTG	--NNT	--GD	--VPGN	--LP	--	--
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	--AIKK	TASQMAEDLLKH	YHG	--DEP	--GQ	--VPGILPG	PPPA	--	--
227	tr J8Q2K7 J8Q2K7_SACAR	--SLQN	NATSLVAYGLMDY	YTG	--NQY	--GE	--TVGM	--FS	--	--
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	--NIEQ	NTALIAKGLLDY	YEG	--EKY	--GG	--TIGM	--FS	--	--
229	tr J6EGN8 J6EGN8_SACK1	--SICD	ATALIQDGMLDY	YDG	--TRY	--GG	--AVGM	--FQ	--	--
230	tr A0A100ISD9 A0A100ISD9_ASPNG	--SIKA	ACHSIARQMLTHY	TG	--NQP	--GD	--NPGN	--LP	--	--
231	tr G8YQC0 G8YQC0_PICSO	--SVCS	AAKKVADGEFNY	YEG	--IKK	--GG	--TVGM	--FT	--	--
232	tr F7VVP8 F7VVP8_SORMK	--NIRAS	AKTLAYDTMLFY	YKG	--NQS	--GE	--IPGILPG	PPSDG	-K	
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	--SICD	ATALIQGGMLDY	YEG	--TRY	--GG	--TVGM	--FQ	--	--
234	tr F8MBP4 F8MBP4_NEUT8	--SVKD	AAATIAYGMLKY	YTG	--NNT	--GD	--TPGN	--LP	--	--
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	--SIKN	AAATAAFNAMS	YTG	--NQT	--GQ	--IPGK	--IN	--	--
236	tr A1CD27 A1CD27_ASPCL	--TLKS	SAGKDLAAPMMDF	YKK	--NQT	--EG	--IPGK	--LT	--	--
237	tr A5DHF3 A5DHF3_PICGU	--SICE	AASYVANGELNY	YEG	--LKY	--GG	--TVGM	--FA	--	--
238	tr D4ATT7 D4ATT7_ARTBC	--SIKKA	ASHIAHGMVSY	YTG	--NHT	--GD	--VPGN	--LP	--	--
239	tr E2PT42 E2PT42_ASPNC	--SIKA	ACHSVARQMLTHY	TG	--NQP	--GD	--NPGN	--LP	--	--
240	tr C7ZPE5 C7ZPE5_NECH7	--DIKK	TAKQLAADLIEY	YHG	--LDP	--GG	--IPGILPG	PPPN	--	--
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	--SIKS	AAKTLARGCKSY	YTG	--DNP	--GD	--VPGN	--LP	--	--
242	tr Q4WKP7 Q4WKP7_ASPFU	--SIKD	AAATTAYNMMSNY	TG	--NQT	--GQ	--IPGK	--LP	--	--
243	tr F8MXT4 F8MXT4_NEUT8	--DIKK	TSADIAFDMMQY	YKG	--NLT	--GQ	--TPGILPG	PPPA	--	--
244	tr A0A177AJ74 A0A177AJ74_9PEZI	--SVKT	AASSIAFDLMSY	YTG	--NQS	--GQYN	IPGLLEQP	PP	--	--
245	tr J7RQR5 J7RQR5_KAZNA	--PVC	SATALVTDGLLDY	YSG	--TRY	--GG	--TVGM	--FQ	--	--
246	tr Q75CW7 Q75CW7_ASHGO	--SICD	ATKLIQVGILDY	YEG	--AKP	--GG	--RAGF	--LV	--	--
247	tr G3AKK4 G3AKK4_SPAPN	--STCA	AAKVVADGIWNY	YEG	--LKY	--GG	--VVG	--FA	--	--
248	tr B2W762 B2W762_PYRTR	--SIKKA	AKDISINLRQY	YTG	--DRP	--GD	--TPGN	--LP	--	--
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	--SLQN	NATSLIAYGLMDY	YTG	--NQY	--GK	--TVGM	--FS	--	--
250	tr C4XWK1 C4XWK1_CLAL4	--SICE	AAWEVIVGELNY	YEG	--TKY	--GG	--TVGM	--FS	--	--

001	tr A0A254U2J9 A0A254U2J9_ASPNG	S	-	-	K	W	E	G	S	A	L	F	T	S	L	L	L	Y	W	Y	Y	T	-	-	G	D	S	T	Y	N	D	E	V	R	Q	G	M	Q	W	Q	A	G	-	-	D	-	C	D	Y	M	
002	sp Q6FLP9 DCW1_CANGA	D	P	Y	Y	W	W	Q	A	G	G	A	W	G	C	M	L	D	Y	W	Y	F	M	-	-	Q	N	D	T	Y	N	D	K	I	M	A	A	L	L	H	Q	T	G	-	D	N	-	N	D	Y	V
003	tr A1CCM5 A1CCM5_ASPCL	T	-	-	K	W	E	G	S	A	L	F	M	A	M	L	Q	Y	W	Y	F	T	-	-	G	D	S	T	Y	N	S	A	V	S	E	G	L	E	W	Q	A	G	-	-	P	-	G	D	Y	M	
004	tr C1H190 C1H190_PARBA	D	P	Y	Y	W	W	E	A	G	A	M	F	G	A	L	I	D	Y	W	Y	Y	T	-	-	G	D	D	Q	Y	N	E	I	V	M	Q	A	M	L	H	Q	T	G	-	P	D	-	F	D	Y	Q
005	tr Q2UR85 Q2UR85_ASPOR	D	-	-	T	W	E	G	G	A	M	F	M	T	L	I	Q	Y	W	F	W	T	-	-	G	D	T	S	Y	N	E	V	T	T	Q	G	M	L	W	Q	K	G	-	H	D	-	-	D	Y	F	

006	tr A5DV30 A5DV30_LODEL	SPNYWWNAGEAFGGGLLDYYIFCDPEN	EQLESRIFFDGMYPH	QAG-EN-YNYI	
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	QPYWWWESALVFGGMIDTWKIC--NN	YTYVSTIQSAISH	QKG-DS-NDFY	
008	tr J3PGN0 J3PGN0_GAGT3	GDYYWWQGGALMGTMIDYWHLT--GD	TEYNKIIISEGILN	QVG-EG-RDFQ	
009	tr A0A124BXU6 A0A124BXU6_ASPNG	N--TWWE	GAMFMTLMQYYHYT--GD	STYNQEV	IQGMQWQSG--D-CDFM
010	tr C5P4A1 C5P4A1_COCP7	LPYYWWEAGAMFGALIDYWFYT--GD	DTWNEMTTTALLH	QAS-ST-KNFM	
011	tr Q0CG55 Q0CG55_ASPTN	DPYYWWEAGAMFNSLVNYWYYT--GD	DTWNDITTTQAILW	QAG-DT-GTFM	
012	tr C4Y2X5 C4Y2X5_CLAL4	APYYWWESGGAWGSMLEYTIYM--EN	DTYTDLIKEALLY	QVG-DD-FNYI	
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	DPYYWWEAGAMFTALVDYWYLT--SD	DTWNNITTTQGITW	QAG-PS-GSFM	
014	tr C7Z068 C7Z068_NECH7	GDYYWWEGGAMMGTYIDYWKL	T--GDSSYNHVIMEGMLH	QTG-DG-HDYM	
015	tr G8BXX2 G8BXX2_TETPH	DPYYWWEAGGAWGCMLDYWFFM--EN	DTYNDIIKQALLY	QVG-TA-NDYV	
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	WPYYWWHAGGAFGSLLDYSYYT--QD	TQYDDLIEQALTY	QVG-EN-YNYI	
017	tr A1CSC2 A1CSC2_ASPL	D--TWWE	GAMFMTLIQYWYWT--GD	PSYNDVVTQGMLW	QKG-ND--DYF
018	sp Q05031 DFG5_YEAST	SPYYWWHAGEAFGGMLENWFLC--EN	DTYQELLYDALLA	QTG-SN-YDYI	
019	tr H2AQJ6 H2AQJ6_KAZAF	SPYYWWEAGEAFGGMLENWFLC--EN	DTFKEIIYDAILA	QTG-SD-YDFM	
020	tr G2XFH9 G2XFH9_VERDV	GDYYWWQGGAMMGTYVDYWHLT--GD	TSYNDVVREGMVH	QVG-DN-RDYM	
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	WPYYWWEAGGAWNTLIDFWFYM--KN	DTYNSIVQQALVY	QSG-EN-HDYV	
022	sp Q5AD78 DCW1_CANAL	WPYYWWEAGGAWGSLIDYTFYF--DN	DTLVPLITDALLY	QTG-DD-DNYI	
023	tr B6H7E8 B6H7E8_PENRW	G--TWWE	GSMFMTLIQYWYLT--GD	SQFNDAIQEGMYW	QKG-DD-NDFF
024	tr B6GZU2 B6GZU2_PENRW	DPYYWWEAGAMFNSLIDYWFYT--GD	DTWNDITTIQAMLW	QAG-EN-KAFM	
025	tr E9E4W7 E9E4W7_METAQ	GDYYWWEAGAMMGTYIDYWKL	T--GDATYNDVVTQGMLF	QVG-PQ-RDYM	
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	DPYYWWEAGAMFNVLIDYWYFT--GD	DTYNDITMQGMLW	QAG-DN-GAFM	
027	tr G2R7G8 G2R7G8_THITE	GPYYWWQAGAMWGTYIDYWFYT--GD	STYNDETVRAMEF	QSE---NSYQ	
028	tr G3JBY1 G3JBY1_CORMM	GPYYWWEAGGMFGHLIDYWYYT--GD	TTYNEIVMQGMMH	QINAPT-GDFL	
029	tr A5DNV5 A5DNV5_PICGU	WPYYWWEAGGAWGSLIEYSYYM--DN	DTLVPLIKEALGY	QVG-EN-YNYV	
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	YPYYWWECEGAMFGSLIDYWLYT--GD	SAHNDLVTTQGMQF	QVG-PN-NDFM	
031	tr Q2TWC1 Q2TWC1_ASPOR	D--TWYI	AGAMFMTLIQYWATS--GVE	QYNKVVS	HDLMFQSG-EN-YDYF
032	tr J5RXY9 J5RXY9_SACK1	DPYYWWEAGGAWGCMLDYWFFM--DN	DTYNDEIIAAMVH	QSG-DD-NDYI	
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	GPYYWWEAGGMFGHLIDYWYYT--GD	ASNAVTMEGMMH	QIT-PK-GDFL	
034	tr A0A254U5V1 A0A254U5V1_ASPNG	D--TWWE	GAMFMTLIQYWYWT--GD	TSYNEVTTQGMLW	QKG-DN--DYF
035	tr A0A061B6H7 A0A061B6H7_CYBFA	NPYYWWEAGGAWGSMIDFWYYM--EN	DTYLNQTYDALLA	QVG-DN-WDYV	
036	tr G8ZQ93 G8ZQ93_TORDC	DPYYWWEAGGAWGCMLDYWWYM--QN	DTYNDIIMQALLY	QVG-TH-NDYV	
037	tr Q9C2J1 Q9C2J1_NEUCS	DPYYWWEAGAMFGAMVDYWWVT--GD	TSYVEVTTQAI	IVHQAG-DA-RDFN	
038	tr C5NZK5 C5NZK5_COCP7	DPYYWWEAGAMFSALIDYWYYT--GD	DQYNDITTTQAMLH	QVG-KE-NNYM	
039	tr Q6C0T7 Q6C0T7_YARLI	QPYWWHAGAAWNAMIEYWHIT--GN	SSLNDITYEALLH	QRG-ER-FDLM	
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	DPYYWWQAGGAWGCMLDYWFYM--QN	DTYNDKIMAALLH	QTG-DN-NDYV	
041	tr F7VZ72 F7VZ72_SORMK	GDYYWWEAGAMWGTLIDYWRFT--GD	ESYNEVTTQALLW	QVG-PG-QDYM	
042	tr A3LN37 A3LN37_PICST	WPYYWWEAGGVWGS	LIDYTYFT--QN	DTLVPLIKEALLY	QTG-DD-NNYV
043	tr Q6CAI2 Q6CAI2_YARLI	FPYYWWEAGAAWNSMLDFWYYT--GN	DTYNDVVKEALLY	QVG-KN-NDYL	
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	WPYYWWEAGGAWGSLIDYTYYM--EN	DTFVPMIKEALEY	QTG-DD-LNYI	
045	tr G9MJS5 G9MJS5_HYPVG	GDYYWWEAGAMWGTLIDYWYLT--ND	STYNDLIMQAIQW	QTG-PD-DDFQ	
046	tr G9MHI4 G9MHI4_HYPVG	DPYYWWEAGAMFGTMIDYWRMT--KD	SSYNTATMQAMLW	QAG-TD-GAFL	
047	tr D4AP27 D4AP27_ARTBC	DPYFWWETGAMFGGLIDYWFYT--GD	SQFNDIVTTQGMLW	QVG-PD-SNFM	
048	tr B6GZT8 B6GZT8_PENRW	D--KWE	GAPLFMALLEYWHFT--GD	TTYNEELSVGLQW	QGG-TD-GDYM
049	sp O74556 YCZ2_SCHPO	PPAYWWEAGAAWNGLLNRYIAT--GN	STYNELVKTSMLY	QSG-ED-SDYM	
050	tr A0A0C4E991 A0A0C4E991_MAGP6	GDYYWWQGGALMGTMIDYWHLT--GD	TEYNKIIITEGILN	QVG-EG-RDFQ	
051	tr B8MHG0 B8MHG0_TALSN	G--TWWE	GGAFFTFLINYHWT--GD	DQYNNLVAEGLSW	QGG-ED-NDFF
052	tr A3LMV8 A3LMV8_PICST	PPNYWWNAGEAFGGIVDFYTF	CNSTNETLRELIIDAMYH	QAG-EN-FNYI	
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	QPYWWWEAGLVFGGMIDTWKFC--NN	DTYVGIIQEAI	IVHQKG-PD-NDFF	
054	sp Q9P6I3 YHG7_SCHPO	A--YWWMT	GAGLNSMTDTYAAT--GN	TTHLDMLISALVANKG	-DN-NDYA
055	tr Q2US57 Q2US57_ASPOR	D--TWWE	GAMFMTLIEYWHFT--GD	ATYNDEVSEGMQW	QAG--D-GDYM
056	tr W7N622 W7N622_GIBM7	DPYYWWTAGGMFGTLIDYWWT--GD	ESYNKITTTQAMLH	QVG-TN-DDYM	
057	sp Q9P6I4 YHG6_SCHPO	S--YWWVT	GATMGALLNYELF--NN	DTYVDLISSSLLYNAG	-SG-FDYQ
058	tr C7GRB3 C7GRB3_YEAS2	SPYYWWHAGEAFGGMLENWFLC--EN	DTYQELLYDALLA	QTG-SN-YDYI	
059	tr C7GP28 C7GP28_YEAS2	DPYYWWEAGGAWGCMLDYWFFM--DN	DTYNDEIIAAMI	HQAG-DD-NDYI	
060	tr G3JGZ5 G3JGZ5_CORMM	GNYYWWQGGAMMGTYIDYWHLT--GD	TSYNDVVMQGILH	QVG-EN-RDFM	
061	tr C4QXV4 C4QXV4_KOMPG	NPYYWWQSGAAFGSMLDYWWYM--EN	DTYHDAIMQAI	IVYQAG-DN-ADFM	
062	tr I1S0Z5 I1S0Z5_GIBZE	GDYYWWEAGAMWGTYMDYWRCT--GD	TKYNDLVMQGMQF	QVG-DD-QDYQ	
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	PPYYWWEAGAMFNALIDYWYLT--GD	STWNAITTTQALTW	QAG-HT-GTFM	
064	tr G0W5X4 G0W5X4_NAUDC	NPYYWWEAGGAWGTMLDYWFFM--EN	DTYNDLIMSALLH	QTG-EN-NDYI	
065	tr G8ZPC1 G8ZPC1_TORDC	SPYYWWQAGEAFGGMIDNWYFC--EN	TTFESLIYDGMLA	QTG-PH-YDYM	
066	tr A0A167CDC5 A0A167CDC5_9ASCO	NPYYWWEAGAAFGSMLDYWFYT--GN	TTYNDVIKAGMLY	QTG-KN-NDYM	
067	tr R9XAT7 R9XAT7_ASHAC	DPYYWWEAGGAWGSMLDYWYYM--EN	STYNNLLTDSL	LLHQAG-ED-LAYT	
068	tr B2B747 B2B747_PODAN	GDYYWWEAGAMWGTLIDYWKFT--GD	DSYNDLITQAMLW	QVG-PD-KDYM	
069	tr G0SE99 G0SE99_CHATD	GDYYWWEAGAMWGTLIDYHWT--GD	SYNDVITQAMLW	QVG-PN-RDYM	
070	tr J8Q6F0 J8Q6F0_SACAR	SPYYWWHAGEAFGGMIENWFLC--EN	TYKELLYDALLA	QTG-SD-YDYI	
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	SPYYWWEAGLVFGGMIENWYLC--EN	TTYEDMLYEALMA	QTG-PD-YDYM	
072	tr J3K6E2 J3K6E2_COCIM	LPYYWWEAGAMFGALIDYWFYT--GD	DTWNEMTTTALLH	QAS-ST-KNFM	
073	tr G4N3E1 G4N3E1_MAGO7	GDYYWWEAGALMGTMIDYWHFT--GH	TEYNDEVTTQGI	LYQVG-EN-ADFQ	
074	tr C5M5J2 C5M5J2_CANTT	APNYWWNAGEAFGGGLVDYYTF	CDPDNSTLEGWIYDAMYH	QAG-EN-YNYI	
075	tr A0A254U0X0 A0A254U0X0_ASPNG	D--TWYI	AGAMFMTLIQFWKAS--GV	DTYNSVVQHDLMF	QAG-EN-YDYF

076	sp Q75DG6 DCW1_ASHGO	DPYYWWEAGGAWGSLIDYWYYM--ENSTYNDLLTDSLHQAQ-ED-LSYT
077	tr Q6BZF0 Q6BZF0_DEBHA	WPYYWWEAGGAWGSLIDYTYYM--ENDTFVPLIKQALEYQTG-DD-LNYI
078	tr E9E3Q1 E9E3Q1_METAQ	GPYYWWEAGGAMMGTYIDYWKLT--GDSYINKVVMEGMLHQVG-EN-KNYM
079	sp P36091 DCW1_YEAST	DPYYWWEAGGAWGCMLDYWFFM--DNDTYNDEIIAAMIHQAG-DD-NDYI
080	tr W0TAH6 W0TAH6_KLUMD	SPSYWWQAGGAWGSLIDYWYYM--QNDTYNSILTQALLYQTG-DN-NDYM
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	DPYYWWEAGEAMGSM LGFWYFC--NNDTYEDVIYDALMAQRG-SN-NDYI
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	SPYYWWEAGAMFGTLVDYWWT--NDTSNNDAVKRALMHQVG-GD-NNYM
083	tr Q2TYU3 Q2TYU3_ASPOR	D--KWWEGSALFLSLLYYWHYT--GDTTYNAEVSQGM EWQAG--N-GDYM
084	tr G2XGF6 G2XGF6_VERDV	DPYFWWTAGAMFGTLVDYWFLT--GDDTYNEITTQAMVHQAG-DE-ADFM
085	tr G3AMT4 G3AMT4_SPAPN	PPYYWWEAGGAWGGLIDYTFFF--ENDTLVPLIKA ALEYQTG-DD-NNYV
086	tr A0A1C1D213 A0A1C1D213_9EURO	APYYWWQAGAMFGSLIDYWYFT--GDASYNEITTQAMLHQVG-PD-NDFM
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	-- -- -- -- --MDYWHYT--GDSYNELITSSMLFHTG-PPLNAYM
088	tr A0A100I5A6 A0A100I5A6_ASPNG	D--TWYIAGAMFMTLIQFWQAS--GVDTYNSV VQHDLMFQAG-EN-YDYF
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	EPYYWWEAGAVFGGML ENWFLC--QNDTYEQVIYDSMIAQAG-PD-YDFM
090	tr G2Q8A7 G2Q8A7_MYCTT	GDYYWWEAGAMWGTFIDYWKLT--GDSTYNDLVTQAMLFQVG-PD-RDYR
091	tr B8NPZ1 B8NPZ1_ASPFN	D--TWYVAGSMFMTLIQYWQAS--GDDTYNAVVSNDLMFQAG-EN-YDYY
092	tr Q7SAB2 Q7SAB2_NEUCR	-- -- -- -- --MDYWHYT--GDSYNELITSSMLFHTG-PPLNAYM
093	tr A2R8R5 A2R8R5_ASPNC	N--TWWEGGAMFMTLMQYYHYT--GDSTYNKEVIQGMQWQAG--D-CDYM
094	tr C5DHG0 C5DHG0_LACTC	QPYYWWEAGEVFGGML ENWFLC--QNDTYHDI IYDAMIHQAG-PD-YDFM
095	tr G0SFA3 G0SFA3_CHATD	DPYFWWEAGAMFGMLVEYWALT--KDDTYNVM TMQALLHQAT-EN-GDFM
096	tr K1WJG5 K1WJG5_MARBU	APYYWWEAGATFGTMVEYWYYT--NDTSYNPTVTAAILSQVG-DD-KDFM
097	tr H2B005 H2B005_KAZAF	DPYYWWEAGGAWGSMIDFWYYM--DNNTYND DIMAALLHQTG-DD-WDYI
098	tr C5DYD7 C5DYD7_ZYGRC	PPYYWWQAGEAFGGMIDNWHFC--KNNTFEELIREAMLTQAG-PH-YDYM
099	tr G8YM23 G8YM23_PICSO	WPYYWWEAGGAWGSLIDYSYYT--ENDTLVP IIKQALTYQTG-ED-DNYI
100	tr W3X8E3 W3X8E3_PESFW	GDYYWWEGGALWGT MIDYWHLT--GDSTYNDVVTQALLWQVG-PN-QDYM
101	tr A0A254U3K7 A0A254U3K7_ASPNG	N--TWWEGGAMFMTLMQYYHYT--GDSTYNKEVIQGMQWQAG--D-CDYM
102	tr Q6CER8 Q6CER8_YARLI	PPYYWWHAGAAWNAMIEYWHIT--GNSTFNNITFEAMKAQRG-DK-YNLM
103	tr G2QT05 G2QT05_THITE	GDYYWWEAGAMWGTLIDYWKLT--GDSTYNDLVTQAMLWQVG-PD-RDYM
104	sp Q5ACZ2 DFG5_CANAL	PPNYWWNAGEAFGG LVD FYTYCQSDNSTLEKLIYNGMYHQAG-EN-YNYI
105	tr G2WKU6 G2WKU6_YEASK	SPYYWWHAGEAFGGML ENWFLC--ENDTYQELLYDALLAQTG-SN-YDYI
106	tr A0A0C7N752 A0A0C7N752_9SACH	APYYWWEAGEVFGGML ENWYLC--QNDSYSQVLYDSMLAQAG-PE-YDYM
107	tr W3WN68 W3WN68_PESFW	DPYYWWEAGAMFGALVDYWAFT--GDDSYNNITFQALQHQVG-DD-ADFM
108	tr G2QFS1 G2QFS1_MYCTT	GDYYWWEGGAMMGTYVDYWFLT--GDPSYNHV VTEGMLHQVG-PN-ADYM
109	tr G3J9G4 G3J9G4_CORMM	GKYYWWEAGGMFGHLIDYWYYT--GDASYNDV TMQGMMHQIT-PK-GDYL
110	tr Q2UJ03 Q2UJ03_ASPOR	PPYYWWEAGAMFNALIDYWFYT--GDDRWNDI IMQAMTWQAG-DD-GTFM
111	tr G8YRT2 G8YRT2_PICSO	YPYYWWHAGAAFGGLLDYYTFCDKDNSTLKKYIYNGMYHQAG-DD-YNYI
112	tr W3X554 W3X554_PESFW	DPYFWWEAGAMFGTMVDYWFFT--GDDTYNAITSQALLHQVG-DD-KDFM
113	tr Q1K7A8 Q1K7A8_NEUCR	DPYYWWEAGAMFGAMVDYWWT--GDTSYVEVTTQAIVHQAG-DA-RDFN
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	D--TWWEGGAMFMTLIQYFWT--GDTSYNEVTTQGMLWQKG-NN--DYF
115	tr A7F3P0 A7F3P0_SCLS1	DPYYWWEAGAMFGIMVDYWYYT--GDETYNEEVKQALLHQAG-ND-YDFM
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	GPYYWWEAGGMFGHLIDYWYYT--GDATYNDIVMQGMMHQINAPA-GDYL
117	tr Q6CIP9 Q6CIP9_KLULA	SPYYWWEAGLVFGGMIENWYLC--QNTSYEDMLYDALMAQTG-SD-YDYM
118	tr H2AMQ5 H2AMQ5_KAZAF	SPYYWWEAGGAFGTLLDFWYFM--DNDTYNDEILAAMVYQAG-ND-WDYI
119	tr G8JMI3 G8JMI3_ERECY	QPYYWWEAGLVFAGML ENWYLC--DNMEYNSTLYESMIAQTG-AN-YDYI
120	tr C5DGT1 C5DGT1_LACTC	NPYYWWEAGGAWGSLIDFWWYT--QNDTYNAMLEEALLYQAG-EN-HDYI
121	tr K1XJR4 K1XJR4_MARBU	DGYYWWEESGAMWGAMIDYWHYT--GDESYN DVVSTGIQAQVG-EN-LDLM
122	tr A0A124BYC5 A0A124BYC5_ASPNG	H--TWWEGGVLFDTMIRYWYFT--NDASNNAAVSQGM YHQRG-AG-NDYM
123	tr A7TLL2 A7TLL2_VANPO	NPYYWWEAGGAWGTMIDYWYYM--QNDTYNDI IMQALQYQTG-VN-NDYV
124	tr Q0CB86 Q0CB86_ASPTN	D--KWWEGSALFLSTLLYWHYT--GDDTYNSVTSQGM EWQAG--N-GDYM
125	tr K1XLH0 K1XLH0_MARBU	DPYYWWEAGAMFGTMINYWYYT--GDTTYNP TTKQALLHQIG-DN-EDFM
126	tr J7S438 J7S438_KAZNA	DPYYWWEAGGAWGSM LDYSWFM--DNDTYDSQIMSAMLHQTG-EH-NNYI
127	tr Q4WG09 Q4WG09_ASPFU	G--TWYVAGAMFMTLIQYWQSS--GDDTYNSIVSHDLMFQSG-EN-YDFF
128	tr H8WZ09 H8WZ09_CANO9	SPNYWWNAGEAFGG LVD FYTYCDSDNSTLEKWIYDGM YHQAG-DN-YNYI
129	tr I1RL09 I1RL09_GIBZE	GDYYWWEGGAMMGTYIDYWKLT--GDSYINKVVMEGMLHQTG-EG-RDYM
130	tr Q4W985 Q4W985_ASPFU	PPYYWWEAGAMFNALIDYWYLT--GDSTWNAIT TQALTWQAG-HT-GTFM
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	SPYYWWHAGEAFGGMV ENWFLC--KNDTYQELLYNALLAQTG-AD-YDYI
132	tr W6QCW3 W6QCW3_PENRF	DPYYWWEAGAMFNALIDYWFYT--GDDTWNNIT IQGMLWQAG-EN-KAFM
133	tr W3WMD3 W3WMD3_PESFW	GDYYWWQGGAMWGTL LDYRHH T--GDKTYDDMTSTAILFQVG-DD-RDFM
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	WPYYWWEAGGAWNTLIDFWFYM--KNDTYNSIVQQALVYQSG-EN-HDYV
135	tr I2GYH1 I2GYH1_TETBL	PPYYWWEAGEAFGGML ENWYLC--QNDTYADLLFDALTAQTG-PD-YDYI
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	SPYYWWQAGEAFGGMLDNWYFC--QNNTFENLIRKAMLAQTG-TK-YDYM
137	tr Q752P3 Q752P3_ASHGO	PPYYWWEAGEVFGGML ENWYLC--DNQEYESVLRESMIAQTG-DK-FDYI
138	tr B8MYP3 B8MYP3_ASPFN	D--TWWEGGAMFMTLIQYFWT--GDTSYNEVTTQGMLWQKG-HD--DYF
139	tr Q2TXL6 Q2TXL6_ASPOR	G--TWWTGGEVFMALVQYWYW T--GDTSYNDVTKQALI WQKG-HN--DYL
140	tr Q0CN17 Q0CN17_ASPTN	D--TWWEGGAMFMTLIQYWYW T--GDTSYNDVTTQGMLWQKG-SN--DYF
141	tr J3NHD5 J3NHD5_GAGT3	GDYYWWEGGALMGQMVEYWHLT--GDTTYNDEVSQGMLHQVG-QD-RDYQ
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	AGYYWWQAGAMWNTMVDYWYIT--GDTSYNEATTQALT FQVG-DD-QDYM
143	tr A1CVB0 A1CVB0_NEOFI	PPYYWWEAGAMFNALIDYWYLT--GDSTWNAIT TQALTWQAG-HT-GTFM
144	tr B8NVI3 B8NVI3_ASPFN	G--TWWTGGEVFMALVQYWYW T--GDTSYNDVTKQALI WQKG-HN--DYL
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	DPYYWWEAGGAWGSLIDYWWT--GNTTYNGILKQALLAQTG-DN-NDYV

tr A1DJ54 A1DJ54_NEOFI	G--TWYVAGAMFMTLIQYWQAS--GD	DTYNSVSVSHDLMFQSG-EN-YDFF
147 tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	GDYYWWEAGGALMGTMIDYWHLT--GD	TTYNDVITQGILHQVG-DN-RDFQ
148 tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	NPYYWWEAGGAWGSMIDYWYYM--EN	DTYLNQTYEALLAQVG-DD-WNYV
149 tr A0A177A618 A0A177A618_9PEZI	DPYYWWEAGAMFGAMIDYWYYT--ND	TTYNDIVEQALVHQAG-DD-RNYM
150 tr Q75CW6 Q75CW6_ASHGO	KPYYWWMAGEAFGGMVENWYLC--NN	TEYQDILLTDAMVSQIG-EN-KDYV
151 tr Q7S4K4 Q7S4K4_NEUCR	GDYYWWEAGGALMGTMIDYWHLT--GD	TTYNDVITQGILHQVG-DN-RDFQ
152 tr G2QB99 G2QB99_MYCTT	DPYFWWEAGAMFGTLVDYWALT--GD	DSYNAITTTQAILHQAT-EK-GDFM
153 tr G0S3F2 G0S3F2_CHATD	GDYYWWEAGGAMMGTYVDYWHLT--GD	PSYNHVIMEGMLHQVG-PN-ADYQ
154 tr A0A1E4SC85 A0A1E4SC85_9ASCO	PPYYWWEAGGAWGSLIDYSYYM--EN	NTYHDVIQQAMEYQTG-DD-YNXI
155 tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	PPYYWWQAGVAFGGMLENWFLC--QN	DTYKDILLMNALVAQTG-PN-YDYI
156 tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	T--KWEEGSALFMAMLQYQYFT--GD	HTYNSDVSLGLQWQAG--D-GDYM
157 tr G2QKJ0 G2QKJ0_MYCTT	GPYYWWQAGAMWGTYIDYWFT--GD	DTYNAETTRSLLFQAE-PPANAYM
158 tr Q4WFX5 Q4WFX5_ASPFU	APYYWWEAGAFFGTLVNYWRYT--GD	DTYNNITMQAILHQAG-T--GDFM
159 tr Q5BGD7 Q5BGD7_EMENI	D--TWYVAGAMFMTMIQFHWVS--GY	NEHNSVSVSHDLMFQAG-RN-YDYF
160 tr A0A0A8L8U3 A0A0A8L8U3_9SACH	SPNYWWQAGGAWGSILDYWFYM--QN	DTYNDMLKQALLAQTG-DN-YDYM
161 tr G8BYN9 G8BYN9_TETPH	PPYYWWEAGEAFGGMLENWYLC--QN	DTFESLISDALIYQAG-SN-YDYI
162 tr F7VVT4 F7VVT4_SORMK	GPYYWWEAGAMWGTTIIDYWHYT--GD	DTYNTLAKQALVFQAD-PPQNSFM
163 tr G8YB80 G8YB80_PICSO	WPYYWWEAGGAWGSLVDYIYYM--EN	DTLTPLVKQALTYQTG-DD-DNYI
164 tr A0A0H5CB47 A0A0H5CB47_CYBJA	YPYYWWQAGHAFGALIDTWYLC--GN	DTYEQLIHDAIMHQTG-ED-NDYM
165 tr G2WRS7 G2WRS7_VERDV	GDYYWWEAGAMWGALIDYWHLT--GD	DTYNGIITQAMLWQVG-EG-RDYM
166 tr A0A124BVB5 A0A124BVB5_ASPNG	DPYYWWEAGAFFGTLVNYWAYT--GD	TTYNELTAQAILHQVG-PN-RDFM
167 tr A7EIG7 A7EIG7_SCLS1	APYYWWEAGAMFGSLIDYWYYT--GD	SSYNDVTTTEAMLWQTG-PD-NDYM
168 tr C5M5U7 C5M5U7_CANTT	NPYYWWEAGGAWGSLIDYTFFF--DN	DTLVPLIKEALLYQTG-DD-DNYI
169 tr B9WAA8 B9WAA8_CANDC	PPNYWWNAGEAFGGGLVDFYTF	CESDNSTLES LIYNGMYHQAG-EN-YNXI
170 tr F8MRU1 F8MRU1_NEUT8	GDYYWWEAGGALMGTMIDYWHLT--GD	TTYNDVITQGILHQVG-DN-RDFQ
171 tr A1DJS0 A1DJS0_NEOFI	T--KWEEGSALFMAMLQYQYFT--GD	HTYNSEVSLGLQWQAG--E-GDYM
172 tr B6HLM8 B6HLM8_PENRW	R--SWWTGAALFLACLNYWHAT--ND	TTYNEDVSI GLQHQQG-AG-GDYM
173 tr A0A117DWS0 A0A117DWS0_ASPNG	D--TWEEGGAMFMTLIQYWYWT--GD	SSYNEVTTQGMLWQKG-DN--DYF
174 tr A0A124BXM1 A0A124BXM1_ASPNG	S--KWEEGSALFTSLLLYWYYT--GD	STYNDEVRRQGMQWQAG--D-CDYM
175 tr J3NZQ6 J3NZQ6_GAGT3	GPYYWWQAGAVWGAMIDYWHYT--GD	STYNKIVTEGIMNQIG-ED-QDFE
176 tr W0THH8 W0THH8_KLUMD	QPYYWWEAGLVFGGMIENWYLC--QN	NSYQNL LYDAMIAQTG-SN-YDFM
177 tr B2AEF2 B2AEF2_PODAN	GDYYWWEAGGAMMGTYVDYWFLT--GD	ESYNKVVTTEGMIHQVG-PN-EDYM
178 tr Q96TX1 Q96TX1_NEUCS	GDYYWWEAGAMWGALIDYWRFT--GD	ASYNDVITQALLWQVG-PG-QDYM
179 tr E9DYA6 E9DYA6_METAQ	GDYYWWEAGAMWGTLIQYWAFT--GD	STYNDNIMQAMQFQVG-EG-QDYM
180 tr B9WAI2 B9WAI2_CANDC	WPYYWWEAGGAWGSLIDYTIFYF--DN	DTLVPLITEALLYQTG-DD-DNYI
181 tr Q6C171 Q6C171_YARLI	APYYWWHAGAAWNAAFIDYWHIT--GN	DTYNNLTWAAMKHQRG-EK-YDLM
182 tr A6ZMV1 A6ZMV1_YEAS7	SPYYWWHAGEAFGGMLENWFLC--EN	DTYQELLYDALLAQTG-SN-YDYI
183 tr B6K7X7 B6K7X7_SCHJY	LTVYWWMTGAGMNALLNNYYLT--GN	STYNDLVTKVLLSNTG-DE-NDFA
184 tr I1RSA2 I1RSA2_GIBZE	DPYYWWEAGAMFGTLVDYWWT--GD	DTYNAITKQAMIHQAG-SD-GDYM
185 tr F7VWZ9 F7VWZ9_SORMK	DPYYWWEAGAMFGALVDYWWT--GD	TSYVEATSQAIVHQAS-ET-RDFM
186 tr A0A1B2J789 A0A1B2J789_PICPA	NPYYWWQSGAAFGSMLDYWWYM--EN	DTYHDAIMQAI IYQAG-DN-ADFM
187 tr G0V8N4 G0V8N4_NAUCC	DPYYWWEAGGAWGTMLDYWYFM--QN	DTYNDIITAALLHQVG-TD-NNXI
188 tr H8WXN8 H8WXN8_CANO9	SPYYWWEAGGAWGGMIDYTIFYF--NN	DTLVPLIKQALEYQVG-DD-DNYI
189 tr A6ZZS0 A6ZZS0_YEAS7	DPYYWWEAGGAWGCMLDYWFFM--DN	DTYNDEIIAAMIHQAG-DD-NDYI
190 tr G9MQD1 G9MQD1_HYPVG	GDYYWWEAGGAMMGTYVDYWHLT--GD	TSYNAVVMEGMLHQVG-DH-ENYM
191 tr A0A1G4JVK5 A0A1G4JVK5_9SACH	NPYYWWEAGGAWGSILDYWWYT--GN	DTFNGILKQALLAQTG-EN-NDYV
192 tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	G--TWYVAGAMFMTLIQYWQSS--GD	DTYNSIVSHDLMFQSG-EN-YDFF
193 tr A0A254UET1 A0A254UET1_ASPNG	DPYYWWEAGAFFGTLVNYWAYT--GD	TTYNDLTAQAILHQVG-PN-RDFM
194 tr A1D587 A1D587_NEOFI	D--TWEEGGAMFMTLIQYWYWT--GD	TSYNEVTTQGMLFQKG-NN--DYF
195 tr Q1K7I4 Q1K7I4_NEUCR	GDYYWWEAGAMWGALIDYWRFT--GD	ASYNDVITQALLWQVG-PG-QDYM
196 tr G8JS88 G8JS88_ERECY	SPYYWWEAGAAWGALIDYWFYM--EN	DTYNDIIKQALLYQVG-KY-EDYV
197 tr A0A136J7D3 A0A136J7D3_9PEZI	DPYFWWEAGAMFGTLVDYWFHT--GD	DTYNAITKQALLHQVG-KD-RDYM
198 tr E9E4X8 E9E4X8_METAQ	GDYYWWEAGAMWGTLIDYWYWT--GD	STYNDEVMSMLFQVG-EN-MDYM
199 tr Q6CP42 Q6CP42_KLULA	SPSYWWQAGGAWGSILDYWFYM--EN	DTYNDILTQALLYQTG-DN-HDYM
200 tr G0WHL4 G0WHL4_NAUDC	PPYYWWEAGEAFGGMIENWYLC--QN	DTYEDMLKSALLAQTG-SN-YDYI
201 tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	DPYYWWEAGGAWGCMLDYWYFM--EN	DTYNDEITAAMIHQAG-DD-NDYI
202 tr F9WYX6 F9WYX6_ZYMTI	QPYYWYNAGNMFNSLIKYWALS--GD	QSIVPTLQSALVFQLG-PD-FNYM
203 tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	GDYYWWEAGGALMGQMI EYWHLT--GD	TTYNDQVMQGMLHQVG-QD-RDYQ
204 tr J3P147 J3P147_GAGT3	GPYYWWQAGAMWAAMIDYWHLT--GD	TYNKLVTDGILAQHG-PD-RDFQ
205 tr I2H842 I2H842_TETBL	DPYYWWEAGGAWGCMLDYWYYM--QN	DTYNDVIYEALQYQTG-DD-NDYI
206 tr S6E3R3 S6E3R3_ZYGB2	SPYYWWQAGEAFGGMLDNWYFC--QN	NTFENLIRKAMLAQTG-TK-YDYM
207 tr E5AD94 E5AD94_LEPMJ	PPYYWWQAGAMFGTLLDYWHYT--GD	DQYNDMIREGLIHQFG-EH-LDLM
208 tr Q6FJM7 Q6FJM7_CANGA	PPYYWWQAGVAFGGMLENWFLC--QN	DTYKDILLMNALVAQTG-PN-YDYI
209 tr A5DUW0 A5DUW0_LODEL	APYYWWEAGGAWGSLIDYTFFF--EN	DTYVPLIKQALLYQVG-DD-NNXI
210 tr G8BF46 G8BF46_CANPC	SPYYWWEAGGAWGGMIDYTIFYF--QN	DTLVPLIKQALEYQVG-DD-NNXI
211 tr W3XAF6 W3XAF6_PESFW	DPYYWWQC GAMFGSLLDYWSYT--GD	DSYNNITYQAMWHQRG-DD-YDYM
212 tr B6H7I2 B6H7I2_PENRW	E--KWEEGSALFLALLQYWHFT--GD	TYNSLMSEGMQWQSG-EE-GDYM
213 tr Q5ATF9 Q5ATF9_EMENI	E--KWEEGSALFMSLMLYWYYT--GD	SQYNSLITQGMQHQAG--N-GDYL
214 tr A0A167CDD3 A0A167CDD3_9ASCO	NPYYWWEAGVAWGTMLDYWYYT--QN	DTYVDIIKS SMLFQTG-KD-WNYM
215 tr G0RXS3 G0RXS3_CHATD	GPYYWWQAGAMWGTLIDYWYYT--GD	TYVNETRRSLVFQAE-GPGYSFE

216	tr B8NX26 B8NX26_ASPFN	D--TWYIAGSMFMTLIIQYWQAS--GD	DSQNAVVS	SHDLMF	QAG-EN-YDFF	
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	DPYYWWEAGAMFSA	LIDYWYYT--GD	DQYNDIT	TQAMLHQVG-KE-NNYM	
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	APYYWWEAGAFFGT	LVNRYWRYT--GD	DAYNNIT	MQAILHQAG-T--GDFM	
219	tr A1CMM3 A1CMM3_ASPCL	DPYYWWEAGAMFGS	LVEYWYYT--GD	SQWNEIIT	QAILHQTG-PS-FNFM	
220	tr B6HU84 B6HU84_PENRW	G--TWWEGGSMFMTLILYWYVT--GD	SQFNDAIQ	EGMYFQ	KG-DD--NFF	
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	GDYYWWEAGAMWGALIDYWRFT--GD	ASYNDVIT	QALLWQ	VG-PG-QDYM	
222	tr G2WHY6 G2WHY6_YEASK	DPYYWWEAGGAWGC	MLDYWFFM--DN	DTYNDEI	IAAMIHQAG-DD-NDYI	
223	tr G0W7G8 G0W7G8_NAUDC	DPYYWWEAGGAWGS	MLDYWYYM--DN	DTYNDEI	MAALLHQTG-DN-NDYV	
224	tr G4NGV8 G4NGV8_MAGO7	DPYFWWEAGAMFGT	MVDYWLLT--GD	ATYNPVT	TQALLHQVG-DN-KDYM	
225	tr W3X855 W3X855_PESFW	DPYYWWEAGAYFGT	LDYWAFT--GD	EQYVDLT	TFQALQHQVG-DD-GDFM	
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	GPYYWWIGGAMWGT	LIDYWHLT--GD	TTYNDRIM	QAMQFQVG-EN-KAYM	
227	tr J8Q2K7 J8Q2K7_SACAR	DPYYWWEAGGAWGC	MLDYWYFM--DN	DSYNDEI	IAAMIHQTG-DD-NDYI	
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	WPYYWWEAGGAWGS	LIDYTYYM--KN	DTLVPLIK	QALEYQVG-DD-YNYI	
229	tr J6EGN8 J6EGN8_SACK1	SPYYWWHAGEAFGG	MLENWFLC--EN	ETYQELLY	DALLAQTG-SN-YDYI	
230	tr A0A100ISD9 A0A100ISD9_ASPNG	DPYYWWEAGAMFTA	LVEYWYLT--SD	DTWNNIT	TQGITWQAG-PS-GSFM	
231	tr G8YQC0 G8YQC0_PICSO	YPYYWWHAGCAFGG	LIDYFTFC	DKDNSTL	KEIITYDGM	YHQAG-DN-YDYI
232	tr F7VVP8 F7VVP8_SORMK	GDYYWWEAGGALMG	TVVDYWHLT--GD	TTYNDVVI	QGILHQVG-DR-RDFQ	
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	SPYYWWHAGEAFGG	MLENWFLC--EN	DTYQELLY	DALLAQTG-SN-YDYI	
234	tr F8MBP4 F8MBP4_NEUT8	DPYYWWEAGAMFGA	MVDYWWLT--GD	TSYVEVT	TQAIVHQAG-DA-RDFN	
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	H--TWWEGGVLFD	TMIRYWYFT--D	DASNNA	AVSQGMYHQRG-AG-NDYM	
236	tr A1CD27 A1CD27_ASPCL	G--TWYVAGAMFMTLIIQYWQAS--GD	DTYNSIV	SHDLMFQ	SG-EN-YDFF	
237	tr A5DHF3 A5DHF3_PICGU	SPNYWWNAGEAFGG	LIDYFTFC	DSGNKSL	EKLIFNGMYH	QAG-DQ-YNYI
238	tr D4ATT7 D4ATT7_ARTBC	DPYYWWEAGAMFGA	LIDYWWT--GD	DAYNKIT	MQAMLHQVG-TE-KNYE	
239	tr E2PT42 E2PT42_ASPNC	DPYYWWEAGAMFTA	LVDYWYLT--SD	DTWNNIT	TQGITWQAG-PS-GSFM	
240	tr C7ZPE5 C7ZPE5_NECH7	GDYYWWEAGGAMWG	TYMDYWKCT--GD	ATYNAKVI	QAMQFQTG-DD-NDYQ	
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	SPYYWWEAGALFGA	FIDYWFYT--GD	DTYNDIT	MQAMIHQAS-ED-RDFM	
242	tr Q4WKP7 Q4WKP7_ASPFU	D--TWWEGGAMFMTLIIQYFWT--GD	TSYNEVT	TQGMLWQ	KG-NN--DYF	
243	tr F8MXT4 F8MXT4_NEUT8	GDYYWWEAGAMWGALIDYWRFT--GD	ASYNDVIT	QALLWQ	VG-PG-QDYM	
244	tr A0A177AJ74 A0A177AJ74_9PEZI	SGYFWWQAGAMWNT	MLDYWHIT--GD	NSYNEAIS	QALVFQVG-ND-RDYM	
245	tr J7RQR5 J7RQR5_KAZNA	PPYYWWHAGEAFGG	MIENWFLC--QN	STYETLL	MDAFIAQTG-NN-YDYI	
246	tr Q75CW7 Q75CW7_ASHGO	KPYYWWMAGEAFGG	MVENWYLC--NN	TEYQDLL	TDAMVSQIG-EN-KDYM	
247	tr G3AKK4 G3AKK4_SPAPN	PPNYWWNAGEAFGG	LVDYYTFC	DPNNDTL	KDLIFNGMYH	QAG-EN-YNYV
248	tr B2W762 B2W762_PYRTR	DPYYWWECEGAMFNA	FIDYWYYT--GD	DTYNGIT	TQALEAQIG-DY-NAFM	
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	DPYYWWEAGGAWGC	MLDYWFFM--DN	DTYNDEI	IAAMIHQAG-DD-NDYI	
250	tr C4XWK1 C4XWK1_CLAL4	PPYYWWNAGEVFG	GWVDYWAFC	AQDN	DTFTELLSNAMYH	QAG-DD-FNYM

001	tr A0A254U2J9 A0A254U2J9_ASPNG	-P	A	N	Y	S	S	Y	L	G	N	D	D	Q	M	F	W	G	L	A	A	I	T	A	A	E	I	E	F	A	D	S	-	-	-	-	S	S	G	Y	S	-	W	L	A	L	A	Q	Q	V	
002	sp Q6FLP9 DCW1_CANGA	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	I	A	V	M	Q	A	A	E	R	K	F	P	N	P	P	-	-	-	D	D	K	P	Q	-	W	L	Y	L	T	Q	A	V	
003	tr A1CCM5 A1CCM5_ASPCL	-P	S	N	Y	S	S	Y	L	G	N	D	D	Q	M	F	W	G	L	A	A	M	L	A	A	E	T	K	F	P	D	R	-	-	-	A	N	G	Y	S	-	W	L	S	L	A	Q	Q	V		
004	tr C1H190 C1H190_PARBA	-P	H	N	Q	S	R	T	S	G	N	D	D	Q	A	F	W	G	M	A	A	M	T	A	A	E	T	K	F	Q	D	P	A	E	-	-	-	D	K	P	Q	-	W	L	A	L	A	Q	Q	V	
005	tr Q2UR85 Q2UR85_ASPOR	-P	A	N	Y	S	N	Y	L	G	N	D	D	Q	V	F	W	G	L	A	A	M	T	A	A	E	L	N	F	P	E	K	-	-	-	D	D	D	S	S	-	W	L	S	L	A	Q	Q	V		
006	tr A5DV30 A5DV30_LODEL	-P	S	N	Q	S	M	T	E	G	N	D	D	Q	G	V	W	G	M	A	I	M	E	A	V	E	R	N	F	T	N	P	D	-	-	-	L	-	-	H	S	-	W	L	E	M	V	Q	A	I	
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	G	V	P	N	Q	T	H	V	E	A	N	D	D	Q	V	F	W	G	F	T	V	L	E	A	A	E	R	D	F	P	A	Y	S	N	-	N	S	G	D	P	S	-	Y	A	D	L	A	L	N	V
008	tr J3PGN0 J3PGN0_GAGT3	-P	A	K	H	R	A	S	L	G	N	D	D	Q	G	F	W	G	M	T	A	M	L	A	A	E	N	K	F	P	D	P	P	K	-	-	-	D	K	P	Q	-	W	L	A	L	A	Q	A	V	
009	tr A0A124BXU6 A0A124BXU6_ASPNG	-P	A	N	W	S	S	Y	I	G	N	D	D	Q	M	F	W	G	L	A	A	M	T	G	A	E	L	N	F	P	D	S	-	-	-	S	D	G	Y	S	-	W	L	S	L	A	Q	Q	V		
010	tr C5P4A1 C5P4A1_COCP7	-P	L	N	Q	T	S	T	L	G	N	D	D	Q	A	F	W	G	L	A	A	L	A	A	A	E	R	N	F	P	N	P	P	A	-	-	-	H	E	P	Q	-	W	L	A	L	A	Q	Q	V	
011	tr Q0CG55 Q0CG55_ASPTN	-P	T	N	Q	T	R	T	E	G	N	D	D	Q	A	F	W	A	F	A	A	M	S	A	A	E	R	N	F	P	N	P	P	E	-	-	-	T	S	P	G	-	W	L	A	M	A	Q	A	V	
012	tr C4Y2X5 C4Y2X5_CLAL4	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	L	A	V	M	S	A	A	E	K	N	F	S	N	P	T	-	-	-	D	H	R	A	A	-	W	L	T	L	A	Q	A	V	
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	-P	A	N	Q	T	R	T	E	G	N	D	D	Q	S	F	W	A	F	A	A	M	S	A	A	E	R	N	F	P	D	P	P	P	-	S	S	G	S	P	G	-	W	L	A	M	A	Q	A	V	
014	tr C7Z068 C7Z068_NECH7	-P	E	N	H	T	A	S	L	G	N	D	D	Q	G	F	W	G	M	S	A	M	L	A	A	E	N	K	F	P	N	P	P	E	-	-	-	D	K	A	Q	-	W	L	A	L	A	Q	A	V	
015	tr G8BXX2 G8BXX2_TETPH	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	I	A	A	M	M	A	A	E	R	N	F	T	N	P	A	-	-	-	E	D	E	P	Q	-	W	L	Y	L	A	Q	A	V	
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	F	A	V	M	A	A	A	E	H	N	F	T	N	P	E	-	-	-	D	P	K	L	A	-	W	L	A	L	A	Q	A	V	
017	tr A1CSC2 A1CSC2_ASPCL	-P	S	N	Y	S	N	Y	L	G	N	D	D	Q	V	F	W	G	L	A	A	M	T	A	A	E	L	N	Y	P	E	Q	-	-	-	N	G	Q	P	S	-	W	V	S	L	A	Q	Q	V		
018	sp Q05031 DFG5_YEAST	-P	S	N	Q	T	M	V	E	G	N	D	D	Q	G	I	W	G	I	T	V	M	G	A	V	E	R	N	F	T	D	P	G	-	-	-	D	G	K	P	G	-	W	L	A	M	V	Q	A	V	
019	tr H2AQJ6 H2AQJ6_KAZAF	-P	E	N	Q	T	M	V	E	G	N	D	D	Q	C	V	W	G	L	T	I	M	D	A	V	E	R	N	F	T	D	P	T	-	-	-	D	G	S	P	G	-	W	L	Y	M	T	Q	A	I	
020	tr G2XFH9 G2XFH9_VERDV	-P	L	N	H	T	A	S	L	G	N	D	D	Q	G	F	W	G	M	T	A	M	L	A	A	E	N	K	F	P	D	P	P	E	-	-	-	D	Q	P	Q	-	W	L	A	L	A	Q	A	V	
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	I	A	V	L	N	A	A	E	R	K	F	P	N	P	P	-	-	-	S	D	E	P	Q	-	W	L	Y	L	A	Q	A	V	
022	sp Q5AD78 DCW1_CANAL	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	I	A	V	M	A	A	A	E	R	N	F	T	N	P	K	-	-	-	D	P	T	K	A	-	W	L	T	L	A	Q	A	V	
023	tr B6H7E8 B6H7E8_PENRW	-P	S	N	Y	S	Q	Y	L	G	N	D	D	Q	V	F	W	G	L	A	A	I	T	A	G	E	L	N	F	P	E	R	-	-	-	S	G	E	P	S	-	W	I	S	L	A	E	G	V		
024	tr B6GZU2 B6GZU2_PENRW	-P	A	N	Q	S	K	T	E	G	N	D	D	Q	V	F	W	A	F	A	A	M	T	A	A	E	R	N	F	P	N	P	P	E	-	-	-	D	K	P	Q	-	W	L	E	M	A	Q	A	V	
025	tr E9E4W7 E9E4W7_METAQ	-P	P	N	Q	T	L	S	L	G	N	D	D	Q	G	F	W	G	L	S	A	L	L	A	A	E	N	K	F	P	D	P	P	P	-	-	-	D	Q	P	Q	-	W	L	E	L	A	Q	A	V	
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	-P	N	N	Q	S	K	T	E	G	N	D	D	Q	A	F	W	G	F	A	A	M	S	A	A	E	R	N	F	P	N	P	P	D	-	-	-	G	K	P	Q	-	W	L	E	M	A	Q	A	V	
027	tr G2R7G8 G2R7G8_THITE	-P	A	N	W	T	L	S	L	G	N	D	D	Q	G	F	W	G	M	A	A	M	L	A	A	E	T	N	F	Q	N	P	P	D	-	-	-	G	Q	T	P	-	W	L	T	L	A	Q	A	V	
028	tr G3JBY1 G3JBY1_CORMM	-P	R	N	Q	S	N	A	M	G	N	D	D	Q	G	F	W	A	M	T	T	M	M	A	A	E	A	A	F	P	N	P	P	S	-	-	-	D	T	P	Q	-	W	L	A	G	G	Q	A	V	
029	tr A5DNV5 A5DNV5_PICGU	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	G	F	W	G	I	A	V	M	A	A	A	E	K	N	M	S	N	P	K	-	-	-	D	E	R	K	A	-	Y	L	A	L	A	Q	A	V	
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	-P	P	N	Q	T	K	T	L	G	N	D	D	Q	C	F	W	A	L	A	A	I	S	A	A	E	S	N	F	P	N	P	P	K	-	-	-	D	K	P	Q	-	W	L	A	L	A	Q	A	V	
031	tr Q2TWC1 Q2TWC1_ASPOR	-S	S	N	Y	S	Q	W	L	G	N	D	D	Q	M	F	W	G	L	A	S	I	T	A	S	E	T	G	F	P	E	V	-	-	-	S	G	K	P	T	-	W	T	S	L	A	R	T	V		
032	tr J5RXY9 J5RXY9_SACK1	-P	L	N	Q	S	T	T	E	G	N	D	D	Q	A	F	W	G	I	A	A	M	T	A	A	E	R	N	F	T	N	P	P	-	-	-	A	D	E	P	Q	-	W	L	Y	L	A	Q	A	V	

033	tr A0A2N6NN14 A0A2N6NN14_BEABA	-PVNQSTNAMGNDDQGFWALTSMMAAEAVFPNP	PS--	-DTPQ	-WLAGVQAV
034	tr A0A254U5V1 A0A254U5V1_ASPNG	-PANDSNYLGNDQVFWGLAAMTAAELNYPEQ	--	-DGQPS	-WLSLAQGV
035	tr A0A061B6H7 A0A061B6H7_CYBFA	-PLNQSTTEGNDDQGFWGIAVMAAAERDFPNP	P--	-EDGPQ	-WVALAQAV
036	tr G8ZQ93 G8ZQ93_TORDC	-PLNQSTTEGNDDQAFWGIAAMTAAERNFTNPP	--	-SDQPQ	-WLYLAQAV
037	tr Q9C2J1 Q9C2J1_NEUCS	-PANQSRTSSNDDVGFWTITAMMAAEDAFDPDP	PP--	-DQPQ	-WLALVQAV
038	tr C5NZK5 C5NZK5_COCP7	-PLNQSTSLGNDDQAFWGMAAMSAAENKFPNP	PD--	-DKPQ	-WLELAQAV
039	tr Q6C0T7 Q6C0T7_YARLI	-VFNFSSSEGNDDQGVWGMTMMTAAERNFTAPT	--	-DEYG	-WLYVAQGA
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	-PLNQSTTEGNDDQAFWGIAVMQAAERKFPNP	PP--	-DDKPQ	-WLYLAQAV
041	tr F7VZ72 F7VZ72_SORMK	-PPNVTASLGNDQGFWMGTAMTAAENNFPDPP	PP--	-DQPQ	-WLGLAQAV
042	tr A3LN37 A3LN37_PICST	-PLNQSTTEGNDDQGFWGIAVMGAAERNFSNP	PD--	-DDSKA	-WLTLAQAV
043	tr Q6CAI2 Q6CAI2_YARLI	-PVNQSTTEGNDDQGFWGITAMAAAEKNFSNP	G--	-KDEPQ	-WLYLAQAV
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	-PLNQSTTEGNDDQGFWGIAVMAAAEKNFSNP	N--	-DEKKQ	-WLALTQAV
045	tr G9MJS5 G9MJS5_HYPVG	-PPNVTLSLGNDQGFWMGTAMLAAEEKFPNP	PPA--	-DKPG	-WLALAQGV
046	tr G9MHI4 G9MHI4_HYPVG	-PTNQSLTEGNDDQGFWAMAAMSAAENVFPNP	PS--	-DQPQ	-WLAMAQAV
047	tr D4AP27 D4AP27_ARTBC	-PPNQTLTEGNDDQAFWAIAMSAAERKFPNP	PS--	-DKPQ	-WLSLVEAV
048	tr B6GZT8 B6GZT8_PENRW	-PGNYSSYLGNDQMFWGLAAMTAAEFGF	PDR--	-STGFS	-WLSLAQGV
049	sp O74556 YCZ2_SCHPO	-PSNYTTSEGNDDQAFWGLTVISAAEANFSNP	AA--	-DEPQ	-WLELAQAV
050	tr A0A0C4E991 A0A0C4E991_MAGP6	-PAKHRASLGNDQGFWMGTAMLAAENKFPD	PPK--	-DQPQ	-WLALAQAV
051	tr B8MHG0 B8MHG0_TALSN	-PSNYSSYLGNDQEFWALAALTAIEQKF	PDN--	-PGHAS	-WLSLAQGA
052	tr A3LMV8 A3LMV8_PICST	-PSNQSMTEGNDDQGVWVMAIMQAVERNFT	NPA--	-D--HS	-WLSMTIAA
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	NVENQTDVEANDQVFWGFTVMEAAERGF	PMYSN-	DENDVN-	YAQLAMNV
054	sp Q9P6I3 YHG7_SCHPO	-PNSEKFDLGNDQGIWGLSAMSAAEVNMT	TGD--	-SSAS	-FTELAQAV
055	tr Q2US57 Q2US57_ASPOR	-PSNWSSYLGNDQMFWGLAAITAAELQF	PEV--	-SDGYS	-WLSLAQGV
056	tr W7N622 W7N622_GIBM7	-PDNQTMTEGNDDQGFWAVAAMSAAEHKY	PDPPA--	-DQPQ	-WLALVQAV
057	sp Q9P6I4 YHG6_SCHPO	-PSFEYFNLGNDDQGMWAAAAMDAAEANFS	PPN--	-STEHS	-WLELTQAA
058	tr C7GRB3 C7GRB3_YEAS2	-PSNQTMVEGNDDQGIWGITVMGAVERNFT	DPG--	-DGKPG	-WLAMVQAV
059	tr C7GP28 C7GP28_YEAS2	-PLNQSTTEGNDDQAFWGIAAMTAAERNFT	NPP--	-ENEPQ	-WLYLAQAV
060	tr G3JGZ5 G3JGZ5_CORMM	-PRNHTASLGNDQGFWMGTAMLAAENKFP	DPPA--	-DKPQ	-WLALAQAV
061	tr C4QXV4 C4QXV4_KOMPG	-PLNQTTTEGNDDQGFWGITAMAAVERNFT	NPP--	-EDEPQ	-WLYLVQAT
062	tr I1S0Z5 I1S0Z5_GIBZE	-PSNVTLSLGNDQGFWGMSSAMLAAELAF	PNPPS--	-DQPG	-WLALAQAV
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	-PTNQTKTEGNDDQAFWAFAMSAAERNFP	DPD-P	-HGPG	-WLAMAQAV
064	tr G0W5X4 G0W5X4_NAUDC	-PLNQSTTEGNDDQAFWGIAAMTAAERNFT	NPP--	-ESSPQ	-WLYLAQAV
065	tr G8ZPC1 G8ZPC1_TORDC	-PENQTMVEGNDDQGVWGLTLMGAVERNFT	NPT--	-NGAPG	-WLAMAQAI
066	tr A0A167CDC5 A0A167CDC5_9ASCO	-PSNQTTTEGNDDQGFWGILVMDAAEKNFS	NP	-DQAQ	-WLSLAQAV
067	tr R9XAT7 R9XAT7_ASHAC	-PWNETTTEGNDDQAFWGMVMAAAERNYPT	PP--	-PDQPQ	-WLALAQAV
068	tr B2B747 B2B747_PODAN	-PPNVTASLGNDQGFWGMSSAMLAAENNFP	DPPE--	-DEPQ	-WLALAQAV
069	tr G0SE99 G0SE99_CHATD	-PPNVTASLGNDQGFWGMSSAMLAAENGFP	DPPE--	-DEPQ	-WLALAQAV
070	tr J8Q6F0 J8Q6F0_SACAR	-PSNQTMVEGNDDQGIWGITVMGAVERNFT	DPT--	-NGAPG	-WLAMVQAV
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	-PSNQTMVEGNDDQGVWALTVMSAVERNFT	NPK--	-DGAPG	-WLAMAQAV
072	tr J3K6E2 J3K6E2_COCIM	-PLNQSTSLGNDDQAFWGLAALAAERNFP	NPPA--	-HEPQ	-WLALAQGV
073	tr G4N3E1 G4N3E1_MAGO7	-PLSKRASLGNDQGFWMGTAMLAAETNFP	NPPK--	-DKPQ	-WLALAQAV
074	tr C5M5J2 C5M5J2_CANTT	-PSNQSMTEGNDDQGVWGMAIMQAVERNFT	TEPE--	-S--HS	-WLEMTQAV
075	tr A0A254U0X0 A0A254U0X0_ASPNG	-SSNYSQWLGNDQMFWGLAAITASETG	FPEV--	-SGKPS	-WTSLARAV
076	sp Q75DG6 DCW1_ASHGO	-PWNETTTEGNDDQFFWGMVMAAAERNFP	NPP--	-ADQPQ	-WLALAQAV
077	tr Q6BZF0 Q6BZF0_DEBHA	-PLNQSTTEGNDDQGFWGLAVMAATEKNFS	NP	-DKH	-WLALTQAV
078	tr E9E3Q1 E9E3Q1_METAQ	-PNNHTMSLGNDQGFWGLSAMLAAENKFP	DP	-PA	-WLALAQAV
079	sp P36091 DCW1_YEAST	-PLNQSTTEGNDDQAFWGIAAMTAAERNFT	NPP--	-ENEPQ	-WLYLAQAV
080	tr W0TAH6 W0TAH6_KLUMD	-PLNQTVTEGNDDQAFWGFVMAAAERNYP	NPP--	-KDQPQ	-WLYLAQAV
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	-PSNQSMTEGNDDQGFWGLAVMEATERNFT	NPP--	-SDQPG	-WLALTQAV
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	-PENQTRS LGNDQGFWALAAMSAAESGF	ENPPS--	-DKPQ	-WLALVQAV
083	tr Q2TYU3 Q2TYU3_ASPOR	-PANYSSYLGNDQGFWGIAAMTAAEIGF	PDV--	-DDGYS	-WLSLAQGV
084	tr G2XGF6 G2XGF6_VERDV	-PLNQTRTLGNDDQGFWTLSAMMATEAGY	PDPPA--	-DQPQ	-WLALVQAC
085	tr G3AMT4 G3AMT4_SPAPN	-PLNQSTTEGNDDQGFWGIAVMGAAERNFS	NPE--	-DPDKA	-WLTLAQSV
086	tr A0A1C1D213 A0A1C1D213_9EURO	-PPNQTKTEGNDDQAFWGMAALSAAENVFP	NPPS--	-DQPQ	-WLALAQAV
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	-PENYTISMGNDQGFWGLSAMLAAEIGF	PDPPA--	-DKPQ	-WLQLAQAV
088	tr A0A100I5A6 A0A100I5A6_ASPNG	-SSNYSQWLGNDQMFWGLAAITASETG	FPEV--	-SGKPS	-WTSLARAV
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	-PSNQSMVEGNDDQGVWGITVMGAVERNFT	NPS--	-TKGAPG	-WLAMVQGV
090	tr G2Q8A7 G2Q8A7_MYCTT	-PPNYTASLGNDQAFWGMSSAMTAAENNFP	NPPA--	-DQPQ	-WLALAQAV
091	tr B8NPZ1 B8NPZ1_ASPFN	-SKNVSDWLGNDDQMFWGLATITASEAG	FPEI--	-SGKPS	-WTSLARVV
092	tr Q7SAB2 Q7SAB2_NEUCR	-PENYTISMGNDQGFWGLSAMLAAEIGF	PDPPA--	-DKPQ	-WLQLAQAV
093	tr A2R8R5 A2R8R5_ASPNC	-PANWSSYIGNDDQMFWGLAAMTGAELN	FDS--	-SDGYS	-WLSLAQGV
094	tr C5DHG0 C5DHG0_LACTC	-PSNQSMVEGNDDQGVWGLTVMSAAERNFT	NPT-	-EDGVP	-WLAMAQGV
095	tr G0SFA3 G0SFA3_CHATD	-PPNQTRTLGNDDQGFWGMMAAMSAAEYNFP	NPPQ--	-DKPQ	-WLALAQSL
096	tr K1WJG5 K1WJG5_MARBU	-PKNQTKSEGNDDQGFWMGTAMSAAEMNY	PNPPD--	-TEPQ	-WLALAQAV
097	tr H2B005 H2B005_KAZAF	-PLNQSTTEGNDDQAFWGIAVMTAAERNFT	NPP--	-DDQPQ	-WLYLAQAV
098	tr C5DYD7 C5DYD7_ZYGRC	-PQNQTMVEGNDDQGVWGLTVMEAVEERNFT	DPG--	-DGKPG	-WLAMAQAV
099	tr G8YM23 G8YM23_PICSO	-PLNQSTTEGNDDQGFWGIAVMAAAEKNFS	NPS--	-DPKKA	-WLTLAQAV
100	tr W3X8E3 W3X8E3_PESFW	-PPNVTASLGNDQGFWMGTAMLAAEENFP	DPK--	-DQPQ	-WLALAQAV
101	tr A0A254U3K7 A0A254U3K7_ASPNG	-PANWSSYIGNDDQMFWGLAAMTGAELN	FDS--	-SDGYS	-WLSLAQGV
102	tr Q6CER8 Q6CER8_YARLI	-VRNYSSEGNDDQGVWAMTMMTAAERNFT	NPD--	-DDYG	-WLYVAQGA

103	tr G2QT05 G2QT05_THITE	-PPNVTASLGNDDQGF	WGM	SAM	LAA	ENN	FPN	PPA	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
104	sp Q5ACZ2 DFG5_CANAL	-PSNQSMTEGNDDQGV	WGM	AIME	AVE	RNF	TEPE	-	-	-	S	-	-	HS	-	WL	E	M	V	Q	A	V		
105	tr G2WKU6 G2WKU6_YEASK	-PSNQTMVEGNDDQGI	WGI	ITV	MGA	VER	NFT	DPG	-	-	-	DGK	P	G	-	WL	A	M	V	Q	A	V		
106	tr A0A0C7N752 A0A0C7N752_9SACH	-PSNQSMVEGNDDQGV	WGL	ITV	MGA	VER	NFT	NPH	-	-	DKN	A	P	G	-	WL	A	M	V	Q	G	V		
107	tr W3WN68 W3WN68_PESFW	-PENQTRSLGNDDQGF	WAM	ASMT	AAE	MNF	FTNP	SS	-	-	-	DEP	Q	-	WL	L	A	L	S	Q	A	V		
108	tr G2QFS1 G2QFS1_MYCTT	-PPNHTASLGNDDQGF	WGM	SAM	LAA	ENK	FPN	PPE	-	-	-	DKP	Q	-	WL	L	A	L	A	Q	A	V		
109	tr G3J9G4 G3J9G4_CORMM	-PVNQTNAMGNDDQGF	WALT	SMM	AAE	AVF	PNP	PPS	-	-	-	DTP	Q	-	WL	A	G	V	Q	A	V			
110	tr Q2UJ03 Q2UJ03_ASPOR	-PPNQTHAEGNDDQAF	WAF	AAMS	AAE	RNF	FPN	PPD	-	-	-	GQP	P	G	-	WL	A	M	A	Q	A	V		
111	tr G8YRT2 G8YRT2_PICSO	-PSNQSMTEGNDDQGI	WGL	TTME	AAE	RNF	FTDP	P	-	-	-	-	-	HS	-	WL	S	M	T	Q	A	I		
112	tr W3X554 W3X554_PESFW	-PQNQTRSEGNDDQGF	WAMA	AAMS	AAE	ENK	FPD	PPA	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
113	tr Q1K7A8 Q1K7A8_NEUCR	-PANQSRSTS	SNDDVGF	WTIT	AMMA	AAE	DAF	PD	PPP	-	-	-	DQP	Q	-	WL	A	L	V	Q	A	V		
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	-PSNYSNYLGNDDQV	FWGL	AAMT	AAE	LN	FPEE	-	-	-	-	DGQ	P	S	-	WV	S	L	A	Q	G	V		
115	tr A7F3P0 A7F3P0_SCLS1	-PLNQTKTEGNDDQGF	WGM	AAMT	AAE	TNF	FTNP	DN	-	-	-	GTP	P	G	-	WV	A	M	A	Q	A	V		
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	-PRNQSNAMGNDDQGF	WAV	TSM	MATE	AAE	FPN	PPS	-	-	-	DTP	Q	-	WL	A	G	A	Q	A	V			
117	tr Q6CIP9 Q6CIP9_KLULA	-PSNQTMVEGNDDQGV	WALT	VMT	AVE	RNF	FTNP	PK	-	-	EDG	T	P	G	-	WL	A	M	V	Q	A	V		
118	tr H2AMQ5 H2AMQ5_KAZAF	-PLNQSTTEGNDDQAF	WGI	AAMT	AAE	RNF	FTNP	PA	-	-	-	SDE	P	Q	-	WL	Y	L	A	Q	A	V		
119	tr G8JMI3 G8JMI3_ERECY	-PSNQSMTEGNDDQGI	WGL	FIM	GAVE	RNF	FPDA	Q	-	-	MGN	V	P	G	-	WL	T	M	A	Q	A	V		
120	tr C5DGT1 C5DGT1_LACTC	-PLNQSTTEGNDDQAF	WGL	VMA	AAE	RNF	FTNP	P	-	-	-	ETE	P	Q	-	WL	Y	L	A	Q	A	V		
121	tr K1XJR4 K1XJR4_MARBU	-PRNWSQSMGNDDQGF	WGL	TAMS	AAE	TNF	FTNP	PS	-	-	-	GSP	P	G	-	WL	L	A	L	A	Q	A	V	
122	tr A0A124BYC5 A0A124BYC5_ASPNG	-PSNWSSYMGNDQV	AWGL	AAMT	AAE	LN	YPED	-	-	-	-	AGM	P	S	-	WV	D	L	A	E	R	V		
123	tr A7TLL2 A7TLL2_VANPO	-PLNQSTTEGNDDQAF	WGI	AVMS	AAE	RNF	FTNP	P	-	-	-	ADR	P	Q	-	WL	Y	L	A	Q	A	V		
124	tr Q0CB86 Q0CB86_ASPTN	-PANYSSYLGNDQV	FWGI	AAMT	AAE	IN	FDP	V	-	-	-	PD	K	Y	S	-	WL	S	L	A	Q	G	V	
125	tr K1XLH0 K1XLH0_MARBU	-PLNQTRTLGNDDQC	FWGM	SAMT	AAE	TG	FED	PPE	-	-	-	GQP	P	G	-	WL	A	L	V	Q	G	V		
126	tr J7S438 J7S438_KAZNA	-PLNQSTTEGNDDQAF	WGI	AAMT	AAE	RNF	FTNP	P	-	-	-	EDK	P	Q	-	WL	Y	L	A	Q	A	V		
127	tr Q4WG09 Q4WG09_ASPFU	-SGNYSQWLGNDDQM	FWGL	ASIT	ASE	TG	FPEI	-	-	-	-	SGK	P	T	-	WT	S	L	A	R	V	V		
128	tr H8WZ09 H8WZ09_CANO9	-PSNQSTTEGNDDQGV	WGM	AIME	AVE	RNF	FTNP	PE	-	-	-	S	-	-	HS	-	WL	E	M	A	Q	A	I	
129	tr I1RL09 I1RL09_GIBZE	-PSNHSASLGNDDQGF	WGM	SAM	LAA	ENK	FPN	PPE	-	-	-	DQA	Q	-	WL	L	A	L	A	Q	A	V		
130	tr Q4W985 Q4W985_ASPFU	-PTNQTKTEGNDDQAF	WAF	AAMS	AAE	RNF	FPD	PDP	-	D	-	HG	P	G	-	WL	A	M	A	Q	A	V		
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	-PANQTMVEGNDDQGI	WGI	ITV	MGA	VER	NFT	DPD	-	-	-	NGA	P	G	-	WL	A	M	V	Q	A	V		
132	tr W6QCW3 W6QCW3_PENRF	-PANQSKTEGNDDQV	FWGF	AAMT	AAE	RNF	FPN	PPA	-	-	-	DQP	Q	-	WL	E	M	A	Q	A	V			
133	tr W3WMD3 W3WMD3_PESFW	-PANWSASMGNDQAF	WALT	SSL	VAA	ETG	F	TD	PPE	-	-	-	DQP	Q	-	WL	S	L	A	Q	A	V		
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	-PLNQSTTEGNDDQAF	WGI	AVLN	AAE	RK	FPN	PP	-	-	-	SDE	P	Q	-	WL	Y	L	A	Q	A	V		
135	tr I2GYH1 I2GYH1_TETBL	-PQNQTMVEGNDDQCI	WGL	ITV	MDA	VER	NFT	NPT	-	-	-	EG	-	P	G	-	WL	A	M	G	Q	A	V	
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	-PDNQTMVEGNDDQGV	WGL	ITV	MGA	VER	NFT	DPG	-	-	-	DGK	P	G	-	WL	A	M	A	Q	A	I		
137	tr Q752P3 Q752P3_ASHGO	-PANQSTTEGNDDQGV	WALT	FIM	GAVE	RNF	FKD	PE	-	-	KEG	M	P	G	-	WL	T	M	V	Q	T	V		
138	tr B8MYP3 B8MYP3_ASPFN	-PANYSNYLGNDQV	FWGL	AAMT	AAE	LN	FPEK	-	-	-	-	DDD	S	S	-	WL	S	L	A	Q	G	V		
139	tr Q2TXL6 Q2TXL6_ASPOR	-PDNYTQDLGNDDQV	FWGL	AAMT	AAE	LN	FPE	D	-	-	-	-	EEV	S	-	WL	L	A	L	A	Q	G	V	
140	tr Q0CN17 Q0CN17_ASPTN	-PTNYSQWLGNDDQV	FWGL	AAMT	AAE	LN	FPEQ	-	-	-	-	DGQ	P	S	-	WL	S	L	A	Q	G	V		
141	tr J3NHD5 J3NHD5_GAGT3	-PSNYTASLGNDDQGF	WGM	SAMT	AAE	LN	FPN	PPK	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	-PTNQTKALGNDDQGM	WGM	AAMT	AAE	YN	FPN	PPK	-	-	-	GQP	Q	-	WL	L	A	L	A	Q	A	V		
143	tr A1CVB0 A1CVB0_NEOFI	-PTNQTKTEGNDDQAF	WAF	AAMS	AAE	RNF	FPD	PDP	-	D	-	HG	P	G	-	WL	A	M	A	Q	A	V		
144	tr B8NVI3 B8NVI3_ASPFN	-PDNYTQDLGNDDQV	FWGL	AAMT	AAE	LN	FPE	D	-	-	-	-	EEV	S	-	WL	L	A	L	A	Q	G	V	
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	-PLNQSVTEGNDDQAF	WGL	VMA	AAE	RNF	FEN	PA	-	-	-	ENE	P	Q	-	WL	Y	L	A	Q	A	V		
146	tr A1DJ54 A1DJ54_NEOFI	-SGNYSQWLGNDDQM	FWGL	ASIT	ASE	TG	FPEI	-	-	-	-	SDK	P	T	-	WT	S	L	A	R	V	V		
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	-PLNFTASLGNDDQGF	WGMT	AML	LAA	ENK	FPN	PPA	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	-PLNQSTTEGNDDQGF	WGI	AVMA	AAE	RNF	FPN	PP	-	-	-	DDQ	P	Q	-	WL	Y	L	A	Q	A	V		
149	tr A0A177A618 A0A177A618_9PEZI	-PINASKSMGNDDQGF	WGMT	AMMA	AAE	RNF	QNP	PK	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
150	tr Q75CW6 Q75CW6_ASHGO	-PSHQKMIEGNDDQGI	WGL	LVM	GAA	RNF	FPDA	P	-	-	-	PER	A	P	G	-	WL	L	A	L	A	Q	A	V
151	tr Q7S4K4 Q7S4K4_NEUCR	-PLNFTASLGNDDQGF	WGMT	AML	LAA	ENK	FPN	PPA	-	-	-	DQP	Q	-	WL	L	A	L	A	Q	A	V		
152	tr G2QB99 G2QB99_MYCTT	-PPNQTRTLGNDDQGF	WGM	AAMS	AAE	ENN	FPN	PPE	-	-	-	DQP	Q											

172	tr B6HLM8 B6HLM8_PENRW	-PS-WAIGVGNDDQMFWGLAAITAAEYKFPNR--	--PSGDT-WLTLAIGV
173	tr A0A117DWS0 A0A117DWS0_ASPNG	-PANDSNYLGNDQVFWGLAAMTAAELNYPEQ--	--DGQPS-WLSLAQGV
174	tr A0A124BXM1 A0A124BXM1_ASPNG	-PANYSSYLGNDQMFWGLAAITAAEIEFADS--	--SSGYS-WLALAQGV
175	tr J3NZQ6 J3NZQ6_GAGT3	-PRNHRASLGNDQGFWGMTAMLAAENKFPDAPG	SGSTVPG-WLGLAQAV
176	tr W0THH8 W0THH8_KLUMD	-PNNQTMVEGNDDQGVWALTIMSAVERNFTNPK-	-KAGAPD-WLAMVQAV
177	tr B2AEF2 B2AEF2_PODAN	-PPNHTASLGNDQGFWGMSAMLAAENKFPNPPE--	--DQPQ-WLALAQAV
178	tr Q96TX1 Q96TX1_NEUCS	-PLNVTASLGNDQGFWGMTAMTAAENNFPNPPE--	--DQPQ-WLGLAQAV
179	tr E9DYA6 E9DYA6_METAQ	-PRNVTASLGNDQAFWGMAAMLAAEIKFPDPPA--	--DRPG-WLGLAQAV
180	tr B9WAI2 B9WAI2_CANDC	-PLNQSTTEGNDDQAFWGIAVMAAAERNFTNPK--	--DSTKA-WLTLAQAV
181	tr Q6C171 Q6C171_YARLI	-VSNFSSSEGNDDQGVWAMTMMTAAEKNFPEPG--	--DEYG-WLYVAQGA
182	tr A6ZMV1 A6ZMV1_YEAS7	-PSNQTMVEGNDDQGIWGITVMGAVERNFTDPG--	--DGKPG-WLAMVQAV
183	tr B6K7X7 B6K7X7_SCHJY	-PSSQTLDLGNDDQAFWALTAMSAAELNFTESSS--	--QSSS-WLEMAQAV
184	tr I1RSA2 I1RSA2_GIBZE	-PDNQTMTEGNDDQGFWAMAAMSAAEHQFPNPPE--	--DSPG-WLAQAQAV
185	tr F7VWZ9 F7VWZ9_SORMK	-PANQSRRTSSNDDVGFWTLTAMSAAEDAFPDPPE--	--DQPQ-WLALVQAV
186	tr A0A1B2J789 A0A1B2J789_PICPA	-PLNQTTTEGNDDQGFWGITAMAAVERNFTNPP--	--EDEPQ-WLYLAQAT
187	tr G0V8N4 G0V8N4_NAUCC	-PLNQSTTEGNDDQAFWGITVMTAAERNFTNPP--	--EDQPQ-WLYLAQAV
188	tr H8WXN8 H8WXN8_CANO9	-PLNQSTTEGNDDQGFWGIAVMGAAERNFSNPD--	--DEKLS-WLGLAQAV
189	tr A6ZZS0 A6ZZS0_YEAS7	-PLNQSTTEGNDDQAFWGIAAMTAAERNFTNPP--	--ENEPQ-WLYLAQAV
190	tr G9MQD1 G9MQD1_HYPVG	-PLNHTASLGNDQGFWGMTAMLAAENKFPNPPE--	--DKPQ-WLALAQAV
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	-PLNQSVTEGNDDQAFWGLVVMAAAERNFENPS--	--SDEPQ-WLYLAQAV
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	-SGNYSQWLGNDQMFWGLASITASETGFPFI--	--SGKPT-WTSLARVV
193	tr A0A254UET1 A0A254UET1_ASPNG	-PANQTRTEGNDDQAFWAFALRAAERNFPNPQP--	--TNRRGYLSR---
194	tr A1D587 A1D587_NEOFI	-PSNYSNYLGNDQVFWGLAAMTAAELNYPEE--	--DGQPS-WVSLAQGV
195	tr Q1K7I4 Q1K7I4_NEUCR	-PLNVTASLGNDQGFWGMTAMTAAENNFPNPPE--	--DQPQ-WLGLAQAV
196	tr G8JS88 G8JS88_ERECY	-PLNQSTTEGNDDQAFWGIAVMSAAEKGFPNPD--	--PDQPQ-WLALAQAV
197	tr A0A136J7D3 A0A136J7D3_9PEZI	-PVNQTRTLGNDDQGFWAMAAMSAAEAKFPDPS--	--DEPQ-WLALAQAV
198	tr E9E4X8 E9E4X8_METAQ	-PRNVTASLGNDQGFWGMAAMSAAENGFPDPPS--	--DKPQ-WLELAQAV
199	tr Q6CP42 Q6CP42_KLULA	-PLNQTVTEGNDDQAFWGFVMAAAERNYPNPPE--	--DDEPQ-WLYLAQAV
200	tr G0WHL4 G0WHL4_NAUDC	-PANQSMVEGNDDQGVWGLTLMGAAERNFTDPD--	--DGTGP-WLAMTQAV
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	-PLNQSTTEGNDDQAFWGIAAMTAAERNFTNPP--	--SDQPQ-WLYLAQAV
202	tr F9WYX6 F9WYX6_ZYMTI	-PPNQSKSLGNDDQAAWALAAMSAAEYDLPVPND	LLSNNIT-WASIADTV
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	-PSNYTASLGNDQGFWGMSAMTAAELNFPNPPK--	--DQPQ-WLSLAQAV
204	tr J3P147 J3P147_GAGT3	-PPAHRASLGNDQGFWGLAVMQAAEMRFPDSPKE--	--GDAH-WLALAQGV
205	tr I2H842 I2H842_TETBL	-PLNQSTTEGNDDQAFWGIAAITAAERNFTNPP--	--PDKPQ-WLYLAQAV
206	tr S6E3R3 S6E3R3_ZYGB2	-PDNQTMVEGNDDQGVWGLTVMGAVERNFTDPG--	--DGKPG-WLAMAQAI
207	tr E5AD94 E5AD94_LEPMJ	-PSNQSKNEGNDDQVFWAFSMIAAAEYKLPDPPA--	--DQPG-WLAMTQSV
208	tr Q6FJM7 Q6FJM7_CANGA	-PANQTTVEGNDDQGVWGLTILDAVERNFSAPI--	--DGKPG-WLAMSQGI
209	tr A5DUW0 A5DUW0_LODEL	-PLNQSTTEGNDDQGFWGIAVMGAAERNFSNPD--	--SDEIS-WLGLAQAV
210	tr G8BF46 G8BF46_CANPC	-PLNQSTTEGNDDQGFWGIAVMGAAERNFSNPN--	--NKELS-WLGLAQAV
211	tr W3XAF6 W3XAF6_PESFW	-PDNQTRSLGNDDQGFWALTAMSAAELNFTNPPD--	--DEPG-WLGLTQAV
212	tr B6H7I2 B6H7I2_PENRW	-PSNYSSYLGNDQMFWGLAAMMAAELKFPDV--	--EDGFS-WLSLAQGV
213	tr Q5ATF9 Q5ATF9_EMENI	-PSNYSSYLGyddQFFWGATAMLAAEIGFPED--	--DVEYS-WLSLAQGV
214	tr A0A167CDD3 A0A167CDD3_9ASCO	-PSNQTTTEGNDDQGYWGVTVMAAAEKNFSDPG--	--PGNPS-WLYLAQGV
215	tr G0RXS3 G0RXS3_CHATD	-PRNWTLSLGNDDQGFWGMAAMLAAETNFPNPAP--	--DEPQ-WLALAQAV
216	tr B8NX26 B8NX26_ASPFN	-SSNYSQWLGNDQMFWGLAAITASEAGFPER--	--DGKPS-WTSLARAV
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	-PLNQTSSTLGNDQAFWGMAAMSAAENKFPNPDP--	--DKPQ-WLELAQAV
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	-PSNQTRTEGNDDQAFWAFTALMAAEHNFPNPPE--	--DSPS-WLAMARAV
219	tr A1CMM3 A1CMM3_ASPCL	-PPNQSRTRLGNDDQAFWGLAAMTAAEVRFNPPE--	--DQPQ-WLALAQAV
220	tr B6HU84 B6HU84_PENRW	-PSNYSQYLGNDQVFWGLAAITAAEFNFPER--	--EGEPS-WVSLAQGV
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	-PLNVTASLGNDQGFWGMTAMTAAENNFPNPPE--	--DQPQ-WLGLAQAV
222	tr G2WHY6 G2WHY6_YEASK	-PLNQSTTEGNDDQAFWGIAAMTAAERNFTNPP--	--ENEPQ-WLYLAQAV
223	tr G0W7G8 G0W7G8_NAUDC	-PLNQSTTEGNDDQAFWGIAVMTAAERNFTNPP--	--EDSPQ-WLYLAQAV
224	tr G4NGV8 G4NGV8_MAGO7	-PVNQTRTLGNDDQGFWALAAMSAAESRYPDPPP--	--GQPQ-WLALAQAV
225	tr W3X855 W3X855_PESFW	-PENQTLTEGNDDQGFWALSAMTAAEMGFQNPGE--	--DGVQ-WLAAAQAV
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	-PNNYTASLGNDQGFWGMSAMLAAEVKFPDPPS--	--DKPS-WLALAQAV
227	tr J8Q2K7 J8Q2K7_SACAR	-PLNQSTTEGNDDQAFWGIAAMTAAERNFTNPP--	--ESDPQ-WLYLAQAV
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	-PLNQSTTEGNDDQGFWGIAVMAAAEKNFSNPE--	--NPKKA-WLSLAQAV
229	tr J6EGN8 J6EGN8_SACK1	-PANQTMVEGNDDQGIWGITVMGAVERNFTDPD--	--EGKPG-WLAMVQAV
230	tr A0A100ISD9 A0A100ISD9_ASPNG	-PANQTRTEGNDDQSFWAFAAMSAAERNFPDPPP--	--SSGSPG-WLAMAQAV
231	tr G8YQC0 G8YQC0_PICSO	-PSNQSMTEGNDDQGIWGLTTMEAERNFTDPP--	---HS-WLSMTQAI
232	tr F7VVP8 F7VVP8_SORMK	-PLNYTASLGNDQGFWGMTAMLAAENKFPNPDP--	--DQPQ-WLALAQAV
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	-PSNQTMVEGNDDQGIWGITVMGAVERNFTDPG--	--DGKPG-WLAMVQAV
234	tr F8MBP4 F8MBP4_NEUT8	-PANQSRRTSSNDDVGFWTITAMMAAEDAYPDPE--	--DQPQ-WLALVQAV
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	-PSNWSSYIANDDQMAWGLAAMTAAELNYPED--	--AGMPS-WVDLAERV
236	tr A1CD27 A1CD27_ASPCL	-SKNYSQWLGNDQMFWGLASITASETGFPDI--	--SGKPT-WTSLARVV
237	tr A5DHF3 A5DHF3_PICGU	-PSNQSLTEGNDDQGVWGMAIIAAVERNFTDPE--	---HS-WLSMAQAI
238	tr D4ATT7 D4ATT7_ARTBC	-PRNQSRSLGNDDQAFWAISAMSAVENKFPDPPP--	--DQPQ-WLYLVQAV
239	tr E2PT42 E2PT42_ASPNC	-PANQTRTEGNDDQSFWAFAAMSAAERNFPDPPP--	--SSGSPG-WLAMAQAV
240	tr C7ZPE5 C7ZPE5_NECH7	-PGNVTLSLGNDDQGFWGMSAMLAAELKFPNPPE--	--DRPG-WLALAQAV

241	tr A0A0A2VM24 A0A0A2VM24_PARBA	-PTNQTKTEGNDQDAFWAMSAMSAAEERNFPNPSS--DQPQ-WLALVQAV
242	tr Q4WKP7 Q4WKP7_ASPFU	-PSNYSNYLGNDQVFWGLAAMTAAELNFPPEE--DGQPS-WVSLAQGV
243	tr F8MXT4 F8MXT4_NEUT8	-PLNVTASLGNDQGFWMGTAMTAAENNFPNPAP--DQPQ-WLGLAQAV
244	tr A0A177AJ74 A0A177AJ74_9PEZI	-PTNQTKALGNDQGFWGMAAMTAAEYNFPNPPK--DQPQ-WLALAQAV
245	tr J7RQR5 J7RQR5_KAZNA	-PENQTMVEGNDQGIWGLTIMDAVERNFTKPT--KDGVPK-WLAMSQAV
246	tr Q75CW7 Q75CW7_ASHGO	-PSHQKMVEGNDQGIWGLLVMGAAERNFPDAP--PERAPG-WLALAQAV
247	tr G3AKK4 G3AKK4_SPAPN	-PSNQSMTEGNDQGVWGMGIMQAAERNFTNPE--D--HS-WISLTQAI
248	tr B2W762 B2W762_PYRTR	-PANQTKTLGNDQAFWGMAAMTAAENKLPDLPD--GKPS-WLSLAQAV
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	-PLNQSTTEGNDQAFWGIAAMTAAERNFTNPP--ENEPQ-WLYLAQAV
250	tr C4XWK1 C4XWK1_CLAL4	-PSNQSMTEGNDQGVWAMAI IQAVERNFTNTG--D--HS-WLYMTQAI

001	tr A0A254U2J9 A0A254U2J9_ASPNG	YNTQI--DRWD--D--STCGGGLRWQIYPYE--AGYAMKNSISNGG
002	sp Q6FLP9 DCW1_CANGA	FNTMA--LRWD--S--ETCGGGLRWQIFVWN--SGYDYKNTVSNGA
003	tr A1CCM5 A1CCM5_ASPCL	YNTQA--GRWD--P--ATCGGGLRWQIWPYQ--AGYTMKNAISNGG
004	tr C1H190 C1H190_PARBA	FNTQA--PRWA--N--HTCNGGLKWSINTFG--SGWTYKNTISNGC
005	tr Q2UR85 Q2UR85_ASPOR	FNTQV--PRWD--T--SSCDGGLRWQIWPYQ--AGYTTKNAISNGG
006	tr A5DV30 A5DV30_LODEL	FNTMN--ARWD--T--AHCNGLRWQIFTWN--SGYNYKNSISNGC
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	YNSMA--DRWD--A--DNCGGGLRWQIMSNM--SGWTYKSTVSNAG
008	tr J3PGN0 J3PGN0_GAGT3	WTTQA--APER--H--DNTCNGGMRWQVPPTN--AGYDYKNTIANGC
009	tr A0A124BXU6 A0A124BXU6_ASPNG	FNTQV--ARWD--N--STCDGGMRWQIYTYE--SGYTMKNAISNGG
010	tr C5P4A1 C5P4A1_COCP7	FNSQA--ARWN--T--ATCGGGLNWQIFQFN--RGFNYKNSISNGC
011	tr Q0CG55 Q0CG55_ASPTN	FNTQA--ARWD--T--ETCGGGLRWQIFTFN--NGFNYKNTISNGC
012	tr C4Y2X5 C4Y2X5_CLAL4	FNTMS--ARWD--L--GECNGLRWQIFQWN--SGYDYKNSVSNGA
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	FNTQA--ARWD--K--STCNGLRWQIFTFN--NGWTYKNTISNGC
014	tr C7Z068 C7Z068_NECH7	WTTQA--HPNR--H--DKECNGLRWQIPFTN--AGYNYKNTIANGC
015	tr G8BXX2 G8BXX2_TETPH	FNTMA--LRWD--T--DSCGGGLRWQIFQWN--SGYDYKNSVSNGA
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	FNTMS--SRWD--M--DHCNGLRWQIFQWN--SGYDYKNSVSNGA
017	tr A1CSC2 A1CSC2_ASPCL	FNTQV--PRWD--T--STCHGGLRWQIWPYQ--DGYRTKNAISNGG
018	sp Q05031 DFG5_YEAST	FNTMY--SRWD--S--EHCGGGLRWQIFTWN--SGYNYKNTVSNAC
019	tr H2AQJ6 H2AQJ6_KAZAF	FNEMW--SRWD--S--ANCGGGLRWQIFTWN--SGYDYKSTISNVC
020	tr G2XFH9 G2XFH9_VERDV	WSTQN--D--R--F--DDHCNGLRWQAVSTN--AGYNYKNTIANGC
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	FNTMA--ARWD--S--ATCGGGLRWQIFTWN--SGYDYKNTVSNGA
022	sp Q5AD78 DCW1_CANAL	FNTMQ--ARWD--T--ETCNGLRWQIFQWN--SGYDYKNSVSNGA
023	tr B6H7E8 B6H7E8_PENRW	FNGQI--PRWD--M--TACNGLRWQIWPYQ--DGYNLKNAISNGG
024	tr B6GZU2 B6GZU2_PENRW	FNTQV--PRWD--T--SSCGGGLRWQIFTWN--NGYNYKNTISSGG
025	tr E9E4W7 E9E4W7_METAQ	WNTQA--DPSR--Y--DETCNGLRWQIPRTN--AGYDYKNTIANGI
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	FNTQA--PRWD--P--ATCGGGLRWQIFTWN--NGYNYKNTISTGG
027	tr G2R7G8 G2R7G8_THITE	FNTQA--WR-W--D--MEYCNGLHWQIPTTN--GGYDYKNSIANVV
028	tr G3JBY1 G3JBY1_CORMM	FNEYV--VRWN--EE--NGLCGGGLRWQVYSFL--KGYDYKNSIANGC
029	tr A5DNV5 A5DNV5_PICGU	FNTMQ--ARWD--H--NDCHGGLRWQIFQWN--SGYDYKNSVSNGC
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	FNTQA--QRWD--K--STCGGGLKWQIYTFN--NGYTYKNTISNGC
031	tr Q2TWC1 Q2TWC1_ASPOR	FNMQI--ERWD--E--TACNGLRWQIWPYQ--AGYTMKNSISNGG
032	tr J5RXY9 J5RXY9_SACK1	FNTMA--LRWD--A--DSCGGGLRWQIFVWN--SGYDYKNTVSNGA
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	FNEYA--VRWE--EE--NGLCGGGLRWQVYSFL--NGWAYKNSISNGC
034	tr A0A254U5V1 A0A254U5V1_ASPNG	FNT-----Q--AGYTTKNAISNGG
035	tr A0A061B6H7 A0A061B6H7_CYBFA	FNTMS--SRWD--H--DNCGGGLRWQIFEWN--SGYDYKNSVSNGC
036	tr G8ZQ93 G8ZQ93_TORDC	FNTMA--LRWD--T--TSCNGLRWQIFMWN--TGYDYKNTVSNGA
037	tr Q9C2J1 Q9C2J1_NEUCS	FNQMA--SRWD--D--LNCGGGLRWAINDFQ--TGKDYKNSISNGI
038	tr C5NZK5 C5NZK5_COCP7	LHSQV--PRWD--D--ETCGGGLRWQIFSFN--KGYNYKNSISNGC
039	tr Q6C0T7 Q6C0T7_YARLI	FNTMA--SRWD--D--ICGGGLHWQIYQWN--SGYSYKNAIASGA
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	FNTMA--LRWD--S--ETCGGGLRWQIFVWN--SGYDYKNTVSNGA
041	tr F7VZ72 F7VZ72_SORMK	FNTQA--HPDR--H--DETCNGLRWQIPPLN--NGYNYKNSIANGC
042	tr A3LN37 A3LN37_PICST	FNTMT--ARWD--S--AECNGLRWQIFQWN--SGYDYKNSVSNGC
043	tr Q6CAI2 Q6CAI2_YARLI	FNTMS--ARWD--T--DNCGGGLRWQIYTWN--NGYDYKNTVSNGC
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	FNTMT--ARWD--H--KECHGGLRWQIFQWN--SGYDYKNSVSNGC
045	tr G9MJS5 G9MJS5_HYPVG	FNTQA--SPER--H--DGTGGGIRWQIPPTN--PGYSYKNSIANGC
046	tr G9MHI4 G9MHI4_HYPVG	FNQYV--GRWD----SHCGGGLRWQIFSWN--KGYDYKNSVANGC
047	tr D4AP27 D4AP27_ARTBC	FNSQI--PRWD--T--ATCGGGLRWQIFRFN--RGFDYKNTISNGA
048	tr B6GZT8 B6GZT8_PENRW	FNTQI--KRWD--T--SSCGGGLRWQIWPYQ--SGYTMKNSISNGG
049	sp O74556 YCZ2_SCHPO	FNQQV--TRWD--T--DHCNGLRWQITEFN--SGYNYKNTVSNGA
050	tr A0A0C4E991 A0A0C4E991_MAGP6	WTTQA--APER--H--DNTCNGGMRWQVPPTN--AGYDYKNTIANGC
051	tr B8MHG0 B8MHG0_TALSN	FNDFV--ARWEVDK--ASCGGGLRWQLYPTL--AGYDSKNAISNGA
052	tr A3LMV8 A3LMV8_PICST	FNTMN--ARWD--T--DHCGGGLRWQIFTWN--SGYSYKNSVSNGC
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	YNNMA--PRWD--D--QNCGGGLRWQILD TM--NGWDYKSMISTGG
054	sp Q9P6I3 YHG7_SCHPO	FNEIM--SRWD--T--SSCGGGVRWQIYSFN--NGYSYKNSISNGI
055	tr Q2US57 Q2US57_ASPOR	FNTQV--KRWD--T--SSCAGGMRWQIW TYE--AGYRMKNSISNGG
056	tr W7N622 W7N622_GIBM7	FNEYV--SRWD--T--QHCDGGMRWQIFTWN--AGYDYKNSISNGC
057	sp Q9P6I4 YHG6_SCHPO	FNRMS--GRWD--S--STCGGGLRWQAF AWL--NGYSYKASVSNAL

058	tr C7GRB3 C7GRB3_YEAS2	FNTMY----	SRWD--S----	EHCGGGGLRWQIFTWN--	SGYNYKNTVSNAC
059	tr C7GP28 C7GP28_YEAS2	FNTMA----	LRWD--A----	DSCGGGLRWQIFVWN--	SGYDYKNTVSNGA
060	tr G3JGZ5 G3JGZ5_CORMM	WATQA----	DPSR--H----	SYCNGGMRWQIPFSN--	AGYDYKNTIANGC
061	tr C4QXV4 C4QXV4_KOMPG	VNTMW----	ERWD--L----	EHcNGGLRWQIFQWN--	AGYNYKNTVSNAC
062	tr I1S0Z5 I1S0Z5_GIBZE	FNTQA----	NPDR--H----	DTcGGGLRWQIPRTN--	NGYDYKNSIANGC
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	FNTQA----	ARWD--E----	DTcGGGLRWQIFSFN--	NGWNYKNTISNGC
064	tr G0W5X4 G0W5X4_NAUDC	FNTMA----	LRWD--A----	QNCGGGLRWQIFVWN--	SGYDYKNSVSNGA
065	tr G8ZPC1 G8ZPC1_TORDC	FNQLN----	SRWD--T----	ANcGGGLRWQIFTWN--	GGYNYKNTISNAC
066	tr A0A167CDC5 A0A167CDC5_9ASCO	FNTMA----	SRWD--T----	DTCHGGLRWQIFTWN--	NGYNYKNSVANGC
067	tr R9XAT7 R9XAT7_ASHAC	FNTMA----	LRWD--T----	DTcNGGLRWQIFRWN--	AGYHYKNSVSNGA
068	tr B2B747 B2B747_PODAN	FNTQA----	SPDR--H----	DETCNGGLRWQIPWSN--	NGYNYKNSIANGC
069	tr G0SE99 G0SE99_CHATD	FNTQA----	SPTR--H----	DTcGGGLRWQIPWSN--	NGYNYKNSIANGC
070	tr J8Q6F0 J8Q6F0_SACAR	FNTMY----	SRWD--S----	ENCGGGLRWQIFTWN--	SGYNYKNTVSNAC
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	FNEMY----	SRWD--T----	ANcGGGLRWQIFTWN--	SGYNYKNTISNGC
072	tr J3K6E2 J3K6E2_COCIM	FNSQA----	ARWN--T----	ATcGGGLNWQIFQFN--	RGFNYKNSISNGC
073	tr G4N3E1 G4N3E1_MAGO7	WTTQN----	SPER--H----	DKcGGGMRWQIPPLN--	AGYDYKNTIANAI
074	tr C5M5J2 C5M5J2_CANTT	FNTMY----	ARWD--D----	EHcGGGLRWQIFTWN--	SGYDYKNSISNGC
075	tr A0A254U0X0 A0A254U0X0_ASPNG	FNMQV----	NRWD--D----	TTCdGGMRWQIWPYQ--	AGYTMKNAISNGG
076	sp Q75DG6 DCW1_ASHGO	FNTMA----	LRWD--M----	ETcNGGLRWQIFRWN--	DGYHYKNSVSNGA
077	tr Q6BZF0 Q6BZF0_DEBHA	FNTMT----	ARWD--H----	KECHGGLRWQIFQWN--	SGYDYKNSVSNGC
078	tr E9E3Q1 E9E3Q1_METAQ	WNTQA----	DPSR--Y----	DETCNGGLRWQIPFSN--	QGYGYKNTIANGI
079	sp P36091 DCW1_YEAST	FNTMA----	LRWD--A----	DSCGGGLRWQIFVWN--	SGYDYKNTVSNGA
080	tr W0TAH6 W0TAH6_KLUMD	FNTMA----	SRWD--S----	DTcGGGLRWQIFTWN--	SGYDYKNSVSNAA
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	FNTMW----	ARWD--D----	QYCGGGLRWQIFTWN--	SGYTYKNTISTAC
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	FNDYA----	SRWD--E----	ASCsGGGLRWQIYTFN--	AGYNYKNTISNGC
083	tr Q2TYU3 Q2TYU3_ASPOR	FNTQV----	ARWD--S----	SNCGGGLRWQIFPYQ--	AGYAMKNSISNGL
084	tr G2XGF6 G2XGF6_VERDV	FNQWA----	QRWI--EA--S	SDSCGGGLRWQIFSFN--	NGFNYKNSISNGC
085	tr G3AMT4 G3AMT4_SPAPN	FNTMT----	ARWD--E----	NEcNGGLRWQIFQWN--	SGYDYKNSVSNGC
086	tr A0A1C1D213 A0A1C1D213_9EURO	FNTQA----	ARWD--T----	MSCGGGLKWQIFAFN--	NGYNYKNTISNGC
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	FHTQA----	DPGR--H----	DDVCGGGLRWQIPMAN--	IGYDYKNSISNGI
088	tr A0A100I5A6 A0A100I5A6_ASPNG	FNMQV----	NRWD--D----	TACdGGMRWQIWPYQ--	AGYTMKNAISNGG
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	FNLMY----	SRWD--D----	QHCGGGLRWQIFTWN--	SGYNYKNTVSNGC
090	tr G2Q8A7 G2Q8A7_MYCTT	FNTQA----	SPEE--H----	DTcNGGLRWQIYLSN--	NGYDYKNSIANGC
091	tr B8NPZ1 B8NPZ1_ASPFN	FNMEV----	ERWD--K----	SACNGGMRWQLWPYQ--	EGYTMKNAISNGG
092	tr Q7SAB2 Q7SAB2_NEUCR	FHTQA----	DPGR--H----	DDVCGGGLRWQIPMAN--	IGYDYKNSISNGI
093	tr A2R8R5 A2R8R5_ASPNC	FNTQV----	ARWD--N----	STcNGGMRWQIYTYE--	SGYTMKNAISNGG
094	tr C5DHG0 C5DHG0_LACTC	FNTMW----	MRWD--S----	EHcGGGVRWQIFTWN--	SGYNYKNTVSNAC
095	tr G0SFA3 G0SFA3_CHATD	FNQWA----	TRWE--E----	STcQGGLRWQIFSFN--	AGYNYKNSISNGC
096	tr K1WJG5 K1WJG5_MARBU	FNEMV----	ARWD--T----	STcGGGMRWQIFTFN--	SGYTYKNSIANGC
097	tr H2B005 H2B005_KAZAF	FNTMA----	LRWD--D----	ATcGGGLRWQIFTWN--	SGYDYKNTVSNGA
098	tr C5DYD7 C5DYD7_ZYGRC	FNEIN----	ERWD--D----	KSCAGGVRWQIFTWN--	SGYNYKNTISNGC
099	tr G8YM23 G8YM23_PICSO	FNTMS----	ARWD--T----	KDCGGGLRWQIFQWN--	SGYDYKNTVSTGC
100	tr W3X8E3 W3X8E3_PESFW	FNTQA----	DPGR--H----	DTcGGGLRWQIPFSN--	NGYNYKNSIANGC
101	tr A0A254U3K7 A0A254U3K7_ASPNG	FNTQV----	ARWD--N----	STcNGGMRWQIYTYE--	SGYTMKNAISNGG
102	tr Q6CER8 Q6CER8_YARLI	FNSMA----	DRWD--T----	QVCDGGLHWQIFQWN--	SGYDYKNAIANGA
103	tr G2QT05 G2QT05_THITE	WTTQA----	SPDR--H----	DNTcGGGLRWQIPVVN--	NGYDYKNSIANGC
104	sp Q5ACZ2 DFG5_CANAL	FNTMN----	ARWD--A----	DNCGGGLRWQIFTWN--	SGYDYKNSISNGC
105	tr G2WKU6 G2WKU6_YEASK	FNTMY----	SRWD--S----	EHcGGGLRWQIFTWN--	SGYNYKNTVSNAC
106	tr A0A0C7N752 A0A0C7N752_9SACH	FNTMY----	SRWD--D----	QHCGGGVRWQIFTWN--	SGYNYKNTVSNAC
107	tr W3WN68 W3WN68_PESFW	FNEYV----	TRWE--DA--E	DTcGGGLRWQIYTFN--	NGYNYKNSISNGC
108	tr G2QFS1 G2QFS1_MYCTT	FHTQA----	APDR--H----	DGTcNGGLRWQVPPTN--	AGYNYKNTIANAC
109	tr G3J9G4 G3J9G4_CORMM	FNEYA----	VRWE--EE--N	GLcGGGLRWQVYSFL--	NGWAYKNSISNGC
110	tr Q2UJ03 Q2UJ03_ASPOR	FNTQA----	PRWN--T----	ETcGGGLRWQIFSFN--	NGYHYKNTIANGC
111	tr G8YRT2 G8YRT2_PICSO	YNTMN----	DRWD-------	SHcGGGLRWQIFTWN--	SGYDYKNSIANGC
112	tr W3X554 W3X554_PESFW	FNEYV----	DRWD--P----	DTcGGGLRWQIFTFN--	NGFNYKNSISNGC
113	tr Q1K7A8 Q1K7A8_NEUCR	FNQMA----	SRWD--D----	LNCGGGLRWAINDFQ--	TGKDYKNSISNGI
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	FNTQV----	PRWD--T----	STCHGGLRWQISTYQ--	DGYRTKNAISNGG
115	tr A7F3P0 A7F3P0_SCLS1	FNDMA----	ARWD--E----	STcGGGLRWQIFTFN--	SGYNYKNSIANGC
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	FNEYV----	IRWQ--EE--N	GLcGGGLRWQVYSFL--	KGYDYKNSIANGC
117	tr Q6CIP9 Q6CIP9_KLULA	FNQMY----	SRWD--S----	DHCNGGLRWQIFTWN--	SGYNYKNTISNGC
118	tr H2AMQ5 H2AMQ5_KAZAF	FNTMA----	LRWD--T----	DSCGGGLRWQIFTWN--	SGYDYKNSVSNGA
119	tr G8JMI3 G8JMI3_ERECY	FNTMY----	ARWD--S----	QHCGGGLRWQIFTWN--	LGYDYKNTIANGC
120	tr C5DGT1 C5DGT1_LACTC	FNTMA----	LRWD--D----	ESCGGGLRWQIFTWN--	SGYDYKNTVSNGA
121	tr K1XJR4 K1XJR4_MARBU	FNSQS-----	-RR--T----	DGECGGGLRWQVFPYL--	IGYDYKNSIANGC
122	tr A0A124BYC5 A0A124BYC5_ASPNG	FDVQS----	ARWD--N----	SSCEGGLRWQIWPYE--	VGYTMKN SMSNGG
123	tr A7TLL2 A7TLL2_VANPO	FNTMA----	LRWD--D----	QTCNGGLRWQIFVWN--	SGYDYKNSVSNGA
124	tr Q0CB86 Q0CB86_ASPTN	YNTQV----	ARWD--T----	SNCGGGLRWQIFPYQ--	AGYNMKNSISNGG
125	tr K1XLH0 K1XLH0_MARBU	FNTQI----	PRWD--T----	TTCGGGLRWQIFPFN--	AGYTYKNSISNGC
126	tr J7S438 J7S438_KAZNA	FNTMA----	LRWD--T----	ESCGGGLRWQIFNWN--	SGYDYKNTVSNGA
127	tr Q4WG09 Q4WG09_ASPFU	FNMQV----	ARWD--K----	VACdGGMRWQIWPYQ--	AGYTMKNAISNGG

tr H8WZ09 H8WZ09_CANO9	FNTMD	---	ARWD	-	T	---	EH	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	N	G	C						
129 tr I1RL09 I1RL09_GIBZE	WTTQA	---	NPER	-	H	---	D	D	E	C	N	G	G	M	R	W	Q	I	P	F	T	N	---	S	G	Y	D	Y	K	N	T	I	A	N	G	C				
130 tr Q4W985 Q4W985_ASPFU	FNTQA	---	ARWD	-	E	---	D	T	C	G	G	G	L	R	W	Q	I	F	S	F	N	---	N	G	W	N	Y	K	N	T	I	S	N	G	C					
131 tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	FNTMY	---	SRWD	-	S	---	E	H	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	V	S	N	A	C					
132 tr W6QCW3 W6QCW3_PENRF	FNTQV	---	PRWD	-	T	---	T	S	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	N	G	Y	N	Y	K	N	T	I	S	T	G	G					
133 tr W3WMD3 W3WMD3_PESFW	FNEQT	---	HEER	-	R	V	P	A	G	S	N	C	E	W	G	L	R	W	Q	V	Y	R	T	N	---	N	G	F	D	Y	I	N	T	I	A	N	A	C		
134 tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	FNTMA	---	ARWD	-	S	---	A	T	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	D	Y	K	N	T	V	S	N	G	A					
135 tr I2GYH1 I2GYH1_TETBL	FNTMW	---	SRWD	-	A	---	A	D	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	D	Y	K	N	T	I	S	N	A	C					
136 tr A0A1S7HIA4 A0A1S7HIA4_9SACH	FNEVW	---	SRWD	-	D	---	E	H	C	G	G	G	V	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	N	A	C					
137 tr Q752P3 Q752P3_ASHGO	FNKME	---	ARWD	-	A	---	E	N	C	G	G	G	L	R	W	Q	I	F	Q	W	N	---	N	G	Y	T	Y	K	N	T	I	A	N	G	C					
138 tr B8MYP3 B8MYP3_ASPFN	FNTQV	---	PRWD	-	T	---	S	S	C	D	G	G	L	R	W	Q	I	W	P	Y	Q	---	A	G	Y	T	T	K	N	A	I	S	N	G	G					
139 tr Q2TXL6 Q2TXL6_ASPOR	FNTQA	---	EKWD	-	P	---	D	T	C	H	G	G	L	R	W	Q	R	N	S	W	N	---	G	G	Y	D	L	K	N	S	V	S	N	G	G					
140 tr Q0CN17 Q0CN17_ASPTN	FNTQV	---	PRWD	-	K	---	T	S	C	E	G	G	L	R	W	Q	I	W	P	Y	Q	---	A	G	Y	T	T	K	N	S	V	S	N	G	G					
141 tr J3NHD5 J3NHD5_GAGT3	FNTQA	---	APGR	-	H	---	D	N	T	C	G	G	G	L	R	W	Q	I	P	P	T	N	---	N	G	Y	D	Y	K	N	S	I	A	N	G	C				
142 tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	FATQA	---	SRWD	-	D	---	A	H	C	S	G	G	L	K	W	Q	I	F	R	F	N	---	V	G	Y	D	Y	K	N	A	I	S	N	G	C					
143 tr A1CVB0 A1CVB0_NEOFI	FNTQA	---	ARWD	-	E	---	D	T	C	G	G	G	L	R	W	Q	I	F	S	F	N	---	N	G	W	N	Y	K	N	T	I	S	N	G	C					
144 tr B8NVI3 B8NVI3_ASPFN	FNTQA	---	EKWD	-	P	---	D	T	C	H	G	G	L	R	W	Q	R	N	S	W	N	---	G	G	Y	D	L	K	N	S	V	S	N	G	G					
145 tr A0A0C7N2N9 A0A0C7N2N9_9SACH	FNTMA	---	LRWD	-	D	---	D	T	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	D	Y	K	N	T	V	S	N	G	A					
146 tr A1DJ54 A1DJ54_NEOFI	FNMQV	---	ARWD	-	K	---	V	A	C	D	G	G	M	R	W	Q	I	W	P	Y	Q	---	A	G	Y	T	M	K	N	S	I	S	N	G	G					
147 tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	WATQA	---	APDR	-	H	---	D	D	T	C	N	G	G	L	R	W	Q	I	P	P	T	N	---	N	G	Y	D	Y	K	N	T	I	A	N	A	I				
148 tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	FNTMA	---	LRWD	-	P	---	D	H	C	G	G	G	L	R	W	Q	I	F	E	W	N	---	S	G	Y	D	Y	K	N	S	V	S	N	G	C					
149 tr A0A177A618 A0A177A618_9PEZI	FSLQS	---	GRWD	-	T	---	E	T	C	G	G	G	L	K	W	Q	V	Y	V	F	N	---	K	G	Y	D	Y	K	N	S	I	S	N	G	A					
150 tr Q75CW6 Q75CW6_ASHGO	FNTMW	---	ARWD	-	P	---	E	N	C	G	G	G	L	R	W	Q	I	F	P	W	N	---	N	G	Y	D	Y	K	N	T	I	S	N	A	C					
151 tr Q7S4K4 Q7S4K4_NEUCR	WATQA	---	APDR	-	H	---	D	D	T	C	N	G	G	L	R	W	Q	I	P	P	T	N	---	N	G	Y	D	Y	K	N	T	I	A	N	A	I				
152 tr G2QB99 G2QB99_MYCTT	FNQYA	---	SRWE	-	E	---	A	T	C	G	G	G	L	R	W	Q	I	F	T	F	N	---	N	G	F	N	Y	K	N	S	I	S	N	G	C					
153 tr G0S3F2 G0S3F2_CHATD	WTTQA	---	SPER	-	H	---	D	G	T	C	N	G	G	L	R	W	Q	I	P	P	T	N	---	A	G	Y	N	Y	K	N	T	I	A	N	A	C				
154 tr A0A1E4SC85 A0A1E4SC85_9ASCO	FNTMS	---	ARWD	-	P	---	E	H	C	G	G	G	L	R	W	Q	I	F	Q	W	N	---	S	G	Y	N	Y	K	N	S	V	S	N	G	C					
155 tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	FNTMY	---	ARWD	-	M	---	Q	S	C	N	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	N	A	C					
156 tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	YNTQV	---	DRWD	-	T	---	S	T	C	G	G	G	L	R	W	Q	I	Y	T	Y	Q	---	A	G	Y	T	M	K	N	A	I	S	N	G	G					
157 tr G2QKJ0 G2QKJ0_MYCTT	FNTQA	---	PR	-	W	-	E	-	T	E	Y	C	A	G	G	L	R	W	Q	V	V	S	T	N	---	G	G	Y	D	Y	K	N	T	I	A	N	A	V		
158 tr Q4WFX5 Q4WFX5_ASPFU	FNEQI	---	ARWD	-	D	---	Q	A	C	G	G	G	L	N	W	Q	I	F	P	F	N	---	N	G	Y	T	Y	R	N	S	I	S	N	G	G					
159 tr Q5BGD7 Q5BGD7_EMENI	FNMQA	---	NRWD	-	E	---	R	A	C	D	G	G	I	T	W	Q	I	H	P	W	Q	---	A	G	Y	T	L	R	N	S	I	S	N	G	G					
160 tr A0A0A8L8U3 A0A0A8L8U3_9SACH	FNTMS	---	ARWD	-	E	---	D	S	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	D	Y	K	N	S	V	S	N	A	A					
161 tr G8BYN9 G8BYN9_TETPH	FNTMY	---	ARWD	-	T	---	A	Y	C	G	G	G	L	R	W	Q	I	Y	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	N	A	C					
162 tr F7VVT4 F7VVT4_SORMK	FNTQA	---	ADDR	-	H	---	D	D	E	C	G	G	G	L	R	W	Q	I	F	H	S	N	---	A	G	Y	D	Y	K	N	T	I	A	N	A	C				
163 tr G8YB80 G8YB80_PICSO	FNTMS	---	ARWD	-	T	---	T	D	C	G	G	G	L	R	W	Q	I	F	Q	W	N	---	S	G	Y	D	Y	K	N	T	V	S	T	G	C					
164 tr A0A0H5CB47 A0A0H5CB47_CYBJA	YNTMW	---	ERWD	-	M	---	E	H	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	T	A	C					
165 tr G2WRS7 G2WRS7_VERDV	FHTQA	---	SPDR	-	H	---	D	E	T	C	G	G	G	L	R	W	Q	I	P	F	A	N	---	N	G	Y	N	Y	K	N	S	I	A	N	G	C				
166 tr A0A124BVB5 A0A124BVB5_ASPNG	FNEQA	---	QRWN	-	T	---	Q	K	C	G	G	G	L	K	W	Q	I	Y	T	F	N	---	S	G	Y	T	Y	R	N	S	I	S	N	G	C					
167 tr A7EIG7 A7EIG7_SCLS1	FNTQA	---	PRWD	-	D	---	S	T	C	G	G	G	L	R	W	Q	I	F	T	F	N	---	N	G	Y	N	Y	K	N	S	I	S	N	G	C					
168 tr C5M5U7 C5M5U7_CANTT	FNTMQ	---	ARWD	-	E	---	E	D	C	N	G	G	L	R	W	Q	I	F	Q	W	N	---	S	G	Y	D	Y	K	N	S	V	S	N	G	C					
169 tr B9WAA8 B9WAA8_CANDC	FNTMN	---	ARWD	-	A	---	D	N	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	D	Y	K	N	S	I	S	N	G	C					
170 tr F8MRU1 F8MRU1_NEUT8	WATQA	---	APDR	-	H	---	D	D	T	C	N	G	G	L	R	W	Q	I	P	P	T	N	---	N	G	Y	D	Y	K	N	T	I	A	N	A	I				
171 tr A1DJS0 A1DJS0_NEOFI	YNTQV	---	DRWD	-	T	---	S	T	C	G	G	G	L	R	W	Q	I	Y	T	Y	Q	---	A	G	Y	T	M	K	N	A	I	S	N	G	G					
172 tr B6HLM8 B6HLM8_PENRW	FNDQKS	P	D	G	K	G	W	D	-	T	---	T	I	C	G	G	G	L	R	W	Q	K	E	I	W	Q	---	S	G	Y	T	M	K	N	A	V	S	N	G	G
173 tr A0A117DWS0 A0A117DWS0_ASPNG	FNTQV	---	PRWD	-	T	---	S	S	C	Q	G	G	L	R	W	Q	I	W	P	Y	Q	---	A	G	Y	T	T	K	N	A	I	S	N	G	G					
174 tr A0A124BXM1 A0A124BXM1_ASPNG	YNTQV	---	DRWD	-	D	---	S	T	C	G	G	G	L	R	W	Q	I	Y	P	Y	E	---	A	G	Y	A	M	K	N	S	I	S	N	G	G					
175 tr J3NZQ6 J3NZQ6_GAGT3	WVRQT	---	LPER	-	Q	---	D	S	E	C	G	G	G	L	R	W	Q	V	P	G	A	G	W	G	P	G	Q	T	Y	K	N	T	I	A	N	A	C			
176 tr W0THH8 W0THH8_KLUMD	FNEMY	---	SRWD	-	P	---	A	H	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	I	S	N	G	C					
177 tr B2AEF2 B2AEF2_PODAN	FHTQA	---	APER	-	H	---	D	N	T	C	N	G	G	L	R	W	Q	V	P	P	T	N	---	A	G	Y	N	Y	K	N	T	I	A	N	G	C				
178 tr Q96TX1 Q96TX1_NEUCS	FNTQA	---	NPDR	-	H	---	D	K	T	C	N	G	G	L	R	W	Q	I	P	P	L	N	---	N	G	Y	N	Y	K	N	S	I	A	N	G	C				
179 tr E9DYA6 E9DYA6_METAQ	FNTQA	---	SPQR	-	H	---	D	G	T	C	G	G	G	L	R	W	Q	I	P	F	T	N	---	N	G	Y	G	Y	K	N	S	I	A	N	G	C				
180 tr B9WAI2 B9WAI2_CANDC	FNTMQ	---	ARWD	-	T	---	E	E	C	N	G	G	L	R	W	Q	I	F	Q	W	N	---	S	G	Y	D	Y	K	N	S	V	S	N	G	A					
181 tr Q6C171 Q6C171_YARLI	FNTMA	---	ARWD	-	T	---	Q	S	C	Y	G	G	L	H	W	Q	I	F	Q	W	N	---	S	G	Y	D	Y	K	N	A	I	A	N	G	A					
182 tr A6ZMV1 A6ZMV1_YEAS7	FNTMY	---	SRWD	-	S	---	E	H	C	G	G	G	L	R	W	Q	I	F	T	W	N	---	S	G	Y	N	Y	K	N	T	V	S	N	A	C					
183 tr B6K7X7 B6K7X7_SCHJY	FNEQV	---	ARWD	-	T	---	T	L	C	D	G	G	L	R	W	Q	I	Y	P	F	N	---	N	G	Y	T	Y	K	N	S	I	T	N	G	L					
184 tr I1RSA2 I1RSA2_GIBZE	FNEYV	---	SRWD	-	D	---	E	Y	C	G	G	G	M	R	W	Q	I	F	K	W	N	---	T	G	Y	D	Y	K	N	A										

001	tr A0A254U2J9 A0A254U2J9_ASPNG	LFQLAARLARYTSND-T-YADWA	AEKIFDWC	CASTPLLNNE	-----TW
002	sp Q6FLP9 DCW1_CANGA	LFHIAARLARYTG	NQ-S-YVDWA	AERVYDWM	EDVHLID-----NG-T-YR
003	tr A1CCM5 A1CCM5_ASPCL	FFQLAARLARYTN	NH-T-YYEW	GEKAWDWS	CTTPLLNK-----TW
004	tr C1H190 C1H190_PARBA	FFNLAARLARYTG	NT-T-YANWA	EASWNWM	SGVGLISP-----TY
005	tr Q2UR85 Q2UR85_ASPOR	LFQLAARLGRYTK	NQ-T-YIDWA	AEKIWDWS	ATTPLLKTA-----DW
006	tr A5DV30 A5DV30_LODEL	LFHIAARLARYTG	NGTV-YVDWA	AEKVWQWM	DDVGFLTEED---N--G-DF
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	VFALGARLARYTG	DN-E-LVKSS	SRILRWM	KKAFFVHAPTDDSDNAGNY
008	tr J3PGN0 J3PGN0_GAGT3	FFNIGARLARYTG	NK-T-YAEHA	AKTFDWL	WAVNYIDN-----ENY
009	tr A0A124BXU6 A0A124BXU6_ASPNG	LFQLSARLARYTS	NQ-T-YYDWA	AERIWDWS	TTTPLLSNS-----TW
010	tr C5P4A1 C5P4A1_COCP7	FFNIAARLARYTG	ND-T-YAEWA	AVRTWDWT	KGVGLLTG-----DY
011	tr Q0CG55 Q0CG55_ASPTN	FFNLAARLAKYT	GNQ-T-YAEWA	DRVWNWT	TSVNLMTN-----DW
012	tr C4Y2X5 C4Y2X5_CLAL4	LFHMAARLARYT	AND-S-YVEWA	AERVFDWM	YGVNLLT-----EGEWW
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	FFNLAARLAKYT	NS-T-YADWA	ADTVWDWT	GEVGFM TD-----TY

014 tr|C7Z068|C7Z068_NECH7
015 tr|G8BXX2|G8BXX2_TETPH
016 tr|A0A0L0P6S8|A0A0L0P6S8_CANAR
017 tr|A1CSC2|A1CSC2_ASPCL
018 sp|Q05031|DFG5_YEAST
019 tr|H2AQJ6|H2AQJ6_KAZAF
020 tr|G2XFH9|G2XFH9_VERDV
021 tr|A0A1S7HQM3|A0A1S7HQM3_9SACH
022 sp|Q5AD78|DCW1_CANAL
023 tr|B6H7E8|B6H7E8_PENRW
024 tr|B6GZU2|B6GZU2_PENRW
025 tr|E9E4W7|E9E4W7_METAQ
026 tr|A0A1S9RH77|A0A1S9RH77_9EURO
027 tr|G2R7G8|G2R7G8_THITE
028 tr|G3JBY1|G3JBY1_CORMM
029 tr|A5DNV5|A5DNV5_PICGU
030 tr|A0A1B8GPA6|A0A1B8GPA6_9PEZI
031 tr|Q2TWC1|Q2TWC1_ASPOR
032 tr|J5RXY9|J5RXY9_SACK1
033 tr|A0A2N6NN14|A0A2N6NN14_BEABA
034 tr|A0A254U5V1|A0A254U5V1_ASPNG
035 tr|A0A061B6H7|A0A061B6H7_CYBFA
036 tr|G8ZQ93|G8ZQ93_TORDC
037 tr|Q9C2J1|Q9C2J1_NEUCS
038 tr|C5NZK5|C5NZK5_COCP7
039 tr|Q6C0T7|Q6C0T7_YARLI
040 tr|A0A0W0ENZ4|A0A0W0ENZ4_CANGB
041 tr|F7VZ72|F7VZ72_SORMK
042 tr|A3LN37|A3LN37_PICST
043 tr|Q6CAI2|Q6CAI2_YARLI
044 tr|A0A0V1PWA0|A0A0V1PWA0_9ASCO
045 tr|G9MJS5|G9MJS5_HYPVG
046 tr|G9MHI4|G9MHI4_HYPVG
047 tr|D4AP27|D4AP27_ARTBC
048 tr|B6GZT8|B6GZT8_PENRW
049 sp|O74556|YCZ2_SCHPO
050 tr|A0A0C4E991|A0A0C4E991_MAGP6
051 tr|B8MHG0|B8MHG0_TALSN
052 tr|A3LMV8|A3LMV8_PICST
053 tr|A0A099P0Y5|A0A099P0Y5_PICKU
054 sp|Q9P6I3|YHG7_SCHPO
055 tr|Q2US57|Q2US57_ASPOR
056 tr|W7N622|W7N622_GIBM7
057 sp|Q9P6I4|YHG6_SCHPO
058 tr|C7GRB3|C7GRB3_YEAS2
059 tr|C7GP28|C7GP28_YEAS2
060 tr|G3JGZ5|G3JGZ5_CORMM
061 tr|C4QXV4|C4QXV4_KOMPG
062 tr|I1S0Z5|I1S0Z5_GIBZE
063 tr|A0A0J5PSQ0|A0A0J5PSQ0_ASPFM
064 tr|G0W5X4|G0W5X4_NAUDC
065 tr|G8ZPC1|G8ZPC1_TORDC
066 tr|A0A167CDC5|A0A167CDC5_9ASCO
067 tr|R9XAT7|R9XAT7_ASHAC
068 tr|B2B747|B2B747_PODAN
069 tr|G0SE99|G0SE99_CHATD
070 tr|J8Q6F0|J8Q6F0_SACAR
071 tr|A0A0A8LDJ2|A0A0A8LDJ2_9SACH
072 tr|J3K6E2|J3K6E2_COCIM
073 tr|G4N3E1|G4N3E1_MAGO7
074 tr|C5M5J2|C5M5J2_CANTT
075 tr|A0A254U0X0|A0A254U0X0_ASPNG
076 sp|Q75DG6|DCW1_ASHGO
077 tr|Q6BZF0|Q6BZF0_DEBHA
078 tr|E9E3Q1|E9E3Q1_METAQ
079 sp|P36091|DCW1_YEAST
080 tr|W0TAH6|W0TAH6_KLUMD
081 tr|A0A0H5CF34|A0A0H5CF34_CYBJA
082 tr|A0A2C5WNS2|A0A2C5WNS2_9PEZI
083 tr|Q2TYU3|Q2TYU3_ASPOR

[illegible]

084	tr G2XGF6 G2XGF6_VERDV	FFNIAARLARYTGNQ	-T-	YAEWA	AERVWD	WEVKAGLITD	-----	-EF	
085	tr G3AMT4 G3AMT4_SPAPN	LFHIAARLARYTSDND	-T-	YVEWA	AERVWD	WMYGVGLLT	-----	EGDWW	
086	tr A0A1C1D213 A0A1C1D213_9EURO	FFNMGARLAVYTGNQ	-T-	YADWA	DRMWD	WVTAIGLRDE	-----	-QY	
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	FFNLGARLARYTGNE	-T-	YAHHA	ARKTWD	WMAVGLMTE	-----	-DG	
088	tr A0A100I5A6 A0A100I5A6_ASPNG	LFELSARLARFTKND	-T-	YAEWA	AEKIFD	WSETTPLLNTN	-----	-ATI	
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	LFQLAARLGRYTGND	-T-	YLDVA	AEKVFD	WMVDVGIVLS	-----	-DT--A	
090	tr G2Q8A7 G2Q8A7_MYCTT	FFNLGARLARYTGNK	-T-	YAEWA	AECTWD	WVRGVGYMDD	-----	-QY	
091	tr B8NPZ1 B8NPZ1_ASPFN	LFELAARLARFTKNE	-T-	YSEWA	ADRIWD	WSASTPLLQTD	-----	-RW	
092	tr Q7SAB2 Q7SAB2_NEUCR	FFNLGARLARYTGNE	-T-	YAHHA	ARKTWD	WMAVGLMTE	-----	-DG	
093	tr A2R8R5 A2R8R5_ASPNC	LFQLSARLARYTSDNQ	-T-	YYDWA	AERIWD	WSVTTPLLSNS	-----	-TW	
094	tr C5DHG0 C5DHG0_LACTC	MFQLAARLGRYTGND	-T-	YLTKA	AETVWD	WLVDVGFFVLN	-----	-ENGNA	
095	tr G0SFA3 G0SFA3_CHATD	FFNIAARLARYTGNQ	-T-	YAEWA	AARIWA	WEEAIGLITS	-----	-DY	
096	tr K1WJG5 K1WJG5_MARBU	LFNIAARLARYTGNQ	-T-	YADWA	AECTWD	WMETMGLISD	-----	-DG	
097	tr H2B005 H2B005_KAZAF	LFHLAARLARYTGNT	-S-	YVDWA	AEKVYN	WMEDTYLIS	-----	-NG-T-QM	
098	tr C5DYD7 C5DYD7_ZYGRC	YFQLAARLGRYTNNQ	-T-	YLEVA	AEKVFD	WLTDVGFFVL	-----	-K-DKG	
099	tr G8YM23 G8YM23_PICSO	LFNIAARLARYTGND	-S-	YSDWA	AEKIWE	WFDSVGLLA	-----	-VSGDS-Y	
100	tr W3X8E3 W3X8E3_PESFW	FFNLGARLARYTNNQ	-T-	YANWA	AETTWN	WVRSVGFMD	-----	-EY	
101	tr A0A254U3K7 A0A254U3K7_ASPNG	LFQLSARLARYTSDNQ	-T-	YYDWA	AERIWD	WSVTTPLLSNS	-----	-TW	
102	tr Q6CER8 Q6CER8_YARLI	LFNLASRLRYTGNT	-T-	YLEWA	AEKIWT	WSEKVKIID	-----	-G-G	
103	tr G2QT05 G2QT05_THITE	FFNLGARLARYTGNS	-T-	YAEWA	AECTWD	WVRAVGFMD	-----	-KY	
104	sp Q5ACZ2 DFG5_CANAL	LFHLAARLARYTGNS	SV-	YVDTA	AEKVWK	WMEDVGFLTEED	---N--G-	DV	
105	tr G2WKU6 G2WKU6_YEASK	LFQIAARLGRYTGNT	-T-	YLEVA	AEQVFD	WLVDVGIVVL	-----	-N-DTA	
106	tr A0A0C7N752 A0A0C7N752_9SACH	LFQLAARLGRYTGNE	-T-	YIAVA	AEKIFD	WLVDVGIVVLQ	-----	-EN--A	
107	tr W3WN68 W3WN68_PESFW	FFNLASRLARYTGND	-T-	YGDWA	AERIFAW	EQGVDFINS	-----	-DW	
108	tr G2QFS1 G2QFS1_MYCTT	FFNLGARLARYTENQ	-T-	YADWA	ASKTFQ	WLWDVGIDH	-----	-ESW	
109	tr G3J9G4 G3J9G4_CORMM	FFNIAARLARFTGNS	-T-	YADWA	AQKIYD	WEVKEGLITS	-----	-DY	
110	tr Q2UJ03 Q2UJ03_ASPOR	FFNLAARLARYTGNQ	-T-	YADWA	AVRVWD	WTESVGFITE	-----	-DY	
111	tr G8YRT2 G8YRT2_PICSO	LFQIAARLARYTGND	-T-	YAKTA	ATKVWD	WMHSVGFIGEDD	---N--H-	ENM	
112	tr W3X554 W3X554_PESFW	FFNVAASRLARFTGNQ	-T-	YADWA	AEKVYD	WMTDTGLITA	-----	-EF	
113	tr Q1K7A8 Q1K7A8_NEUCR	FFNLGARLARFTGNS	-S-	YGEWA	ASRTWD	WERSINLITD	-----	-EY	
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	LFQLAARLARYTNNQ	-T-	YSQWA	AERIWD	WSATTPLLKES	-----	-DW	
115	tr A7F3P0 A7F3P0_SCLS1	FFNLGARLARYTGNS	-S-	YAEWA	AERIFT	WEQSVGLIGD	-----	-GY	
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	FFNVAARLARFTRND	-T-	YAEWA	ATKIYE	WQEGIGLIGK	-----	-NY	
117	tr Q6CIP9 Q6CIP9_KLULA	LFQLAARLGRYTGNT	-T-	YLEVA	AEKVFD	WLVDIDFVVM	-----	-K-EEA	
118	tr H2AMQ5 H2AMQ5_KAZAF	LFHLAARLARYTGND	-S-	YVDWA	AEKVYN	WMEDTGFIT	-----	-TG-N-TL	
119	tr G8JMI3 G8JMI3_ERECY	LFQLAARLGRYTGNS	-T-	YLDIA	ANETFS	WLVDVKFVVL	-----	-K-DEA	
120	tr C5DGT1 C5DGT1_LACTC	LFHLAARLARYTGNN	-T-	YVDWA	AEKVFD	WMKGIGLLT	-----	-EN--DNS	
121	tr K1XJR4 K1XJR4_MARBU	FFNIAARLARYTDNS	-T-	YYDHA	AVQTWD	WITSVGFIDK	-----	-DY	
122	tr A0A124BYC5 A0A124BYC5_ASPNG	LFQLAARLARYTNNQ	-T-	YVDWA	ANTIWD	WSTV-IILVNQE	-----	-SW	
123	tr A7TLL2 A7TLL2_VANPO	LFHIGARLARFTGND	-S-	YVEWA	AEKVYD	WMESVNIE	-----	-DD-GKGA	
124	tr Q0CB86 Q0CB86_ASPTN	LFQLAARLARYTNNQ	-T-	YADWA	ADKIWD	WSASTPLLNNQ	-----	-TW	
125	tr K1XLH0 K1XLH0_MARBU	LFNIAARLALYTGNQ	-T-	YADWA	AVKTWD	WMSAVGLLNE	-----	-KY	
126	tr J7S438 J7S438_KAZNA	LFHMASRLARYTGNO	-S-	YVDWA	AEKVYD	WMWDVHVIS	-----	-NS--TYM	
127	tr Q4WG09 Q4WG09_ASPFU	LFELSARLARFTKNE	-T-	YAEWA	ANKIWD	WSASTPLLQTD	-----	-RW	
128	tr H8WZ09 H8WZ09_CANO9	LFHIAARLARYTGNE	SA-	YLPTA	AERVWN	WMEEVNFLT	EED---N--G-	NL	
129	tr I1RL09 I1RL09_GIBZE	FFNLGARLARYTGNE	-T-	YAKHA	EETWE	WLWGVNYIDH	-----	-DRW	
130	tr Q4W985 Q4W985_ASPFU	FFHLAARLARYTGNR	-T-	YAEWA	AERVWD	WTVDVGFITD	-----	-DW	
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	LFQIAARLGRYTGNT	-T-	YLDVA	AEQVFD	WLVDVGIVVL	-----	-N-DTA	
132	tr W6QCW3 W6QCW3_PENRF	FFNLASRLAKYTNNQ	-T-	YHDWA	AEKAWN	WTAEVGFMT	-----	-EY	
133	tr W3WMD3 W3WMD3_PESFW	YFNIGARLARYTNNQ	-T-	YMELA	AGRTFD	IMEKLGIVDA	-----	-DW	
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	LFHIAARLAHFTGNN	-T-	YAEWA	AEKVYN	WMSDVKLIN	-----	-SG--DTK	
135	tr I2GYH1 I2GYH1_TETBL	LFQMAARLGRYTGNN	-T-	YLDVA	ANDVFD	WLTGVGYVNM	-----	-D-NAA	
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	LFQLAARLGRYTNNQ	-T-	YLHIA	EEVF	DLHDVGIVVL	-----	-S-DKG	
137	tr Q752P3 Q752P3_ASHGO	LFQLAARLGRYTGNO	-T-	YLDVA	ASKVFQ	WLVDVKYIVL	-----	-E-DKA	
138	tr B8MYP3 B8MYP3_ASPFN	LFQLAARLGRYTKNQ	-T-	YIDWA	AEKIWD	WSATTPLLKTA	-----	-DW	
139	tr Q2TXL6 Q2TXL6_ASPOR	FFQLAARLARYTKNE	-T-	YTEWA	AEKAFT	WATSVP	LIIEK--	-----	-GW
140	tr Q0CN17 Q0CN17_ASPTN	LFQLAARLGRYTNNH	-T-	YTDWA	AEKIWD	WSATTPLLRTT	-----	-DW	
141	tr J3NHD5 J3NHD5_GAGT3	FFNLGARLARYTTNK	-T-	YADWA	AEKQWD	WVTSVGLMDA	-----	-QY	
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	FFNIGSRLARYTGND	-T-	YANWA	AECTWD	WTQKIGLIDA	-----	-QY	
143	tr A1CVB0 A1CVB0_NEOFI	FFHLAARLARYTGNS	-T-	YAEWA	AERVWD	WTVDVGFITD	-----	-DW	
144	tr B8NVI3 B8NVI3_ASPFN	FFQLAARLARYTKNE	-T-	YTEWA	AEKAFT	WATSVP	LIIEK--	-----	-GW
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	LFHIAARLARYTGNT	-S-	YVDWA	AEKVFE	WMKGIHLE	-----	-E--DGTN	
146	tr A1DJ54 A1DJ54_NEOFI	LFELSARLARFTKND	-T-	YAQWA	ADKIWD	WSASTPLLQMD	-----	-RW	
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	FFNMGARLARYTRND	-T-	YATWA	ATKQFQ	WIYDVNYIDH	-----	-DSW	
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	LFHIGARLARYTGNT	-S-	YVDWC	DKVHD	WMYDTGLIH	-----	-ES--DWY	
149	tr A0A177A618 A0A177A618_9PEZI	MFNIASRLAVYTGNS	-T-	YAEWA	AECTWD	WVESIGLIND	-----	-KG	
150	tr Q75CW6 Q75CW6_ASHGO	LFQIAARLARFTGNS	-T-	YQDVA	AETVFD	WLVD SKLVQL	-----	-E-NDA	
151	tr Q7S4K4 Q7S4K4_NEUCR	FFNMGARLARYTRND	-T-	YATWA	ATKQFQ	WIYDVNYIDH	-----	-DSW	
152	tr G2QB99 G2QB99_MYCTT	FFNIAARLARYTGNE	-T-	YAEWA	AEKIFAW	EESVGLIDA	-----	-NL	

153	tr G0S3F2 G0S3F2_CHATD	FFDLGARLARLYTKNN-T-YAEWA	EKIFDWLYAVGYIDH	-----ETW
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	LFHIGARLARLYTGND-S-YADWSE	EKVWDWMIDSDLII	-----PGPYW
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	LFQIAARLGRYTGNT-T-YLDVA	ERVFDWLVGVGYIVL	-----S-EKG
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	LFQLAARLARLYTNNH-T-YYEWA	EKVWDWSCSSPLLNNK	-----TW
157	tr G2QKJ0 G2QKJ0_MYCTT	FMNIAARLARLYTNN-D-T-YALWA	TKAWDWIEAMGYVTD	-----KY
158	tr Q4WFX5 Q4WFX5_ASPFU	LFNIAARLARLYTGDA-T-YAKWA	NKVWDWVTETGLIGR	-----EY
159	tr Q5BGD7 Q5BGD7_EMENI	LFQLAARLGRTNNQ-T-YFDWA	EKIWDWAAASPLIDTN	-----TW
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	LFHLAARLARLYTGND-T-YTEWA	EKVYDWMQDIDLLI	-----DEEPNQY
161	tr G8BYN9 G8BYN9_TETPH	LFQIAARLGRYTGNN-T-YLEVA	EKVFDWMVDVEYIVL	-----K-DVA
162	tr F7VVT4 F7VVT4_SORMK	FFNLGARLARLYTDNS-T-YSKWA	EKTYNWIRDVGYIDK	-----NW
163	tr G8YB80 G8YB80_PICSO	LFNIAARLARLYTGND-S-YSDWA	EKVWEWFDSAGLLA	-----DSGDS-Y
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	LFNLAGRLARFTRND-T-YVETA	AQTAYDWLVDIGFIV	-----DSN-GMA
165	tr G2WRS7 G2WRS7_VERDV	FFNIGARLARLYTKNS-T-YADWA	DKTWDWMTNVGFIDS	-----EY
166	tr A0A124BVB5 A0A124BVB5_ASPNG	FFNMAARLARLYTGNA-T-YADWA	NKVWDWVTDVGLIGP	-----EY
167	tr A7EIG7 A7EIG7_SCLS1	FFNLGARLAKYTGND-T-YAQWA	AERTWDMENVGLMDE	-----NY
168	tr C5M5U7 C5M5U7_CANTT	LFHLAARLARLYTGND-S-YVVWA	AERVWDWMYGVGLLT	-----EGNWW
169	tr B9WAA8 B9WAA8_CANDC	LFHLAARLARLYTGNS-SV-YVDTA	EKVWKMEDVGFLTTEEQ	---N-G-DV
170	tr F8MRU1 F8MRU1_NEUT8	FFNMGARLARLYTRNE-T-YATWA	TKQFQWIYDVNYIDH	-----DSW
171	tr A1DJS0 A1DJS0_NEOFI	LFQLAARLARLYTNNH-T-YYEWA	EKVWDWSCSSPLLNNK	-----TW
172	tr B6HLM8 B6HLM8_PENRW	FFMLAARLAWYTQDE-E-YATWA	EKVWDWSTSVNLVNNE	-----TW
173	tr A0A117DWS0 A0A117DWS0_ASPNG	LFQLAARLGRYTNN-D-T-YTDWA	EKIWDWSATTPLLQTE	-----DW
174	tr A0A124BXM1 A0A124BXM1_ASPNG	LFQLAARLARLYTSND-T-YADWA	EKIFDWCASTPLLNNQ	-----TW
175	tr J3NZQ6 J3NZQ6_GAGT3	YFNIGARLTRYTGNS-T-YAEHA	EKIWDWMWALGYIDH	-----ESW
176	tr W0THH8 W0THH8_KLUMD	LFQLAARLGRYTGNT-Q-T-YLDVA	EKVFNWLTDVNYVVL	-----K-DEA
177	tr B2AEF2 B2AEF2_PODAN	FFDLGARLAAYTFNQ-T-YADWA	DKTFQWLWDVGYIDN	-----KDW
178	tr Q96TX1 Q96TX1_NEUCS	FFNLGARLARLYTGNK-T-YADWA	EETTWDWMRGVGLMDD	-----NY
179	tr E9DYA6 E9DYA6_METAQ	FFNMAARLARLYTGDA-K-YSDWA	EKTWDWVEKIGFIDP	-----QNY
180	tr B9WAI2 B9WAI2_CANDC	LFHLAARLARLYTGND-S-YVVWA	AERVWDWMYGVGLLT	-----EQNWW
181	tr Q6C171 Q6C171_YARLI	LFNLGARLYRYTGNV-T-YLEWA	AERIWDWSTQIEIVD	-----N-G
182	tr A6ZMV1 A6ZMV1_YEAS7	LFQIAARLGRYTGNT-T-YLEVA	EQVFDWLVDVGYYVVL	-----N-DTA
183	tr B6K7X7 B6K7X7_SCHJY	LFQLAARIARFTAND-T-CAEFA	EKVWDWTTTVGFLNE	-----STF
184	tr I1RSA2 I1RSA2_GIBZE	FFNVASRLARLYTGND-T-YADWA	GKVFDWHIKAGIITD	-----KF
185	tr F7VWZ9 F7VWZ9_SORMK	FFNLGARLSRFTGNS-S-YGDWA	ATRTWDWERSINLITD	-----EY
186	tr A0A1B2J789 A0A1B2J789_PICPA	LFQLSARLARLYTAND-T-YITLA	EEAFDWMYGAGFLT	-----EG--DWW
187	tr G0V8N4 G0V8N4_NAUCC	LFHMAARLARLYTGNT-Q-S-YVDWA	AERVYDWMSDVGLIS	-----NG-S-YK
188	tr H8WXN8 H8WXN8_CANO9	LFHLAARLARLYTQND-S-YVEWA	AQKVWDWMYGVGLLT	-----EGDHW
189	tr A6ZZS0 A6ZZS0_YEAS7	LFHIAARLARLYTGNT-Q-T-YVDWA	EKVYEWVMVGVNLI	-----NG-T-YK
190	tr G9MQD1 G9MQD1_HYPVG	FFNIGARLARLYTRNE-T-YAKRA	EEDTWNWLWVGYYIDH	-----KSW
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	LFHIAARLARLYTGNT-Q-S-YVDWA	EKVFDWMKGINLLT	-----ENTGGNN
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	LFELSARLARFTKNE-T-YAEWA	NKIWDWSASTPLLQTD	-----RW
193	tr A0A254UET1 A0A254UET1_ASPNG	FFNIAARLARLYTGNA-T-YADWA	NKVWDWVTDVGLIGP	-----EY
194	tr A1D587 A1D587_NEOFI	LFQLAARLARLYTNN-E-T-YSQWA	AERIWDWSATTPLLKES	-----DW
195	tr Q1K7I4 Q1K7I4_NEUCR	FFNLGARLARLYTGNT-K-T-YADWA	EETTWDWMRGVGLMDD	-----NY
196	tr G8JS88 G8JS88_ERECY	LFHLAARLARLYTGNT-Q-S-YADWA	EKVYDWMASVQLIN	-----QT-GDWT
197	tr A0A136J7D3 A0A136J7D3_9PEZI	FFNLASRLARLYTGNS-T-YADWA	GRVWTWLEGHELIDK	-----DF
198	tr E9E4X8 E9E4X8_METAQ	FFNMGARLARLYTGNT-T-YSDWA	DKTWDWMWNIGFIDN	-----KNY
199	tr Q6CP42 Q6CP42_KLULA	LFHLSARLARLYTGNS-S-YTEWA	EKIYDWMQDIQLLV	-----DEEPTQY
200	tr G0WHL4 G0WHL4_NAUDC	LFQLAARLGRYTGNT-T-YLEVA	EKVFTWLVDVGYYVLL	-----D-DTA
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	LFHIAARLARLYTGNT-Q-T-YVDWA	EKVYEWVMVGVDLI	-----NG-T-YK
202	tr F9WYX6 F9WYX6_ZYMTI	FASLAARLHRFTGNS-T-FYGEWA	ATRAVEWTRSIGLMNQ	-----GQT-
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	FFNLGARLARLYTLNT-T-YADWA	EKQWDWITSVGLMDS	-----QY
204	tr J3P147 J3P147_GAGT3	YFNLGARLARLYTGNT-Q-T-YLKEA	AERTWDLWAVKYIDH	-----DTY
205	tr I2H842 I2H842_TETBL	LFHIAARLARLYTGNT-Q-S-YVDWSE	EKVYDWMMLGTGLIS	-----NG-TTYK
206	tr S6E3R3 S6E3R3_ZYGB2	LFQLAARLGRYTNNQ-T-YLHIA	EEVFDFWLHDVGYYVVL	-----S-DKG
207	tr E5AD94 E5AD94_LEPMJ	LFHLGARLAMYTKN-D-T-YAQWA	EKAFDWLSQSPILPG	-----DG
208	tr Q6FJM7 Q6FJM7_CANGA	LFQIAARLGRYTGNT-T-YLDVA	ERVFDWLVGVGYIVL	-----S-EKG
209	tr A5DUW0 A5DUW0_LODEL	LFHLAARLARFTGND-S-YVEWA	AERVWNWMYGAGLLT	-----EGNEW
210	tr G8BF46 G8BF46_CANPC	LFHLAARLARLYTQND-S-YVEWA	AQKTWDWMYGVGLLT	-----EGDHW
211	tr W3XAF6 W3XAF6_PESFW	FFNIASRLARLYTGND-T-YREWAD	KIFDWELNVGFISD	-----TW
212	tr B6H7I2 B6H7I2_PENRW	LFQLAARLARLYTDDD-K-YSKWA	EKIWDWSTSSPLVNNK	-----TW
213	tr Q5ATF9 Q5ATF9_EMENI	LFQLAARLARLYTNN-D-T-YAEKA	QMVWDWVVSSPLVNNK	-----TW
214	tr A0A167CDD3 A0A167CDD3_9ASCO	LFHIGARLARLYTGNN-S-YVDWA	AERVWNWAEGVNFLN	-----QTFVNQT
215	tr G0RXS3 G0RXS3_CHATD	FMNIASRLARLYTRND-T-YARWA	ETAWDWTQGVGYITK	-----EY
216	tr B8NX26 B8NX26_ASPFN	LFELAARLARFTKNE-T-YTEWA	EKIWDWSAKSGLMDVN	-----KW
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	FINLAARLALYTKNE-T-YVELA	EKHWDWMTAIGLISP	-----TS
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	LFNIAARLARLYTGDA-T-YAEWA	NKVWDWVTETGLIGP	-----EY
219	tr A1CMM3 A1CMM3_ASPCL	FFNLASRLAVYTGNA-T-YAEWA	EKTWDWVVRIGLTNE	-----AY
220	tr B6HU84 B6HU84_PENRW	LLQLSARLALYTGNT-K-T-YADWA	AERIWDWSATTPLLKQK	-----NW
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	FFNLGARLARLYTGNT-K-T-YADWA	EETTWDWMRGVGLMDD	-----NY

222	tr G2WHY6 G2WHY6_YEASK	L F H I A A R L A R Y T G N Q - T - Y V D W A E K V Y E W M V G V N L I S - - - - - N G - T - Y K
223	tr G0W7G8 G0W7G8_NAUDC	L F H M A A R L A R Y T G N Q - S - Y V D W A E K V Y D W M Y E V G L V S - - - - - N G - S - S M
224	tr G4NGV8 G4NGV8_MAGO7	F F N L A A R L A R F T G N Q - T - Y A E W A N K I W D W E R S T N L M T E - - - - - - - - - K F
225	tr W3X855 W3X855_PESFW	F F N I A A R L A R Y T G N T - T - Y A D W A E T V Y D W M E T V G F I D D - - - - - - - - - E Y
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	F F N M G A R L G R Y T G N Q - T - Y I D W A E K T F D W V T G V G Y I D P - - - - - - - - - Q T H
227	tr J8Q2K7 J8Q2K7_SACAR	L F H I A A R L A R Y T G N S - T - Y V D W A E K V Y E W M V S V N L I S - - - - - N G - T - Y K
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	L F N I A A R L A R Y T S N D - S - Y V E W A E K V W D W M Y G I G L L S - - - - - - - E G D W W
229	tr J6EGN8 J6EGN8_SACK1	L F Q I A A R L G R Y T G N S - T - Y L D V A E Q V F D W L V D V G Y V V L - - - - - N - D T A
230	tr A0A100ISD9 A0A100ISD9_ASPNG	F F N L A A R L A K Y T G N S - T - Y A D W A D T V W D W T G E V G F M T D - - - - - - - - - T Y
231	tr G8YQC0 G8YQC0_PICSO	L F Q I A A R L A R Y T G N D - T - Y A K T A E K V W D W M S D V G F I G E D D - - - N - - H E N M
232	tr F7VVP8 F7VVP8_SORMK	F F N L G A R L A R Y T R N E - T - Y A T W A T K Q F E W L Y N V N Y I D H - - - - - - - - - E S W
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	L F Q I A A R L G R Y T G N T - T - Y L E V A E Q V F D W L V D V G Y V V L - - - - - N - D T A
234	tr F8MBP4 F8MBP4_NEUT8	F F N L G A R L A R F T G N S - S - Y G E W A S R T W D W E R S I N L I T D - - - - - - - - - E Y
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	L F Q L A A R L A R Y T N N Q - T - Y V D W A N T I W D W S T S - H L V Q E G - - - - - - - - - S W
236	tr A1CD27 A1CD27_ASPL	L F E L S A R L A R F T K N G - T - Y A D W A E K I W D W S V T T P L L Q T D - - - - - - - - - R W
237	tr A5DHF3 A5DHF3_PICGU	L F H I S A R L A R Y L K N D - T - Y A K T A E K V W D W M E E L F F - - H E D - - - D - - G - E L
238	tr D4ATT7 D4ATT7_ARTBC	F F H L A A R L A R Y T N D E - S - Y S K W A D K A R W M T A I K L I S P - - - - - - - - - T F
239	tr E2PT42 E2PT42_ASPNC	F F N L A A R L A K Y T G N S - T - Y A D W A D T V W D W T G E V G F M T D - - - - - - - - - T Y
240	tr C7ZPE5 C7ZPE5_NECH7	F F N L G A R L A R Y T G N T - T - Y A D W A E T T W E W M E S V G F L D P - - - - - - - - - T S Y
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	F F N M A A R L A T Y T G N A - T - Y A E W A E I A W D W M E S V G F V T E - - - - - - - - - N E
242	tr Q4WKP7 Q4WKP7_ASPFU	L F Q L A A R L A R Y T N N E - T - Y S Q W A E R I W D W S A T T P L L K E S - - - - - - - - - D W
243	tr F8MXT4 F8MXT4_NEUT8	F F N L G A R L A R Y T G N K - T - Y A D W A E T T W D W M R G V G L M D D - - - - - - - - - N Y
244	tr A0A177AJ74 A0A177AJ74_9PEZI	F F N L G S R L A R Y T G N D - T - Y S N W A E K T W E W T Q S I G L I D A - - - - - - - - - Q F
245	tr J7RQR5 J7RQR5_KAZNA	L F N L A A R L G R Y T G N S - T - Y L D V A D Q V F D W M V S V G Y V V L - - - - - G - D T A
246	tr Q75CW7 Q75CW7_ASHGO	L F Q I A A R L A R F T G N S - T - Y Q D V A E T V F D W L V D V K L V E L - - - - - E - N D A
247	tr G3AKK4 G3AKK4_SPAPN	L F H L A A R L A R Y T G N S T V - Y L D V A E K V W D W M D E V G F L T E E D - - - N - G - N F
248	tr B2W762 B2W762_PYRTR	F F N I A A R L Y K Y L G N E - T - Y A D W A E K A W K W E L S V G L M S A - - - - - - - - - D Y
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	L F H I A A R L A R Y T G N Q - T - Y V D W A E K V Y E W M V G V N L I S - - - - - N G - T - Y K
250	tr C4XWK1 C4XWK1_CLAL4	L F H L A A R L Y R F T G E K - V - Y L E T A E K V W N W M W D V G F I R - - D - - - S - - P - E F

039	tr Q6C0T7 Q6C0T7_YARLI	EVYDGIETGSGN---	VP-NCTN-RTPYIWTYNSGIF---	MHGAAALY
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	YVYDGVVS----I---	ND-NCTT-VTKYQWTYNQGGLM-----	LSGSAYLF
041	tr F7VZ72 F7VZ72_SORMK	NIHDGAH----V---	EK-NCTD-VNGAQFSYNFGVF-----	TLGAAHMY
042	tr A3LN37 A3LN37_PICST	FVYDGLT----I---	DN-NCSN-ITKYQWTYNQGGLM-----	LSGCAYLY
043	tr Q6CAI2 Q6CAI2_YARLI	YHIRDGV-----D---	VN-NCTN-ITPYVWTYNAGLF-----	IGGS AFLY
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	YVYDGVVT----V---	GG-NCSD-ITKLQWTYNQGGLM-----	MSGCAFLY
045	tr G9MJS5 G9MJS5_HYPVG	AIYDGDAD----V---	SN-NCTQ-INKAEFSYNSAVW-----	VLTGTAYMY
046	tr G9MHI4 G9MHI4_HYPVG	QIYDGLS----INTD--	NTCGHMDQNQWSYNAGIY-----	MHGAAAMY
047	tr D4AP27 D4AP27_ARTBC	QFFDGLSS----D---	TL-NCTE-LNRLQWTYNAGVY-----	LLGAAAMY
048	tr B6GZT8 B6GZT8_PENRW	NVADSAVV-----	GD-NCAT-QGNAQWTYNYGTY-----	LMGAAYMY
049	sp O74556 YCZ2_SCHPO	TVFDGLSS----I---	KD-NCSS-IEITQWTYNIGLY-----	MAGAAYMY
050	tr A0A0C4E991 A0A0C4E991_MAGP6	KVYDGGH----V---	NN-NCTD-IHKAQFSYNI AVL-----	LQGC AFMY
051	tr B8MHG0 B8MHG0_TALSN	KIYDMTRI-----	SE-NC KD-QDVMQWSYNYGSY-----	VTGAAYMY
052	tr A3LMV8 A3LMV8_PICST	TLYDGAN----I---	DT-NCTD-LTKKQWTYTYGIF-----	MAGAAYMY
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	NVNDGAE----I---	VHGACPV-VNGALWSYNYALM-----	LMGSAYLY
054	sp Q9P6I3 YHG7_SCHPO	TVYDGLASV-----	TS-NCSS-ITNEQWSYNVGVY-----	LAGT AFLY
055	tr Q2US57 Q2US57_ASPOR	NVADSTDV-----	DD-NCTS-QGNTQWTYNYGAY-----	IGGAAYMY
056	tr W7N622 W7N622_GIBM7	QVRDGVH----FEGK	CP---SSMDTNQWTYNSGVY-----	LYGAAAMY
057	sp Q9P6I4 YHG6_SCHPO	AVYDGDADT-----	ST-NCTT-LDPSQWSYNIGIF-----	MVGAAYMY
058	tr C7GRB3 C7GRB3_YEAS2	NVFDGAE----I---	DT-NCTD-ITKIEWTYNHGIV-----	LGGLAYMY
059	tr C7GP28 C7GP28_YEAS2	YVYDGVVS----I---	DD-NCTK-VTSYQWTYNQGLL-----	LAGSAYLY
060	tr G3JGZ5 G3JGZ5_CORMM	LVYDGGGR----V---	PF-NCTD-INKATFSYNAA IL-----	TQGA AFMY
061	tr C4QXV4 C4QXV4_KOMPG	FVYDGAFF----V---	DD-NCTE-IVMLQWTYNAGLM-----	VSGCAYLA
062	tr I1S0Z5 I1S0Z5_GIBZE	KIYDGGH----V---	GK-NCTD-INKAQFSYNSGVF-----	LQGA AFMY
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	LFYDGDAD----V---	LL-NCSD-LNRIEWTYNSGVY-----	LLGAANMY
064	tr G0W5X4 G0W5X4_NAUDC	YVYDGTST----I---	ET-NCTT-ITSYQWSYNQGGLM-----	LAGAAYLY
065	tr G8ZPC1 G8ZPC1_TORDC	NVFDGAE----I---	DN-NCTS-INKLEWSYNHGVV-----	LGGLAFMY
066	tr A0A167CDC5 A0A167CDC5_9ASCO	QLIDGAD----I---	ST-NCTT-HTPYEWTYNYGLF-----	TAGAAYLY
067	tr R9XAT7 R9XAT7_ASHAC	FVFDGTA----I---	DH-NCTV-IDRLQWSYNHGLI-----	MAGCAFIY
068	tr B2B747 B2B747_PODAN	NIYDGGH----V---	EH-NCTD-INRAQFSYNNGVF-----	LLGAAYMY
069	tr G0SE99 G0SE99_CHATD	NIYDGGH----V---	EQ-NCTD-INRAQFSYNNAIF-----	LLGAAYMY
070	tr J8Q6F0 J8Q6F0_SACAR	NVFDGAE----I---	DT-NCTD-ITKIEWTYNHGIV-----	LGGIAYMY
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	NVYDGAT----I---	ED-NCTD-IVKYEWSYNHGVV-----	LGGCAYMY
072	tr J3K6E2 J3K6E2_COCIM	QFFDGLSD----E---	KL-NCSE-VNRIQWTYNAGVY-----	LLGAASMY
073	tr G4N3E1 G4N3E1_MAGO7	KVYDGGH----V---	EH-NCTD-INKAQFSYSI AVL-----	TQATAFLW
074	tr C5M5J2 C5M5J2_CANTT	RMYDGAKE----I---	EN-NCSS-VTDLRWSYTYGVF-----	MAGCAYLY
075	tr A0A254U0X0 A0A254U0X0_ASPNG	NVADSTSN-----	EA-NC KD-VGNNQWTYNYGTY-----	LSGA AFMY
076	sp Q75DG6 DCW1_ASHGO	VVYDGLTD----I---	ND-NCTN-LNKLQWTYNHGLI-----	MAGCAFIY
077	tr Q6BZF0 Q6BZF0_DEBHA	YVYDGVVT----V---	GG-NCTD-ITKLQWSYNQALM-----	LSGC AFLY
078	tr E9E3Q1 E9E3Q1_METAQ	HAYDGAH----V---	AQ-NCTD-INKATFSYNAGVL-----	VQGA AFLY
079	sp P36091 DCW1_YEAST	YVYDGVVS----I---	DD-NCTK-VTSYQWTYNQGLL-----	LAGSAYLY
080	tr W0TAH6 W0TAH6_KLUMD	FVYDGLAS----I---	EN-NCTK-IVKYQWTYNQGLI-----	LSGSAYMY
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	RVYDGLAS----I---	TD-NCTS-ITGLEWTYNI GLL-----	MAGSAYAY
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	EVEDGAH----T---	DD-ECKD-MTKVLWSYNFGIH-----	MHAAANMY
083	tr Q2TYU3 Q2TYU3_ASPOR	NVADSTD I-----	EG-GCKS-QGNNQWSYNYGTY-----	LIGAAYMY
084	tr G2XGF6 G2XGF6_VERDV	KVYDGLVE----VVEN	SP-DCPK-IDKNQWSYNAGIY-----	LHGA AVMH
085	tr G3AMT4 G3AMT4_SPAPN	FVYDGAKE----V---	AN-NCSN-ITKYQWTYNQGGLM-----	LAGCAYLY
086	tr A0A1C1D213 A0A1C1D213_9EURO	NFFDGLSD----D---	NL-NCTE-LDHIQWTYNAGTF-----	LVGAANMY
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	NVYDGAH----T---	GV-NCTD-VFKAQFSYNAAIF-----	LQGA AFMY
088	tr A0A100I5A6 A0A100I5A6_ASPNG	NVADSTSN-----	EA-NC KD-IGNNQWTYNYGTY-----	LSGA AFMY
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	NVYDGAT----I---	DE-NCTD-ITKLEWSYNQGIV-----	LGGAAYMY
090	tr G2Q8A7 G2Q8A7_MYCTT	NIYDGGH----V---	QH-NCTD-INRAQFSYNNAVY-----	LLGA AFMY
091	tr B8NPZ1 B8NPZ1_ASPFN	YIADSTSN-----	LN-NCSD-AGDQQWSYNYGTY-----	LAGA AFMY
092	tr Q7SAB2 Q7SAB2_NEUCR	NVYDGAH----T---	GV-NCTD-VFKAQFSYNAAIF-----	LQGA AFMY
093	tr A2R8R5 A2R8R5_ASPNC	NVADSTST-----	TN-DCST-QGDNQWSYNYGAY-----	LGGAAYMY
094	tr C5DHG0 C5DHG0_LACTC	NVYDGAN----I---	DD-NCTD-IVKNEWSYNQGIV-----	LGGAAYMY
095	tr G0SFA3 G0SFA3_CHATD	QVHDGVH----INTN	DN-TCSQ-TDTNQWTYNAGIF-----	LHGA AVMY
096	tr K1WJG5 K1WJG5_MARBU	KVYDGLSS----D---	TE-NCTS-IDHLQFSYNHGIF-----	LFGA AVMY
097	tr H2B005 H2B005_KAZAF	FVYDGLAS----V---	TD-NCTT-VTKYQWTYNQGLL-----	LSGSAYLY
098	tr C5DYD7 C5DYD7_ZYGRC	NVFDGLAV----T---	DD-NCTQ-VTELEWTYNHGII-----	LGGSAYMY
099	tr G8YM23 G8YM23_PICSO	YIYDGLVE----A---	-K-ACEN-ITKYQWTYNQGLL-----	LSGC AFLY
100	tr W3X8E3 W3X8E3_PESFW	NIYDGAH----I---	ET-NCTD-INKAQFSYNNGVY-----	LLGAAYMY
101	tr A0A254U3K7 A0A254U3K7_ASPNG	NVADSTST-----	TN-DCST-QGDNQWSYNYGAY-----	LGGAAYMY
102	tr Q6CER8 Q6CER8_YARLI	HVYDGI-----H---	LP-NCSD-RSPYLWTYNSGIY-----	MSGAASIY
103	tr G2QT05 G2QT05_THITE	NIYDGGH----V---	EQ-NCTD-INRAQFSYNNAVY-----	LLGAAYMY
104	sp Q5ACZ2 DFG5_CANAL	RIYDGAKE----I---	TN-NCSS-VTDLRWSYTYGVF-----	MAGCAYLY
105	tr G2WKU6 G2WKU6_YEASK	NVFDGAE----I---	DT-NCTD-ITKIEWTYNHGIV-----	LGGLAYMY
106	tr A0A0C7N752 A0A0C7N752_9SACH	NVYDGAN----I---	DD-NCTD-ITKLEWSYNQGIV-----	LGGLAYLY
107	tr W3WN68 W3WN68_PESFW	NVL DGLAG-----N-AD	DA-NCTE-INAAQFTYNAGIY-----	LIGA AHMY
108	tr G2QFS1 G2QFS1_MYCTT	KVYDGGH----V---	EH-NCTD-INKAQFSYNAA LL-----	LHGS AFMY

109	tr G3J9G4 G3J9G4_CORMM	VIYD	IGIE	----	VDDA	TQ-	SKN-	IHQV	QFTY	NAGVW	-----	LQ	GAA	AMY
110	tr Q2UJ03 Q2UJ03_ASPOR	LFLD	GAD	----	E---	LK-	NCSE	-FNYL	QWTY	NSGVY	-----	LF	GAA	SMY
111	tr G8YRT2 G8YRT2_PICSO	YIYD	GAN	----	I---	GK-	NCSD	-VTKT	KWSY	IYGVF	-----	IA	GCA	YMH
112	tr W3X554 W3X554_PESFW	QVFD	G AQ	----	V---	TS-	NCSD	-IDKA	QWTY	NAGIF	-----	LH	GAA	VMY
113	tr Q1K7A8 Q1K7A8_NEUCR	DVKD	G AH	----	FDVT	TH-	VCRND	SGPH	VWSY	NIGVF	-----	LQ	GAA	AFMY
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	TIAD	TTSP	----	---	ET-	GCTD	-HGDL	QWTY	NYGTY	-----	IS	GAA	YMY
115	tr A7F3P0 A7F3P0_SCLS1	EVYD	GTS	----	D---	TD-	NCSS	-IDHI	QWTY	NAGIY	-----	LL	GS	AMMY
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	EVYD	GYN	----	VLDD	TK-	TCGH	-LHDV	QFSY	NAGIW	-----	LA	GTA	AMY
117	tr Q6CIP9 Q6CIP9_KLULA	NVFD	GAT	----	I---	DD-	NCTS	-IVKY	EWSY	NHGVV	-----	LG	GCA	YMY
118	tr H2AMQ5 H2AMQ5_KAZAF	YVYD	GAS	----	T---	ED-	NCSS	-VTKY	QWTY	NQGLI	-----	LS	GAA	YLY
119	tr G8JMI3 G8JMI3_ERECY	NVFD	G AH	----	I---	GD-	NCQD	-IKGI	EWTY	NHGV I	-----	LG	GCA	YMY
120	tr C5DGT1 C5DGT1_LACTC	YVYD	GAK	----	V---	DG-	NCST	-ITKL	IWTY	NQGLL	-----	MS	GCA	YLY
121	tr K1XJR4 K1XJR4_MARBU	NVFD	G GH	----	I---	GH-	NCTD	-INRV	QFSY	NIAIW	-----	LL	GAA	NMY
122	tr A0A124BYC5 A0A124BYC5_ASPNG	NVAD	STDA	----	---	AN-	GCTN	-LGNN	QWSY	NYAVY	-----	LS	GAA	YMY
123	tr A7TLL2 A7TLL2_VANPO	FVYD	G VN	----	I---	GD-	NCTK	-VTNL	QWTY	NHGLI	-----	LS	GCA	YLY
124	tr Q0CB86 Q0CB86_ASPTN	NVAD	STDI	----	---	ET-	GCKT	-QGNN	QWSY	NYGTY	-----	LT	GAA	YMY
125	tr K1XLH0 K1XLH0_MARBU	EVFD	GSQ	----	N---	TD-	NCTE	-KDHN	KWTY	NTGIF	-----	IM	GAA	ATMY
126	tr J7S438 J7S438_KAZNA	FVYD	GVD	----	A---	ND-	NCST	-VTQY	QWTY	NHGLL	-----	LA	GS	AYLY
127	tr Q4WG09 Q4WG09_ASPFU	YIAD	STSN	----	---	EA-	NCKD	-AGNT	QWSY	NYGTY	-----	LS	GAS	FMY
128	tr H8WZ09 H8WZ09_CANO9	RIYD	G AN	----	I---	EE-	NCTD	-VTDL	RWSY	TYGVF	-----	MT	GCA	YLY
129	tr I1RL09 I1RL09_GIBZE	LVYD	G GH	----	V---	GK-	NCTD	-INKA	TFSY	NAAIL	-----	IQ	GAA	AFMY
130	tr Q4W985 Q4W985_ASPFU	LFYD	GAD	----	V---	LL-	NCSD	-LNR I	EWTY	NSGVY	-----	LL	GAA	NMY
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	NVFD	GAE	----	I---	DT-	NCTD	-ITKI	EWTY	NHGIV	-----	LG	GM	AYMY
132	tr W6QCW3 W6QCW3_PENRF	RFWD	GAS	----	D---	LT-	ECKP	-INQ I	EWTY	NAGVY	-----	LL	GAA	NMY
133	tr W3WMD3 W3WMD3_PESFW	NVYD	G AH	----	L---	P-	DCTD	-INKA	QFSY	NSAML	-----	MQ	GAA	FLY
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	YVYD	GVD	----	I---	TN-	NCSG	AVHKY	QWTY	NQGLL	-----	MS	GCA	YLY
135	tr I2GYH1 I2GYH1_TETBL	EVYD	GAD	----	I---	SE-	NCTD	-ITKY	QWSY	NNGCM	-----	LG	GAA	YMY
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	NVYD	GAE	----	I---	DS-	NCTE	-VTR L	EWTY	NHGIV	-----	LG	G L	AYMY
137	tr Q752P3 Q752P3_ASHGO	RVFD	G AH	----	I---	HE-	NCSD	-IKGI	EWSY	NHGII	-----	LA	GAA	YMY
138	tr B8MYP3 B8MYP3_ASPFN	NIAD	TTTS	----	---	EA-	NCKD	-HGDL	QWTY	NYGTY	-----	LS	GAA	YMY
139	tr Q2TXL6 Q2TXL6_ASPOR	TIND	LVTV	----	---	ES-	NCQA	-PNQM	QWSY	NYGIY	-----	FN	GAA	YMY
140	tr Q0CN17 Q0CN17_ASPTN	TIAD	TTTT	----	---	QS-	NCKA	-HGDL	QWTY	NYGMY	-----	LS	GAA	YMY
141	tr J3NHD5 J3NHD5_GAGT3	NVYD	G GH	----	I---	GK-	NCTD	-INRA	QFSY	NNAVW	-----	LL	GAA	HMY
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	NIYD	GTD	----	D---	LS-	NCTT	-INHV	QYSY	NAGVW	-----	IL	GAA	HMY
143	tr A1CVB0 A1CVB0_NEOFI	LFYD	GAD	----	V---	LL-	NCSD	-FNRI	EWTY	NSGVY	-----	LL	GAA	NMY
144	tr B8NVI3 B8NVI3_ASPFN	TIND	LVTV	----	---	ES-	NCQA	-PNQM	QWSY	NYGIY	-----	FN	GAA	YMY
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	LVFD	GAK	----	V---	EG-	NCSN	-ITKL	VWTY	NQGLL	-----	MS	GCA	YLW
146	tr A1DJ54 A1DJ54_NEOFI	YIAD	STSN	----	---	EA-	NCKD	-AGNT	QWSY	NYGTY	-----	LS	GAS	FMY
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	KVYD	G GH	----	V---	EH-	NCTD	-INKA	QFSY	SAAIL	-----	VQ	GAA	AFMY
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	FVYD	G AN	----	I---	DD-	NCTE	-ITKL	QWTY	NQGLM	-----	MS	GS	AYLY
149	tr A0A177A618 A0A177A618_9PEZI	DVFD	GSS	----	D---	LL-	NCTE	-LDHT	QWSY	NAGIW	-----	LH	GAA	NMY
150	tr Q75CW6 Q75CW6_ASHGO	KVLD	G AH	----	I---	SE-	RCED	-HSKL	EWTY	NHGVV	-----	LA	GCA	YMY
151	tr Q7S4K4 Q7S4K4_NEUCR	KVYD	G GH	----	V---	EH-	NCTD	-INKA	QFSY	SAAIL	-----	VQ	GAA	AFMY
152	tr G2QB99 G2QB99_MYCTT	TVRD	G VH	----	VSLE	DG-	SCNS	-RDEN	QWTY	NAGIF	-----	LH	GAA	VMY
153	tr G0S3F2 G0S3F2_CHATD	AVYD	G GH	----	V---	EH-	NCTD	-INRA	QFSY	NAALL	-----	LH	GAA	AFMW
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	KIVD	G LE	----	V---	-D-	NCSD	-ISPY	QWTY	NHGLF	-----	LA	GCA	YLY
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	NVYD	GAK	----	V---	ED-	NCTD	-ITAI	EWTY	NHGVV	-----	LG	G L	AYMY
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	SVAD	STNI	----	---	ED-	GCES	-QGNN	QWSY	NYGTY	-----	LM	GAA	YMW
157	tr G2QKJ0 G2QKJ0_MYCTT	DVLD	G AH	----	I---	GH-	NCTD	-LNPV	QFSY	ANAAML	-----	IH	ATA	VMY
158	tr Q4WFX5 Q4WFX5_ASPFU	YVFD	GTY	----	E---	SD-	NCSA	-LNRV	EWTY	NNGVF	-----	LH	GAA	HMW
159	tr Q5BGD7 Q5BGD7_EMENI	FVAD	STSG	----	---	SN-	DCVD	-ADRM	QWSY	NYGTF	-----	IA	GAA	YMY
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	YVYD	GAS	----	I---	DE-	NCSD	-IVKY	QWTY	NQGLM	-----	LG	GS	AYLY
161	tr G8BYN9 G8BYN9_TETPH	NVYD	GAT	----	T---	NY-	NCSD	-IVQY	EWSY	NHGVV	-----	LG	G L	AYMY
162	tr F7VVT4 F7VVT4_SORMK	NIYD	G GH	----	V---	PH-	NCTD	-INKV	QWSY	ANAAIM	-----	IH	GVA	AIMY
163	tr G8YB80 G8YB80_PICSO	YIYD	GVE	----	A---	-K-	ACEN	-ITKY	QWTY	NQGLL	-----	LA	GCA	FLY
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	RVYD	GAD	----	I---	AE-	NCTD	-IVHI	EWTY	NIGIM	-----	IG	GAA	YLY
165	tr G2WRS7 G2WRS7_VERDV	NIYD	G GH	----	V---	EH-	NCTD	-INKA	QFSY	NNAVF	-----	LL	GAA	HMY
166	tr A0A124BVB5 A0A124BVB5_ASPNG	HVFD	GTS	----	E---	EN-	NCTD	-LNHI	EWTY	NNGVF	-----	LL	GAA	HMW
167	tr A7EIG7 A7EIG7_SCLS1	YVYD	GSD	----	D---	TI-	NCTR	-LNHI	QWTY	NAGAF	-----	LL	GAA	NMY
168	tr C5M5U7 C5M5U7_CANTT	FVYD	G VK	----	I---	YN-	NCSN	-ITKY	QWTY	NQGLM	-----	LA	GCA	YLY
169	tr B9WAA8 B9WAA8_CANDC	RIYD	GAK	----	I---	AN-	NCSS	-VTDL	RWSY	TYGVF	-----	MT	GCA	YLY
170	tr F8MRU1 F8MRU1_NEUT8	KVYD	G GH	----	V---	EH-	NCTD	-INKA	QFSY	SAAIL	-----	VQ	GAA	AFMY
171	tr A1DJS0 A1DJS0_NEOFI	SVAD	STNI	----	---	ED-	GCES	-QGNN	QWSY	NYGAY	-----	LM	GAA	YMW
172	tr B6HLM8 B6HLM8_PENRW	AVAD	SVRQGTGG	----	---	PN-	GCTL	-PDHT	RWTY	NYGTY	-----	LS	GAG	YMY
173	tr A0A117DWS0 A0A117DWS0_ASPNG	YIAD	TTTT	----	---	EA-	NCKD	-HGDI	QWTY	NYGTY	-----	LS	GAA	YMY
174	tr A0A124BXM1 A0A124BXM1_ASPNG	NVAD	STDV	----	---	DN-	NCKT	-QGNN	QWSY	NYGVF	-----	LT	GAA	YMY
175	tr J3NZQ6 J3NZQ6_GAGT3	RVYD	G AN	----	S---	NH-	NCTN	-INHV	QYMY	NPGIL	-----	MQ	GAA	AMY
176	tr W0THH8 W0THH8_KLUMD	NVYD	G AN	----	I---	EE-	NCTN	-IIKY	EWSY	NHGVV	-----	LG	GCA	YMY
177	tr B2AEF2 B2AEF2_PODAN	RVYD	G GH	----	V---	EH-	NCTD	-INKA	QFSY	NAALL	-----	LH	GS	AFMY

178	tr Q96TX1 Q96TX1_NEUCS	SIH	DGAH	-	-	-	V	-	-	-	ET	-	NCTD	-	INNAQFS	SYNSGVF	-	-	-	-	-	VL	GAAHMY																				
179	tr E9DYA6 E9DYA6_METAQ	AIY	DGAN	-	-	-	V	-	-	-	AN	-	QCKD	-	INKVQYS	SAFALRVPDCAKRRL	TP	RI	PT																								
180	tr B9WAI2 B9WAI2_CANDC	FVY	DGVK	-	-	-	V	-	-	-	AN	-	NCSN	-	ITKYQWS	YNQGLM	-	-	-	-	-	LA	GCAYLY																				
181	tr Q6C171 Q6C171_YARLI	RVY	DGI	-	-	-	H	-	-	-	LP	-	NCSD	-	RSPYLWT	YNAGIY	-	-	-	-	-	LS	GAAALY																				
182	tr A6ZMV1 A6ZMV1_YEAS7	NVF	DGAE	-	-	-	I	-	-	-	DT	-	NCTD	-	ITKIEWT	YNHGIV	-	-	-	-	-	LG	GLAYMY																				
183	tr B6K7X7 B6K7X7_SCHJY	TVY	DGAT	-	-	-	T	-	-	-	AD	-	NCST	-	LTETQWT	YNVGVF	-	-	-	-	-	LG	GAAAYLY																				
184	tr I1RSA2 I1RSA2_GIBZE	EIY	DGVH	-	-	-	I	G	K	E	K	S	-	SV	CDDIDKT	QWSYNSGVF	-	-	-	-	-	LH	GAAHMY																				
185	tr F7VWZ9 F7VWZ9_SORMK	DVK	DGAH	-	-	-	F	D	V	T	TH	-	V	CR	ND	SGPHVWS	YNVGVF	-	-	-	-	-	LQ	GAAAFMY																			
186	tr A0A1B2J789 A0A1B2J789_PICPA	FVY	DGAF	-	-	-	V	-	-	-	ED	-	NCTE	-	IVMLQWT	YNAGLM	-	-	-	-	-	VS	GCAYLA																				
187	tr G0V8N4 G0V8N4_NAUCC	FVY	DGAS	-	-	-	T	-	-	-	TD	-	NCTV	-	LTTYQWT	YNQGLL	-	-	-	-	-	LS	GAAYLY																				
188	tr H8WXN8 H8WXN8_CANO9	FVY	DGVK	-	-	-	I	-	-	-	AN	-	NCSN	-	ITKYQWT	YNQGLM	-	-	-	-	-	LA	GCAYLY																				
189	tr A6ZZS0 A6ZZS0_YEAS7	YVY	DGVS	-	-	-	I	-	-	-	DD	-	NCTK	-	VTSYQWT	YNQGLL	-	-	-	-	-	LAG	SAYLY																				
190	tr G9MQD1 G9MQD1_HYPVG	LVY	DGGH	-	-	-	V	-	-	-	EK	-	NCTD	-	INLATFS	YNAAVL	-	-	-	-	-	TH	GAAAFMY																				
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	FVY	DGAK	-	-	-	V	-	-	-	EG	-	NCSN	-	ITRLVWT	YNQGLL	-	-	-	-	-	MS	GSAYLY																				
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	YI	A	D	S	T	S	N	-	-	-	-	EA	-	NCKD	-	AGNTQWS	YNYGTY	-	-	-	-	LS	GASFGMY																			
193	tr A0A254UET1 A0A254UET1_ASPNG	HV	F	D	G	T	S	-	-	-	E	-	EN	-	NCTD	-	LNHIEWT	YNNGVF	-	-	-	-	LL	GAAHMY																			
194	tr A1D587 A1D587_NEOFI	TI	A	D	T	T	T	P	-	-	-	-	DT	-	ECTD	-	HGDLQWT	YNYGTY	-	-	-	-	LS	GAAYMY																			
195	tr Q1K7I4 Q1K7I4_NEUCR	SIH	DGAH	-	-	-	V	-	-	-	ET	-	NCTD	-	INNAQFS	SYNSGVF	-	-	-	-	-	VL	GAAHMY																				
196	tr G8JS88 G8JS88_ERECY	FVY	DGAF	-	-	-	I	-	-	-	HD	-	NCSR	-	PSVLQWS	YNHGLI	-	-	-	-	-	LG	GCAYLT																				
197	tr A0A136J7D3 A0A136J7D3_9PEZI	NVF	DGAG	-	-	-	I	-	-	-	VD	-	Q	C	A	T	-	H	D	K	A	Q	W	T	Y	N	A	G	I	Y	-	-	-	-	-	L	H	G	A	A	A	L	Y
198	tr E9E4X8 E9E4X8_METAQ	AIY	DGAK	-	-	-	V	-	-	-	AN	-	GCKD	-	INKAEFS	SYNNAVF	-	-	-	-	-	AE	G	V	A	F	M	Y															
199	tr Q6CP42 Q6CP42_KLULA	YVY	DGAS	-	-	-	I	-	-	-	EE	-	NCTD	-	TVKYQWT	YNQGLM	-	-	-	-	-	LS	GS	A	A	L	Y																
200	tr G0WHL4 G0WHL4_NAUDC	NVF	DGAE	-	-	-	I	-	-	-	DS	-	NCTD	-	MTQIEWT	YNHGIV	-	-	-	-	-	LG	G	A	A	L	Y																
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	YVY	DGVS	-	-	-	I	-	-	-	GD	-	NCTK	-	VTSYQWT	YNQGLL	-	-	-	-	-	LAG																					

248	tr B2W762 B2W762_PYRTR	HFYDGTD-----D--LQ-NCTT-INHIQWTYNAGVH-----MAGAAALW
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	YVYDGVVS-----I--DD-NCTK-VTSYQWTYNQGLL-----LAGSAYLY
250	tr C4XWK1 C4XWK1_CLAL4	I IYDGAD-----D--TE-NCTD-LTIHKWSYTYGVY-----LAGCAYLY

001	tr A0A254U2J9 A0A254U2J9_ASPNG	NYT-----G--K-AKWKTAAEGLLNVT---LDTEFFPAKY-----G
002	sp Q6FLP9 DCW1_CANGA	NM--T-----G--SDLWHERTHAFLNAS---RVFFN-----
003	tr A1CCM5 A1CCM5_ASPCL	NYT-----N-GTN-TKWESAVNGLLNVT---LNQFFPTAY-----G
004	tr C1H190 C1H190_PARBA	KY--T-----N-G--DKKWEERVVRGILKGI---EIFFQN-----
005	tr Q2UR85 Q2UR85_ASPOR	NLT-----D-GG--EKWKEAIDGLLGTT---IAKFFFPHEY-----G
006	tr A5DV30 A5DV30_LODEL	NF--T-----G--DEVWKTTRTEEIVDAS---LSYFFT-----S--
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	NA--T-----G--DDAWGSELDKFLGGI---EHYFLS-----P-N
008	tr J3PGN0 J3PGN0_GAGT3	NYT-----DG-----GAIWKERVEKLTESI---FRDFFA-----
009	tr A0A124BXU6 A0A124BXU6_ASPNG	NYT-----N-GTS-TKWMNAVDGLLNRT---LNKFFFPASH-----G
010	tr C5P4A1 C5P4A1_COCP7	NY--T-----D-G--EQIWRERVQGIIDGL---RP-FFP-----E
011	tr Q0CG55 Q0CG55_ASPTN	NL--T-----G-G--SPVWKERTENILNAT---DVFFQN-----Q
012	tr C4Y2X5 C4Y2X5_CLAL4	NY--T-----L--DEKWLNRMTMNL LHAT---QVFVYN-----KSA
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	NL--T-----E--SPVWKARTEGILNAS---FVFFQD-----D
014	tr C7Z068 C7Z068_NECH7	NYT-----NG-----SEVWETRVNNLTDSL---LKNFFP-----
015	tr G8BXX2 G8BXX2_TETPH	NY--T-----G--SELWHTRTKNFLSSS---SVFFN-----
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	NY--T-----E--DEKWHNRTLNFLKSA---EVFFYK-----KLE
017	tr A1CSC2 A1CSC2_ASPCL	NYT-----N-GG--SKWKKGLDGLLGTT---FSLFFPFQN-----
018	sp Q05031 DFG5_YEAST	NA--T-----N-G--TGEWETSLTKILNGA---KSYFFK-----
019	tr H2AQJ6 H2AQJ6_KAZAF	NA--T-----N-G--SVWESRVTDIVEGA---KSIFFS-----
020	tr G2XFH9 G2XFH9_VERDV	NYT-----E-----DAKWLDRVEKLLDRC---LTDFFP-----
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	KT--N-----G--QEKWYNRTEDFVKSS---GVFYN-----
022	sp Q5AD78 DCW1_CANAL	NY--T-----E--EEKWYNNTIKLLESA---QVFFKN-----I-S
023	tr B6H7E8 B6H7E8_PENRW	NYT-----N-GTE-GKWLEGVTGLLKTS---EQFFPKSG-----D
024	tr B6GZU2 B6GZU2_PENRW	NL-----T-E--DPKWKERTQKIIDAS---GVFFSH-----D
025	tr E9E4W7 E9E4W7_METAQ	NHT-----S-----DKWKTRLDSSLDAC---LKSFFP-----
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	NV-----T-E--DPKWKERVQKILDSS---EIFFAK-----T
027	tr G2R7G8 G2R7G8_THITE	NYT-----NG-S-----EKWRARVAGLLNHT---IDFFFP-----
028	tr G3JBY1 G3JBY1_CORMM	DY--T-----K---QDIWKTHVDGLLNFT---TRMFVS-----H
029	tr A5DNV5 A5DNV5_PICGU	NY--T-----Q--DEVWHERTKKLLAGS---VVFFNS-----T--
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	NY--T-----G-G--TDTWRRHTSGLLKTT---SSTFFR-----D
031	tr Q2TWC1 Q2TWC1_ASPOR	NYT-----N-GD--EKWLKRVNGLLEST---FATFFPSTY-----
032	tr J5RXY9 J5RXY9_SACK1	NF--T-----G--SDLWHTRTKEFLNSS---QVFFQ-----
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	DY--N-----K---TDVWKGHVNGILNNT---ERTFFQ-----N
034	tr A0A254U5V1 A0A254U5V1_ASPNG	NLT-----N-GA--DKWKKGLDGLLAST---FSRFFPKEY-----G
035	tr A0A061B6H7 A0A061B6H7_CYBFA	NF--T-----E--EEKWRNRTRLRYLQSS---SVFFN-----
036	tr G8ZQ93 G8ZQ93_TORDC	NM--T-----K--DEKWGNRTFEYLNSA---SIFYN-----
037	tr Q9C2J1 Q9C2J1_NEUCS	NV--S-----T-GAEQETWKTRVDGLLGAV---EAKFLT-----N
038	tr C5NZK5 C5NZK5_COCP7	NF--T-----K-G--ADIWEKRLNGLIDAA---GIFFSK-----D
039	tr Q6C0T7 Q6C0T7_YARLI	NG--TLLRNGTGS-----EKWQGQVRVGIFNTAVSP LHF FGG-----P
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	NM--T-----G--SDLWHERTHAFLNAS---RVFFN-----
041	tr F7VZ72 F7VZ72_SORMK	NYT-----DG-----SPVWKDRLDKLVNAT---LVRFFP-----
042	tr A3LN37 A3LN37_PICST	NY--T-----E--DQKWLDRTLNL LNAS---QVFFMK-----I-G
043	tr Q6CAI2 Q6CAI2_YARLI	NA--T-----S--DEQWATRAKNIWTGC---EDLFFQ-----
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	NY--T-----E--DELWYNRTMRLLSGA---QVFFKD-----
045	tr G9MJS5 G9MJS5_HYPVG	NHT-----N-----SAVWKTRLQKLVDHG---LETFFP-----
046	tr G9MHI4 G9MHI4_HYPVG	NF--T-----G---NATWKANLDGIISQT---VSKFV-----
047	tr D4AP27 D4AP27_ARTBC	NY--T-----D-G--SPKWAERVQGILDGL---RP-FFH-----P
048	tr B6GZT8 B6GZT8_PENRW	NYT-----N-GTIQAQWLTAVNGLLNAT---FNNFFLT-----
049	sp O74556 YCZ2_SCHPO	NY--T-----N---STVWKTRVEGFANKT---AKTFFF-----
050	tr A0A0C4E991 A0A0C4E991_MAGP6	NFT-----DG-----GAIWKDRVEKLTDSI---FRDFFA-----
051	tr B8MHG0 B8MHG0_TALSN	NYT-----N-GG--TKWKEGVDGLLNTT---WNTFFPTEY-----
052	tr A3LMV8 A3LMV8_PICST	DY--T-----K--DEKWLNRVNDLMETS---ASTFFP-----K-S
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	NA--T-----G--EQFWDNELGLFLGGL---EHYMVN-----T-T
054	sp Q9P6I3 YHG7_SCHPO	NYT-----N-GS--SVWQTHMEGLMNKA---LDYYFTSD-----
055	tr Q2US57 Q2US57_ASPOR	NYT-----N-GS--TTWLTAVNGLLNAT---IDGFFPTKY-----G
056	tr W7N622 W7N622_GIBM7	NI--T-----G---KASWKTRVDGLLEDI---KTVFV-----
057	sp Q9P6I4 YHG6_SCHPO	NYT-----GE--TVWRERLDGLISHA---TSYFFT-D-----
058	tr C7GRB3 C7GRB3_YEAS2	NA--T-----N-G--TGEWETSLTKILNGA---KSYFFK-----
059	tr C7GP28 C7GP28_YEAS2	NF--T-----G--SDLWHTRTKEFLNAS---QVFFH-----
060	tr G3JGZ5 G3JGZ5_CORMM	NYT-----DG-----DKKWGDRVDKLLDSM---LKNFIR-----
061	tr C4QXV4 C4QXV4_KOMPG	NY--T-----G--DEMWLDRTENFLHGI---QVFTN-----
062	tr I1S0Z5 I1S0Z5_GIBZE	NHT-----DG-----EQKWEQRLDKLVDAT---ISNFFP-----
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	NCYKT-----E-G--DSRWEARTKHILQAT---DAFFAE-----D
064	tr G0W5X4 G0W5X4_NAUDC	NF--T-----G--STIWLNRMTMDLLDAT---SVFFN-----

065	tr G8ZPC1 G8ZPC1_TORDC	NA	-	T	-	-	-	-	-	N	-	G	-	S	SEW	ETRV	TQIL	GGA	-	-	ESY	FFK	-	-	-	-																			
066	tr A0A167CDC5 A0A167CDC5_9ASCO	DF	-	T	-	-	-	-	-	N	-	D	T	KWL	TRA	QLLM	ERG	-	-	SSI	YFD	-	-	-	-	-																			
067	tr R9XAT7 R9XAT7_ASHAC	NH	-	T	-	-	-	-	-	Q	-	D	E	VWH	QRT	LRFL	ESA	-	-	AVF	IK	-	-	-	-	-																			
068	tr B2B747 B2B747_PODAN	NYT	-	-	-	-	-	-	NG	-	-	-	-	S	DVW	RER	LDGL	T	D	A	T	-	-	-	-	-																			
069	tr G0SE99 G0SE99_CHATD	NYT	-	-	-	-	-	-	NG	-	-	-	-	S	DVW	AAR	VNGL	V	N	S	T	-	-	-	-	-																			
070	tr J8Q6F0 J8Q6F0_SACAR	NA	-	T	-	-	-	-	-	N	-	G	-	T	SEW	KTR	V	T	QV	L	N	G	A	-	-	-																			
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	NA	-	T	-	-	-	-	-	N	-	G	-	S	SEW	QTR	TE	QIL	G	G	A	-	-	-	-	-																			
072	tr J3K6E2 J3K6E2_COCIM	NY	-	T	-	-	-	-	-	D	-	G	-	E	QIW	RER	V	QGM	I	D	G	L	-	-	-	E																			
073	tr G4N3E1 G4N3E1_MAGO7	NIT	-	-	-	-	-	-	S	-	-	-	-	-	NEK	WKT	A	T	T	K	L	T	E	S	L	-																			
074	tr C5M5J2 C5M5J2_CANTT	NF	-	T	-	-	-	-	-	G	-	D	T	VWQ	TRA	L	E	I	V	E	A	S	-	-	-	S																			
075	tr A0A254U0X0 A0A254U0X0_ASPNG	NYT	-	-	-	-	-	-	-	N	-	G	S	-	T	KWE	ERV	NNLL	T	S	L	-	-	-	-																				
076	sp Q75DG6 DCW1_ASHGO	NH	-	T	-	-	-	-	-	Q	-	D	E	LWH	QRT	LRFL	D	S	A	-	-	-	-	-	-																				
077	tr Q6BZF0 Q6BZF0_DEBHA	NY	-	T	-	-	-	-	-	E	-	D	E	MWY	NRT	MRL	L	S	G	A	-	-	-	-	-																				
078	tr E9E3Q1 E9E3Q1_METAQ	NYT	-	-	-	-	-	-	DG	-	-	-	-	A	QKW	KTA	L	D	G	L	D	A	T	-	-																				
079	sp P36091 DCW1_YEAST	NF	-	T	-	-	-	-	-	G	-	S	D	LWH	TRT	K	E	F	L	N	A	S	-	-	-	-																			
080	tr W0TAH6 W0TAH6_KLUMD	NH	-	T	-	-	-	-	-	G	-	S	E	LWH	KRT	K	D	F	L	N	S	A	-	-	-	T																			
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	NA	-	T	-	-	-	-	-	E	-	D	D	VWL	ERTE	A	L	V	Q	G	A	-	-	-	-																				
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	NV	-	T	-	-	-	-	-	G	-	-	D	Q	T	W	L	D	A	T	N	G	I	L	K	K																			
083	tr Q2TYU3 Q2TYU3_ASPOR	NMT	-	-	-	-	-	-	-	E	-	-	K	E	T	W	K	T	A	V	D	G	L	L	G	V	T																		
084	tr G2XGF6 G2XGF6_VERDV	NV	-	T	-	-	-	-	-	G	-	-	D	K	W	R	T	R	T	V	G	L	L	D	E	T																			
085	tr G3AMT4 G3AMT4_SPAPN	SY	-	T	-	-	-	-	-	Q	-	D	K	K	W	Y	N	T	H	R	L	L	D	G	S	-																			
086	tr A0A1C1D213 A0A1C1D213_9EURO	NY	-	T	-	-	-	-	-	D	-	G	-	S	Q	V	W	A	D	R	I	S	G	I	I	S	R																		
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	SFTKG	-	-	-	-	-	-	PE	-	-	-	-	-	Q	V	L	W	K	N	R	V	Q	A	L	L	D	R	T																
088	tr A0A100I5A6 A0A100I5A6_ASPNG	NYT	-	-	-	-	-	-	-	N	-	G	S	-	T	KWE	Q	R	V	NNLL	T	S	I	-	-	-																			
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	NA	-	T	-	-	-	-	-	N	-	G	-	S	S	T	WE	T	K	L	S	Q	L	V	G	G	S																		
090	tr G2Q8A7 G2Q8A7_MYCTT	NYT	-	-	-	-	-	-	NG	-	-	-	-	S	D	K	W	R	E	A	I	E	G	L	A	D	A	T																	
091	tr B8NPZ1 B8NPZ1_ASPFN	NHT	-	-	-	-	-	-	-	N	-	G	S	-	D	K	W	L	K	R	V	D	G	L	L	N	S	V																	
092	tr Q7SAB2 Q7SAB2_NEUCR	SFTKG	-	-	-	-	-	-	PE	-	-	-	-	-	Q	V	L	W	K	N	R	V	Q	A	L	L	D	R	T																
093	tr A2R8R5 A2R8R5_ASPNC	NYT	-	-	-	-	-	-	-	N	-	G	T	S	-	T	K	W	M	N	A	V	D	G	L	L	N	R	T																
094	tr C5DHG0 C5DHG0_LACTC	NA	-	T	-	-	-	-	-	Q	N	D	-	-	T	S	V	W	K	S	R	L	D	Q	I	V	G	G	S																
095	tr G0SFA3 G0SFA3_CHATD	NL	-	T	-	-	-	-	-	S	-	-	-	N	P	I	W	R	E	R	V	E	G	L	L	R	T	T																	
096	tr K1WJG5 K1WJG5_MARBU	EF	-	K	-	-	-	-	-	N	-	Q	-	S	S	V	W	Q	G	R	L	E	S	L	I	K	N	-																	
097	tr H2B005 H2B005_KAZAF	NF	-	T	-	-	-	-	-	E	-	S	E	L	W	H	E	R	T	K	G	F	L	N	A	S	-																		
098	tr C5DYD7 C5DYD7_ZYGRC	NA	-	T	-	-	-	-	-	N	-	G	-	S	S	K	W	E	E	R	V	N	Q	I	L	N	G	A																	
099	tr G8YM23 G8YM23_PICSO	NH	-	T	-	-	-	-	-	E	-	D	E	V	W	H	N	R	T	K	A	L	L	K	G	S	-																		
100	tr W3X8E3 W3X8E3_PESFW	NYT	-	-	-	-	-	-	NG	-	-	-	-	-	N	E	T	W	K	E	R	L	D	G	L	I	D	G	T																
101	tr A0A254U3K7 A0A254U3K7_ASPNG	NYT	-	-	-	-	-	-	-	N	-	G	T	S	-	T	K	W	M	N	A	V	D	G	L	L	N	R	T																
102	tr Q6CER8 Q6CER8_YARLI	NA	-	T	M	L	R	N	G	T	N	A	T	L	E	-	A	S	V	W	M	N	R	T	Y	Q	L	W	N	A	D	I	M	K	G	I	F	F	G	G	-	-	-	-	N
103	tr G2QT05 G2QT05_THITE	NYT	-	-	-	-	-	-	NG	-	-	-	-	-	S	D	T	W	K	E	R	L	E	G	L	V	D	A	T																
104	sp Q5ACZ2 DFG5_CANAL	NF	-	T	-	-	-	-	-	G	-	D	D	V	W	L	T	R	T	N	E	I	V	Q	A	S	-	-																	
105	tr G2WKU6 G2WKU6_YEASK	NA	-	T	-	-	-</																																						

134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	KT--N-----G--QEKWYNRTEDFVKSS---NVFYN-----
135	tr I2GYH1 I2GYH1_TETBL	NA--T-----N-G--SSIWETRVTQLLNGA---KAYFFQ-----
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	NA--T-----N-G--STWEERVVSQILGGA---TDYFFE-----
137	tr Q752P3 Q752P3_ASHGO	NA--T-----G-G--SPEWESHVTKILAGA---ADFYFR-----
138	tr B8MYP3 B8MYP3_AS PFN	NLT-----D-GG--EKWKEAIDGLLGTT---IAKFFFPHEY----G
139	tr Q2TXL6 Q2TXL6_AS POR	NLT-----N-GD--TKWKKNVVEGLLNTT---WRNFFFPQEY----G
140	tr Q0CN17 Q0CN17_AS PTN	NLT-----N-GD--TKWKKGIDGLLQTT---FSQFFFPKY----G
141	tr J3NHD5 J3NHD5_GAGT3	NYT-----DG-----SEVWKGRVQSLLDAT---LKTFPP-----
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	NI--T-----K--SDVWKARVQGILLDQS---IKIFSD-----N
143	tr A1CVB0 A1CVB0_NEOFI	NF--T-----E-G--DPRWEARTKRILQAT---DVFFVK-----D
144	tr B8NVI3 B8NVI3_AS PFN	NLT-----N-GD--TKWKNVVEGLLNTT---WRNFFFPQEY----G
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	NM--T-----Q--DEKWHVRTWEFLNAS---SIFFN-----
146	tr A1DJ54 A1DJ54_NEOFI	NYT-----N-GE--DKWLKRVNGLLDSDL---IGTFCKPKDK-----
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	NYT-----EGDAAT---QDMWKTRIEKLTEGL---FRDFFP-----
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	NF--T-----E--EEKWRTRTL DYLRSS---TVFFN-----
149	tr A0A177A618 A0A177A618_9PEZI	KF--T-----N-N--SEKWLERSTRILNKA---STQFFW-----K
150	tr Q75CW6 Q75CW6_ASHGO	NI--T-----N-G--SAVWGDRLNKVLNGS---LLF-FP-----
151	tr Q7S4K4 Q7S4K4_NEUCR	NYT-----EGDAAT---QDMWKTRIEKLTEGL---FRDFFP-----
152	tr G2QB99 G2QB99_MYCTT	NH--T-----N-A--SAIWRE RVDGLLAST---TATFVD-----P
153	tr G0S3F2 G0S3F2_CHATD	NYT-----ED-----QKWKDRV DNLLTGI---LRDFFK-----
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	NY--T-----E--DEKWRNR TLELLKGA---VVFFGT-----E--
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	NA--T-----N-G--SVWQSRLTSVLGGA---TAYFFQ-----
156	tr A0A0J5PQ80 A0A0J5PQ80_AS PFM	NYT-----N-GTN-SKWATAVDGLLNNT---LDIFFPAKY----G
157	tr G2QKJ0 G2QKJ0_MYCTT	NYT-----TG--ET---RAKWR RHVAGLLNHT---IDHFFP-----
158	tr Q4WFX5 Q4WFX5_AS PFU	NY--T-----Q-G--NSSWKARIDGLLDAQ---RTFLSP-----NKS
159	tr Q5BGD7 Q5BGD7_EMENI	NAT-----K--D--VKWRNRTEGLVDHV---FKHFFPTKYTTAIG
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	NY--T-----E--SEVWHERTKNFLNSS---GVFFN-----
161	tr G8BYN9 G8BYN9_TETPH	NA--T-----N-G--TSKWETALTQVLSGA---TTYFFK-----
162	tr F7VVT4 F7VVT4_SORMK	NMT-----ED--A-----EKWGKPLTGLVNRT---LEFFFP-----
163	tr G8YB80 G8YB80_PICSO	NY--T-----E--DEVWHNR TKLLNGS---RVFFQ-----
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	NF--T-----E--EQYWLDVTAHLVEGA---KVFFR-----
165	tr G2WRS7 G2WRS7_VERDV	NFT-----DG-----DAKWKTRIDGLLQRT---IEVFFP-----
166	tr A0A124BVB5 A0A124BVB5_AS PNG	NY--T-----N-G--DEKWRERV NGLLQAQ---NSFLST-----NES
167	tr A7EIG7 A7EIG7_SCLS1	NY--T-----N-G--SDIWRGRVEGLLNG---THVFFQ-----N
168	tr C5M5U7 C5M5U7_CANTT	NY--T-----E--EEKWYN YTIK LLESS---QVFFMN-----I-T
169	tr B9WAA8 B9WAA8_CANDC	NF--T-----G--DDVWL TRTNEIVQAS---LSYFFA-----
170	tr F8MRU1 F8MRU1_NEUT8	NYT-----EGDAAT---QDMWKTRIEKLTEGL---FRDFFP-----
171	tr A1DJS0 A1DJS0_NEOFI	NYT-----N-GTN-TKWATAVDGLLNNT---LDLFFPTKY----G
172	tr B6HLM8 B6HLM8_PENRW	AYT-----N-D--TKWLDITNNLLDSL---FTTFFLPEH-----
173	tr A0A117DWS0 A0A117DWS0_AS PNG	NLT-----N-GA--DKWKKGLDGLLAST---FSRFFFPKEY----G
174	tr A0A124BXM1 A0A124BXM1_AS PNG	NYT-----G---K-AKWKNAAEGLLNVT---LDTFFPAKY----G
175	tr J3NZQ6 J3NZQ6_GAGT3	DHT-----KK-----DKWKNIV TNLAKAA---IESFFA-----
176	tr W0THH8 W0THH8_KLUMD	NA--T-----N-G--SEEWKS RTSQ ILGGA---TSFFFD-----
177	tr B2AEF2 B2AEF2_PO DAN	NYT-----NG-----SEIWKTRVDKLWEGM---HRDFFE-----
178	tr Q96TX1 Q96TX1_NEUCS	NYTNTTQT DG-----SPVWKDR LDK LLNAT---LVRFFP-----
179	tr E9DYA6 E9DYA6_METAQ	AQT-----NG-----DQKWKD RV DKLINYG---LKTFPP-----
180	tr B9WAI2 B9WAI2_CANDC	NY--T-----E--EEKWYN YTIK LLESA---QVFFKN-----I-T
181	tr Q6C171 Q6C171_YARLI	NA--TVLRNH TD P--N--HGKWLD RANELWN STKGPNLF FGG-----P
182	tr A6ZMV1 A6ZMV1_YEAS7	NA--T-----N-G--TGEWETS LT KILNGA---KSYFFK-----
183	tr B6K7X7 B6K7X7_SCHJY	NY--T-----N-G--STVWQQ RL DGLITKA---SNHFFS-----
184	tr I1RSA2 I1RSA2_GIBZE	NI--T-----S---SDKWKKRAD GLLDDV---FNKFV-----
185	tr F7VWZ9 F7VWZ9_SORMK	NV--T-----T-GEEQ NT WKTRVDG LLGSI---ESKFLT-----N
186	tr A0A1B2J789 A0A1B2J789_PICPA	NY--T-----G--DDLWLD RT ENFLHGI---QVFTN-----
187	tr G0V8N4 G0V8N4_NAUCC	NY--T-----G--SDTWLLRTNQ LLNAS---SVFFN-----
188	tr H8WXN8 H8WXN8_CANO9	NY--T-----E--DQKWYN YTIQL LTS A---QVFFKN-----M-S
189	tr A6ZZS0 A6ZZS0_YEAS7	NF--T-----G--SDLWHT RT KEFLNAS---QVFFH-----
190	tr G9MQD1 G9MQD1_HYPVG	NYT-----DG-----SEIWKN RV DRLLDSL---LARFFP-----
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	NM--T-----E--DEKWHTR TW EFLNSS---NIFFN-----
192	tr A0A0J5PNX6 A0A0J5PNX6_AS PFM	NYT-----N-GE--DKWLKRVNGLLDSDL---IGTFCKPKDK-----
193	tr A0A254UET1 A0A254UET1_AS PNG	NY--T-----N-G--DENWRKRV NGLLQAQ---SSFLST-----NES
194	tr A1D587 A1D587_NEOFI	NFT-----N-GG--DKWKKGLDGLLNTT---FKRFFFPQN-----
195	tr Q1K7I4 Q1K7I4_NEUCR	NYT-----DG-----SPVWKDR LDK LLNAT---LVRFFP-----
196	tr G8JS88 G8JS88_ERECY	DF--T-----G--SDVWHNR TLRL LRSA---SIFFP-----
197	tr A0A136J7D3 A0A136J7D3_9PEZI	DY--T-----N-G--SALWEGRV NGLIKRT---REHFFN-----
198	tr E9E4X8 E9E4X8_METAQ	NYT-----NG-----NATWKAR LD GLIKHG---METFLP-----
199	tr Q6CP42 Q6CP42_KLULA	NF--T-----E--SETWHERTKNFLNSS---GIFFN-----
200	tr G0WHL4 G0W	

001	tr A0A254U2J9 A0A254U2J9_ASPNG	G-NIMSEILCEP-----TEVC--NDNEILFKGLVTGWLGLVALVMP-S
002	sp Q6FLP9 DCW1_CANGA	-NSILYEAAQCQGP-----NTC--NTDQRSFKAYFARFLGSTAELVP-E
003	tr A1CCM5 A1CCM5_ASPCL	G-NIMSEVLCEP-----SELC--NNNEVLFKGLVSSWLAFSTALLVP-S
004	tr C1H190 C1H190_PARBA	--DVMTEVACERN-----GKC--NVDQRSFKAYLSRWMAMTVKVAP-F
005	tr Q2UR85 Q2UR85_ASPOR	G-DIMSEISCEQ-----SMMF--DRNQDCFKGFLSSWLTFTTTIIAP-F
006	tr A5DV30 A5DV30_LODEL	-DKIMQETTQCP-----HNLC--NNDQRSFRSLFSRCLGLTKLIIP-E
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	ATDVLYEYQCLQW-----GRC--NNDQRSFRAIVARALGDVVVLAP-S
008	tr J3PGN0 J3PGN0_GAGT3	D-GAAYELPCEQK-----KGGC--TADMLSFKGYVHRWLAVASQVAP-F
009	tr A0A124BXU6 A0A124BXU6_ASPNG	G-EILTEILCEP-----TEVC--NDNEIIFKGLVSAWLAYTALLVP-S
010	tr C5P4A1 C5P4A1_COCP7	QADIMVNESSCEPH-----DNC--LTDQRSFKAFLSRWMAETTQLAP-F
011	tr Q0CG55 Q0CG55_ASPTN	--VLYERACEPI-----NTC--KVDQRSFKGYLASWMAQTTQMAP-F
012	tr C4Y2X5 C4Y2X5_CLAL4	DAHIIEYEAACSSP-TEN--KFTC--NNDQRSFKAYFCRCLGATSVLVP-Q
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	--VMYERACEPV-----STC--QVDQRSFKGYLARWMAATTQMAP-F
014	tr C7Z068 C7Z068_NECH7	K-GIAWEVPCEGR-----KGAC--STDMLSFKGYVHRWLSVVTQIVP-H
015	tr G8BXX2 G8BXX2_TETPH	-DTILYEAAQCQNQ-----ETC--NTDQRSFKAYFSRFLGVTAQLVP-E
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	DAHIMYEAAQCQSP-TSN--QYSC--NQDQRSFKAYFSRFLGLTSILVP-E
017	tr A1CSC2 A1CSC2_ASPCL	G-TVMSDEVACEP-----NMMC--DRNQDCFKGFLSSWLTFTTTIIAP-Y
018	sp Q05031 DFG5_YEAST	-DSIMYESACQD-----YGTC--NTDQRTFKSIFSRMLGLTSVMAP-F
019	tr H2AQJ6 H2AQJ6_KAZAF	-DGIMYESACQD-----YDTC--NNDQRSFKSIFSRMLGFTSVLAP-F

020	tr G2XFH9 G2XFH9_VERDV	R-GVAYEVP CERD	-----PGR	C--SADMLT	FKGYMHRWLAVVTQLVP	-S
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	-NSILYEATCQPY	-----NSC	--NTDQRS	FKAYFTTMLAATAQLIP	-D
022	sp Q5AD78 DCW1_CANAL	GSMVMYEAA CQPS	-----NSC	--NNDQRS	FKAYFSRFLGLTSVLVP	-Q
023	tr B6H7E8 B6H7E8_PENRW	H-QIISDITCEP	-----IDMC	--DRNQKT	FKTYFTSWIGFMSLIVP	-T
024	tr B6GZU2 B6GZU2_PENRW	PPNVMYERACE SV	-----NTC	--MVDQRS	FKGYFSRWMAQTAQMAP	-F
025	tr E9E4W7 E9E4W7_METAQ	R-QVAYELSCEFS	----LGGSVC	--KTDMLS	YKGYLVRWLAVVTQLAP	-H
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	PQNVMYERACE TV	-----NTC	--MVDQRS	FKGYLARWMAATTQMAP	-F
027	tr G2R7G8 G2R7G8_THITE	D-GIMVERACE LP	----DRVQC	--NTDQHS	FKGYMHRALATTAVVAP	-F
028	tr G3JBY1 G3JBY1_CORMM	NGSHLWEPPCETQ	-----PRGC	--DQNQVS	FKGYLLRCLAYTSKMAP	-W
029	tr A5DNV5 A5DNV5_PICGU	-NDIMYEAA CQNG	-KGGQGAGSC	--NQDQRS	FKAYFSRFLGLTAIMAP	-E
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	--KVMFEPA CEL	-----DDTC	--NTDQLS	FKAYLSRWMAATTKLAP	-F
031	tr Q2TWC1 Q2TWC1_ASPOR	GGNVLS EVA CEP	-----IMSC	--DRNQLG	FKGYTAMWLAHTAILVP	-S
032	tr J5RXY9 J5RXY9_SACK1	-DGIVYEAA CQGP	-----NSC	--NTDQRS	FKAYFARFLGVTAQLVP	-E
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	--DIMYEPA CET	-----VHTC	--DQNMVS	FKGYLIRFLAATAKMAP	-W
034	tr A0A254U5V1 A0A254U5V1_ASPNG	S-NIMSEIS CEP	-----NMMC	--DRNQDC	FKGFLSSWLTFTTTIAP	-Y
035	tr A0A061B6H7 A0A061B6H7_CYBFA	-NSVLYEAA CQGA	-----GNC	--NNDQRS	FKAYFSRFLGLTAYMVP	-E
036	tr G8ZQ93 G8ZQ93_TORDC	-NSVLYEVA CQTV	-----NKC	--NTDQRS	FKAYFTRFLGVTAELIP	-Q
037	tr Q9C2J1 Q9C2J1_NEUCS	DTKIIEK EWCESGFS	DRGHPYQC	--NIDQQT	FKGYLLRWLSSTSQVAP	-Y
038	tr C5NZK5 C5NZK5_COCP7	PPDVMT EVA CEGN	-----GKC	--NIDQRS	FKAYLSRWMAMTIKLAP	-Y
039	tr Q6C0T7 Q6C0T7_YARLI	EGNIMREIACQQA	-----TITC	--DADQRT	FKGIYSSLLGQTAQLVP	-D
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	-NSILYEAA CQGP	-----NTC	--NTDQRS	FKAYFARFLGSTAELVP	-E
041	tr F7VZ72 F7VZ72_SORMK	E-KIAYEPA CED	-----GMTC	--TTDMLS	FKGYVHRWLSTTTQIAP	-H
042	tr A3LN37 A3LN37_PICST	--SVMYEAA CQPS	-----NNC	--NNDQRS	FKAYFSRFLGMTAVMVP	-Q
043	tr Q6CAI2 Q6CAI2_YARLI	-DDKMVEIS CQQT	-----RITC	--NNDQRC	FKAIFSRFIGYAALFLP	-D
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	--NVMYEAA CQDS	-----NNC	--NQDQRS	FKAYFSRFLGLTSVMVP	-D
045	tr G9MJS5 G9MJS5_HYPVG	N-GIAYEPS CEG	-----VNTC	--TTDMVS	FKGYLHRWYATTTQLAP	-F
046	tr G9MHI4 G9MHI4_HYPVG	QNGVIYE QFCEPR	-----GFC	--SVDQST	FKGYLVRYMASTMQLAS	-H
047	tr D4AP27 D4AP27_ARTBC	ETYIMSEISCEEQ	-----GNC	--ETDQRS	FKAYLSRWMAASTQFAP	-F
048	tr B6GZT8 B6GZT8_PENRW	G-GIIEDYY CEP	-----TETC	--NNNEIL	FKGLTSSWLALTALLVP	-S
049	sp O74556 YCZ2_SCHPO	-KDIMFEPVCEIA	-----LSC	--NYDQTS	FKGFLTRFMVYTAQMAP	-F
050	tr A0A0C4E991 A0A0C4E991_MAGP6	D-GAAYELPCEQK	-----KGGC	--TADMLS	FKGYVHRWLAVASQVAP	-F
051	tr B8MHG0 B8MHG0_TALSN	GGNILSEVA CDP	-----ILTC	--NRDQVC	FKGLMANWLSTIALIVP	-Y
052	tr A3LMV8 A3LMV8_PICST	NGGYMTEIQ CFF	-----SNTC	--NNDQRS	SFRSLFSRCIGLTMKLVN	-S
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	GGNTLYEYQCEKW	-----QRC	--NNDQRA	FRAVVARTLGEIYQLAP	-Q
054	sp Q9P6I3 YHG7_SCHPO	--KIIYEPS CEP	-----TESC	--NSDQTA	FKGMLARFLGYTMQLAP	-Y
055	tr Q2US57 Q2US57_ASPOR	G-NIMSDYT CET	-----TEVC	--NNNEII	FKGLLSMWLAFTALLVP	-S
056	tr W7N622 W7N622_GIBM7	KNGVIYE QFCEEH	-----KLC	--NLDQQT	FKGYLARWMAATALVAP	-H
057	sp Q9P6I4 YHG6_SCHPO	--DIAWD PQCEY	-----FDDC	--NSDQTA	FKGIFMQSFGNTIRLAP	-Y
058	tr C7GRB3 C7GRB3_YEAS2	-DSIMYESA CQD	-----YGTC	--NTDQRT	FKSIFSRMLGLTSVMAP	-F
059	tr C7GP28 C7GP28_YEAS2	-DGIVYEAA CQGP	-----NSC	--NTDQRS	FKAYFARFLGVTAQLVP	-E
060	tr G3JGZ5 G3JGZ5_CORMM	D-GAMYE LPCEGT	-----PGTC	--TADMLT	FKGYVHRWLSVVAQIVP	-H
061	tr C4QXV4 C4QXV4_KOMPG	-QSVFFEAA CQGS	-----GNC	--NTDQRS	FKAYLARFLGLTAQMVP	-S
062	tr I1S0Z5 I1S0Z5_GIBZE	K-DIAVEIACEN	-----HDTC	--TTDMYS	FKGYVHRWMSQATHLAP	-F
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	PAMVMYERACE LV	-----DTC	--QVDQRA	FKGFLARWMAAATQVAP	-F
064	tr G0W5X4 G0W5X4_NAUDC	-NSIMYEVA CQPT	-----GTC	--NNDQRS	FKSYLARFLGLTAQLVT	-I
065	tr G8ZPC1 G8ZPC1_TORDC	-DSIMYESL CQGD	----GTSTC	--NSDQRS	FKAIFSRMLGLTSVLVP	-S
066	tr A0A167CDC5 A0A167CDC5_9ASCO	-DGVMFEAA CQNT	-----GRC	--NNDQRS	FKAIYSRFLGLTAQLAP	-P
067	tr R9XAT7 R9XAT7_ASHAC	-NDTLYEAA CQDT	-----NTC	--NVDQRS	FKAYFSRFLGLTAQLVP	-E
068	tr B2B747 B2B747_PODAN	D-NIAYEVA CEE	-----HMSC	--TTDMLS	FKGYVARWMATATQVAP	-F
069	tr G0SE99 G0SE99_CHATD	D-NVAYEVA CEE	-----HMSC	--TTDMLS	FKGYLHRWLATATEVAP	-F
070	tr J8Q6F0 J8Q6F0_SACAR	-DGIMYESA CQD	-----YSTC	--NTDQRT	FKSIFSRMLGLTSVMAP	-F
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	-NSIMYESTCQGS	-----GKC	--NTDQRV	FKAIFSRMLGYTAVLAP	-F
072	tr J3K6E2 J3K6E2_COCIM	QADIMVEYSCEPH	-----DNC	--LTDQRS	FKAFLSRWMAETTQLAP	-F
073	tr G4N3E1 G4N3E1_MAGO7	D-GAAFE LP CETT	-----PGGC	--TADMLS	FKGYVHRWMSVVTQVAP	-F
074	tr C5M5J2 C5M5J2_CANTT	-DKIMQETT CQP	-----YNLC	--NNDQRS	SFRSLFSRCLGLTMLLMP	-E
075	tr A0A254U0X0 A0A254U0X0_ASPNG	NGTVLS EVT CEP	-----ILSC	--DRNQLG	FKGYVAMWLAFTALLVP	-S
076	sp Q75DG6 DCW1_ASHGO	-NDTLYEAGCQGG	-----DNC	--NIDQRS	FKAYFSRFLGLTAQLVP	-E
077	tr Q6BZF0 Q6BZF0_DEBHA	--DIMYEAA CQGS	-----NNC	--NQDQRS	FKAYFSRLLGLTSVMVL	-D
078	tr E9E3Q1 E9E3Q1_METAQ	K-GIAFEVA CERD	---NGSGTC	--TPDMLS	YKGFLHRWLAVTSQIAP	-Y
079	sp P36091 DCW1_YEAST	-DGIVYEAA CQGP	-----NSC	--NTDQRS	FKAYFARFLGVTAQLVP	-E
080	tr W0TAH6 W0TAH6_KLUMD	-NNIMYEAA CQGS	-----GYC	--NNDQRS	FKAYFSRFLGLTAQLVP	-E
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	-DDIMYERACQGS	-----SSGC	--NTDQRS	FKSIFSRCLGQTAVMAP	-T
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	RENIVYEPA CEPF	-----GTC	--NNDQRS	FKTYLLRWMAQTAQLVS	-S
083	tr Q2TYU3 Q2TYU3_ASPOR	--YIMSEVL CEP	-----NEVC	--NDNEIL	FKGLVSGWLAFTALLVP	-S
084	tr G2XGF6 G2XGF6_VERDV	QNGVMFEQRC EPF	-----KQC	--NIDQSS	FKGYLARWMAGTAQVVP	-D
085	tr G3AMT4 G3AMT4_SPAPN	GSMVMYEAA CQPS	-----GTC	--NNDQRS	FKAYFSRFLGLTSVLVP	-D
086	tr A0A1C1D213 A0A1C1D213_9EURO	--NIMKE TACEAV	-NADGSDTC	--NVDQRS	FKAYLARWIAATIVRAP	-F
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	A-GPMIELS CETP	----TVILC	--KTDMLS	FKGYTHRWLATTTQLAP	-F
088	tr A0A100I5A6 A0A100I5A6_ASPNG	NGTVLS EVT CEP	-----ILSC	--DRNQLG	FKGYVAMWLAFTALLVP	-S
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	-NKIMYEATCQD	-----SNSC	--NNDQRS	FKSIFSRMLS LTKILAP	-F

090	tr G2Q8A7 G2Q8A7_MYCTT	D-GVAYE	E	I	A	C	E	Q	-	-	-	-	-	-	-	G	M	T	C	-	-	T	A	D	M	L	S	F	K	G	Y	L	H	R	W	L	A	T	A	T	Q	V	A	P	-	I		
091	tr B8NPZ1 B8NPZ1_ASPFN	NGVVLS	E	V	A	C	E	P	-	-	-	-	-	-	-	-	I	L	T	C	-	-	D	R	N	Q	L	C	F	K	G	Y	V	A	M	W	L	A	F	T	A	I	L	V	P	-	S	
092	tr Q7SAB2 Q7SAB2_NEUCR	A-GPMI	E	L	S	C	E	T	P	-	-	-	-	-	-	T	V	I	L	C	-	-	K	T	D	M	L	S	F	K	G	Y	T	H	R	W	L	A	T	T	T	Q	L	A	P	-	F	
093	tr A2R8R5 A2R8R5_ASPNC	G-EILT	E	I	L	C	E	P	-	-	-	-	-	-	-	-	T	E	V	C	-	-	N	D	N	E	I	I	F	K	G	L	V	S	A	W	L	A	Y	T	A	L	L	V	P	-	S	
094	tr C5DHG0 C5DHG0_LACTC	-DKVMY	E	S	T	C	Q	D	-	-	-	-	-	-	-	-	S	K	T	C	-	-	N	Q	D	Q	R	S	F	K	S	I	F	S	R	M	L	S	L	T	R	S	L	A	P	-	Y	
095	tr G0SFA3 G0SFA3_CHATD	RTPVMH	E	Q	L	C	E	P	L	-	-	-	-	-	-	-	-	S	L	C	-	-	N	I	D	Q	R	S	F	K	G	Y	L	T	R	W	L	A	N	T	A	Q	L	A	P	-	F	
096	tr K1WJG5 K1WJG5_MARBU	--GVMF	E	A	A	C	E	T	V	-	-	N	N	N	-	-	A	G	A	C	-	-	N	T	D	Q	L	S	F	K	A	Y	F	S	R	W	L	A	A	T	A	K	L	I	P	-	A	
097	tr H2B005 H2B005_KAZAF	-NSIMY	E	A	A	C	Q	G	P	-	-	-	-	-	-	-	-	Q	T	C	-	-	N	T	D	Q	R	S	F	K	A	Y	F	S	R	F	L	G	A	T	A	Q	L	V	P	-	E	
098	tr C5DYD7 C5DYD7_ZYGRC	-DNVMY	E	S	G	C	Q	H	G	-	-	-	-	-	-	-	D	S	S	N	C	-	-	N	N	D	Q	R	S	F	K	S	I	F	S	R	M	L	G	L	T	S	V	L	V	P	-	S
099	tr G8YM23 G8YM23_PICSO	-NDTMY	E	A	A	C	Q	P	-	-	-	-	-	-	-	-	A	N	T	C	-	-	S	Q	D	Q	R	S	F	K	A	Y	F	S	R	F	L	G	L	T	A	V	M	A	P	-	E	
100	tr W3X8E3 W3X8E3_PESFW	N-DIAF	E	V	A	C	E	E	-	-	-	-	-	-	-	-	H	M	T	C	-	-	T	T	D	M	L	S	F	K	G	Y	V	H	R	W	M	S	T	I	T	Q	I	A	P	-	Y	
101	tr A0A254U3K7 A0A254U3K7_ASPNG	G-EILT	E	I	L	C	E	P	-	-	-	-	-	-	-	-	T	E	V	C	-	-	N	D	N	E	I	I	F	K	G	L	V	S	A	W	L	A	Y	T	A	L	L	V	P	-	S	
102	tr Q6CER8 Q6CER8_YARLI	QSNIMV	E	I	A	C	Q	Q	V	-	-	-	-	-	-	-	T	I	T	C	-	-	N	Q	D	Q	R	T	F	K	G	I	Y	S	S	L	L	G	Q	T	A	Q	M	M	P	-	S	
103	tr G2QT05 G2QT05_THITE	N-NIAY	E	I	A	C	E	E	-	-	-	-	-	-	-	-	Q	M	S	C	-	-	T	T	D	M	L	S	F	K	G	Y	L	H	R	W	M	A	T	A	T	Q	I	A	P	-	F	
104	sp Q5ACZ2 DFG5_CANAL	-NKIMQ	E	T	T	C	Q	P	-	-	-	-	-	-	-	-	Q	N	K	C	-	-	N	N	D	Q	R	S	F	R	C	L	F	S	R	C	L	G	L	T	T	Q	L	A	P	-	E	
105	tr G2WKU6 G2WKU6_YEASK	-DSIMY	E	S	A	C	Q	D	-	-	-	-	-	-	-	-	Y	G	T	C	-	-	N	T	D	Q	R	T	F	K	S	I	F	S	R	M	L	G	L	T	S	V	M	A	P	-	F	
106	tr A0A0C7N752 A0A0C7N752_9SACH	-DKIMY	E	V	T	C	Q	D	-	-	-	-	-	-	-	-	S	N	S	C	-	-	N	N	D	Q	R	S	F	K	S	I	F	S	R	M	L	S	L	T	S	I	L	A	P	-	Y	
107	tr W3WN68 W3WN68_PESFW	ENGIMY	E	P	P	C	E	S	-	-	-	-	-	-	-	-	-	T	S	C	-	-	N	T	D	Q	Q	A	F	K	G	H	L	A	R	W	M	A	Y	T	A	K	L	A	P	-	F	
108	tr G2QFS1 G2QFS1_MYCTT	D-GVAF	E	L	P	C	E	G	R	-	-	-	-	-	-	-	K	G	A	C	-	-	T	A	D	M	L	S	F	K	G	Y	V	H	R	W	M	A	V	V	T	K	L	V	P	-	D	
109	tr G3J9G4 G3J9G4_CORMM	--GIMY	E	P	A	C	E	T	-	-	-	-	-	-	-	-	V	H	T	C	-	-	D	Q	N	M	V	S	F	K	G	Y	L	I	R	F	L	A	Y	T	S	K	V	A	P	-	W	
110	tr Q2UJ03 Q2UJ03_ASPOR	--VLYE	R	A	C	E	P	I	-	-	-	-	-	-	-	-	-	N	T	C	-	-	G	V	D	Q	R	S	F	K	G	Y	L	A	R	W	M	A	A	S	A	Q	V	A	P	-	F	
111	tr G8YRT2 G8YRT2_PICSO	-NGVMY	E	A	S	C	E	P	-	-	-	-	-	-	-	-	-	D	R	C	-	-	N	N	D	Q	T	A	F	K	A	L	F	A	R	S	L	G	Q	T	A	V	L	I	P	-	E	
112	tr W3X554 W3X554_PESFW	DNGIMV	E	K	P	C	E	E	G	-	-	-	-	-	-	-	-	G	F	C	-	-	D	I	D	Q	Q	S	F	K	A	Y	L	A	R	W	L	A	G	T	S	Q	L	A	P	-	F	
113	tr Q1K7A8 Q1K7A8_NEUCR	DTKI	I	K	E	W	Y	C	E	S	G	F	S	D	R	G	H	P	Y	Q	C	-	-	N	I	D	Q	Q	T	F	K	G	Y	L	L	R	W	L	S	S	T	S	Q	V	A	P	-	Y
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	G-MVMS	E	I	A	C	E	P	-	-	-	-	-	-	-	-	N	M	K	C	-	-	D	R	N	Q	D	C	F	K	G	F	L	S	S	W	L	T	F	M	T	T	I	V	P	-	Y	
115	tr A7F3P0 A7F3P0_SCLS1	--NVML	E	V	A	C	E	T	T	-	-	-	-	-	-	-	-	T	V	G	C	-	-	D	T	D	E	Q	S	F	K	A	Y	L	S	R	W	M	A	A	T	T	K	I	A	P	-	F
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	EGGHLW	E	P	P	C	E	K	T	-	-	-	-	-	-	-	-	A	R	G	C	-	-	D	Q	N	Q	V	S	F	K	G	Y	V	I	R	F	L	A	F	T	A	Q	M	A	P	-	W
117	tr Q6CIP9 Q6CIP9_KLULA	-DDIMY	E	S	T	C	Q	A	S	-	-	-	-	-	-	-	-	N	K	C	-	-	N	T	D	Q	R	V	F	K	A	I	F	S	R	M	L	G	Y	T	S	V	L	A	P	-	F	
118	tr H2AMQ5 H2AMQ5_KAZAF	-NSIMC	E	A	A	C	Q	P	A	-	-	-	-	-	-	-	-	G	T	C	-	-	D	T	D	Q	R	S	F	K	A	Y	F	A	R	F	L	G	V	T	A	Q	L	V	P	-	S	
119	tr G8JMI3 G8JMI3_ERECY	-NNIMY	E	R	A	C	Q	D	-	-	-	-	-	-	-	-	-	N	K	I	C	-	-	N	N	D	Q	R	S	F	K	S	I	F	S	R	M	L	A	V	T	S	V	L	A	P	-	F
120	tr C5DGT1 C5DGT1_LACTC	-NSILY	E	R	T	C	Q	E	A	-	-	-	-	-	-	-	-	N	T	C	-	-	N	T	D	Q	R	S	F	K	A	Y	F	S	R	F	L	G	L	T	A	Q	L	V	P	-	A	
121	tr K1XJR4 K1XJR4_MARBU	N-DIAY	E	V	A	C	E	P	-	-	-	-	-	-	-	-	-	K	L	T	C	-	-	S	I	D	M	F	S	F	K	S	Y	V	M	R	W	L	A	A	T	S	M	V	A	P	-	F
122	tr A0A124BYC5 A0A124BYC5_ASPNG	GGEIMS	E	V	L	C	E	P	-	-	-	-	-	-	-	-	A	N	V	C	-	-	N	D	N	E	I	T	F	K	G	L	L	A	E	S	L	T	L	T	S	M	L	A	P	-	Y	
123	tr A7TLL2 A7TLL2_VANPO	-RTVMF	E	A	A	C	Q	G	V	-	-	-	-	-	-	-	-	H	T	C	-	-	T	T	D	Q	R	T	F	K	A	Y	F	S	R	F	L	G	M	T	A	Q	L	V	P	-	D	
124	tr Q0CB86 Q0CB86_ASPTN	G-NVMS	E	I	L	C	E	P	-	-	-	-	-	-	-	-	-	Q	E	L	C	-	-	N	N	N	E	I	L	F	K	G	L	V	T	S	W	L	G	F	V	A	M	I	V	P	-	S
125	tr K1XLH0 K1XLH0_MARBU	PEGIMY	E	-	P	C	E	S	V	-	-	-	-	-	-	-	-	K	S	C	-	-	N	V	D	Q	R	S	F	K	A	Y	F	S	R	Q	L	A	A	T	A	I	L	A	P	-	Y	
126	tr J7S438 J7S438_KAZNA	-TGIMY	E	A	A	C	Q	A	A	-	-	-	-	-	-	-	-	L	T	C	-	-	N	N	D	Q	R	S	F	K	A	Y	F	S	R	F	L	G	L	T	A	Q	L	V	P	-	E	
127	tr Q4WG09 Q4WG09_ASPFU	GGNVLS	E	V	A	C	E	P	-	-	-	-	-	-	-	-	-	I	M	T	C	-	-	D	R	N	Q	I	G	F	K	G	Y	T	A	M	W	L	A	H	T	A	I	L	V	P	-	S
128	tr H8WZ09 H8WZ09_CANO9	TNKVMQ	E	T	T	C	Q	P	-	-	-	-	-	-	-	-	-	S	K	K	C	-	-	N	N	D	Q	R	S	F	R	S	L	F	S	R	C	L	G	L	T	M	V	I	I	P	-	D
129	tr I1RL09 I1RL09_GIBZE	K-NIMW	E	V	P	C	E	G	R	-	-	-	-	-	-	-	-	K	G	A	C	-	-	S	T	D	M	L	S	F	K	G	Y	V	H	R	W	L	A	V	T	T	Q	V	A	P	-	F
130	tr Q4W985 Q4W985_ASPFU	PAMVMY	E	R	A	C	E	L	V	-	-	-	-	-	-	-	-	D	T	C	-	-	Q	V	D	Q	R	A	F	K	G	F	L	A	R	W	M	A	A	A	T	Q	V	A	P	-	F	
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	-DGIMY	E	S	A	C	Q	D	-	-	-	-	-	-	-	-	-	Y	G	T	C	-	-	N	T	D	Q	R	T	F	K	S	I	F	S	R	M	L	G	L	T	S	V	M	A	P	-	F
132	tr W6QCW3 W6QCW3_PENRF	PPDVMY	E	R	A	C	E	T	V	-	-	-	-	-	-	-	-	-	N	T	C	-	-	M	V	D	Q	R	S	F	K	G	Y	L	S	R	W	M	A	Q	T	A	Q	M	A	P	-	F
133	tr W3WMD3 W3WMD3_PESFW	D-GIAF	E	P	A	C	E	P	-	-	-	-	-	-	-	-	-	G	-	N	C	-	-	N	A	D	M	R	S	F	K	G	F	L	H	R	W	M	A	S	T	A	M	M	A	P	-	F
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	-NSILY	E	A	T	C	Q	P	Y	-	-	-	-	-	-	-	-	-	N	S	C	-	-	N	T	D	Q	R	S	F	K	A	Y	F	T	T	M	L	A	A	T	A	Q	L	I	P	-	D
135	tr I2GYH1 I2GYH1_TETBL	-DDIMY	E	S	A	C	Q	T	-	-																																						

159	tr Q5BGD7 Q5BGD7_EMENI	PGTIFSDVACEP-----LQTC--DRNMLNFKGWSSMWLMAAAIMVP-E
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	-NSVMYEAA CQGG-----NYC--NNDQRSFKAYFSRFLGLTAQLVP-E
161	tr G8BYN9 G8BYN9_TETPH	-NNIMYESA CQE-----AKTC--NTDQRSFKSIFSRMLGYTRVLAP-Y
162	tr F7VVT4 F7VVT4_SORMK	K-GVMVERACELS-----DRLLC--NVDQHSFKGYLLRSLATAALMAPDL
163	tr G8YB80 G8YB80_PICSO	-NDTMYEAA CQP-----ANSC--SQDQRSFKAYFARFLGLTAVMAP-E
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	-DGIMYERA CQDG-----DNC--NNDQRF FKGVFARCLGLTALLVP-D
165	tr G2WRS7 G2WRS7_VERDV	N-NIAYEVA CEK-----KMSC--TTDMLSFKGYVARWLSTMTQVAP-F
166	tr A0A124BVB5 A0A124BVB5_ASPNG	SKNVLYEAA CEMV-----GTC--QTDQFSFKAYLVRWMADTARLAP-F
167	tr A7EIG7 A7EIG7_SCLS1	--NIMMEVA CEN-----NGKC--NIDQQSFKAYLARWLAATTKMAP-F
168	tr C5M5U7 C5M5U7_CANTT	GSMVMYEAA CQPS-----GTC--NNDQRSFKAYFSRFLGLTSVLVP-Q
169	tr B9WAA8 B9WAA8_CANDC	-NKIMQETT CQP-----QNKC--NNDQRSFRCLFSRCLGLTTQLVP-E
170	tr F8MRU1 F8MRU1_NEUT8	K-GIAFELACEGR-----QGAC--TPDMVSFKGYVHRWMAMVTQIAP-F
171	tr A1DJS0 A1DJS0_NEOFI	G-NIMSEVL CEP-----NEVC--NNNEILFKGLVASWLAFTALLVP-S
172	tr B6HLM8 B6HLM8_PENRW	G-GVVTDWRCEA-----VGQCYKDANGPLFKGLTVSWLSDIALIIP-S
173	tr A0A117DWS0 A0A117DWS0_ASPNG	S-NVMS EVS CEP-----NMMC--DRNQDCFKGFLSSWLTFTTTIAP-Y
174	tr A0A124BXM1 A0A124BXM1_ASPNG	G-NIMSEIIL CEP-----AEVC--NDNEILFKGLVTGWLGLVALIMP-S
175	tr J3NZQ6 J3NZQ6_GAGT3	D-GAAL EAS CEPY-----GPTNC--TSDMVMFKGFVHRWMAQTLLAP-F
176	tr W0THH8 W0THH8_KLUMD	-NNIMYEST CQSE-----KQMT C--NTDQRVFKAIFSRMLGYTAVLAP-F
177	tr B2AEF2 B2AEF2_PODAN	D-DIAYEIP CEGR-----KGAC--TADMLSFKGYVHRWLSVVTKVAP-H
178	tr Q96TX1 Q96TX1_NEUCS	D-NIAFEPA CED-----QMSC--TTDMLSFKGYVHRWLSITTQIAP-Y
179	tr E9DYA6 E9DYA6_METAQ	H-DVAVEIS CEL-----NDGC--KTDMFTYKGFVHRWYATTTQIAP-F
180	tr B9WAI2 B9WAI2_CANDC	GSMVMYEAA CQPS-----NSC--NNDQRSFKAYFSRFLGLTSVLVP-Q
181	tr Q6C171 Q6C171_YARLI	QKNIMVEIA CQQK-----TITC--NSDQRTFKGIFSSLLGQTAQMVP-T
182	tr A6ZMV1 A6ZMV1_YEAS7	-DSIMYESA CQD-----YGTC--NTDQRTFKSIFSRMLGLTSVMAP-F
183	tr B6K7X7 B6K7X7_SCHJY	-GGVIYDPO CETS-----NSC--NTDQPSFKGYLARFMMYAMQLAP-Y
184	tr I1RSA2 I1RSA2_GIBZE	KNEIIYEQF CEPH-----KQC--SQDQQSFKGYLARWLAATTQLYP-E
185	tr F7VWZ9 F7VWZ9_SORMK	DTKIIKEWYCESGF TDRGHPYQC--NIDQQTFKGYLLRWLSSTSQVAP-F
186	tr A0A1B2J789 A0A1B2J789_PICPA	-QSVFFEAA CQGS-----GNC--NTDQRSFKAYLARFLGLTAQMVP-S
187	tr G0V8N4 G0V8N4_NAUCC	-NSILYEAA CQKS-----GTC--NNDQRSFKAYFLRFLGQTAQLVS-E
188	tr H8WXN8 H8WXN8_CANO9	GSSVMYEAA CQPS-----NTC--NNDQRSFKAYFSRFLALTAQLVP-E
189	tr A6ZZS0 A6ZZS0_YEAS7	-DGIVYEAA CQGP-----NSC--NTDQRSFKAYFARFLGVTAQLVP-E
190	tr G9MQD1 G9MQD1_HYPVG	E-GIAYEVA CEPR-----QGAC--STDMLSFKGYVHRWLSVVAQIAP-H
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	-NSVLYEVT CQAG-----KTC--NTDQRSFKAYYSRFLGLTAQLVP-A
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	GGNVLS EVA CEP-----IMTC--DRNQIGFKGYTAMWLAHTAILVP-S
193	tr A0A254UET1 A0A254UET1_ASPNG	SKNVLYEAA CETV-----GTC--QTDQFSFKAYLVRWMADTARLAP-F
194	tr A1D587 A1D587_NEOFI	G-MVMS EIA CEP-----NMMC--DRNQDCFKGFLSSWLTFMTTIVP-Y
195	tr Q1K7I4 Q1K7I4_NEUCR	D-NIAFEPA CED-----QMSC--TTDMLSFKGYVHRWLSITTQIAP-Y
196	tr G8JS88 G8JS88_ERECY	-NGVVTEVA CQEA-----GTC--NTDQRSFKAYFMRFLGLTAQLIP-E
197	tr A0A136J7D3 A0A136J7D3_9PEZI	EDGILVERACE TF-----GTC--NNDQQSFRGYLMRWFGATMQLAP-F
198	tr E9E4X8 E9E4X8_METAQ	K-GIAVEIS CEN-----AGTC--TTDMLTFKGFLHRWYSTITQLAP-Y
199	tr Q6CP42 Q6CP42_KLULA	-NSIMYEAA CQGS-----GYC--NNDQRSFKAYFSRFLGLTAQLVP-E
200	tr G0WHL4 G0WHL4_NAUDC	-NGIMFENAC QE-----YNTC--NNDQRSFKSIFSRMLGLTSVLAP-F
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	-DGIVYEAA CQGP-----NSC--NTDQRSFKAYFARFLGATAQLVP-E
202	tr F9WYX6 F9WYX6_ZYMTI	--NIIYEVA CEP-----SDNC--NIDQTGFRAQFVRSLATYRDMAP-A
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	D-DVAYEIA CET-----GNTC--TTDMLSFKGYLHRWLSTATRIAP-F
204	tr J3P147 J3P147_GAGT3	K-GAAFEIS CEHL-----DNQC--TQDMVMFKGFVARWMPVVVQLVP-E
205	tr I2H842 I2H842_TETBL	-NSVIYEAA CQPN-----KNC--NNDQRSFKAYFARFLGVTAQLLP-S
206	tr S6E3R3 S6E3R3_ZYGB2	-NDIMYESGC QHN-----D SSTC--NNDQRSFRSIFSRMLGLTSVLVP-S
207	tr E5AD94 E5AD94_LEPMJ	KGGVMVE-VCEAK-----TLC--NADQESFKAYLARWLGNAIQMAP-F
208	tr Q6FJM7 Q6FJM7_CANGA	-DDIMYESA CQP-----YKTC--NNDQRCFKSIFSRMLGFTSVLAP-F
209	tr A5DUW0 A5DUW0_LODEL	GSMVMYEAA CQPG-----YTC--NNDQRSFKAYFSRFLGLTSVLVP-E
210	tr G8BF46 G8BF46_CANPC	GSSVMYEAA CQPS-----NTC--NNDQRSFKAYFSRFLALTAQLVP-E
211	tr W3XAF6 W3XAF6_PESFW	TDGIMWE LPCEAT-----ATSC--NNDQRMFKGFLSRWMAGAAKLAP-T
212	tr B6H7I2 B6H7I2_PENRW	G-DVIEDVT CEP-----IKMC--NFNEILFKGLTSSWLAFTTLLAP-D
213	tr Q5ATF9 Q5ATF9_EMENI	GGKIFSEYL CEP-----KALC--NYNEILFKGIVSTWITFVGLIVP-E
214	tr A0A167CDD3 A0A167CDD3_9ASCO	-DGIMYEAA CQPS-----NQC--NTDQRCFKGIYSRFLGLTMLLVP-S
215	tr G0RXS3 G0RXS3_CHATD	E-GIMVERACE LP-----DRMQC--NVDQHSFKGYMHRALATVAVVAP-F
216	tr B8NX26 B8NX26_ASPFN	GGNTMSEVA CEP-----IMSC--DRNQIGFKGYLSMWLAFTAILVP-S
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	PPDVMT EVA CEGN-----GKC--NIDQRSFKAYLSRWMAMTIKLAP-Y
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	SENVLYEYAC ETI-----NTC--RTDQYSFKAYLARWMGDVAQLAP-W
219	tr A1CMM3 A1CMM3_ASPCL	-GNIMSEVA CEST-----GKC--NVDQRSFKAYLSRWMAATVQIAP-F
220	tr B6HU84 B6HU84_PENRW	G-QILSDIT CEG-----SGNC--DRNQITFKAYFTNWLGMLTITIVP-G
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	D-NIAFEPA CED-----QMSC--TTDMLSFKGYVHRWLSITTQIAP-Y
222	tr G2WHY6 G2WHY6_YEASK	-DGIVYEAA CQGP-----NSC--NTDQRSFKAYFARFLGVTAQLVP-E
223	tr G0W7G8 G0W7G8_NAUDC	-NSIMYEAA CQGP-----NSC--NTDQRSFKAYFSRFLGATAQLVP-E
224	tr G4NGV8 G4NGV8_MAGO7	QNQVLVE PGCE FV-----GTC--NNDQRSFKGYLTRWLGGTSQLAP-F
225	tr W3X855 W3X855_PESFW	NESVMYE PPCE PQ-----SNC--ATDSFSFKAYLVRWMAKT TQLMS-S
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	KEDIA YEPA CEG-----VKTC--TTDMLSFKGYLHRWWSASAQMAP-Y
227	tr J8Q2K7 J8Q2K7_SACAR	-DGIVYEAA CQGP-----NSC--NTDQRSFKAYFARFLGVTAQLVP-E
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	--SVMYEAA CQNS-----NNC--NQDQRSFKAFFSRFLGLTSVLIP-Q

229	tr J6EGN8 J6EGN8_SACK1	-DTIMYESA	CQD	-	-	-	-	-	-	YSTC	-	-	NTDQRTFKS	IFSR	RMLGLT	SVMAP	P	-	F																																		
230	tr A0A100ISD9 A0A100ISD9_ASPNG	-	-	-	VMYERA	CEPV	-	-	-	-	-	-	NTC	-	-	QVDQRS	FKGYLAR	WMAATTQ	MAP	-	F																																
231	tr G8YQC0 G8YQC0_PICSO	-	ND	TMYESA	CEP	-	-	-	-	-	-	-	DRC	-	-	NNDQTS	SFKSLFAR	SLAQTA	VLIP	P	-	E																															
232	tr F7VVP8 F7VVP8_SORMK	K	-	GIAFEL	PCETR	-	-	-	-	-	-	-	HGAC	-	-	TPDMLS	SFKGYVHR	WMATVAQ	VAP	P	-	F																															
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	-	DSIMYESA	CQD	-	-	-	-	-	-	-	-	YGTC	-	-	NTDQRTFKS	IFSR	RMLGLT	SVMAP	P	-	F																															
234	tr F8MBP4 F8MBP4_NEUT8	D	T	K	I	I	K	E	W	Y	C	E	S	G	F	S	N	N	G	H	P	Y	Q	C	-	-	N	I	D	Q	Q	T	F	K	G	Y	L	L	R	W	L	S	S	T	S	Q	V	A	P	-	Y		
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	G	G	E	I	M	S	E	I	L	C	E	P	-	-	-	-	-	-	-	-	T	E	V	C	-	-	N	D	D	E	I	I	F	K	G	L	L	A	E	S	L	T	F	T	S	M	V	A	P	-	Y	
236	tr A1CD27 A1CD27_ASPCL	G	G	N	V	L	S	E	V	A	C	E	P	-	-	-	-	-	-	-	-	I	M	T	C	-	-	D	R	N	Q	L	G	F	K	G	Y	V	A	M	W	L	A	Q	T	S	I	L	V	P	-	S	
237	tr A5DHF3 A5DHF3_PICGU	-	N	K	I	M	Q	E	T	Q	C	W	N	N	-	-	-	-	-	-	-	G	D	P	K	C	-	-	N	N	D	Q	R	S	F	R	S	L	F	S	R	S	I	Q	L	T	S	V	L	V	P	-	D
238	tr D4ATT7 D4ATT7_ARTBC	P	K	G	V	M	I	E	V	A	C	E	A	Q	-	-	-	-	-	-	-	-	G	N	C	-	-	N	N	D	Q	R	S	F	K	A	Y	M	A	R	W	M	A	A	T	M	I	I	A	P	-	D	
239	tr E2PT42 E2PT42_ASPNC	-	-	-	VMYERA	CEPV	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	S	T	C	-	-	Q	V	D	Q	R	S	F	K	G	Y	L	A	R	W	M	A	A	T	T	Q	M	A	P	-	F		
240	tr C7ZPE5 C7ZPE5_NECH7	D	-	G	I	A	V	E	I	A	C	E	N	-	-	-	-	-	-	-	-	H	G	T	C	-	-	T	T	D	M	L	S	F	K	G	Y	V	H	R	W	M	S	Q	V	T	H	L	A	P	-	I	
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	E	T	N	I	M	F	E	V	P	W	E	P	N	-	-	-	-	-	-	-	-	G	K	W	-	-	E	K	D	Q	R	S	F	K	A	Y	L	A	R	W	M	A	A	T	T	Q	V	A	P	-	F	
242	tr Q4WKP7 Q4WKP7_ASPFU	G	-	M	V	M	S	E	I	A	C	E	P	-	-	-	-	-	-	-	-	-	N	M	K	C	-	-	D	R	N	Q	D	C	F	K	G	F	L	S	S	W	L	T	F	M	T	T	I	V	P	-	Y
243	tr F8MXT4 F8MXT4_NEUT8	D	-	N	I	A	F	E	P	A	C	E	D	-	-	-	-	-	-	-	-	Q	M	S	C	-	-	T	T	D	M	L	S	F	K	G	Y	V	H	R	W	L	S	I	T	T	Q	I	A	P	-	H	
244	tr A0A177AJ74 A0A177AJ74_9PEZI	A	T	G	I	I	I	E	R	D	C	E	L	T	-	-	E	-	-	-	-	-	P	A	S	C	-	-	N	T	D	Q	K	S	F	K	A	Y	I	T	R	W	M	A	A	T	S	K	V	A	P	-	F
245	tr J7RQR5 J7RQR5_KAZNA	-	DSIMFESA	CQD	-	-																																															

001	tr A0A254U2J9 A0A254U2J9_ASPNG	TYD	----	SI	-LPKLQGS	SAEAA	-AAS	CSGMS	-----	-
002	sp Q6FLP9 DCW1_CANGA	TRQ	----	QI	-MTWLNT	SALAA	-AKS	CSGGT	-----	-
003	tr A1CCM5 A1CCM5_ASPCL	TYS	----	TI	-LPKLQGS	SAVAA	-AQS	CTGNG	-----	-
004	tr C1H190 C1H190_PARBA	SRP	----	LI	-MPKLRA	SAEAA	-AKQ	CSGP	-----	-
005	tr Q2UR85 Q2UR85_ASPOR	TQD	----	QI	-LPKIQAS	SAQAA	-AKQ	CSGGD	-----	-
006	tr A5DV30 A5DV30_LODEL	FKE	----	KI	-EPYLEAS	SAQAA	-AQS	CSGGT	-----	-
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	FAD	----	RA	-NKIIDAS	SAKGA	-ASA	CSGGS	-----	-
008	tr J3PGN0 J3PGN0_GAGT3	MAD	----	KI	-RPVLRK	SAEMA	-VKQ	CTGGA	-----	T
009	tr A0A124BXU6 A0A124BXU6_ASPNG	TYN	----	RI	-LPKLQGS	SAQGA	-AAT	CTGYG	-----	-
010	tr C5P4A1 C5P4A1_COCP7	TRE	----	LI	-MPKLRA	SAMAA	-AKA	CTGGN	-----	-
011	tr Q0CG55 Q0CG55_ASPTN	TFD	----	WV	-MPRLRV	SAKAA	-AET	CTGGS	-----	-
012	tr C4Y2X5 C4Y2X5_CLAL4	TYD	----	RI	-RRWLVD	SANAA	-AYS	CSGGS	-----	-
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	TYD	----	LV	-MPKLRA	SAKAA	-AET	CTGGE	-----	-
014	tr C7Z068 C7Z068_NECH7	LKE	----	KI	-LPVLQT	STEAA	-VKQ	CTGGK	-----	S
015	tr G8BXX2 G8BXX2_TETPH	TRD	----	NI	-MKWIDA	SAYGA	-AES	CSGGT	-----	-
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	TYD	----	HI	-RRWLVD	SANAA	-AWS	CVGGS	-----	-
017	tr A1CSC2 A1CSC2_ASPCL	TAD	----	EI	-VPRIQQ	SALAA	-AKQ	CSGGD	-----	-
018	sp Q05031 DFG5_YEAST	TRD	----	TI	-DDLIKT	SAEAA	-AKS	CNGGT	-----	-
019	tr H2AQJ6 H2AQJ6_KAZAF	TAS	----	TI	-DPLIKS	SAAAC	-ALS	CDGGT	-----	-
020	tr G2XFH9 G2XFH9_VERDV	TAS	----	KI	-LPVLQN	STQAA	-IRQ	CTGGA	-----	S
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	LRD	----	QI	-MGWINT	TAQAA	-AQS	CVGGY	-----	-
022	sp Q5AD78 DCW1_CANAL	TEP	----	VI	-TKWLVD	SANGA	-AGS	CSGGS	-----	-
023	tr B6H7E8 B6H7E8_PENRW	NVT	----	AEV	-MGKFKAS	SAVAA	-GQQ	CSGGS	-----	-
024	tr B6GZU2 B6GZU2_PENRW	TYD	----	TV	-MKRLKAS	SAKAA	-AKT	CTGGV	-----	-
025	tr E9E4W7 E9E4W7_METAQ	TAG	----	NI	-LGPLRKS	SGEAA	-AGQ	CTGGA	-----	S
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	TFD	----	QI	-MPKLKAS	SGQAA	-AKT	CTGGE	-----	-
027	tr G2R7G8 G2R7G8_THITE	TRE	----	QI	-VRVLR	SSTEGA	-VSS	CLADG	-----	T
028	tr G3JBY1 G3JBY1_CORMM	TAD	----	GI	-RALIQQA	AARDA	-AAA	CTGPVGGTGP	-----	VGGTFQGI
029	tr A5DNV5 A5DNV5_PICGU	TSD	----	TI	-MNWLEK	SAIAA	-AQS	CSGGT	-----	-
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	TYG	----	TI	-KPLLRA	SAEAA	-MSH	CNGGN	-----	-
031	tr Q2TWC1 Q2TWC1_ASPOR	TAE	----	RI	-TPKLQGS	SAEAI	-AKQ	CSGES	-----	-
032	tr J5RXY9 J5RXY9_SACK1	TRN	----	QI	-MTWLNT	SAIAA	-AKS	CSGGT	-----	-
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	TAD	----	SI	-TKLISTA	AAKDA	-VKV	CNGAN	-----	GPKFPGP
034	tr A0A254U5V1 A0A254U5V1_ASPNG	TAD	----	QI	-LPKIQQ	SALAA	-AKQ	CSGGD	-----	-
035	tr A0A061B6H7 A0A061B6H7_CYBFA	TYE	----	TI	-YSLIEAS	SARAA	-SNS	CSGGS	-----	-
036	tr G8ZQ93 G8ZQ93_TORDC	ARD	----	QA	-MKWINAS	SALGA	-AQS	CSGGY	-----	-
037	tr Q9C2J1 Q9C2J1_NEUCS	TYE	----	RI	-NPWIRA	TAAAA	-VAT	CTGPVGAAAPQVDSGGIQPGFKGI	-----	-
038	tr C5NZK5 C5NZK5_COCP7	TRD	----	RL	-LPKLQAS	SATAA	-ALQ	CSGP	-----	-
039	tr Q6C0T7 Q6C0T7_YARLI	LAP	----	EI	-MKYLVPS	SAMAG	-ART	CSGGR	-----	-
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	TRQ	----	QI	-MTWLNT	SALAA	-AKS	CSGGT	-----	-
041	tr F7VZ72 F7VZ72_SORMK	TAP	----	II	-LPVLKV	SAEAA	-VQT	CTGEA	-----	S
042	tr A3LN37 A3LN37_PICST	TYD	----	GI	-RQWLVD	SANAA	AKNS	CTGGT	-----	-
043	tr Q6CAI2 Q6CAI2_YARLI	IRD	----	DI	-MKKLRA	SATAA	-LAT	CSGGS		

046	tr G9MHI4 G9MHI4_HYPVG	TVQ-----TL-MPLITGSSASAA-AAICTGPA-----SAAFKGI
047	tr D4AP27 D4AP27_ARTBC	STD-----FI-MRRLRACARGA-AKACTGGE-----
048	tr B6GZT8 B6GZT8_PENRW	TFD-----SI-LAKLQKSGQAA-AASCTGHN-----
049	sp O74556 YCZ2_SCHPO	TAP-----LL-EPLLISTAKAA-AGACCGGY-----
050	tr A0A0C4E991 A0A0C4E991_MAGP6	IAP-----KI-RPVLRKSAEMA-VKQCTGGP-----T
051	tr B8MHG0 B8MHG0_TALSN	TYS-----TI-LPKLQGSAVGA-GAQCSGPN-----
052	tr A3LMV8 A3LMV8_PICST	TQE-----VL-GPLIEQSAKGA-AASCSGGA-----
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	YSE-----RV-LALIDS SAAGA-AASCSGGS-----
054	sp Q9P6I3 YHG7_SCHPO	TVE-----TI-LPYIQSSAEAA-ALACSGGS-----
055	tr Q2US57 Q2US57_ASPOR	TYS-----TI-IPKLQGSGVAA-AETCTGHN-----
056	tr W7N622 W7N622_GIBM7	TSE-----YI-TATLLSTAKKA-AVSCSGSP-----SSGFAGQ
057	sp Q9P6I4 YHG6_SCHPO	TYD-----TL-YPLIQTSAAAA-AKQCCGGY-----
058	tr C7GRB3 C7GRB3_YEAS2	TRD-----TI-DDLIKTSAEAA-AKSCNNGT-----
059	tr C7GP28 C7GP28_YEAS2	TRN-----QI-MSWLNTSAIAA-AKSCSGGT-----
060	tr G3JGZ5 G3JGZ5_CORMM	TAD-----KI-LPALAKSAKAA-ANQCTGRE-----S
061	tr C4QXV4 C4QXV4_KOMPG	TAE-----TI-MNWMNTSAVAV-AQSCSGGT-----
062	tr I1S0Z5 I1S0Z5_GIBZE	IRP-----KI-LPVLEKSAQAA-IAQCTGGD-----T
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	TYD-----WV-MPRLRASAAAA-ARTCTGGP-----
064	tr G0W5X4 G0W5X4_NAUDC	TQS-----RI-MNWLNTSAIAV-AHSCSGGT-----
065	tr G8ZPC1 G8ZPC1_TORDC	TQS-----KV-DDLLKQSAALAA-AASCSGGT-----
066	tr A0A167CDC5 A0A167CDC5_9ASCO	MAD-----QI-MNLLSSSAAAA-AISCSGGT-----
067	tr R9XAT7 R9XAT7_ASHAC	SRD-----TV-MRWLRGSAMGA-AASCSGGT-----
068	tr B2B747 B2B747_PODAN	LAP-----KV-LPVLQNSAKAA-IASCVGEK-----N
069	tr G0SE99 G0SE99_CHATD	IRD-----TV-LPVLRTSAEAA-VSTCTGEE-----N
070	tr J8Q6F0 J8Q6F0_SACAR	SSD-----TI-DDLIKTSAEAA-AKSCDGGT-----
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	TED-----SI-TPLVDA SAAAA-AVSCSGGS-----
072	tr J3K6E2 J3K6E2_COCIM	TRE-----LI-MPKLRA SAMAA-AKACTGGN-----
073	tr G4N3E1 G4N3E1_MAGO7	TAA-----TV-IPALKKSAEAC-IKQCTGGP-----T
074	tr C5M5J2 C5M5J2_CANTT	SRD-----TI-LPYMEAS AAGA-AESCSGGS-----
075	tr A0A254U0X0 A0A254U0X0_ASPNG	THD-----LI-LPKLQGSAAAI-SKQCSGTG-----
076	sp Q75DG6 DCW1_ASHGO	SRE-----TI-VRWIRAS AQGA-AASCSGGR-----
077	tr Q6BZF0 Q6BZF0_DEBHA	TEP-----VI-TEWLVAS ANAAAKNSCTGGS-----
078	tr E9E3Q1 E9E3Q1_METAQ	TKD-----KI-LPVLRKSTEAA-IKQCTGGP-----T
079	sp P36091 DCW1_YEAST	TRN-----QI-MSWLNTSAIAA-AKSCSGGT-----
080	tr W0TAH6 W0TAH6_KLUMD	TRD-----DI-MKLLKASAKGA-AQSCSGGT-----
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	TYN-----TI-YTWLVAS AEGA-AKSCCTGGY-----
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	TAS-----TI-QPLLLASGKAA-AATCTGTA-----DGFKGI
083	tr Q2TYU3 Q2TYU3_ASPOR	TYD-----EI-LPKLQAS AQGA-AASCSGMS-----
084	tr G2XGF6 G2XGF6_VERDV	LFD-----RI-MGLLRPNAQSV-ASVCVGD P-----DQ-GFPGI
085	tr G3AMT4 G3AMT4_SPAPN	TEP-----II-TKWLVD SANSA-AGSCSGGT-----
086	tr A0A1C1D213 A0A1C1D213_9EURO	TYP-----LL-KPILEQSA AAAA-AATCTGGV-----
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	TRD-----AI-EKALRNSTAAA-VDSCRGPL-----H
088	tr A0A100I5A6 A0A100I5A6_ASPNG	THD-----II-LPKLQGSAAAI-SKQCSGTG-----
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	TAE-----KI-DPLLTAS AEGA-AKSCSGGT-----
090	tr G2Q8A7 G2Q8A7_MYCTT	LSE-----KI-LPVLKTSTEAA-VSTCTGEA-----N
091	tr B8NPZ1 B8NPZ1_ASPFN	TRE-----LI-TPKLQGSAAAI-SKQCSGGD-----
092	tr Q7SAB2 Q7SAB2_NEUCR	TRD-----AI-EKALRNSTAAA-VDSCRGPL-----H
093	tr A2R8R5 A2R8R5_ASPNC	TYD-----RI-LPKLQGS AQGA-AATCTGYG-----
094	tr C5DHG0 C5DHG0_LACTC	TAE-----TI-DPLMEAS AEGA-AKSCCTGGT-----
095	tr G0SFA3 G0SFA3_CHATD	TFQ-----TI-QPLLLADAAAA-AQACTGSVRTLV PNIPGARPEPPFRGE
096	tr K1WJG5 K1WJG5_MARBU	YHD-----TI-MPLLKTSAAAA-AAQCSGGT-----
097	tr H2B005 H2B005_KAZAF	TRD-----TV-MHWVNTSAVAA-ATSCSGGT-----
098	tr C5DYD7 C5DYD7_ZYGRC	VSD-----KV-DQLLNAS AEGA-AKSCSGGI-----
099	tr G8YM23 G8YM23_PICSO	TSD-----TI-MTWLTASAKAA-AQSCSGGT-----
100	tr W3X8E3 W3X8E3_PESFW	TAD-----KI-LPVLKTSATAG-IQQCTGGD-----N
101	tr A0A254U3K7 A0A254U3K7_ASPNG	TYD-----RI-LPKLQGS AQGA-AATCTGYG-----
102	tr Q6CER8 Q6CER8_YARLI	LAG-----SI-LSYLRPSAYAA-ARTCSGGF-----
103	tr G2QT05 G2QT05_THITE	ISA-----KV-LPVLKTSTAAA-VDTCTGGT-----N
104	sp Q5ACZ2 DFG5_CANAL	TKD-----RI-REVLEAS AEGA-AKSCSGGS-----
105	tr G2WKU6 G2WKU6_YEASK	TRD-----TI-DDLIKTSAEAA-AKSCNNGT-----
106	tr A0A0C7N752 A0A0C7N752_9SACH	TAD-----TI-NPLMEASAAAA-AKSCSGGS-----
107	tr W3WN68 W3WN68_PESFW	TYD-----TI-VPLLKSSATAA-AEQCSGSP-----TSGFKGH
108	tr G2QFS1 G2QFS1_MYCTT	TAG-----TI-LPVLRTSAEAA-ARQCTGGD-----T
109	tr G3J9G4 G3J9G4_CORMM	TSD-----SI-TKLISTAASDA-VKVCNGAN-----GASYPGP
110	tr Q2UJ03 Q2UJ03_ASPOR	ILD-----QV-MPKLRTSASAA-ARTCTGGS-----
111	tr G8YRT2 G8YRT2_PICSO	TSD-----DI-MKLLDKSASAA-GESCSGGT-----
112	tr W3X554 W3X554_PESFW	TFD-----TI-MPLLQSTATAA-AQQCNGSP-----AAALYKGP
113	tr Q1K7A8 Q1K7A8_NEUCR	TYE-----RI-NPWIRATAAAA-VATCTGPVGAAAPQVDSGGIQPGFKGI
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	TSS-----EV-VPRIQQSALAA-AKQCSGGQ-----
115	tr A7F3P0 A7F3P0_SCLS1	TEP-----TI-MTYINAS AQGA-AAQCSGGS-----

116	tr A0A2N6NK60 A0A2N6NK60_BEABA	TSD	----	GI	-KTILQQ	AAKDA	-AAAC	CIGPR	-----	NEKYQGA	-
117	tr Q6CIP9 Q6CIP9_KLULA	TTD	----	TI	-TPLIDTS	SAAAA	-AKS	CSGGT	-----		-
118	tr H2AMQ5 H2AMQ5_KAZAF	TRT	----	QI	-MTWMNA	SAYAV	-AES	CSGGT	-----		-
119	tr G8JMI3 G8JMI3_ERECY	TRE	----	TI	-DPLLEA	SAAAA	-AAS	CSGGK	-----		-
120	tr C5DGT1 C5DGT1_LACTC	ARN	----	QI	-MDWIRA	SAKGA	-AQSC	SGGT	-----		-
121	tr K1XJR4 K1XJR4_MARBU	TRD	----	RI	-MPKLRK	SAQAA	-AAQ	CTGGA	-----		T
122	tr A0A124BYC5 A0A124BYC5_ASPNG	TSS	----	DI	-LPRLQG	SAVGA	-AKQ	CSGGS	-----		-
123	tr A7TLL2 A7TLL2_VANPO	TQH	----	TI	-MTYMNA	SAYGA	-AQSC	SGGY	-----		-
124	tr Q0CB86 Q0CB86_ASPTN	TYD	----	QI	-LPKLQG	SAQSA	-AQT	CTGMG	-----		-
125	tr K1XLH0 K1XLH0_MARBU	THD	----	GI	-MKEIKT	SAMAA	-IQT	CTAGD	-----		-
126	tr J7S438 J7S438_KAZNA	TRT	----	QI	-MGWLNT	SAVAA	-GKS	CSGGY	-----		-
127	tr Q4WG09 Q4WG09_ASPFU	TAA	----	RI	-FPVLQG	SALAL	-SKQ	CSKAP	-----		-
128	tr H8WZ09 H8WZ09_CANO9	FED	----	MI	-RPYLET	SAEAA	-AQSC	SGGS	-----		-
129	tr I1RL09 I1RL09_GIBZE	TAK	----	KI	-LPVLKT	STEAA	-VKQ	CTGGD	-----		S
130	tr Q4W985 Q4W985_ASPFU	TYD	----	WV	-MPRLRA	SAAAA	-ART	CTGGP	-----		-
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	TSD	----	TI	-DALIKT	SAEAA	-AKS	CDGGR	-----		-
132	tr W6QCW3 W6QCW3_PENRF	TYD	----	IV	-MKRLKA	SAQAA	-AKT	CTGGI	-----		-
133	tr W3WMD3 W3WMD3_PESFW	TYD	----	TI	-MPVLRT	SVEAA	-VKQ	CTGGD	-----		N
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	LRD	----	QI	-MGWINT	TAQAA	-AQSC	VGGY	-----		-
135	tr I2GYH1 I2GYH1_TETBL	TRD	----	II	-DPLILA	SAQGA	-ART	CNGGT	-----		-
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	VSD	----	QI	-DTLLTA	SAAAA	-AKS	CSGGT	-----		-
137	tr Q752P3 Q752P3_ASHGO	TRG	----	TI	-DELIYA	SATAA	-SKS	CEQGT	-----		-
138	tr B8MYP3 B8MYP3_ASPFN	TQD	----	QI	-LPKIQAS	SAQAA	-AKQ	CSGGD	-----		-
139	tr Q2TXL6 Q2TXL6_ASPOR	TLD	----	QI	-LPKLQG	SAEGA	-AKQ	CSGPS	-----		-
140	tr Q0CN17 Q0CN17_ASPTN	TSG	----	EI	-LPKIQAS	SAQAA	-AKQ	CSGGS	-----		-
141	tr J3NHD5 J3NHD5_GAGT3	IHD	----	KT	-LAVLRK	SAEAA	-VAT	CTGGS	-----		S
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	TAD	----	QI	-SKFLLI	SAKAA	-AAQ	CTGGD	-----		-
143	tr A1CVB0 A1CVB0_NEOFI	TYD	----	WV	-MPRLRA	SAAAA	-ART	CTGGP	-----		-
144	tr B8NVI3 B8NVI3_ASPFN	TLD	----	QI	-LPKLQG	SAEGA	-AKQ	CSGPS	-----		-
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	ARD	----	KI	-MSWIHA	SAKGA	-AQSC	SGGT	-----		-
146	tr A1DJ54 A1DJ54_NEOFI	TAA	----	RI	-FPVLQG	SALAI	-SKQ	CSKAP	-----		-
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	TRD	----	TI	-LPVLKT	SAEAA	-AKQ	CTGGA	-----		T
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	TYD	----	DI	-YSLLDAS	SARAA	-SNS	CSGGT	-----		-
149	tr A0A177A618 A0A177A618_9PEZI	LAP	----	KI	-MGLLQS	SAVAA	-ANQ	CIPGP	-----		-
150	tr Q75CW6 Q75CW6_ASHGO	TKA	----	RI	-EPNIRK	SAEAA	-AKS	CSGGR	-----		-
151	tr Q7S4K4 Q7S4K4_NEUCR	TRD	----	TI	-LPVLKT	SAEAA	-AKQ	CTGGA	-----		T
152	tr G2QB99 G2QB99_MYCTT	TLD	----	AV	-RPLLEAD	DAAAA	-ARACT	GD	-----	APPVFRGH	-
153	tr G0S3F2 G0S3F2_CHATD	TKD	----	RI	-LPVLRT	SAEAA	-VKQ	CVGPP	-----		T
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	TEA	----	VI	-RPWLVN	AANAA	-ANS	CSGGT	-----		-
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	TSD	----	TI	-DPLLKA	SAMAA	-AGS	CDGGT	-----		-
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	TYD	----	RI	-LPKLQG	SAVAA	-GAT	CTGNG	-----		-
157	tr G2QKJ0 G2QKJ0_MYCTT	TYE	----	TI	-TKTLRS	STEGC	-VSS	CLADG	-----		T
158	tr Q4WFX5 Q4WFX5_ASPFU	TRD	----	TI	-ATRLRA	SATAA	-AAQ	CVGGK	-----		-
159	tr Q5BGD7 Q5BGD7_EMENI	LRE	----	KI	-TPKLQG	SALAI	-GRS	CDGSS	-----		-
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	TRD	----	NI	-LTLLKA	SALGA	-AEAC	SGGT	-----		-
161	tr G8BYN9 G8BYN9_TETPH	TAA	----	TL	-DTLIETS	SAKAA	-AGS	CDGGT	-----		-
162	tr F7VVT4 F7VVT4_SORMK	VRE	----	PI	-IKTFKT	NIEGV	-IDS	CLADG	-----		T
163	tr G8YB80 G8YB80_PICSO	TSD	----	TI	-MTWLTA	SAKAA	-AAS	CSGGT	-----		-
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	LYD	----	DI	-MPLLDTS	SASAA	-AQSC	VGGI	-----		-
165	tr G2WRS7 G2WRS7_VERDV	TAD	----	TI	-LPVLRN	SAEKA	-IAQ	CTGGP	-----		E
166	tr A0A124BVB5 A0A124BVB5_ASPNG	TYE	----	TI	-MARLQP	TAKAA	-AAQ	CVGGK	-----		-
167	tr A7EIG7 A7EIG7_SCLS1	TYD	----	AV	-MAYLRP	SAAGA	-AAS	CVGGD	-----		-
168	tr C5M5U7 C5M5U7_CANTT	TEP	----	VI	-RKWLVD	SANAA	-AGS	CSGGS	-----		-
169	tr B9WAA8 B9WAA8_CANDC	TKD	----	KI	-REVLEAS	SAEGA	-AKS	CSGGS	-----		-
170	tr F8MRU1 F8MRU1_NEUT8	TRD	----	TI	-LPVLKT	SAEAA	-AKQ	CTGGS	-----		T
171	tr A1DJS0 A1DJS0_NEOFI	TYD	----	RI	-LPKLQG	SAVAA	-GAT	CTGNG	-----		-
172	tr B6HLM8 B6HLM8_PENRW	LKE	----	KI	-LPRLQV	SAEGA	-AKS	CTGDG	-----		-
173	tr A0A117DWS0 A0A117DWS0_ASPNG	TSD	----	QI	-IPKIQQ	SALAA	-AKQ	CSGGD	-----		-
174	tr A0A124BXM1 A0A124BXM1_ASPNG	TSS	----	LI	-LPKLQG	SAEAA	-AAS	CNGMG	-----		-
175	tr J3NZQ6 J3NZQ6_GAGT3	VKD	----	EI	-RPVLRK	SAAQA	-LKT	CTGAP	-----		T
176	tr W0THH8 W0THH8_KLUMD	TLD	----	TV	-NPLIEAS	SATAA	-AKS	CSGGT	-----		-
177	tr B2AEF2 B2AEF2_PODAN	TRD	----	KI	-LPVLRT	STEAA	-VKQ	CTGGD	-----		T
178	tr Q96TX1 Q96TX1_NEUCS	TAP	----	VI	-LPVLKT	SAEAA	-VQT	CTGEQ	-----		N
179	tr E9DYA6 E9DYA6_METAQ	TAE	----	RI	-LPVLQK	SAQAA	-VTQ	CTGGA	-----		N
180	tr B9WAI2 B9WAI2_CANDC	TEP	----	VI	-TKWLVD	SANAA	-AGS	CTGGS	-----		-
181	tr Q6C171 Q6C171_YARLI	LAS	----	DI	-MWYLEAS	SAYAA	-ART	CSGGR	-----		-
182	tr A6ZMV1 A6ZMV1_YEAS7	TRD	----	TI	-DDLIKT	SAEAA	-AKS	CNGGT	-----		-
183	tr B6K7X7 B6K7X7_SCHJY	TYD	----	TI	-MPLMKT	SATAA	-ALAC	SGGS	-----		-
184	tr I1RSA2 I1RSA2_GIBZE	TSA	----	RI	-IPLLKT	SAQAA	-AKV	CTGS	-----	GDGYKGP	-

185	tr F7VWZ9 F7VWZ9_SORMK	TYE-----RI-NPWIRST	TAIAA-AAVCTG	FPV	VGAAAPQVDSGGIQPGFRGI
186	tr A0A1B2J789 A0A1B2J789_PICPA	TAE-----TI-MNWMNTS	SAVAV-AQSCS	GGT	
187	tr G0V8N4 G0V8N4_NAUCC	TQP-----TI-MKWLNAS	SA YAA-AQSCS	GGT	
188	tr H8WXN8 H8WXN8_CANO9	TKG-----TI-YKWLVDS	SANAA-AGACSG	GGF	
189	tr A6ZZS0 A6ZZS0_YEAS7	TRN-----QI-MSWLNTS	SAIAA-AKSCS	GGT	
190	tr G9MQD1 G9MQD1_HYPVG	TKD-----KI-LPILRKST	EAA-VKQCT	GGD	S
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	ARE-----KI-MDWIRTS	SAQGA-AQSCS	GGY	
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	TAA-----RI-FPVLQGS	SALAL-SKQCS	KAP	
193	tr A0A254UET1 A0A254UET1_ASPNG	TYD-----TI-MARLQPT	AKAA-AAQCV	GGK	
194	tr A1D587 A1D587_NEOFI	TSS-----EI-VPRIQQS	SALAA-AKQCS	GGS	
195	tr Q1K7I4 Q1K7I4_NEUCR	TAP-----VI-LPVLKTS	SAEAA-VQTCT	GEQ	N
196	tr G8JS88 G8JS88_ERECY	SRE-----IA-MKWIRD	SAKGA-ALSCS	GGR	
197	tr A0A136J7D3 A0A136J7D3_9PEZI	TYD-----TL-MPILRTN	AAAA-ARQCT	GSP	PASKYRGP
198	tr E9E4X8 E9E4X8_METAQ	TTE-----TI-RPVLQTS	SAEAA-VKQCT	GAA	L
199	tr Q6CP42 Q6CP42_KLULA	TRD-----TV-LALLQAS	SAQGA-ADSCS	GGT	
200	tr G0WHL4 G0WHL4_NAUDC	TSD-----TV-DPLIKTS	SAAAA-AKSCT	GGT	
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	TRN-----QI-MSWLNTS	SAIAA-AKSCS	GGT	
202	tr F9WYX6 F9WYX6_ZYMTI	SDDSVYRQNI	-TAVLRA	SATAA-AAACV	YGE
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	IHD-----KT-LAVLRKS	STAAA-VETCT	GGD	N
204	tr J3P147 J3P147_GAGT3	MRG-----TI-MPVMRKS	SEAG-LRQCT	GGP	R
205	tr I2H842 I2H842_TETBL	TRD-----TI-MQWIDTS	SA YAA-AQACSG	GGT	
206	tr S6E3R3 S6E3R3_ZYGB2	VSD-----QI-DTLLTAS	SAAAA-AKSCS	GGT	
207	tr E5AD94 E5AD94_LEPMJ	THD-----LI-MPKLQVS	SAKAA-AQTCE	GPSQHN	G
208	tr Q6FJM7 Q6FJM7_CANGA	TSD-----TI-DPLLKAS	SAMAA-AGSCD	GGT	
209	tr A5DUW0 A5DUW0_LODEL	TEP-----TI-NKWLVDS	SANGA-AGSCS	GGS	
210	tr G8BF46 G8BF46_CANPC	TSG-----II-YKWLVDS	SANAA-AGACSG	GGI	
211	tr W3XAF6 W3XAF6_PESFW	VYD-----TI-MPLLQTS	TAVAA-AEHCVS	-H	TSGFKGH
212	tr B6H7I2 B6H7I2_PENRW	TAA-----QI-KPKLAS	SAEAA-AKSCT	GNN	
213	tr Q5ATF9 Q5ATF9_EMENI	TYD-----RI-FAKLQTS	SAQAA-ALSCS	GAG	
214	tr A0A167CDD3 A0A167CDD3_9ASCO	LQD-----EI-MPLIQS	SAKAA-AASC	SGGT	
215	tr G0RXS3 G0RXS3_CHATD	TRD-----TI-MRTLRS	STEGC-VSSCL	ADG	T
216	tr B8NX26 B8NX26_ASPFN	TKD-----QI-VPKLQGS	SVESI-SKMCN	GQS	
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	TRD-----RL-LPKLQAS	SATAA-ALQCS	GSP	
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	TRD-----TI-ATRLRA	SATAA-AAQCV	GGK	
219	tr A1CMM3 A1CMM3_ASPCL	TYD-----LL-MPKMRA	SAEAA-ALQCS	GSP	
220	tr B6HU84 B6HU84_PENRW	TYD-----LI-YPQLKTS	SAQAA-AKQCS	GGD	
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	TAP-----VI-LPVLKTS	SAEAA-VQTCT	GEQ	N
222	tr G2WHY6 G2WHY6_YEASK	TRN-----QI-MSWLNTS	SAIAA-AKSCS	GGT	
223	tr G0W7G8 G0W7G8_NAUDC	TRS-----KI-LPLINAS	SAIAA-GKSCS	GGT	
224	tr G4NGV8 G4NGV8_MAGO7	TFN-----TI-QPILRN	NAEAA-AVACSG	GSP	AA-GFSGK
225	tr W3X855 W3X855_PESFW	TYD-----TI-YPLLEAS	SAQGA-AKQCD	G TG	EGTAGRTL
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	IAD-----KV-LPVLRKST	AAC-VKQCT	GGE	G
227	tr J8Q2K7 J8Q2K7_SACAR	TRK-----QI-MSWLNTS	SAIAA-AKSCS	GGT	
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	TYD-----II-HSWLVDS	SANAA-ASSCS	GGL	
229	tr J6EGN8 J6EGN8_SACK1	TSD-----TI-DDLIKTS	SAEAA-AKSCN	G GT	
230	tr A0A100ISD9 A0A100ISD9_ASPNG	TYD-----LV-MPKLRA	SAKAA-AETCT	GGE	
231	tr G8YQC0 G8YQC0_PICSO	TSD-----TI-MNLLDKS	SASAA-GESCS	GGT	
232	tr F7VVP8 F7VVP8_SORMK	MKD-----TI-LPVLKTS	SAEAA-AKQCT	GGA	T
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	TRD-----TI-DDLIKTS	SAEAA-AKSCN	G GT	
234	tr F8MBP4 F8MBP4_NEUT8	TYE-----RI-NPWIRAT	AAAA-VATCT	GPV	VGAAAPQVDSGGIQPGFKGI
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	TSS-----DI-LPRLQGS	SAVGA-AKQCS	GGS	
236	tr A1CD27 A1CD27_ASPCL	TAD-----RI-YPVLQGS	SALAI-SKQCS	-AP	
237	tr A5DHF3 A5DHF3_PICGU	MYD-----EM-RPYVEAS	AE GA-AKSCS	GGS	
238	tr D4ATT7 D4ATT7_ARTBC	TIE-----KI-NPLLRES	SAKAA-ALQCN	G GP	
239	tr E2PT42 E2PT42_ASPNC	TYD-----LV-MPKLRA	SAKAA-AETCT	GGE	
240	tr C7ZPE5 C7ZPE5_NECH7	VSD-----KI-LPVLQTS	SAEAA-IKQCT	GGT	T
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	TAE-----QL-MPKLQAS	SALAA-AKVCT	GST	
242	tr Q4WKP7 Q4WKP7_ASPFU	TSS-----EV-VPRIQQS	SALAA-AKQCS	GGQ	
243	tr F8MXT4 F8MXT4_NEUT8	TAP-----VI-LPVLKTS	SAEAA-VQTCT	GEQ	N
244	tr A0A177AJ74 A0A177AJ74_9PEZI	TAD-----TI-SKFL LTS	SAKAA-AAQCT	G GV	
245	tr J7RQR5 J7RQR5_KAZNA	TAS-----TI-DPLMQS	SAQAA-AQSCS	GGT	
246	tr Q75CW7 Q75CW7_ASHGO	TKA-----RI-EPNIRKS	SAEAA-AKSCS	GGR	
247	tr G3AKK4 G3AKK4_SPAPN	TES-----VI-APYLEAS	SA AAA-AQSCS	GGS	
248	tr B2W762 B2W762_PYRTR	TRP-----QI-DPLLQTS	SAKAA-AKQCN	AGV	
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	TRN-----QI-MSWLNTS	SAIAA-AKSCS	GGT	
250	tr C4XWK1 C4XWK1_CLAL4	HFD-----QIYN	NWIVP	SAKAA-AASC	SGGS

002	sp Q6FLP9 DCW1_CANGA	D--GHTCG-LNWFR--D-----DWDG-- -- --MY--GLGEQMAALEVMVNTQ
003	tr A1CCM5 A1CCM5_ASPCL	N---SSCG-VRWHT--S-----SWDG-- -- --Q--TGMEEQISVTDVLSVNL
004	tr C1H190 C1H190_PARBA	---DNSCG-LRWTK--Q---AEFDG-- -- --ET--GVGEQMSALEVIQANL
005	tr Q2UR85 Q2UR85_ASPOR	S--KTD CG-RSWYK-QD-----KWDG-- -- --S-KSLES DMSALS SVLSSTM
006	tr A5DV30 A5DV30_LODEL	D--GMTCG-EDWST--S-----GWDG-- -- --VY--GLPEQISALEVIMSLV
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	D--GHTCG-MSWSD--G-----DWDG-- -- --FY--GLGEQIAALEIIQNSV
008	tr J3PGN0 J3PGN0_GAGT3	---GRACG-FYWTG--G-----EYIDPAID-KTS--GAGERMNVLA AVSSLL
009	tr A0A124BXU6 A0A124BXU6_ASPNG	N---NTCG-VRWWN--S-----TWDG-- -- --W-SGLEEQMSVTSVFSSTM
010	tr C5P4A1 C5P4A1_COCP7	D--GKSCG-LKWST--G-----GFDG-- -- --SV--GIGEQMAALEVIQSNL
011	tr Q0CG55 Q0CG55_ASPTN	E--HGMCA-LKWTE--R-----KWDG-- -- --ME--DVGLQIGALEVIQSTL
012	tr C4Y2X5 C4Y2X5_CLAL4	D--GHTCG-LSWTN--G-----TWDG-- -- --NY--GLGEQMSALEVMNNLR
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	L--QATCG-LKWTD--R-----KWDG-- -- --MD--DVGIQMAALEVMQSTL
014	tr C7Z068 C7Z068_NECH7	---GRACG-FYWSD--G-----EFVDPAVD-ETS--GAGEQMNVLA AVSSLL
015	tr G8BXX2 G8BXX2_TETPH	D--GHTCG-INW FY--G-----GWDG-- -- --KY--GLGEQMAALEILVNTR
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	D--GHTCG-LSWTN--G-----TWDG-- -- --VY--GLGEQMNALEVIQNLL
017	tr A1CSC2 A1CSC2_ASPCL	S--HTQCG-RRWHQ--D-----TWDG-- -- --T-TSLEEDMSALS SVFSSTM
018	sp Q05031 DFG5_YEAST	D--GHTCG-LNWQK--Q-----TNDG-- -- --YY--GLGEQMSALEVIQNLL
019	tr H2AQJ6 H2AQJ6_KAZAF	D--GHTCG-LNWQD--K-----TCDG-- -- --NY--GLGEQMSALEVIQNLL
020	tr G2XFH9 G2XFH9_VERDV	---GRVCG-FYWDR--G-----EFIDPAVD-KTT--GAGEQMNVLA AVSSLL
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	D--GHTCG-LNWQA--K-----GWDG-- -- --KY--GLGEQMSALEALVVTQ
022	sp Q5AD78 DCW1_CANAL	D--GVTCG-LSWTDWSQ-----GWDG-- -- --KW--GLGEQMSALEVMQNLM
023	tr B6H7E8 B6H7E8_PENRW	D--GKHCG-IRWTM-KS-----EWDG-- -- --T-MGLEQQMSV LGVLNAV M
024	tr B6GZU2 B6GZU2_PENRW	N--KNVCG-LKWTE--Q-----KWDK-- -- --TK--DFGQQMAALEVIQANL
025	tr E9E4W7 E9E4W7_METAQ	---GRQCG-FYWTE--G-----KFIDPRVD-KTS--GAGEAMDVLA AVSSML
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	L--GTTCG-LKWTD--Q-----KWDH-- -- --TK--DFGQQMAALEVVQANL
027	tr G2R7G8 G2R7G8_THITE	-----CG-FRWNT--G-----SYDGDNV-- --VG--PAGQEMSALAALSTLL
028	tr G3JBY1 G3JBY1_CORMM	D--GTACG-FSWLE--P-----KYDG-- -- --LA--GVGPQMNALAAVFYNL
029	tr A5DNV5 A5DNV5_PICGU	D--GHTCG-INWFL--G-----HWDG-- -- --VY--GLGEQMCAL EVITALR
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	N--GRTC CG-LKWSN--W---GNWDG-- -- --SN--GIGQQMAALEVLQSNL
031	tr Q2TWC1 Q2TWC1_ASPOR	---ENL CG-ETW GK--D-----TWDG-- -- --M-KGLEVQMAALGGITSNL
032	tr J5RXY9 J5RXY9_SACK1	D--GHTCG-LNWFN--G-----TWDG-- -- --MY--GLGEQMAALEVMVNTR
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	D--GTGCG-FSWLN--G-----TNDG-- -- --LM--GVGPQMDALA AVFYNL
034	tr A0A254U5V1 A0A254U5V1_ASPNG	S--GTQCG-RRWYQ--A-----QWDG-- -- --E-TSLES DMSALS SVFSSTM
035	tr A0A061B6H7 A0A061B6H7_CYBFA	D--GHTCG-MNWWH--S-----GWDG-- -- --FY--GLGEQMSALEIMQNLR
036	tr G8ZQ93 G8ZQ93_TORDC	D--GHTCG-INW FY--D-----GWDG-- -- --MY--GLGEQMSALEALVNTQ
037	tr Q9C2J1 Q9C2J1_NEUCS	D--GTACG-FKWTQ-- -- --TFDG-- -- --WA--GVGAQMNALSAVMYTL
038	tr C5NZK5 C5NZK5_COCP7	---DNACG-LRWTK--G---EAYDG-- -- --ST--GVGEQMAALEIIQSNL
039	tr Q6C0T7 Q6C0T7_YARLI	D--GHTCS-LDWLT--G-----KYDD-- -- --QYIGLGEQLSVLGAILNTQ
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	D--GHTCG-LNWFR--D-----DWDG-- -- --MY--GLGEQMAALEVMVNTQ
041	tr F7VZ72 F7VZ72_SORMK	---GRMCG-FNWRK--K-----KYDG-- -- --SH--GVGQQMNALGAVSALL
042	tr A3LN37 A3LN37_PICST	D--GHTCG-LNWFN-ST-----GWDG-- -- --YW--GLGEQISALEVIQNLR
043	tr Q6CAI2 Q6CAI2_YARLI	D--GHTCS-IDWLT--G-----SYSN-- -- --DWIGLGEQMSALEVLQNNL
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	D--WHTCG-LNWQN--G-----TWDG-- -- --YY--GLGEQMSALEVVTNLR
045	tr G9MJS5 G9MJS5_HYPVG	---GRQCG-IKWNT--G-----KYDG-- -- --RT--GAGQEMNVVGAVSSLL
046	tr G9MHI4 G9MHI4_HYPVG	D--GTGCG-FAWLP--K--GTYDG-- -- --KN--GVGSQMNALDAVMYTM
047	tr D4AP27 D4AP27_ARTBC	E--GTTCG-LKWTT--G-----KFDG-- -- --ST--GVGEQLAAMEIFQSNL
048	tr B6GZT8 B6GZT8_PENRW	N---GT CG-VQWYR--S-----TWDG-- -- --W-MGMEEQISATKVFIANL
049	sp O74556 YCZ2_SCHPO	D--GVTCG-VQWWW--N--NDTWDG-- -- --LY--GLGEQMSALEAIQAPL
050	tr A0A0C4E991 A0A0C4E991_MAGP6	---GRACG-FYWTS--G-----EYIDPAID-KTS--GAGERMNVLA AVSSLL
051	tr B8MHG0 B8MHG0_TALSN	---SACG-MQWFS--Q-----KYDG-- -- --T-SAIEQEMSAMSIFSNAL
052	tr A3LMV8 A3LMV8_PICST	D--GITCG-MN WAS--G-----NWDG-- -- --IY--GLGEQTSALEVMNALI
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	D--GKT CG-INWGI--N-----GWDG-- -- --LY--GLGEQISALEIIQNSL
054	sp Q9P6I3 YHG7_SCHPO	D--GVTCG-YMWYWNNG-----TWDD-- -- --H-YGLGEQISAVETFQALL
055	tr Q2US57 Q2US57_ASPOR	N---NSCG-VVWYN--S-----TWDG-- -- --W-SGLEEQMSVTSILAANM
056	tr W7N622 W7N622_GIBM7	A--GTACG-FTWLT--N-----GFDG-- -- --IV--GVGPQMSSLQSIMYTL
057	sp Q9P6I4 YHG6_SCHPO	S--GTS CG-IYWFWNNG-----TWDD-- -- --N-YGVQE QFSALQAVQMLM
058	tr C7GRB3 C7GRB3_YEAS2	D--GHTCG-LNWQK--Q-----TNDG-- -- --YY--GLGEQMSALEVIQNLL
059	tr C7GP28 C7GP28_YEAS2	D--GHTCG-LNWFN--G-----TWDG-- -- --MY--GLGEQMSALEVMVNTR
060	tr G3JGZ5 G3JGZ5_CORMM	---GRTC CG-FYW SG--G-----KFVDPSVD-KTT--GAGEYMSALA AVSSLL
061	tr C4QXV4 C4QXV4_KOMPG	D--GHTCG-MN WLA--D-----GWDG-- -- --FY--GLGEQMSALETLQNTR
062	tr I1S0Z5 I1S0Z5_GIBZE	---GRVCG-FKWAS--G-----KYDG-- -- --KT--GAGQQMNVLAAVSSLL
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	D--GAACG-LKWTT--G-----VWDG-- -- --SE--DVGLQMSALEVIQNLL
064	tr G0W5X4 G0W5X4_NAUDC	D--GHTCG-LNWFD--N-----GWDG-- -- --YY--GLGEQMSALDVLVNTR
065	tr G8ZPC1 G8ZPC1_TORDC	D--GHTCG-LNWNK--Q-----ANDG-- -- --YY--GLGEQMSALEVMQQLL
066	tr A0A167CDC5 A0A167CDC5_9ASCO	D--GHTCG-LNWNV--G-----KWDG-- -- --VW--GLGEQMSALEVMQNTR
067	tr R9XAT7 R9XAT7_ASHAC	D--GHTCG-INWLT--N-----SWDG-- -- --TW--GLGEQMAALEVMQNLR
068	tr B2B747 B2B747_PODAN	G--QRA CG-FKWST--G-----TFDG-- -- --SQ--GAGQTMNVLGAVSSLL
069	tr G0SE99 G0SE99_CHATD	---GRTC CG-FQWVK--R-----QYDG-- -- --SK--GAGQQMNVLGAVSALL
070	tr J8Q6F0 J8Q6F0_SACAR	D--GHTCG-LNWEK--Q-----TNDG-- -- --YY--GLGEQMNALEVIQNLL
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	D--GHTCG-LDWST-TS-----GWDG-- -- --LY--GLGEQASALEIMNQLL

072	tr J3K6E2 J3K6E2_COCIM	D--GKS	CG-LKW	ST--G	----	GFDG--	----	SV--	GIGE	QMAA	LEVI	QSNL	
073	tr G4N3E1 G4N3E1_MAGO7	---GRA	CG-FYW	TR--G	----	TYIDPS	V	D-K	TT--	GAGER	MDALA	AAVSSLL	
074	tr C5M5J2 C5M5J2_CANTT	D--GVT	CG-EDW	ST--G	----	SWDG--	----	VY--	GLGE	QMS	SLEV	IMSLI	
075	tr A0A254U0X0 A0A254U0X0_ASPNG	DGLENL	CG-VRW	HQ--A	----	TWDG--	----	S-M	GLE	VQMS	ALGG	VTGAL	
076	sp Q75DG6 DCW1_ASHGO	D--GHT	CG-LNW	LI--N	----	GWDG--	----	KW--	GLGE	QMAA	LEII	QNL	
077	tr Q6BZF0 Q6BZF0_DEBHA	D--WHT	CG-LNW	QN--G	----	TWDG--	----	YY--	GLGE	QMC	ALEV	VTNLR	
078	tr E9E3Q1 E9E3Q1_METAQ	---QRQ	CG-FYW	SL--G	----	RFVDPA	A	D-K	TT--	GAGE	QMN	VLA	AAVSSLL
079	sp P36091 DCW1_YEAST	D--GHT	CG-LNW	FN--G	----	TWDG--	----	MY--	GLGE	QMS	ALEV	MVNTR	
080	tr W0TAH6 W0TAH6_KLUMD	D--GHT	CG-LNW	FY--D	----	GWDG--	----	KY--	GLGE	QMAA	LEV	MQNLR	
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	D--GHT	CG-LNW	QL--G	----	SNDG--	----	NY--	GLGE	QIS	ALEV	FDNLL	
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	A--NTA	CG-QKW	YT--G	----	SFDG--	----	IN--	GVGE	QMAS	LA	AVMYNL	
083	tr Q2TYU3 Q2TYU3_ASPOR	N--NT	CG-VRW	HE--S	----	KWDG--	----	W-V	GMEE	QIS	ATD	VLS	SVL
084	tr G2XGF6 G2XGF6_VERDV	G--GTA	CG-LAW	QP--R	----	GQFDG--	----	LV--	GVGE	QMS	ALS	AVMYTL	
085	tr G3AMT4 G3AMT4_SPAPN	D--GHT	CG-LDW	TNWTR	----	GWDG--	----	MY--	GLGE	QMS	ALEV	MQNLM	
086	tr A0A1C1D213 A0A1C1D213_9EURO	N--GTS	CG-SKW	WK--G	----	AIFDG--	----	ET--	GVG	QQMC	ALEV	IQSNL	
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	D--GRA	CG-FRW	TT--G	----	GYDG--	----	LT--	GAG	QEMS	VLA	ALSSLL	
088	tr A0A100I5A6 A0A100I5A6_ASPNG	DGLENL	CG-VRW	HQ--D	----	TWDG--	----	S-M	GLE	VQMS	ALGG	VTGAL	
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	D--GHT	CG-INW	EK--G	----	SWDG--	----	MY--	GLGE	QAS	ALEV	IQSLL	
090	tr G2Q8A7 G2Q8A7_MYCTT	---GRT	CG-FQW	SK--R	----	QYDG--	----	SH--	GAG	QQMN	VLA	AAVSSLL	
091	tr B8NPZ1 B8NPZ1_ASPFN	---QNL	CG-ERW	YS--T	----	EPVG--	----	P-T	GLE	VQMA	ALGG	ITSNL	
092	tr Q7SAB2 Q7SAB2_NEUCR	D--GRA	CG-FRW	TT--G	----	GYDG--	----	LT--	GAG	QEMS	VLA	ALSSLL	
093	tr A2R8R5 A2R8R5_ASPNC	N--NT	CG-VRW	WN--S	----	TWDG--	----	W-S	GLE	EQMS	VT	SVFSSNM	
094	tr C5DHG0 C5DHG0_LACTC	D--GHT	CG-LNW	QL--G	----	YHDG--	----	YY--	GLGE	QAC	ALE	GIIQSLM	
095	tr G0SFA3 G0SFA3_CHATD	P--GTA	CG-FRW	TV--N	----	GFDG--	----	QT--	GVGE	QMN	ALA	AVMYSM	
096	tr K1WJG5 K1WJG5_MARBU	D--GVT	CG-GHW	YL--	----	DYDG--	----	NY--	GLG	QQMS	ALS	SVIQAML	
097	tr H2B005 H2B005_KAZAF	D--GHT	CG-INW	FY--G	----	GWDG--	----	YY--	GLGE	QMS	ALE	IMVNTR	
098	tr C5DYD7 C5DYD7_ZYGR	D--GHT	CG-LNW	FK--G	----	SWDN--	----	QY--	GLGE	QMS	ALEV	IQNTL	
099	tr G8YM23 G8YM23_PICSO	D--GHT	CG-TNW	FK--H	----	SWDG--	----	NY--	GLGE	QMC	ALEV	IQNLR	
100	tr W3X8E3 W3X8E3_PESFW	---GRT	CG-FGW	AS--G	----	TFDG--	----	SV--	GAG	QTM	NVL	GAISSLL	
101	tr A0A254U3K7 A0A254U3K7_ASPNG	N--NT	CG-VRW	WN--S	----	TWDG--	----	W-S	GLE	EQMS	VT	SVFSSNM	
102	tr Q6CER8 Q6CER8_YARLI	D--GHT	CS-INW	LT--G	----	EYNV--	----	SYI	GLGE	QLS	ALEV	ILNTQ	
103	tr G2QT05 G2QT05_THITE	---QRT	CG-FKW	ST--K	----	AFDG--	----	SV--	GAG	QQMN	VLA	AAVSSLL	
104	sp Q5ACZ2 DFG5_CANAL	D--GVT	CG-ENW	AI--D	----	KWDG--	----	VY--	GLGE	QTS	ALEV	MMALI	
105	tr G2WKU6 G2WKU6_YEASK	D--GHT	CG-LNW	QK--Q	----	TNDG--	----	YY--	GLGE	QMS	ALEV	IQNLL	
106	tr A0A0C7N752 A0A0C7N752_9SACH	D--GHT	CG-LNW	QN--G	----	SWDG--	----	WY--	GLGE	QAS	ALEV	IQSSL	
107	tr W3WN68 W3WN68_PESFW	T--GTA	CG-FSW	LD--N	----	STWDG--	----	NS--	GVGE	QLN	AMS	IVMVNL	
108	tr G2QFS1 G2QFS1_MYCTT	---GRR	CG-FYW	SE--G	----	VFVDPA	V	D-K	TS--	GAGE	AMS	VLA	AAVSSLL
109	tr G3J9G4 G3J9G4_CORMM	D--GTG	CG-FSW	LN--G	----	TNDG--	----	LM--	GVGP	QMD	ALA	AVFYNL	
110	tr Q2UJ03 Q2UJ03_ASPOR	D--HST	CG-MKW	TS--G	----	QWDQ--	----	SD--	DVG	VQMS	VLEV	MQATL	
111	tr G8YRT2 G8YRT2_PICSO	D--GVT	CG-TSW	TE--G	----	KWDN--	----	SW--	GLG	QQMS	ALE	TMLGTI	
112	tr W3X554 W3X554_PESFW	A--GTA	CG-FQW	TK--S	----	PEFDN--	----	MV--	GVGE	QQS	ALA	AVMYNL	
113	tr Q1K7A8 Q1K7A8_NEUCR	D--GTA	CG-FKW	TQ--	----	TFDG--	----	WA--	GVGA	QMN	ALS	AVMYTL	
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	N--RTL	CG-RRW	HQ--D	----	TFDG--	----	T-S	SLEE	QMS	ALS	VFSSSM	
115	tr A7F3P0 A7F3P0_SCLS1	S--GRA	CG-EHW	TA--G	----	SAYDG--	----	KY--	GVGE	QMS	ALS	SVIQATL	
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	D--GTA	CG-FTW	LE--G	----	KKYDG--	----	LA--	GVGP	QMN	ALA	AVFYNL	
117	tr Q6CIP9 Q6CIP9_KLULA	D--GHT	CG-LNW	FT--G	----	DWDG--	----	QY--	GLGE	QAC	ALE	IINQLL	
118	tr H2AMQ5 H2AMQ5_KAZAF	D--GHT	CG-LNW	FL--G	----	SWDG--	----	YY--	GLGE	QMS	ALEV	LVNTR	
119	tr G8JMI3 G8JMI3_ERECY	D--GET	CG-LDW	TT--G	----	SWDG--	----	YY--	GLGE	QAS	ALEV	IQNLL	
120	tr C5DGT1 C5DGT1_LACTC	D--GHT	CG-LNW	FY--D	----	GWDG--	----	KW--	GLGE	QMAA	LEI	MQNVL	
121	tr K1XJR4 K1XJR4_MARBU	---GRT	CG-LSW	SS--G	----	VYDG--	----	TV--	GVG	QQMA	AMS	ALFVNI	
122	tr A0A124BYC5 A0A124BYC5_ASPNG	S--KTT	CG-QHW	YK--S	----	SWDG--	----	T-E	GIEE	EEMS	ATS	IFTSNL	
123	tr A7TLL2 A7TLL2_VANPO	D--GHT	CG-INW	FY--G	----	GWDG--	----	YY--	GLGE	QMC	ALEV	MINTR	
124	tr Q0CB86 Q0CB86_ASPTN	N--NS	CG-VRW	YG--N	----	KWDG--	----	W-H	GMEE	QIS	VADV	ISSAM	
125	tr K1XLH0 K1XLH0_MARBU	S--GTQ	CG-LQW	TL--K	----	QNDG--	----	SL--	GVGE	QMA	VLEV	IQSNL	
126	tr J7S438 J7S438_KAZNA	D--GHT	CG-LNW	FK--D	----	GWDG--	----	MY--	GLGE	QMS	ALE	CILNTR	
127	tr Q4WG09 Q4WG09_ASPFU	---DNT	CG-VRW	WQ--P	----	TWDG--	----	FTP	GLET	QMA	ALAG	ITANL	
128	tr H8WZ09 H8WZ09_CANO9	D--GVT	CG-ENW	SA--S	----	GWDG--	----	VY--	GLGE	QMS	SLEV	ILSLI	
129	tr I1RL09 I1RL09_GIBZE	---GRA	CG-FYW	SG--G	----	EFVDVA	V	D-G	TS--	GAGE	QMN	VLA	AAVSSLL
130	tr Q4W985 Q4W985_ASPFU	D--GAA	CG-LKW	TT--G	----	VWDG--	----	SE--	DVGL	QMS	ALEV	IQNLL	
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	D--GHT	CG-LDW	EK--Q	----	TNDG--	----	YY--	GLGE	QMN	ALEV	IQNLL	
132	tr W6QCW3 W6QCW3_PENRF	N--KNV	CG-MKW	TA--Q	----	KWDK--	----	TK--	DFG	QQMA	ALEV	IQANL	
133	tr W3WMD3 W3WMD3_PESFW	---GRF	CG-FHW	TT--G	----	TFDG--	----	KT--	GAG	QQMN	NVL	GGLTSL	
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	D--GHT	CG-LNW	QA--K	----	GWDG--	----	KY--	GLGE	QMS	ALE	ALVVTQ	
135	tr I2GYH1 I2GYH1_TETBL	D--GHT	CG-LNW	QA--S	----	THDG--	----	YY--	GLGE	QMS	ALEV	IQNLL	
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	D--GHT	CG-LNW	FK--G	----	AWDG--	----	QY--	GLGE	QMS	ALEV	IQNTL	
137	tr Q752P3 Q752P3_ASHGO	H--GYT	CG-LNW	HK--G	----	TYDG--	----	VY--	GLGE	QAS	ALE	IIQNVL	
138	tr B8MYP3 B8MYP3_ASPFN	S--KTD	CG-RSW	YK--Q	D----	KWDG--	----	S-K	SLES	DMS	ALS	SVLSSTM	
139	tr Q2TXL6 Q2TXL6_ASPOR	P--DKI	CG-QRW	FL--D	----	KYDG--	----	V-T	GLRE	HMC	ALS	SVFTANM	
140	tr Q0CN17 Q0CN17_ASPTN	Q--QSE	CG-RRW	YQ--A	----	TYDG--	----	T-N	SLET	DMS	ALS	VFSSNM	
141	tr J3NHD5 J3NHD5_GAGT3	---QRA	CG-FKW	ST--R	----	AFDG--	----	SV--	GAG	QQMN	NVL	GAVTSL	

142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	K--GRA	CG	LRWVH	--DGQIG	VWDG	--	--	--TT	GVGGEEMSALEVIQSTL	
143	tr A1CVB0 A1CVB0_NEOFI	D--GAA	CG	LKWTT	--G----	VWDG	--	--	--SE	DVGLQMSALEVMQNLL	
144	tr B8NVI3 B8NVI3_ASPFN	P--DKI	CG	QRWFL	--D----	KYDG	--	--	--V-T	GLREHMCALSVFTANM	
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	D--GHT	CG	QNWFY	--N----	GWDG	--	--	--YY	GLGEQMAALEIMQNTF	
146	tr A1DJ54 A1DJ54_NEOFI	--DNT	CG	IRWWQ	--P----	TWDG	--	--	--FTP	GLSQMAALAGITANL	
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	--GRV	CG	FYWSG	--G----	VFVDPA	V	D	-KTT	GAGEAMDVLAADVSSLL	
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	D--GHT	CG	LNWWY	--S----	GWDG	--	--	--YY	GLGEQMSALEVMQNLR	
149	tr A0A177A618 A0A177A618_9PEZI	V--GAE	CG	LRWYE	--G----	ATDG	--	--	--RE	GVGQQMGAMDVMGALL	
150	tr Q75CW6 Q75CW6_ASHGO	D--GHT	CG	MVWRN	--Y----	TWDG	--	--	--KY	GLGEQLSALEIIQNTL	
151	tr Q7S4K4 Q7S4K4_NEUCR	--GRV	CG	FYWSG	--G----	VFVDPA	V	D	-KTT	GAGEAMDVLAADVSSLL	
152	tr G2QB99 G2QB99_MYCTT	P--GTA	CG	FRWTT	--G----	AFDG	--	--	--SA	GVGEQMNALSAVMYPL	
153	tr G0S3F2 G0S3F2_CHATD	--GRR	CG	FYWKS	--G----	KFVDPS	V	D	-HTS	GAGEAMSVLAADVSSLL	
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	D--GVT	CG	LNWFT	--G----	GWDG	--	--	--KF	GLGEQMCALAMTNLQ	
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	D--GHT	CG	LDWQL	--K----	TNDG	--	--	--YY	GLGEQMSALEVIQQLL	
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	N--NS	CG	VRWYT	--S----	KWDG	--	--	--W-T	GMEEQISVTDVLSVSL	
157	tr G2QKJ0 G2QKJ0_MYCTT	-----	CG	FRWNT	--G----	EYDGD	T	A	--AG	PAGQEMSALAALSTML	
158	tr Q4WFX5 Q4WFX5_ASPFU	T--GTY	CG	MRWTT	--G----	EFDG	--	--	--TT	GVGQQLSALEVQQANL	
159	tr Q5BGD7 Q5BGD7_EMENI	EGKSNL	CG	SRWYQ	--E----	TWDG	--	--	--I-Q	GLEVQQAALGGITANL	
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	D--GHT	CG	LNWFY	--G----	GWDG	--	--	--KY	GLGEQMAALEVMQNVR	
161	tr G8BYN9 G8BYN9_TETPH	D--GVT	CG	LSWFD	--S----	TNDG	--	--	--YY	GLGEQMSALEIINQLL	
162	tr F7VVT4 F7VVT4_SORMK	-----	CG	YRWNV	--G----	KYDGD	V	D	--NG	PAGQGMSALAAFSTYL	
163	tr G8YB80 G8YB80_PICSO	D--GHT	CG	TNWFK	--N----	YWDG	--	--	--NY	GLGEQMSALEVMQNLR	
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	D--GHT	CG	LNWEY	--D----	GWDG	--	--	--YY	GLGEQVSALETIQSHL	
165	tr G2WRS7 G2WRS7_VERDV	--GRT	CG	FQWYS	--G----	QYDG	--	--	--KT	GAGQTMNVLSAVSSLL	
166	tr A0A124BVB5 A0A124BVB5_ASPNG	T--GTY	CG	MQWTT	--G----	AYDG	--	--	--TV	GVGQQMAALEVVQANL	
167	tr A7EIG7 A7EIG7_SCLS1	N--GRT	CG	LNWRT	--G----	GFDG	--	--	--FY	GVGEQMSALEVIQSNL	
168	tr C5M5U7 C5M5U7_CANTT	D--GVT	CG	LSWTNWS	Q-----	GWDG	--	--	--KW	GLGEQMSALEVMQNLM	
169	tr B9WAA8 B9WAA8_CANDC	D--GVT	CG	ENWAL	--G----	SWDG	--	--	--VY	GLGEQTSALEVMMALI	
170	tr F8MRU1 F8MRU1_NEUT8	--GRV	CG	FYWSG	--G----	VFVDPA	V	D	-KTT	GAGEAMDVLAADVSSLL	
171	tr A1DJS0 A1DJS0_NEOFI	N--NS	CG	VRWYT	--S----	KWDG	--	--	--W-T	GMEEQISVTDVLSVNL	
172	tr B6HLM8 B6HLM8_PENRW	K--DL	CG	NRWYG	-----	GYDG	--	--	--Q-N	SMENAIISGSQMMSAVM	
173	tr A0A117DWS0 A0A117DWS0_ASPNG	S--GTQ	CG	RRWYQ	--A----	QWDG	--	--	--E-T	SLETDMSALS VFSSNM	
174	tr A0A124BXM1 A0A124BXM1_ASPNG	N--NT	CG	VRWYP	--K----	KWDG	--	--	--W-N	GMEEEI AVTNVLASAL	
175	tr J3NZQ6 J3NZQ6_GAGT3	--GRA	CG	FFWTK	--G----	EFVDPA	L	T	S	DTL	GVGTRLDALS AVMSLL
176	tr W0THH8 W0THH8_KLUMD	D--GHT	CG	LNWFK	--G----	SWDG	--	--	--QY	GLGEQASALEIMNQLL	
177	tr B2AEF2 B2AEF2_PODAN	--GRR	CG	FYWRE	--G----	VYVDPA	V	D	-KTS	GAGEQMNVLAAVSSLL	
178	tr Q96TX1 Q96TX1_NEUCS	--GRM	CG	FSWKK	--K----	KYDG	--	--	--SH	GVGQQMNVLGAVSSLL	
179	tr E9DYA6 E9DYA6_METAQ	--GRQ	CG	LKWAD	--G----	KYDG	--	--	--KT	GAGQEMSVLAAVQSLL	
180	tr B9WAI2 B9WAI2_CANDC	D--GVT	CG	LSWTNWSE	-----	GWDG	--	--	--KW	GLGEQMSALEVMQNLM	
181	tr Q6C171 Q6C171_YARLI	D--GHT	CS	LNWLT	--G----	QYSN	--	--	--DYI	GLGEQLSAMEVIQNTQ	
182	tr A6ZMV1 A6ZMV1_YEAS7	D--GHT	CG	LNWQK	--Q----	TNDG	--	--	--YY	GLGEQMSALEVIQNLL	
183	tr B6K7X7 B6K7X7_SCHJY	D--AVT	CG	TRWYW	--N--NG	TWDS	--	--	--NY	GVGQQLSALEVIQSLL	
184	tr I1RSA2 I1RSA2_GIBZE	A--GTA	CG	FSWTT	--N----	TFDG	--	--	--SL	GVGPQMNALDIFMYTL	
185	tr F7VWZ9 F7VWZ9_SORMK	D--GTA	CG	FKWAE	-----	PFDG	--	--	--WS	GVGAQMNALS AVMYTL	
186	tr A0A1B2J789 A0A1B2J789_PICPA	D--GHT	CG	LNWLY	--D----	GWDG	--	--	--FY	GLGEQMSALETLQNTR	
187	tr G0V8N4 G0V8N4_NAUCC	D--GHT	CG	LNWFS	--G----	GWDG	--	--	--MY	GLGEQMSALEALVNTR	
188	tr H8WXN8 H8WXN8_CANO9	D--GHT	CG	LSWTNWT	V-----	GYDG	--	--	--YY	GLGEQMSALEVIQSVM	
189	tr A6ZZS0 A6ZZS0_YEAS7	D--GHT	CG	LNWFN	--G----	TWDG	--	--	--MY	GLGEQMSALEVMVNTR	
190	tr G9MQD1 G9MQD1_HYPVG	--GRR	CG	FYWSS	--G----	KYIDTA	V	D	-HTS	GAGEAMNVLAADVSSLL	
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	D--GHT	CG	QNWFY	--G----	GWDG	--	--	--FY	GLGEQMAALEIMQNVL	
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	--DNT	CG	VRWWQ	--P----	TWDG	--	--	--FTP	GLETQMAALAGITANL	
193	tr A0A254UET1 A0A254UET1_ASPNG	T--GTY	CG	MQWTT	--G----	TYDG	--	--	--TV	GVGQQMAALEVVQANL	
194	tr A1D587 A1D587_NEOFI	N--HTL	CG	RRWHQ	--D----	TFDG	--	--	--T-S	SLEGQMSALS VFSSSM	
195	tr Q1K7I4 Q1K7I4_NEUCR	--GRM	CG	FSWKK	--K----	KYDG	--	--	--SH	GVGQQMNVLGAVSSLL	
196	tr G8JS88 G8JS88_ERECY	D--GHT	CG	MNYSR	--G----	SYDE	--	--	--SY	GLGEQLAALETIQNLR	
197	tr A0A136J7D3 A0A136J7D3_9PEZI	A--GTA	CG	FTWTG	-----	TFDN	--	--	--LV	GVGPQMSALSALQYTL	
198	tr E9E4X8 E9E4X8_METAQ	--GRQ	CG	FKWAS	--G----	VYDG	--	--	--KT	GAGQEMSVLSAVMSLL	
199	tr Q6CP42 Q6CP42_KLULA	D--GHT	CG	LNWFY	--G----	GWDG	--	--	--KY	GLGEQMAALEVMQNLR	
200	tr G0WHL4 G0WHL4_NAUDC	D--GHT	CG	LNWAK	--G----	SHDG	--	--	--YY	GLGEQMCALAMVNLL	
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	D--GHT	CG	LNWFN	--G----	TWDG	--	--	--MY	GLGEQMAALEVMVNTR	
202	tr F9WYX6 F9WYX6_ZYMTI	N--GTS	CA	NSWYDP	V-----	DTE	D	--	--YYG	QINQNLNALEVFLANL	
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	--KRT	CG	FKWST	--K----	AFDG	--	--	--SV	GAGQQMNVLGAVTSLL	
204	tr J3P147 J3P147_GAGT3	--GRT	CG	FYWSK	--G----	VFRDP	N	A	D	-GTT	GVGEQMNVLSGVMSLL
205	tr I2H842 I2H842_TETBL	D--GHT	CG	MDWNW	--N----	GWDG	--	--	--YY	GLGEQMAALEFMVNTR	
206	tr S6E3R3 S6E3R3_ZYGB2	D--GHT	CG	LNWFK	--G----	AWDG	--	--	--QY	GLGEQMSALEVIQNTL	
207	tr E5AD94 E5AD94_LEPMJ	G--DHQ	CG	MKWWS	--P----	GFDG	--	--	--IG	GVGPQMTALNVISVLN	
208	tr Q6FJM7 Q6FJM7_CANGA	D--GHT	CG	LDWQL	--K----	TNDG	--	--	--YY	GLGEQMSALEVIQQLL	
209	tr A5DUW0 A5DUW0_LODEL	D--GHT	CG	LSWTNWT	V-----	GWDG	--	--	--LY	GLGEQMSALEVMQNLM	
210	tr G8BF46 G8BF46_CANPC	D--GHT	CG	LSWTNWT	V-----	GYDG	--	--	--YY	GLGEQMSALEVIQSVM	
211	tr W3XAF6 W3XAF6_PESFW	A--GTA	CT	FSWYD	--D----	STYED	--	--	--ST	GVGEQMNAMSVIYAML	

029	tr A5DNV5 A5DNV5_PICGU	V-HD-----RPPPYTA--NNGGSS-KGNP---GGGYG
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	I--HA-----T----KGPVRE--EDGGTS-QGNP---SAGGD
031	tr Q2TWC1 Q2TWC1_ASPOR	MLLE-----SKSPQTI--DTNPDA-AEHH---I----
032	tr J5RXY9 J5RXY9_SACK1	A-LD-----KPAPYTA--DEGGSS-AGDG----AAGTQ
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	LPNV-----EAPATE--QDRGTS-KGDA----GAGST
034	tr A0A254U5V1 A0A254U5V1_ASPNG	ITHRQSQG-----G---QSQGPLTS--DTGGTS-QSNP----NAGTG
035	tr A0A061B6H7 A0A061B6H7_CYBFA	I-RD-----RPAPCSN--FTCGTS-IGDP----AAGTY
036	tr G8ZQ93 G8ZQ93_TORDC	A-LN-----RPPPYNS--TNGGSS-TGDG----AAGTE
037	tr Q9C2J1 Q9C2J1_NEUCS	THKGV-----GKAAKGPVTT--AQGGTS-KGDP----GAGVT
038	tr C5NZK5 C5NZK5_COCP7	I--DL-----V----AGPA-D--NSTGIS-RGNP----SAGTG
039	tr Q6C0T7 Q6C0T7_YARLI	I-MS-----SEGPLAD--ITGGDS-KGNG----SAGTV
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	A-LK-----RAPPYNA--TNGGNS-TGDG----AAGTK
041	tr F7VZ72 F7VZ72_SORMK	INQQG-----TKAPVTN--TTGGTS-KGNP----NAGSQ
042	tr A3LN37 A3LN37_PICST	V-RD-----FPPPLTA--NTGGSS-KGNP----AAGYS
043	tr Q6CAI2 Q6CAI2_YARLI	I-FNPKMAPKPTTNENGTTSGGVGPLTH--DTGGTS-EGSP----DAGSS
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	V-HD-----LPPPYTA--NNGGSS-KGNP----SGGYA
045	tr G9MJS5 G9MJS5_HYPVG	IGQA-----HVPVTN--STGGTS-QGNP----NAGSK
046	tr G9MHI4 G9MHI4_HYPVG	VLQA-----PSPATA--DKGGTS-KGNP----GAGMG
047	tr D4AP27 D4AP27_ARTBC	I--RK-----V----VPPVTQ--SSGGIS-LGGP----GSGSK
048	tr B6GZT8 B6GZT8_PENRW	INFNK-----TAPVTS--TTGGNS-TSNP----TAGED
049	sp O74556 YCZ2_SCHPO	LLKS-----LQVFKA--SNGGSS-TGDP----NAGLY
050	tr A0A0C4E991 A0A0C4E991_MAGP6	IDQT-----PAPVTN--STGGTS-AGDP----LAGTI
051	tr B8MHG0 B8MHG0_TALSN	VGFSAPTANSYGSSYTAP---EAPAPVTA--DTGGNS-TSNP----SGGQS
052	tr A3LMV8 A3LMV8_PICST	V---EEPLKRSAADPEVAD-KIDY----NAGLN
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	I-PQ-----VPPPC EE--DTCGEKLASDV----SIVVV
054	sp Q9P6I3 YHG7_SCHPO	AQQS-----ATILTL--DTGASS-ESNPD----AGTD
055	tr Q2US57 Q2US57_ASPOR	IGLNT-----SGAPVTS--TTGGNS-TSDP----TAGES
056	tr W7N622 W7N622_GIBM7	APAA-----KAPVTT--KTGGTS-KGNP----GGGQT
057	sp Q9P6I4 YHG6_SCHPO	IEYA-----PEIATL--ASSTDN-RSNSTYASNVIN
058	tr C7GRB3 C7GRB3_YEAS2	I-HDR-----PAPYKE--DNGGTS-KGDA----NAGMN
059	tr C7GP28 C7GP28_YEAS2	A-LD-----KPAPYTA--ENGGSS-VGDG----AAGTQ
060	tr G3JGZ5 G3JGZ5_CORMM	IKDA-----NPPLITN--STGGTS-KGSP----NGGFA
061	tr C4QXV4 C4QXV4_KOMPG	A-LV-----RPAPYTA--QTGGSS-QGDP----AAGLG
062	tr I1S0Z5 I1S0Z5_GIBZE	MENT-----PPPVTA--KKGGTS-KGNP----NAGNV
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	V--DR-----V----DPPVTD--ATGGTS-VGDP----SGGME
064	tr G0W5X4 G0W5X4_NAUDC	V-LD-----RPAPYNS--TNGGSS-IGDG----AAGTE
065	tr G8ZPC1 G8ZPC1_TORDC	I-HQK-----AAPFTE--QTGGSS-AGDA----NAGLN
066	tr A0A167CDC5 A0A167CDC5_9ASCO	A-LQ-----IAKPLTA--HTGGSS-KGNP----AAGSD
067	tr R9XAT7 R9XAT7_ASHAC	C-LE-----RPGPYTA--MNGGTS-PGDP----AAGTK
068	tr B2B747 B2B747_PODAN	IGQS-----RPPVTN--STGGTS-KGDP----NAGSQ
069	tr G0SE99 G0SE99_CHATD	VEYS-----KPPLTN--GTGGTS-KGDP----MAGSR
070	tr J8Q6F0 J8Q6F0_SACAR	I-HDR-----PAPYTE--NTGGTS-QGDA----NAGMN
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	I-HQR-----PGPLTG--STGGSS-EGDA----NAGLN
072	tr J3K6E2 J3K6E2_COCIM	I--GS-----V----DPPVTE--KDGGTS-KGNP----GAGV-
073	tr G4N3E1 G4N3E1_MAGO7	VSEV-----AAPLTG--KDGGTS-QGNP----NAGIP
074	tr C5M5J2 C5M5J2_CANTT	V---EDPISV--KTGGTN-RTNY----AAGTD
075	tr A0A254U0X0 A0A254U0X0_ASPNG	MMLGDG-----SSGPQTI--DENPDA-AEHH---I----
076	sp Q75DG6 DCW1_ASHGO	C-LE-----RPAPYTA--MNGGTS-PGDP----AAGTK
077	tr Q6BZF0 Q6BZF0_DEBHA	V-HD-----LPPPYTA--NNGGSS-KGNP----AGGYA
078	tr E9E3Q1 E9E3Q1_METAQ	IESA-----EQPVTN--KTGGTS-VGNP----NAGGK
079	sp P36091 DCW1_YEAST	A-LD-----KPAPYTA--ENGGSS-VGDG----AAGTQ
080	tr W0TAH6 W0TAH6_KLUMD	C-LD-----RPAPLTA--NTGGTS-VGNP----AAGTE
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	I-KD-----RPAPYTN--ETGGTS-EGDA----SAGTS
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	YPSS-----HLPLSA--ASGGTS-TGNV----DAGGS
083	tr Q2TYU3 Q2TYU3_ASPOR	VTEKK-----GSGPLTS--TTGGNS-TSNP----NAGSG
084	tr G2XGF6 G2XGF6_VERDV	VQKER-----T----APVTA--QTGGTS-KGNP----QAGM-
085	tr G3AMT4 G3AMT4_SPAPN	A-HK-----RPAPYTA--SNGGSS-LSNP----AAGYG
086	tr A0A1C1D213 A0A1C1D213_9EURO	I--TE-----V----SGPVTE--GSGGTS-QGDP----NAGTS
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	VFNEN--ASG---GDGKAGNKGAPLTG--ETGGTS-KGNP----HAGTG
088	tr A0A100I5A6 A0A100I5A6_ASPNG	MMLGDG-----SSGPQTI--DENPDA-AEHH---I----
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	F-DQR-----PAPYTD--SDGGTS-VGDS----SAGLN
090	tr G2Q8A7 G2Q8A7_MYCTT	IDQS-----PPPYTN--MSGGTS-KGDP----LAGTN
091	tr B8NPZ1 B8NPZ1_ASPFN	MLFE-----AQSPKTI--ESNPNA-TETE---I----
092	tr Q7SAB2 Q7SAB2_NEUCR	VFNEN--ASG---GDGKAGNKGAPLTG--ETGGTS-KGNP----HAGTG
093	tr A2R8R5 A2R8R5_ASPNC	IAFSNS-----SAAPLTS--STGGNS-TSNP----SAGTD
094	tr C5DHG0 C5DHG0_LACTC	Y-SKR-----PAPYTS--DNGGSS-VGDT----EAALN
095	tr G0SFA3 G0SFA3_CHATD	VVDPN-----LSQFAPVPLTA--VSGGTS-KGDP----SAGTS
096	tr K1WJG5 K1WJG5_MARBU	I--DE-----A----PQLLTN--TTGGTS-QGDV----NAGTG
097	tr H2B005 H2B005_KAZAF	A-LH-----KPGPYTS--ENGGSS-MGDG----AAGTQ
098	tr C5DYD7 C5DYD7_ZYGRG	I-HNS-----DPPYRE--STGGSS-KGDS----AAGLN

099	tr G8YM23 G8YM23_PICSO	V-HD	-----LPPPYTA--HEGGS--IGNP----	AAGYS
100	tr W3X8E3 W3X8E3_PESFW	IGES	-----KVPVTN--TTGGTS--GGNY----	NAGQD
101	tr A0A254U3K7 A0A254U3K7_ASPNG	IAFSNS	-----SAAPLTS--STGGNS--TSNP----	SAGTD
102	tr Q6CER8 Q6CER8_YARLI	I-LN	-----STGPLTD--ITGGTS--KGNG----	SAGTQ
103	tr G2QT05 G2QT05_THITE	IDQV	-----PPPVTN--STGGTS--KGDP----	TAGMH
104	sp Q5ACZ2 DFG5_CANAL	V----	-----EPPLSV--KTGGTN--RTDY----	SAGTN
105	tr G2WKU6 G2WKU6_YEASK	I-HDR	-----PAPYKE--DNGGTS--KGDA----	NAGMN
106	tr A0A0C7N752 A0A0C7N752_9SACH	Y-NQR	-----PAPFTS--DNGGSS--VGDA----	NAGLN
107	tr W3WN68 W3WN68_PESFW	LADS	-----PSPYTS--TTGGSS--VGNA----	NAGAS
108	tr G2QFS1 G2QFS1_MYCTT	IDEA	-----PPPATN--AT-GIS--RGDP----	NAGSR
109	tr G3J9G4 G3J9G4_CORMM	LPNV	-----KVPATE--QDRGTS--KGDA----	GAGSS
110	tr Q2UJ03 Q2UJ03_ASPOR	V--GG-----V----	APPVTE--DNGGTS--KGDP----	SAGTE
111	tr G8YRT2 G8YRT2_PICSO	V-EH	-----FHAPYTR--HTGGTS--KSEP----	NAGLN
112	tr W3X554 W3X554_PESFW	VKRTT-----Q----	APVTA--DTGGTS--KGDV----	NAGAS
113	tr Q1K7A8 Q1K7A8_NEUCR	THKGV-----GKAAKGPVTT--	AQGGTS--KGDP----	GAGVT
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	IAHRM-----QAQA	APLTV--DTGGTS--KSNA----	SAGTG
115	tr A7F3P0 A7F3P0_SCLS1	I--GT-----A----	PNLVTN--DTGGTS--VGNA----	AGGSG
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	LEDA	-----KPPATS--EDRGTS--KGNP----	NAGSN
117	tr Q6CIP9 Q6CIP9_KLULA	V-HDL	-----PAPLTE--STGGSS--SGDA----	SAGLN
118	tr H2AMQ5 H2AMQ5_KAZAF	A-LD	-----KPGPYNS--TNGGSS--QGDG----	AAGTE
119	tr G8JMI3 G8JMI3_ERECY	I-HTK	-----APPLTK--NAGGTS--NGDP----	GAGLG
120	tr C5DGT1 C5DGT1_LACTC	C-LD	-----RPAPYTA--ATGGSS--VGNG----	AAGTE
121	tr K1XJR4 K1XJR4_MARBU	LPLQE	-----VPPPLTN--NTGGTS--VGNP----	DAGAR
122	tr A0A124BYC5 A0A124BYC5_ASPNG	VAYKHL	-----SPATK--TTATNV-T-SSAGT-DTGNG	
123	tr A7TLL2 A7TLL2_VANPO	A-LH	-----VPGPYNS--TNGGSS--TGDG----	AAGTE
124	tr Q0CB86 Q0CB86_ASPTN	LKFQ	-----AKAPLTS--STGGTS--SSDP----	NAGGT
125	tr K1XLH0 K1XLH0_MARBU	V--DQ-----A----	QPWKSL-VAGTGTS--EGNV----	NAGQD
126	tr J7S438 J7S438_KAZNA	A-LM	-----KPAPYTA--TNGGSS--KGES----	SRWYG
127	tr Q4WG09 Q4WG09_ASPFU	MYYK	-----SSAPKTI--QSNPDG--KEHQ----	I----
128	tr H8WZ09 H8WZ09_CANO9	A----	-----EEPISV--QTGGTN--RSNY----	AAGTD
129	tr I1RL09 I1RL09_GIBZE	IEDA	-----EPPATN--KTGGIS--KGDP----	NAGKD
130	tr Q4W985 Q4W985_ASPFU	V--DR-----V----	DPPVTD--ATGGTS--VGDP----	SGGME
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	I-HDR	-----PAPYKE--DNGGTS--KGDA----	NAGMN
132	tr W6QCW3 W6QCW3_PENRF	I--TH-----V----	AAPVTN--DDGGTS--KGNP----	NAGDK
133	tr W3WMD3 W3WMD3_PESFW	AA--	-----TPPLTN--TTGGTS--VGDP----	NAGSE
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	A-LK	-----RPAPYNS--TDGGNS--TGNG----	AAGLE
135	tr I2GYH1 I2GYH1_TETBL	I-HDR	-----PAPYTS--IDGGSS--EGDA----	NAGLN
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	V-HTR	-----PAPFTA--SDGGSS--VGDA----	SAGLN
137	tr Q752P3 Q752P3_ASHGO	I-HTK	-----APPLTA--DTGGQS--EGNP----	DAGLD
138	tr B8MYP3 B8MYP3_ASPFN	IAHKKE	-----HQAPLTA--ETGGTS--KSNP----	SAGSG
139	tr Q2TXL6 Q2TXL6_ASPOR	VPFKTG-----N--RD	QGPLTA--DTGGTS--KGDP----	SAGTG
140	tr Q0CN17 Q0CN17_ASPTN	IKFKKG	-----DVSPLTA--DTGGKS--KSNP----	DAGTG
141	tr J3NHD5 J3NHD5_GAGT3	VDDV	-----RPPLTN--DTGGTS--QGDP----	NAGTH
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	I--GS-----T----	AGPLTN--TTGGTS--QGNP----	NAGSN
143	tr A1CVB0 A1CVB0_NEOFI	V--DR-----V----	DPPVTD--ATGGTS--VGDP----	SGGME
144	tr B8NVI3 B8NVI3_ASPFN	VPFKTG-----N--RD	QGPLTA--DTGGTS--KGDP----	SAGTG
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	C-LD	-----RPAPYTA--QDGGTS--QGNG----	AAGTE
146	tr A1DJ54 A1DJ54_NEOFI	MYYK	-----STAPNTI--QTNPEG--KEHQ----	I----
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	IDEA	-----DPPVTN--TTGGTS--KGDP----	NAGTG
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	I-RD	-----RPAPCTN--STCGTS--QGDG----	AAGTQ
149	tr A0A177A618 A0A177A618_9PEZI	V--TG-----A----	RDLVTN--KTGGTS--LGDY----	GDSVD
150	tr Q75CW6 Q75CW6_ASHGO	I-HTK	-----PPPTTA--NEGAKS--KGDP----	SAGLY
151	tr Q7S4K4 Q7S4K4_NEUCR	IDEA	-----DPPVTN--TTGGTS--KGDP----	NAGTG
152	tr G2QB99 G2QB99_MYCTT	VVRGA	-----AAPPLTA--DTGGTS--KGNP----	GGGAA
153	tr G0S3F2 G0S3F2_CHATD	IEYA	-----EPPATN--ET-GIS--RGDP----	NAGMR
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	A--N	-----KFPIYTA--TTGGSS--QGNP----	AGGYA
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	I-HER	-----PAPYRA--DNGGTS--VGDA----	AAGLN
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	ITTK	-----HKGPVTS--TTGGNS--TSNP----	TAGSG
157	tr G2QKJ0 G2QKJ0_MYCTT	LEQEK	-----VLKGPLTN--TTGGTS--QGDP----	NAGQK
158	tr Q4WFX5 Q4WFX5_ASPFU	Y--NT-----V----	AGPLTN--STGGTS--KRNS----	AVERG
159	tr Q5BGD7 Q5BGD7_EMENI	MLLT	-----DVVAKTI--DTNPGA--KEQF----	L----
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	C-LD	-----RPAPYTA--NTGGSS--IGNP----	AAGTE
161	tr G8BYN9 G8BYN9_TETPH	V-SSS-----GAAAE	AASSGSASSPYTS--SSGGNS--TGDA----	SAGLN
162	tr F7VVT4 F7VVT4_SORMK	ITEEQ	-----AVFKPLVTN--STGGES--RGNP----	NAGEK
163	tr G8YB80 G8YB80_PICSO	V-HD	-----LPPPYTA--DDGGSS--IGNA----	AAGYS
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	T-PI	-----SPAPLTQ--ENEGVA--VGDV----	NAGLN
165	tr G2WRS7 G2WRS7_VERDV	INSA	-----KPPLTN--STGGTS--KGDF----	NAGAD
166	tr A0A124BVB5 A0A124BVB5_ASPNG	L--QA-----N----	PGPLTH--TTGGSS--QGNS----	AAGTQ
167	tr A7EIG7 A7EIG7_SCLS1	I--EQ-----V----	AGPVTN--TTGGTS--KGDP----	SAGSG
168	tr C5M5U7 C5M5U7_CANTT	S-NS	-----RPAPYTA--DTGGSS--IGNP----	AAGYG

169	tr B9WAA8 B9WAA8_CANDC	V---	EPPLSV--KTGGTN--RTNY----	AAGTGN
170	tr F8MRU1 F8MRU1_NEUT8	IDEA-----	DPVVTN--TTGGTS--KGD-----	NAGTGN
171	tr A1DJS0 A1DJS0_NEOFI	ITTK-----	HKGPTS--TTGNS--TSDP-----	TAGSG
172	tr B6HLM8 B6HLM8_PENRW	LKFLGS-----	SKPVST--ATGNG--TSDP-----	NAGTR
173	tr A0A117DWS0 A0A117DWS0_ASPNG	ITHRQSQG-----G--QS	QGPLTS--DTGGTS--QSNP-----	NAGTGN
174	tr A0A124BXM1 A0A124BXM1_ASPNG	YSTK-----	QTAPVTS--STGNS--TSDP-----	NAGTGN
175	tr J3NZQ6 J3NZQ6_GAGT3	VEFDD-----	AQSPTTG--STGNS--AGDT-----	GAGLR
176	tr W0THH8 W0THH8_KLUMD	I-HER-----	PPLTE--KTGSA--RGDP-----	NAGLN
177	tr B2AEF2 B2AEF2_PODAN	IDDA-----	PPPANN--IT-GLS--KPNY-----	DAGSR
178	tr Q96TX1 Q96TX1_NEUCS	IEQPG-----	KAPVTN--STGTS--KGNP-----	NAGSQ
179	tr E9DYA6 E9DYA6_METAQ	IGKA-----	RPPVTH--DSGTS--TGNT-----	DGGQG
180	tr B9WAI2 B9WAI2_CANDC	V-HK-----	RPAPYTA--DTGSS--IGNP-----	AAGYG
181	tr Q6C171 Q6C171_YARLI	V-LK-----	SSGPLTD--ITGTS--KNG-----	SAGAL
182	tr A6ZMV1 A6ZMV1_YEAS7	I-HDR-----	PAPYKE--DNGTS--KGD-----	NAGMN
183	tr B6K7X7 B6K7X7_SCHJY	VDKM-----	QVPLTQ--STGTS--ASNP-----	NAGSA
184	tr I1RSA2 I1RSA2_GIBZE	VGNA-----	KAPYTS--KTGTS--KGD-----	GGGNT
185	tr F7VWZ9 F7VWZ9_SORMK	THKGL-----GKA	AKGPATE--KLGTS--KGD-----	GAGVT
186	tr A0A1B2J789 A0A1B2J789_PICPA	A-LV-----	RPAPYTA--QSGSS--QGD-----	AAGLG
187	tr G0V8N4 G0V8N4_NAUCC	V-LH-----	LPGPYNS--TDGSS--IGNS-----	AAGTE
188	tr H8WXN8 H8WXN8_CANO9	IYHD-----	APAPYTA--ETGSS--KSQP-----	ASGYG
189	tr A6ZZS0 A6ZZS0_YEAS7	A-LD-----	KPAPYTA--ENGSS--VGDG-----	AAGTQ
190	tr G9MQD1 G9MQD1_HYPVG	INEA-----	PPVVTN--SSGGIS--KGNP-----	NAGHG
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	C-LQ-----	KPAPYTA--KDGGTS--IGNG-----	AAGTE
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	MYYK-----	SSAPKTI--QSNPDG--KEHQ-----	I----
193	tr A0A254UET1 A0A254UET1_ASPNG	L--QA-----N--	PGPLTH--TTGSS--QGNS-----	AAGTQ
194	tr A1D587 A1D587_NEOFI	IAHKM-----	QAQAPLTA--DTGGTS--KGNA-----	SAGTGN
195	tr Q1K7I4 Q1K7I4_NEUCR	IEQPG-----	KAPVTN--STGTS--KGNP-----	NAGSQ
196	tr G8JS88 G8JS88_ERECY	C-LE-----	KGSPLTA--SEGGTS--IGNP-----	GAGGN
197	tr A0A136J7D3 A0A136J7D3_9PEZI	IKKNN-----R--	AAVPATA--NNGTS--IGNV-----	NAGAR
198	tr E9E4X8 E9E4X8_METAQ	IPQA-----	KAPVTE--KDGGTS--KGNP-----	NAGGS
199	tr Q6CP42 Q6CP42_KLULA	C-LD-----	RPAPYTA--DDGGTS--AGNP-----	AAGTE
200	tr G0WHL4 G0WHL4_NAUDC	I-HDR-----	PAPLTG--NTGSS--KGD-----	LAGLN
201	tr A0A0L8RFQ5 A0A0L8RFQ5_SACEU	A-LD-----	RPAPYTA--EDGSS--VGDG-----	AAGTQ
202	tr F9WYX6 F9WYX6_ZYMTI	PSGDIKM-----	--GV-G--STNGTG--VGGSLGGG	ANGTE
203	tr A0A0C4DZP6 A0A0C4DZP6_MAGP6	IDDV-----	SPPLTN--ATGTS--IGDP-----	NAGTH
204	tr J3P147 J3P147_GAGT3	HEFPN-----NPV	EGPVTL--ETGTS--RGDP-----	NAGLR
205	tr I2H842 I2H842_TETBL	A-LD-----	LPPPYTS--SNGSS--TSGG-----	AAGTE
206	tr S6E3R3 S6E3R3_ZYGB2	V-HTR-----	PAPFTA--SDGSS--VGDA-----	SAGLN
207	tr E5AD94 E5AD94_LEPMJ	V--DR-----V--	PPPYSS--VTGSS--QGNP-----	NLGTE
208	tr Q6FJM7 Q6FJM7_CANGA	I-HER-----	PAPYRA--DNGTS--VGDA-----	AAGLN
209	tr A5DUW0 A5DUW0_LODEL	S-TK-----	RPAPYTA--NDGSS--KSHP-----	ASGYG
210	tr G8BF46 G8BF46_CANPC	IYHD-----	APAPYTA--ETGSS--ESQP-----	DSGYG
211	tr W3XAF6 W3XAF6_PESFW	VDDA-----	ITPYTT--TTGSS--SSNV-----	DGGTS
212	tr B6H7I2 B6H7I2_PENRW	INYNTG-----	SFGPTS--KTGNS--ASDP-----	NAGSE
213	tr Q5ATF9 Q5ATF9_EMENI	VSEK-----	SNPPLTT--KTGNS--TSDP-----	NAGTS
214	tr A0A167CDD3 A0A167CDD3_9ASCO	I-NT-----	RPGPLTE--KSGPS--GGFG-----	AAGLN
215	tr G0RXS3 G0RXS3_CHATD	LDQPH-----	VLRGPLTN--DTGGTS--RGDP-----	NAGLV
216	tr B8NX26 B8NX26_ASPFN	MLMA-----	AESPNTI--DTNPDA--KEHN-----	V----
217	tr A0A0E1RZZ1 A0A0E1RZZ1_COCIM	I--DL-----V--	AGPA-D--NSTGIS--RGNP-----	SAGTGN
218	tr A0A0J5SN29 A0A0J5SN29_ASPFM	Y--NT-----V--	AGPLTT--STGTS--KRNS-----	AVERG
219	tr A1CMM3 A1CMM3_ASPCL	VDYV-----E--	KRVT-A--SSGGIS--QGD-----	NAGTGN
220	tr B6HU84 B6HU84_PENRW	IPFS-----	GKAPLTA--SSGTS--KSNP-----	NAGTN
221	tr A0A0B0E7F0 A0A0B0E7F0_NEUCS	IEQPG-----	KAPVTN--STGTS--KGNP-----	NAGSQ
222	tr G2WHY6 G2WHY6_YEASK	A-LD-----	KPAPYTA--ENGSS--VGDG-----	AAGTQ
223	tr G0W7G8 G0W7G8_NAUDC	C-LH-----	KPAPYTS--TNGSS--VGDG-----	AAGTE
224	tr G4NGV8 G4NGV8_MAGO7	IRNNT-----K--	EVVPTS--DTGGTS--KGD-----	TAGII
225	tr W3X855 W3X855_PESFW	LDGA-----	PTPYTS--VTGTS--TGDS-----	SGGTA
226	tr A0A2N6NY19 A0A2N6NY19_BEABA	YPNA-----	KPAVTA--DTGGTS--KSDP-----	NAGVG
227	tr J8Q2K7 J8Q2K7_SACAR	A-LD-----	KPAPYTA--EDGSS--TGDG-----	AAGTQ
228	tr A0A1E4RBW2 A0A1E4RBW2_9ASCO	V-RD-----	VDPPTYTA--NTGSS--IGDP-----	ASGYS
229	tr J6EGN8 J6EGN8_SACK1	I-HDR-----	PAPYKE--SNGTS--KGD-----	NAGMN
230	tr A0A100ISD9 A0A100ISD9_ASPNG	I--SR-----V--	DPPVTQ--DTGGTS--KGNP-----	AGGEP
231	tr G8YQC0 G8YQC0_PICSO	I-EH-----	FHAPYTD--RTGTS--KSEP-----	NAGID
232	tr F7VVP8 F7VVP8_SORMK	IEQA-----	DPPATN--QTGGIS--KGD-----	NAGTGN
233	tr A0A0L8VJB2 A0A0L8VJB2_9SACH	I-HDR-----	PAPYKE--DNGTS--KGD-----	NAGLN
234	tr F8MBP4 F8MBP4_NEUT8	THTGV-----GNV	TKGPVTA--AQGGTS--KGD-----	GAGVT
235	tr A0A254TXZ7 A0A254TXZ7_ASPNG	VVYKHQ-----	-SLATK--ATATNG--TSGSTGT--	ESGNG
236	tr A1CD27 A1CD27_ASPCL	MFFK-----	SAAPKTI--ESNPDG--KEHQ-----	I----
237	tr A5DHF3 A5DHF3_PICGU	A-RD-----	VDPPTYTS--KNGTS--KSQP-----	NAGID
238	tr D4ATT7 D4ATT7_ARTBC	WYKKQ-----A--	SGPA-N--SKTGT--KGD-----	NAGND

239	tr E2PT42 E2PT42_ASPNC	I--SR-----V-----DPPVTQ--DTGGTS-QGNP----AGGEP
240	tr C7ZPE5 C7ZPE5_NECH7	MEHT-----EPPVTA--KKGGIS-KGNP----NAGS-
241	tr A0A0A2VM24 A0A0A2VM24_PARBA	I--SK-----T-----PPPLTN--LTGGTS-PGDP----NAGLQ
242	tr Q4WKP7 Q4WKP7_ASPFU	IAHRM-----QAQAPLTV--DTGGTS-KSNA----SAGTG
243	tr F8MXT4 F8MXT4_NEUT8	IEQPG-----KAPVTN--STGGTS-KGNP----NAGSQ
244	tr A0A177AJ74 A0A177AJ74_9PEZI	I--GQ-----T-----PGPVTN--TTGGTS-QGNY----KTGSN
245	tr J7RQR5 J7RQR5_KAZNA	V-HDR-----PAPYTA--AAGGSS-VGDA----SAGLN
246	tr Q75CW7 Q75CW7_ASHGO	I-HTK-----PPPTTA--NEGAKS-QGDP----GAGLQ
247	tr G3AKK4 G3AKK4_SPAPN	V-----QEPLSV--KTGGSN-KTDF----DAGTK
248	tr B2W762 B2W762_PYRTR	Y--PT-----V-----PGPATQ--QSGGIS-KSNP----DAGSD
249	tr A0A0L8VLX1 A0A0L8VLX1_9SACH	A-LD-----KPAPTYTA--ENGSS-VGDG----AAGTQ
250	tr C4XWK1 C4XWK1_CLAL4	T-SK-----N-TPLTP--KTGGS-DAGASV----DAGNS

001	tr A0A254U2J9 A0A254U2J9_ASPNG	DNTGSS-GST---	ESKIT	TGDKAG-----	AS-IL	TIAF-VGMW	GGMIA	
002	sp Q6FLP9 DCW1_CANGA	PHPT-----	NLAPLHIT	TGGSRAG-----	AG-II	TAII-GISI	IACAL	
003	tr A1CCM5 A1CCM5_ASPCL	DKSGNT-PT---	THTVT	TGDKAG-----	AS-IL	TVGF-AVGW	VSLMA	
004	tr C1H190 C1H190_PARBA	DGRNDL---PKL--	SEITM	GDKVG-----	AG-FM	TSII-LLGV	LGGA	
005	tr Q2UR85 Q2UR85_ASPOR	HKDQQT-GT---	PKPIT	TGDRAG-----	AS-IV	TFFF-ACGW	MASVS	
006	tr A5DV30 A5DV30_LODEL	SQDS-----	VNKNEIT	VTGKDKAG-----	AG-VL	TAIV-LAIL	LLGSI	
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	PAPT-----	VTRHFTKT	STDVHT-----	ET-HT	TSVVTELF	YTAK--	
008	tr J3PGN0 J3PGN0_GAGT3	----P---SKD	RTFKEIT	TGDRAG-----	AA-IL	TMLL-LGSAT	STFG	
009	tr A0A124BXU6 A0A124BXU6_ASPNG	DSSDDK-TI---	LSTIT	TGDRAG-----	AG-IV	TVAF-AGGW	IGLMA	
010	tr C5P4A1 C5P4A1_COCP7	STGDA----	IGM-QRGI	KTADKAG-----	AW-IT	TMIL-IGVI	LGGFY	
011	tr Q0CG55 Q0CG55_ASPTN	PAPP---T-P	GILTMNIT	TADRAG-----	AG-IL	TVLM-SAFV	IGSTG	
012	tr C4Y2X5 C4Y2X5_CLAL4	KAAT-----	NASPLVLD	SGDKAG-----	AG-II	TAVI-GAAI	VGASV	
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	GPPAP-V-P	EGLRLEIT	KADRAG-----	AG-MM	TVML-SMIV	IGSTG	
014	tr C7Z068 C7Z068_NECH7	---SH---D-	MPEPDPI	TQADKAG-----	AG-VL	TFLI-LSSA	LGTfV	
015	tr G8BXX2 G8BXX2_TETPH	TSAT-----	NLSPLNIT	AGSRAG-----	AG-II	TAVI-GISI	VACAC	
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	IAKT-----	DAEPLDLH	AKDKAG-----	AS-II	TAII-GVAI	IGTAT	
017	tr A1CSC2 A1CSC2_ASPCL	TEH-SA-PQ---	ALPVT	TGDRAG-----	AG-VL	TVVF-LGAW	IGAVT	
018	sp Q05031 DFG5_YEAST	SST---TNV	LQNNLN	IKKGD	RAG-----	AA-II	TAVI-LSVL	TGGAV
019	tr H2AQJ6 H2AQJ6_KAZAF	TTTS--SADV	LENNLTIT	SKDRAG-----	AA-II	TAVI-LGCM	MAGGAI	
020	tr G2XFH9 G2XFH9_VERDV	----S---NP	FQDPEPI	TADRAG-----	AG-IV	TFII-LASGL	SSWY	
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	QAAT-----	NLSPLHIT	AGSRAG-----	AG-II	TCVV-GISI	IASLL	
022	sp Q5AD78 DCW1_CANAL	KLTS-----	DATPLSID	GGD	KAG-----	AG-II	TAII-GASL	VGSCV
023	tr B6H7E8 B6H7E8_PENRW	PEGSTG-PA----	PITSG	DRVG-----	AG-IL	TAVF-VLVW	VGAIS	
024	tr B6GZU2 B6GZU2_PENRW	PQKA---I-P	PALDYDIT	TGDKAG-----	AG-VL	TVLF-LIGI	GGSTG	
025	tr E9E4W7 E9E4W7_METAQ	----D---NG	ERPVKPVT	AAGKAG-----	APPVL	TILL-LVGAV	SLFV	
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	DPEP---K-P	KALSYKIT	TADRAG-----	AG-IL	TIMM-LATL	GGSCG	
027	tr G2R7G8 G2R7G8_THITE	----F---NA	LAPPKPIT	TADRAG-----	AG-IL	TAVV-LATF	LGGLV	
028	tr G3JBY1 G3JBY1_CORMM	AA-----Q	PLPQPRAI	VTSDRAG-----	AA-IL	TMLC-TVGI	VGGSF	
029	tr A5DNV5 A5DNV5_PICGU	ATNI-----	NATPLNLG	SGDKAG-----	AG-II	TAII-GMSI	VGATV	
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	GRGNKD-VKAG	PWSQPAGT	GDRVG-----	AG-AL	TAVM-AAGV	VCGVW	
031	tr Q2TWC1 Q2TWC1_ASPOR	DNNENS-SKD	PTKAKPIE	TADRAG-----	AW-IL	TVMI-AAGA	IGAVG	
032	tr J5RXY9 J5RXY9_SACK1	AQPT-----	NLAPLNIT	KGS	KAG-----	AG-II	TAVI-GISI	VACAL
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	QA-----Q	PLPEPRAI	TGGDRAG-----	AA-IL	TILI-AAGI	LAGSF	
034	tr A0A254U5V1 A0A254U5V1_ASPNG	PKDAPN-KL---	P-DIT	TGDRAG-----	AS-IL	TVLF-VGSW	AGAIT	
035	tr A0A061B6H7 A0A061B6H7_CYBFA	EDST-----	NLSPLSID	GGD	KAG-----	AA-II	TCII-GASL	VGSA
036	tr G8ZQ93 G8ZQ93_TORDC	ESPT-----	NLSPLDIT	AGSRAG-----	AG-II	TCVI-GISI	ILSCAL	
037	tr Q9C2J1 Q9C2J1_NEUCS	DPAS---RG	GLAALKPI	TMADR	VG-----	AG-IV	TAIL-AISI	VGGSV
038	tr C5NZK5 C5NZK5_COCP7	--SDPT---F	EL--SDIT	TGDRVG-----	AG-FL	TSVV-LIGI	LGGA	
039	tr Q6C0T7 Q6C0T7_YARLI	FESPYAN--T	IAAPLTIG	PGDRAG-----	AG-II	TAVV-CIVLI	IGSGW	
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	PHPT-----	NLAPLHIT	TGGSRAG-----	AG-II	TAII-GISI	IACAL	
041	tr F7VZ72 F7VZ72_SORMK	S---D---S	FGAHNRPI	STADRAG-----	AG-IV	TFLL-SAGA	LGTFA	
042	tr A3LN37 A3LN37_PICST	TLHT-----	ITSPLELE	TKDL	AG-----	AG-II	TAVV-GVSL	VAAGV
043	tr Q6CAI2 Q6CAI2_YARLI	SANP-----	LLMNLDIK	GS	DKAG-----	AG-VI	TAVL-LIGV	LGSGW
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	VTNT-----	NTSPLNLD	KGDT	AG-----	AS-II	TVII-AASLI	IGSGV
045	tr G9MJS5 G9MJS5_HYPVG	P---N---S	FQRPLTPV	TAGDRAG-----	AG-IV	TIVI-IGSL	CTGLA	
046	tr G9MHI4 G9MHI4_HYPVG	SIDP---AG	DLDKITKPT	TGGKVG-----	AA-IL	TLLL-LLSV	LGSA	
047	tr D4AP27 D4AP27_ARTBC	GKDNG---P	KI-LRPIT	GADTAG-----	AA-IL	TLLM-FCSL	SGVSY	
048	tr B6GZT8 B6GZT8_PENRW	DTDGSN-QM---	RTTTTT	TADKAG-----	AG-IV	TAVF-VAGW	IGLMS	
049	sp O74556 YCZ2_SCHPO	TAPV-----	SFANKNFEN	LRKHW-----	ML-LG----	FFLL	VPTL	
050	tr A0A0C4E991 A0A0C4E991_MAGP6	----P---SR	DRTFKEIT	TADRAG-----	AG-IL	TMLL-LGSAT	GTFG	
051	tr B8MHG0 B8MHG0_TALSN	STGYQP-PK---	AMKIT	TGGDRAG-----	AA-IL	TLVF-ATAW	IGMMV	
052	tr A3LMV8 A3LMV8_PICST	SHDT-----	VNQNEIT	VTGKDRAG-----	AG-VL	TAVV-LGIL	LGGA	
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	PTPT-----	IFRTFTTT	TNTVHKT-----	AT-LI	HSSVEN	VFYTG	TTS
054	sp Q9P6I3 YHG7_SCHPO	DGDTV	T-----	ITPATKSD	KGW-----	AG-FL	TFAF-SFV	FLLSI
055	tr Q2US57 Q2US57_ASPOR	DNEGSS-ST---	TTKVTT	TGDVVG-----	AS-IV	TIVL-VAVPI	AMV	

056	tr W7N622 W7N622_GIBM7	NMDV---	DVS	KPKYAKIT	TMDKVG	-----	AG-AIT	TCMI-	LACVIGGSV																														
057	sp Q9P6I4 YHG6_SCHPO	DTNTTT---	---	TIVVKEKDRGG	-----	AG-FLT	FLS-	AIFILGASI																															
058	tr C7GRB3 C7GRB3_YEAS2	SST---	TNV	LQNNLN	IKKGD	RAG-----	AA-IIT	TAVI-	LSVLTGGAV																														
059	tr C7GP28 C7GP28_YEAS2	AQPT-----		NLAPLN	ITKGS	SKAG-----	AG-IIT	TAVI-	GISIVACAL																														
060	tr G3JGZ5 G3JGZ5_CORMM	G--GD---	NG	LKIPKP	ITVAD	KAG-----	AG-ILT	TFML-	VFGASGTF																														
061	tr C4QXV4 C4QXV4_KOMPG	VKTE-----		AVPPLK	LTNAD	VAG-----	AA-IIT	TAII-	GLSVIAGAI																														
062	tr I1S0Z5 I1S0Z5_GIBZE	GD--G---	KI	DKTYGP	LTTAD	KAG-----	AG-ILT	TFLV-	LSFACGMFG																														
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	QPDP---	R-P	PVLTMT	ITGAD	RAG-----	AG-LLT	TAML-	GVLMI	GTTG																													
064	tr G0W5X4 G0W5X4_NAUDC	ATPT-----		NLAPLH	ITKGS	SKVG-----	AA-FIT	TAAI-	GLSIVFTSI																														
065	tr G8ZPC1 G8ZPC1_TORDC	TSS---	ANV	LQNQLD	IQT	KDRAG-----	AA-IIT	TAVI-	LMVLVGGSI																														
066	tr A0A167CDC5 A0A167CDC5_9ASCO	SGTS----	NF	LTNSLD	ISSK	DRAG-----	AG-ILT	TALL-	AVGIVGTGW																														
067	tr R9XAT7 R9XAT7_ASHAC	TKAE-----		NLPPLN	IKAGD	RAG-----	AG-IIT	TALI-	GSSLLACTL																														
068	tr B2B747 B2B747_PODAN	S---D---	NF	KDKYLP	PPT	TGDKAG-----	AG-ILT	TVLI-	LVS	VGT	FG																												
069	tr G0SE99 G0SE99_CHATD	S---D---	TL	TQEVKP	PPT	GGDKAG-----	AA-ILT	TFVV-	LATT	VGAFT																													
070	tr J8Q6F0 J8Q6F0_SACAR	SST---	TNV	LQNNLN	ITKGG	RAG-----	AA-IIT	TAVV-	LSVLVGGAV																														
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	VST---	TNV	LQKKIN	ITTGS	RAG-----	AS-IIT	TAIV-	LGVIILGGV																														
072	tr J3K6E2 J3K6E2_COCIM	STGNA----	IG	M-QRG	IKTAD	KAG-----	AW-IT	TMIL-	IAVILGGFY																														
073	tr G4N3E1 G4N3E1_MAGO7	----	K---	SNDR	TLSE	ITSGDKAG-----	AS-FLT	TILT-	MVSAASLFG																														
074	tr C5M5J2 C5M5J2_CANTT	SEDS-----	VN	QNELN	ITGK	DKAG-----	AG-ALT	TAIV-	LAIILGGTI																														
075	tr A0A254U0X0 A0A254U0X0_ASPNG	DTSEDE--	KD	PTAKEK	IDT	GD	RAG-----	AW-ILT	TVVL-	AGLMLGAVG																													
076	sp Q75DG6 DCW1_ASHGO	TKAE-----		NLPPLD	IKAGD	RAG-----	AG-IIT	TALI-	GSSFLACTL																														
077	tr Q6BZF0 Q6BZF0_DEBHA	VTNT-----		NSSPLN	LDKGD	TAG-----	AS-IIT	TVII-	AASLVGSGV																														
078	tr E9E3Q1 E9E3Q1_METAQ	----	D---	NGER	PVKP	ITTAD	KAG-----	AG-ILT	TFLL-	LGGAAGMFV																													
079	sp P36091 DCW1_YEAST	AQPT-----		NLAPLN	ITKGS	SKAG-----	AG-IIT	TAVI-	GISIVACAL																														
080	tr W0TAH6 W0TAH6_KLUMD	QSPS-----		NLSPLT	ITHGD	KAG-----	AG-IIT	TAVI-	GITII	IGSVV																													
081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	STTT-----	QL	DTAALN	ITGGD	KAG-----	AG-IIT	TAIV-	LAVII	IAGCV																													
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	STN---	AD	GKTL	SKITT	GD	KVG-----	AA-FLT	TMLV-	ISGLLGGMF																													
083	tr Q2TYU3 Q2TYU3_ASPOR	DDSSSD-K	SQ--	LKS	ITTGD	KAG-----	AS-IVT	IAF-	VGIW	GGLIA																													
084	tr G2XGF6 G2XGF6_VERDV	-E-----	PH	SWEFKP	IQQA	ERVA-----	AG-FLT	TVGI-	LTS	LMAGVF																													
085	tr G3AMT4 G3AMT4_SPAPN	TSIT-----		NTSPLH	LGP	GD	KAG-----	AG-IIT	TAIV-	GVSLIGTCV																													
086	tr A0A1C1D213 A0A1C1D213_9EURO	SPIGP----	G	DLHR	HEVT	AAD	KAG-----	AG-ILT	TVLL-	IFFIVGGAW																													
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	KG--V---	VD	PTEL	REL	TKGD	KVG-----	AW-ILT	TAFT-	LGT	LGAA	AFV																											
088	tr A0A100I5A6 A0A100I5A6_ASPNG	DTSEDE--	KD	PTAKEK	IE	TGD	RAG-----	AW-ILT	TVLL-	AGLMLGAVG																													
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	KTT---	TNV	LQSDLK	ITHK	DRV	G-----	AA-VLT	TAVV-	LAILLGGSI																													
090	tr G2Q8A7 G2Q8A7_MYCTT	S---R---	ST	LNEPRQ	ITGGD	RAG-----	AG-ILT	TILL-	IAAG	VGTFG																													
091	tr B8NPZ1 B8NPZ1_ASPFN	DHHSDE--	EP	NKPKP	ITTGD	RAG-----	AG-IIT	TVVV-	AI	AVAGTVV																													
092	tr Q7SAB2 Q7SAB2_NEUCR	KG--V---	VD	PTEL	REL	TKGD	KVG-----	AW-ILT	TAFT-	LGT	LGAA	AFV																											
093	tr A2R8R5 A2R8R5_ASPNC	DSSDDK-T	I----	L	ST	ITTGD	RAG-----	AG-IVT	TVVF-	AGGWVGLMA																													
094	tr C5DHG0 C5DHG0_LACTC	STS---	TNV	LQSDIK	ITHK	DKVG-----	AS-IIT	TAVI-	LAVLVGGSI																														
095	tr G0SFA3 G0SFA3_CHATD	PRGAE	E-ER	KRL	LDGE	ITIKDQVA-----	AG-FLT	TS	AI-	VL	GV	VM	GCI																										
096	tr K1WJG5 K1WJG5_MARBU	DDASE-----		I	V	V	I	T	P	A	T	T	A	D	KAG-----	AA-ILT	TAAV-	LAGLVGGVG																					
097	tr H2B005 H2B005_KAZAF	ATAT-----		NLSPLN	VTPG	S	KAA-----	AG-IIT	TAVI-	GISIFACVL																													
098	tr C5DYD7 C5DYD7_ZYGRC	TTT---	TNV	LQKKLK	IQS	KDRAG-----	AA-IIT	TGVV-	LMVVVGGSV																														
099	tr G8YM23 G8YM23_PICSO	VTTA-----		NASPLK	LDGGD	KAG-----	AA-IIT	TTTT-	SAS	LIGSGI																													
100	tr W3X8E3 W3X8E3_PESFW	S---D---	S	FTNS	LTP	ITQGD	RAG-----	AG-ILT	TVLI-	LASAAGTFG																													
101	tr A0A254U3K7 A0A254U3K7_ASPNG	DSSDDK-T	I----	L	ST	ITTGD	RAG-----	AG-IVT	TVVF-	AGGWVGLMA																													
102	tr Q6CER8 Q6CER8_YARLI	GDNPFGTSQD		DASPLD	I	HGGD	RAG-----	AG-IIT	TAAV-	GIMLILVFW																													
103	tr G2QT05 G2QT05_THITE	S---D---	T	FQNT	LAP	PTA	AD	KAG-----	AS-ILT	TILI-	LCS	AV	GTFG																										
104	sp Q5ACZ2 DFG5_CANAL	SEDN-----		ANKNEL	TITG	KDKAG-----	AG-VLT	TAIV-	LAVILGGAI																														
105	tr G2WKU6 G2WKU6_YEASK	SST---	TNV	LQNNLN	IKKGD	RAG-----	AA-IIT	TAVI-	LSVLTGGAV																														
106	tr A0A0C7N752 A0A0C7N752_9SACH	KST---	TNV	LHSNLK	ITHK	DKVG-----	AA-ILT	TAVV-	IAILVGGSI																														
107	tr W3WN68 W3WN68_PESFW	DS-----	SK	IATARA	IT	TGD	RAG-----	AG-ILT	TALV-	LASLLSSLV																													
108	tr G2QFS1 G2QFS1_MYCTT	SRGP-----		SEPLAP	ITTAD	RAG-----	AG-ILT	TMLI-	LGGFI	GTWS																													
109	tr G3J9G4 G3J9G4_CORMM	QA-----	Q	P	LPEP	RAIT	AG	DRAG-----	AA-ILT	TILI-	AAGILAGSF																												
110	tr Q2UJ03 Q2UJ03_ASPOR	SGGP---	R-P	HSRR	TD	TSTAN	RAG-----	AG-ILT	TILM-	AMLIF	YTVW																												
111	tr G8YRT2 G8YRT2_PICSO	THET-----		ANRN	LLK	ISGK	DKAG-----	AG-VLT	TAVI-	LAI	VL	GCAV																											
112	tr W3X554 W3X554_PESFW	TE-----		D	K	M	P	V	L	E	P	I	T	L	K	E	K	V	A-----	AG-FIT	SAL-	ALSVLGGSI																	
113	tr Q1K7A8 Q1K7A8_NEUCR	DPAS---	R	G	G	L	A	A	L	K	P	I	T	M	A	D	R	V	G-----	AG-IVT	AIL-	AISIVGGSV																	
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	SEQ-AA-P	Q----	P	Q	P	V	T	T	G	D	RAG-----	AG-IVT	TVVF-	LSGWIAAVT																								
115	tr A7F3P0 A7F3P0_SCLS1	TSTNS----	D	G	T	V	E	K	P	V	T	S	G	D	RAG-----	AG-FVT	TALI-	ISGV	LGGVG																				
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	VA-----		Q	P	LPEP	R	K	L	V	T	A	D	RAG-----	AA-ILT	TMLV-	AAGIVGGSL																						
117	tr Q6CIP9 Q6CIP9_KLULA	VST---	TNV	LQKNIN	ITAGD	KAG-----	AG-IIT	TTIV-	LGVLV	LGTV																													
118	tr H2AMQ5 H2AMQ5_KAZAF	TTAT-----		NLAPLN	ITGG	S	KAA-----	AA-IIT	TAVI-	GISLI	ACTL																												
119	tr G8JMI3 G8JMI3_ERECY	TPM---	D-NV	LHSRLN	IGS	K	DRIK-----	AA-LIT	TAVC-	LGLLVGTAL																													
120	tr C5DGT1 C5DGT1_LACTC	TSAT-----		NLSPLD	IT	TGS	R	AA-----	AG-IIT	TAVI-	GLSIVACTV																												
121	tr K1XJR4 K1XJR4_MARBU	S---V---	T	D	L	R	M	L	L	P	P	S	E	Q	D	KAG-----	AA-VIT	TAVW-	VAASVVLFG																				
122	tr A0A124BYC5 A0A124BYC5_ASPNG	TTTAST-A	A	G	S	N	S	V	V	P	T	N	G	A	N	S	L	S	C	G	L	F	G	V	A	L	A	-	I	V	A	G	A	F	-	A	I	A	-----
123	tr A7TLL2 A7TLL2_VANPO	QHPT-----		NLSPLN	ITSG	S	RAG-----	AG-IIT	TAII-	GISII	ACGL																												
124	tr Q0CB86 Q0CB86_ASPTN	DSNSNS-G	N----	L	K	P	I	T	T	G	D	KAG-----	AG-ILT	TVVF-	VGLWGGMVA																								

125	tr K1XLH0 K1XLH0_MARBU	SPEDV----	AALTEMKITSGDRVG-----	AG-ILTALV-ICGVIASG
126	tr J7S438 J7S438_KAZNA	SIPN-----	QLVP IAYHKRVSGR-----	CR-YHHCHHRYFNRIVCIV
127	tr Q4WG09 Q4WG09_ASPFU	DTHEDE---	APDALAPINTGDRAG-----	AW-ILTVII-VVAVGGSVG
128	tr H8WZ09 H8WZ09_CANO9	TEDT-----	TNKNKIDITGKDRAG-----	AG-VLTAIV-LAILLGGGI
129	tr I1RL09 I1RL09_GIBZE	---SH---D-	NPEPTPIT TADRAG-----	AG-ILTFLV-LAGGLGTFV
130	tr Q4W985 Q4W985_ASPFU	QPDP---R-	PPVLTMTITGADRAG-----	AG-LLTAML-GVLMIGTTG
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	SST-----	TSVLQNNLNITKGDRAG-----	AA-IITAVI-LSVLIGGAV
132	tr W6QCW3 W6QCW3_PENRF	PPKA---I-	PPGLDYDITSGDKAG-----	AG-ILTVLF-MLAIGGSTG
133	tr W3WMD3 W3WMD3_PESFW	E-----	STLTPLAEIT TADRAG-----	AG-ILTCII-LASSLGAF
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	QAAT-----	NLNPLHITAGSRAG-----	AG-IITCVV-GISIIASLL
135	tr I2GYH1 I2GYH1_TETBL	KTS-----	KNVLSNDLDIKTGDKVG-----	AA-ILTAGV-LAILVGGA
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	SST-----	TNVLQNKLSINTGDRAG-----	AA-VVTAVV-LMIVVGGSV
137	tr Q752P3 Q752P3_ASHGO	HDS---QPA	ILRGMHNITGKDRVG-----	AS-LIT AIS-LGLMLGCAL
138	tr B8MYP3 B8MYP3_ASPFN	HKDQQT-GT-	---PKPIT TADRAG-----	AS-IVTFFF-ACGWMASVS
139	tr Q2TXL6 Q2TXL6_ASPOR	SRKPEK-DK-	---PREIT TADRAG-----	AS-IATVVV-VGIWLGIAA
140	tr Q0CN17 Q0CN17_ASPTN	TGEP PA-SE-	---PAPIT TADRAG-----	AS-ILTILF-ISGWVAAVT
141	tr J3NHD5 J3NHD5_GAGT3	S---T---PH-	QTFAPITGGDRAG-----	AA-FLTIAV-LGGAAFMFG
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	AKTAA----	SLDDWTPTTGDKAG-----	AG-IVTAVI-LAGVIGGVA
143	tr A1CVB0 A1CVB0_NEOFI	QHDP---R-	PQVLTMTITGADRAG-----	AG-LLTAML-GVLMIGTTG
144	tr B8NVI3 B8NVI3_ASPFN	SRKPEK-DK-	---PREIT TADRAG-----	AS-IATVVV-VGIWLGIAA
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	TQAT-----	NLSPLTIT TGSRAA-----	AG-IITAVI-GLSIVACTV
146	tr A1DJ54 A1DJ54_NEOFI	DTHEDA---	APDALDPINTGDRAG-----	AW-ILTVII-VVAVGGSVG
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	SRHA-----	TEPAKPIT TADKAG-----	AA-MCTILL-IAGGIAIWI
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	QSQT-----	NLSPLTIDKGDEAG-----	AA-IITVVV-GATIVGAFA
149	tr A0A177A618 A0A177A618_9PEZI	TAAKVR----	---GSIEVT TAGRAA-----	AG-MLTAGM-CALVGVA VW
150	tr Q75CW6 Q75CW6_ASHGO	DNSP--N-II	TLSTRQIGLKDRVG-----	AG-FMTVVL-LIPFITGSI
151	tr Q7S4K4 Q7S4K4_NEUCR	SRHA-----	TEPAKPIT TADKAG-----	AA-MCTILL-IAGGIAIWI
152	tr G2QB99 G2QB99_MYCTT	PAHEE-----	VGIGYAEITLQDRVA-----	AG-FVTSAL-ALGVVAGSA
153	tr G0S3F2 G0S3F2_CHATD	SRGA-----	AQHFREINAGDRVG-----	AA-ILTMLI-LGGA IATWS
154	tr A0A1E4SC85 A0A1E4SC85_9ASCO	VTNT-----	NASPLKLGAGDTAG-----	AG-IITAI I-GVSI VASGV
155	tr A0A0W0CYJ3 A0A0W0CYJ3_CANGB	STT-----	TNVLKNNLKITGGDRAG-----	AA-IVTTIV-LGIIIGGAA
156	tr A0A0J5PQ80 A0A0J5PQ80_ASPFM	DKSGQT-TP-	---TRKITMGDKAG-----	AS-ILTIGL-VVGWVSLIA
157	tr G2QKJ0 G2QKJ0_MYCTT	----F---EG	VSPPREITAGDRAG-----	AG-ILTAVV-LASFLGSLV
158	tr Q4WFX5 Q4WFX5_ASPFU	SSERIA---V	SDDSSI VTMADRVV-----	AW-VATGGL-GVLLGGYLY
159	tr Q5BGD7 Q5BGD7_EMENI	DTYNDD---	TPDALPHIT TADRAG-----	SW-ILTVLW-GSGIVAAGW
160	tr A0A0A8L8U3 A0A0A8L8U3_9SACH	QSPT-----	NLAPLTITKG DQAG-----	AG-IITAVI-GITIIGSVV
161	tr G8BYN9 G8BYN9_TETPH	TTS-----	TNVLQNNLDIESKDKAG-----	AA-VATAVI-LAVLVGGSV
162	tr F7VVT4 F7VVT4_SORMK	----P---AT	LLSMSELTGKDKAG-----	AG-VLTA FV-VCSILGT YF
163	tr G8YB80 G8YB80_PICSO	VTTE-----	NASPLKLDGGDKAG-----	AA-IITIVI-SASLIGSGI
164	tr A0A0H5CB47 A0A0H5CB47_CYBJA	TTRN-----	ELTYS DMNIDSGDVAG-----	AA-IVTAVV-LLLAVGGAI
165	tr G2WRS7 G2WRS7_VERDV	S---Q---DI	MDHHKPIKMGDRAG-----	AG-ILTFMV-LATALSTWA
166	tr A0A124BVB5 A0A124BVB5_ASPNG	TSDSSR---D	PSELRPIT TSDRIG-----	AG-VVTGVT-ACLVLGVTA
167	tr A7EIG7 A7EIG7_SCLS1	KSSSS-----	FSSTKPVT TADKVG-----	AG-ILTTVL-LLAFIGSIW
168	tr C5M5U7 C5M5U7_CANTT	TPIT-----	DARPLSLDSGDKAG-----	AG-ILTAVI-GATLVGTCI
169	tr B9WAA8 B9WAA8_CANDC	SEDN-----	ANKNELTITGKDKAG-----	AG-VLTAIV-LAVILGGAI
170	tr F8MRU1 F8MRU1_NEUT8	SRHA-----	TEPAKPIT TADKAG-----	AA-MCTILL-IAGGIAIWI
171	tr A1DJS0 A1DJS0_NEOFI	DKSGQT-TP-	---TRKIT TADRAG-----	AS-ILTIGF-AVGWVGLMA
172	tr B6HLM8 B6HLM8_PENRW	GRSGSS-----	---ALPPIT TADKAG-----	AG-ILAVVF-VGAMVGGVV
173	tr A0A117DWS0 A0A117DWS0_ASPNG	PTDAPN-KL-	---P-DIT TADRAG-----	AS-ILTVLF-VGGWAGAIT
174	tr A0A124BXM1 A0A124BXM1_ASPNG	DNTD-S-GST	---ESKIT TADRAG-----	AS-ILTIAF-VGMWGGMIA
175	tr J3NZQ6 J3NZQ6_GAGT3	----H---TP	ATGYSDIT TADRAG-----	AA-ILTTVI-IGLALGTFG
176	tr W0THH8 W0THH8_KLUMD	VST-----	TNVLQKKLTITGSDKAG-----	AG-VITAVV-IGVLVLGSV
177	tr B2AEF2 B2AEF2_PODAN	SRGP-----	SEPLAPITNGDRAG-----	AA-ILTILI-LGSAVGSWA
178	tr Q96TX1 Q96TX1_NEUCS	S---D---SFT	-HDKPVT TADKAG-----	AG-IVTFLL-AASTLGTFV
179	tr E9DYA6 E9DYA6_METAQ	D---G---SV	MPNQKPV TAGDKVG-----	AS-IVTILL-LGGACGMFG
180	tr B9WAI2 B9WAI2_CANDC	KLTS-----	DATPLSIDGGDRAG-----	AG-IITAI I-GASLVGSCV
181	tr Q6C171 Q6C171_YARLI	GFHPFGE--D	EAHPLNIGGGDRAG-----	AA-IITVIV-GVMMIAAGW
182	tr A6ZMV1 A6ZMV1_YEAS7	SST-----	TNVLQNNLNIKKGDRAG-----	AA-IITAVI-LSVLTGGAV
183	tr B6K7X7 B6K7X7_SCHJY	SASS-----	TVVISPATTKDKHW-----	SR-FFTALL-VFLLLSCSI
184	tr I1RSA2 I1RSA2_GIBZE	DAND---NDG	TPKRKAITGADKAG-----	AG-ILTFLF-VAGIIGGVS
185	tr F7VWZ9 F7VWZ9_SORMK	DPAS---RGG	LAALKPIT TADRVG-----	AG-IVTAIL-AISIVGGSV
186	tr A0A1B2J789 A0A1B2J789_PICPA	TKTE-----	AVPPLKLTNADVAG-----	AA-IITAI I-GLSVIAGAI
187	tr G0V8N4 G0V8N4_NAUCC	QEAT-----	NLAPLHITDGSKAA-----	AA-IITSVV-GISIVAAGV
188	tr H8WXN8 H8WXN8_CANO9	TTAT-----	NAKPLSLDSGDKAG-----	AG-IITAI I-GASLVGCGV
189	tr A6ZZS0 A6ZZS0_YEAS7	AQPT-----	NLAPLNITKGSKAG-----	AG-IITAVI-GISIVACAL
190	tr G9MQD1 G9MQD1_HYPVG	---GD---NG	EVELKPIGAADKAG-----	AV-IVTLIL-LGSATAAFI
191	tr A0A1G4JVK5 A0A1G4JVK5_9SACH	TQAS-----	NLSPLTITAGSRAA-----	AG-IITAVI-GLSIVACTV
192	tr A0A0J5PNX6 A0A0J5PNX6_ASPFM	DTHEDE---	APDALAPINTGDRAG-----	AW-ILTVII-VVAVGGSVG
193	tr A0A254UET1 A0A254UET1_ASPNG	ASDSSR---D	PSELRPIT TSDRIG-----	AG-VVTGVT-ACLVLGVTA
194	tr A1D587 A1D587_NEOFI	TEQ-AA-PQ-	---PQPVTAGDRAG-----	AG-ILTVVF-LSGWIAAVT

001	tr A0A254U2J9 A0A254U2J9_ASPNG	WMVIGG	-----
002	sp Q6FLP9 DCW1_CANGA	WLVY--	-----
003	tr A1CCM5 A1CCM5_ASPCL	FMVIGG	-----
004	tr C1H190 C1H190_PARBA	WMMA--	-----
005	tr Q2UR85 Q2UR85_ASPOR	WMVYGG	-----
006	tr A5DV30 A5DV30_LODEL	WMIF--	-----
007	tr A0A1E3NP59 A0A1E3NP59_9ASCO	-----	-----
008	tr J3PGN0 J3PGN0_GAGT3	WMSWDR	-----
009	tr A0A124BXU6 A0A124BXU6_ASPNG	WLMLG-	-----
010	tr C5P4A1 C5P4A1_COC7	FSVS--	-----
011	tr Q0CG55 Q0CG55_ASPTN	WVLYE-	-----

012	tr C4Y2X5 C4Y2X5_CLAL4	WLVL--
013	tr A0A254TKQ9 A0A254TKQ9_ASPNG	WMVYE-
014	tr C7Z068 C7Z068_NECH7	WMCADF-
015	tr G8BXX2 G8BXX2_TETPH	WLVI--
016	tr A0A0L0P6S8 A0A0L0P6S8_CANAR	WLVI--
017	tr A1CSC2 A1CSC2_ASPCL	WMVYGR-
018	sp Q05031 DFG5_YEAST	WMLF--
019	tr H2AQJ6 H2AQJ6_KAZAF	WMLF--
020	tr G2XFH9 G2XFH9_VERDV	WMSFWD-
021	tr A0A1S7HQM3 A0A1S7HQM3_9SACH	WLIL--
022	sp Q5AD78 DCW1_CANAL	WLIL--
023	tr B6H7E8 B6H7E8_PENRW	WMLVG GK-
024	tr B6GZU2 B6GZU2_PENRW	WLIWD-
025	tr E9E4W7 E9E4W7_METAQ	WMSFFDPVVP-
026	tr A0A1S9RH77 A0A1S9RH77_9EURO	WLIWD-
027	tr G2R7G8 G2R7G8_THITE	WMGMGWSEGS-
028	tr G3JBY1 G3JBY1_CORMM	FLMSDLYD-
029	tr A5DNV5 A5DNV5_PICGU	WLLL--
030	tr A0A1B8GPA6 A0A1B8GPA6_9PEZI	WIVC--
031	tr Q2TWC1 Q2TWC1_ASPOR	WLIK TQ-
032	tr J5RXY9 J5RXY9_SACK1	WLVF--
033	tr A0A2N6NN14 A0A2N6NN14_BEABA	FVTSNVFSYA-
034	tr A0A254U5V1 A0A254U5V1_ASPNG	WLVYGG-
035	tr A0A061B6H7 A0A061B6H7_CYBFA	WLLL--
036	tr G8ZQ93 G8ZQ93_TORDC	WLIL--
037	tr Q9C2J1 Q9C2J1_NEUCS	FLT I-
038	tr C5NZK5 C5NZK5_COCP7	WMVS--
039	tr Q6C0T7 Q6C0T7_YARLI	WLLTDGMYKGFKI-----KRRDPSE-----
040	tr A0A0W0ENZ4 A0A0W0ENZ4_CANGB	WLVY--
041	tr F7VZ72 F7VZ72_SORMK	WMGLG K-
042	tr A3LN37 A3LN37_PICST	WLVI--
043	tr Q6CAI2 Q6CAI2_YARLI	WMLA--
044	tr A0A0V1PWA0 A0A0V1PWA0_9ASCO	WLIL--
045	tr G9MJS5 G9MJS5_HYPVG	WMSLG P-
046	tr G9MHI4 G9MHI4_HYPVG	VLLL--
047	tr D4AP27 D4AP27_ARTBC	WMSV--
048	tr B6GZT8 B6GZT8_PENRW	FMMLG G-
049	sp O74556 YCZ2_SCHPO	VLY--
050	tr A0A0C4E991 A0A0C4E991_MAGP6	WMSWDR-
051	tr B8MHG0 B8MHG0_TALSN	WLVIGGAN-
052	tr A3LMV8 A3LMV8_PICST	WMIF--
053	tr A0A099P0Y5 A0A099P0Y5_PICKU	SA--
054	sp Q9P6I3 YHG7_SCHPO	WLYF--
055	tr Q2US57 Q2US57_ASPOR	LLIFT-
056	tr W7N622 W7N622_GIBM7	FATI--
057	sp Q9P6I4 YHG6_SCHPO	WALVEDEEGKIPSRGKKGIAIS-S-
058	tr C7GRB3 C7GRB3 YEAS2	WMLF--
059	tr C7GP28 C7GP28 YEAS2	WLVF--
060	tr G3JGZ5 G3JGZ5_CORMM	WMGF MV-
061	tr C4QXV4 C4QXV4_KOMPG	WLLL--
062	tr I1S0Z5 I1S0Z5_GIBZE	WMSLG K-
063	tr A0A0J5PSQ0 A0A0J5PSQ0_ASPFM	WLLYE-
064	tr G0W5X4 G0W5X4_NAUDC	WIIL--
065	tr G8ZPC1 G8ZPC1_TORDC	WMLF--
066	tr A0A167CDC5 A0A167CDC5_9ASCO	WMMKQ-
067	tr R9XAT7 R9XAT7_ASHAC	WLI I-
068	tr B2B747 B2B747_PODAN	WMSTGV-
069	tr G0SE99 G0SE99_CHATD	WMSMGE-
070	tr J8Q6F0 J8Q6F0_SACAR	WMLF--
071	tr A0A0A8LDJ2 A0A0A8LDJ2_9SACH	WMI Y-
072	tr J3K6E2 J3K6E2_COCIM	FSVS--
073	tr G4N3E1 G4N3E1_MAGO7	WMSWDS R-
074	tr C5M5J2 C5M5J2_CANTT	WMMF--
075	tr A0A254U0X0 A0A254U0X0_ASPNG	WLVKAQ-
076	sp Q75DG6 DCW1_ASHGO	WLI I-
077	tr Q6BZF0 Q6BZF0_DEBHA	WLIL--
078	tr E9E3Q1 E9E3Q1_METAQ	WMSAFD-
079	sp P36091 DCW1_YEAST	WLVF--
080	tr W0TAH6 W0TAH6_KLUMD	WLVL--

081	tr A0A0H5CF34 A0A0H5CF34_CYBJA	WMVI--
082	tr A0A2C5WNS2 A0A2C5WNS2_9PEZI	FVFYESM-
083	tr Q2TYU3 Q2TYU3_ASPOR	FMVLTG-
084	tr G2XGF6 G2XGF6_VERDV	FVMKEETK-
085	tr G3AMT4 G3AMT4_SPAPN	WLIL--
086	tr A0A1C1D213 A0A1C1D213_9EURO	WIIL--
087	tr A0A0B0DST1 A0A0B0DST1_NEUCS	LMASGSFEKLEGGATPATAAAGEKKKKKEKKEKLSIWYKIARESNNFFRHQG
088	tr A0A100I5A6 A0A100I5A6_ASPNG	WLVKAQ-
089	tr A0A1G4JM92 A0A1G4JM92_9SACH	WMVF--
090	tr G2Q8A7 G2Q8A7_MYCTT	WMSTGV-
091	tr B8NPZ1 B8NPZ1_ASPFN	WMIVP-
092	tr Q7SAB2 Q7SAB2_NEUCR	LMASGSFEKLEGGATPATAAAGEKKKKKEKKEKLSIWYKIARESNNFFRHQG
093	tr A2R8R5 A2R8R5_ASPNC	WLMLG-
094	tr C5DHG0 C5DHG0_LACTC	WMIF--
095	tr G0SFA3 G0SFA3_CHATD	FVII--
096	tr K1WJG5 K1WJG5_MARBU	CLVLG-
097	tr H2B005 H2B005_KAZAF	WLIY--
098	tr C5DYD7 C5DYD7_ZYGRC	WMLF--
099	tr G8YM23 G8YM23_PICSO	WLIL--
100	tr W3X8E3 W3X8E3_PESFW	WMSLGM-
101	tr A0A254U3K7 A0A254U3K7_ASPNG	WLMLG-
102	tr Q6CER8 Q6CER8_YARLI	FLIV--
103	tr G2QT05 G2QT05_THITE	WMSVGE-
104	sp Q5ACZ2 DFG5_CANAL	WMIF--
105	tr G2WKU6 G2WKU6 YEASK	WMLF--
106	tr A0A0C7N752 A0A0C7N752_9SACH	WMMF--
107	tr W3WN68 W3WN68_PESFW	LMIKD-
108	tr G2QFS1 G2QFS1_MYCTT	WMSIGD-
109	tr G3J9G4 G3J9G4_CORMM	FVTSDVLSFA-
110	tr Q2UJ03 Q2UJ03_ASPOR	WGINE-
111	tr G8YRT2 G8YRT2_PICSO	WMAI--
112	tr W3X554 W3X554_PESFW	FVMK--
113	tr Q1K7A8 Q1K7A8_NEUCR	FLT I--
114	tr A0A0J5PM92 A0A0J5PM92_ASPFM	WMVYGR-
115	tr A7F3P0 A7F3P0_SCLS1	FMVIG-
116	tr A0A2N6NK60 A0A2N6NK60_BEABA	FLMSNLF D-
117	tr Q6CIP9 Q6CIP9_KLULA	WMVL--
118	tr H2AMQ5 H2AMQ5_KAZAF	WIIF--
119	tr G8JMI3 G8JMI3_ERECY	WMVI--
120	tr C5DGT1 C5DGT1_LACTC	WLVL--
121	tr K1XJR4 K1XJR4_MARBU	WMSVGES-
122	tr A0A124BYC5 A0A124BYC5_ASPNG	- - - - -
123	tr A7TLL2 A7TLL2_VANPO	WLIV--
124	tr Q0CB86 Q0CB86_ASPTN	WLF IG G-
125	tr K1XLH0 K1XLH0_MARBU	FMIIGS-
126	tr J7S438 J7S438_KAZNA	ASVLRKQE-
127	tr Q4WG09 Q4WG09_ASPFU	WLIK T-
128	tr H8WZ09 H8WZ09_CANO9	WMIF--
129	tr I1RL09 I1RL09_GIBZE	WMCAFD-
130	tr Q4W985 Q4W985_ASPFU	WLLYE-
131	tr A0A0L8RDQ3 A0A0L8RDQ3_SACEU	WMLF--
132	tr W6QCW3 W6QCW3_PENRF	WLIWD-
133	tr W3WMD3 W3WMD3_PESFW	WMNLET M-
134	tr A0A1S7HZQ4 A0A1S7HZQ4_9SACH	WLIL--
135	tr I2GYH1 I2GYH1_TETBL	WMLF--
136	tr A0A1S7HIA4 A0A1S7HIA4_9SACH	WMLF--
137	tr Q752P3 Q752P3_ASHGO	WMIF--
138	tr B8MYP3 B8MYP3_ASPFN	WMVYGG-
139	tr Q2TXL6 Q2TXL6_ASPOR	FMVTGG-
140	tr Q0CN17 Q0CN17_ASPTN	WLVYGG-
141	tr J3NHD5 J3NHD5_GAGT3	WMNKGD-
142	tr A0A1B8GPI3 A0A1B8GPI3_9PEZI	WMSMAD-
143	tr A1CVB0 A1CVB0_NEOFI	WLLYE-
144	tr B8NVI3 B8NVI3_ASPFN	FMVTGG-
145	tr A0A0C7N2N9 A0A0C7N2N9_9SACH	WLVI--
146	tr A1DJ54 A1DJ54_NEOFI	WLIK T-
147	tr A0A0B0DFT3 A0A0B0DFT3_NEUCS	FMNLGD-
148	tr A0A0H5C0E8 A0A0H5C0E8_CYBJA	WTVL--
149	tr A0A177A618 A0A177A618_9PEZI	FMVGSDGWVA-
150	tr Q75CW6 Q75CW6_ASHGO	WMSI--

[illegible]

038 tr|C5NZK5|C5NZK5_COCP7
039 tr|Q6C0T7|Q6C0T7_YARLI
040 tr|A0A0W0ENZ4|A0A0W0ENZ4_CANGB
041 tr|F7VZ72|F7VZ72_SORMK
042 tr|A3LN37|A3LN37_PICST
043 tr|Q6CAI2|Q6CAI2_YARLI
044 tr|A0A0V1PWA0|A0A0V1PWA0_9ASCO
045 tr|G9MJS5|G9MJS5_HYPVG
046 tr|G9MHI4|G9MHI4_HYPVG
047 tr|D4AP27|D4AP27_ARTBC
048 tr|B6GZT8|B6GZT8_PENRW
049 sp|O74556|YCZ2_SCHPO
050 tr|A0A0C4E991|A0A0C4E991_MAGP6
051 tr|B8MHG0|B8MHG0_TALSN
052 tr|A3LMV8|A3LMV8_PICST
053 tr|A0A099P0Y5|A0A099P0Y5_PICKU
054 sp|Q9P6I3|YHG7_SCHPO
055 tr|Q2US57|Q2US57_ASPOR
056 tr|W7N622|W7N622_GIBM7
057 sp|Q9P6I4|YHG6_SCHPO
058 tr|C7GRB3|C7GRB3_YEAS2
059 tr|C7GP28|C7GP28_YEAS2
060 tr|G3JGZ5|G3JGZ5_CORMM
061 tr|C4QXV4|C4QXV4_KOMPG
062 tr|I1S0Z5|I1S0Z5_GIBZE
063 tr|A0A0J5PSQ0|A0A0J5PSQ0_ASPFM
064 tr|G0W5X4|G0W5X4_NAUDC
065 tr|G8ZPC1|G8ZPC1_TORDC
066 tr|A0A167CDC5|A0A167CDC5_9ASCO
067 tr|R9XAT7|R9XAT7_ASHAC
068 tr|B2B747|B2B747_PODAN
069 tr|G0SE99|G0SE99_CHATD
070 tr|J8Q6F0|J8Q6F0_SACAR
071 tr|A0A0A8LDJ2|A0A0A8LDJ2_9SACH
072 tr|J3K6E2|J3K6E2_COCIM
073 tr|G4N3E1|G4N3E1_MAGO7
074 tr|C5M5J2|C5M5J2_CANTT
075 tr|A0A254U0X0|A0A254U0X0_ASPNG
076 sp|Q75DG6|DCW1_ASHGO
077 tr|Q6BZF0|Q6BZF0_DEBHA
078 tr|E9E3Q1|E9E3Q1_METAQ
079 sp|P36091|DCW1_YEAST
080 tr|W0TAH6|W0TAH6_KLUMD
081 tr|A0A0H5CF34|A0A0H5CF34_CYBJA
082 tr|A0A2C5WNS2|A0A2C5WNS2_9PEZI
083 tr|Q2TYU3|Q2TYU3_ASPOR
084 tr|G2XGF6|G2XGF6_VERDV
085 tr|G3AMT4|G3AMT4_SPAPN
086 tr|A0A1C1D213|A0A1C1D213_9EURO
087 tr|A0A0B0DST1|A0A0B0DST1_NEUCS
088 tr|A0A100I5A6|A0A100I5A6_ASPNG
089 tr|A0A1G4JM92|A0A1G4JM92_9SACH
090 tr|G2Q8A7|G2Q8A7_MYCTT
091 tr|B8NPZ1|B8NPZ1_ASPFN
092 tr|Q7SAB2|Q7SAB2_NEUCR
093 tr|A2R8R5|A2R8R5_ASPNC
094 tr|C5DHG0|C5DHG0_LACTC
095 tr|G0SFA3|G0SFA3_CHATD
096 tr|K1WJG5|K1WJG5_MARBU
097 tr|H2B005|H2B005_KAZAF
098 tr|C5DYD7|C5DYD7_ZYGRC
099 tr|G8YM23|G8YM23_PICSO
100 tr|W3X8E3|W3X8E3_PESFW
101 tr|A0A254U3K7|A0A254U3K7_ASPNG
102 tr|Q6CER8|Q6CER8_YARLI
103 tr|G2QT05|G2QT05_THITE
104 sp|Q5ACZ2|DFG5_CANAL
105 tr|G2WKU6|G2WKU6_YEASK
106 tr|A0A0C7N752|A0A0C7N752_9SACH

107 tr|W3WN68|W3WN68_PESFW

108 tr|G2QFS1|G2QFS1_MYCTT
109 tr|G3J9G4|G3J9G4_CORMM
110 tr|Q2UJ03|Q2UJ03_ASPOR
111 tr|G8YRT2|G8YRT2_PICSO
112 tr|W3X554|W3X554_PESFW
113 tr|Q1K7A8|Q1K7A8_NEUCR
114 tr|A0A0J5PM92|A0A0J5PM92_ASPFM
115 tr|A7F3P0|A7F3P0_SCLS1
116 tr|A0A2N6NK60|A0A2N6NK60_BEABA
117 tr|Q6CIP9|Q6CIP9_KLULA
118 tr|H2AMQ5|H2AMQ5_KAZAF
119 tr|G8JMI3|G8JMI3_ERECY
120 tr|C5DGT1|C5DGT1_LACTC
121 tr|K1XJR4|K1XJR4_MARBU
122 tr|A0A124BYC5|A0A124BYC5_ASPNG
123 tr|A7TLL2|A7TLL2_VANPO
124 tr|Q0CB86|Q0CB86_ASPTN
125 tr|K1XLH0|K1XLH0_MARBU
126 tr|J7S438|J7S438_KAZNA
127 tr|Q4WG09|Q4WG09_ASPFU
128 tr|H8WZ09|H8WZ09_CANO9
129 tr|I1RL09|I1RL09_GIBZE
130 tr|Q4W985|Q4W985_ASPFU
131 tr|A0A0L8RDQ3|A0A0L8RDQ3_SACEU
132 tr|W6QCW3|W6QCW3_PENRF
133 tr|W3WMD3|W3WMD3_PESFW
134 tr|A0A1S7HZQ4|A0A1S7HZQ4_9SACH
135 tr|I2GYH1|I2GYH1_TETBL
136 tr|A0A1S7HIA4|A0A1S7HIA4_9SACH
137 tr|Q752P3|Q752P3_ASHGO
138 tr|B8MYP3|B8MYP3_ASPFN
139 tr|Q2TXL6|Q2TXL6_ASPOR
140 tr|Q0CN17|Q0CN17_ASPTN
141 tr|J3NHD5|J3NHD5_GAGT3
142 tr|A0A1B8GPI3|A0A1B8GPI3_9PEZI
143 tr|A1CVB0|A1CVB0_NEOFI
144 tr|B8NVI3|B8NVI3_ASPFN
145 tr|A0A0C7N2N9|A0A0C7N2N9_9SACH
146 tr|A1DJ54|A1DJ54_NEOFI
147 tr|A0A0B0DFT3|A0A0B0DFT3_NEUCS
148 tr|A0A0H5C0E8|A0A0H5C0E8_CYBJA
149 tr|A0A177A618|A0A177A618_9PEZI
150 tr|Q75CW6|Q75CW6_ASHGO
151 tr|Q7S4K4|Q7S4K4_NEUCR
152 tr|G2QB99|G2QB99_MYCTT
153 tr|G0S3F2|G0S3F2_CHATD
154 tr|A0A1E4SC85|A0A1E4SC85_9ASCO
155 tr|A0A0W0CYJ3|A0A0W0CYJ3_CANGB
156 tr|A0A0J5PQ80|A0A0J5PQ80_ASPFM
157 tr|G2QKJ0|G2QKJ0_MYCTT
158 tr|Q4WFX5|Q4WFX5_ASPFU
159 tr|Q5BGD7|Q5BGD7_EMENI
160 tr|A0A0A8L8U3|A0A0A8L8U3_9SACH
161 tr|G8BYN9|G8BYN9_TETPH
162 tr|F7VVT4|F7VVT4_SORMK
163 tr|G8YB80|G8YB80_PICSO
164 tr|A0A0H5CB47|A0A0H5CB47_CYBJA
165 tr|G2WRS7|G2WRS7_VERDV
166 tr|A0A124BVB5|A0A124BVB5_ASPNG
167 tr|A7EIG7|A7EIG7_SCLS1
168 tr|C5M5U7|C5M5U7_CANTT
169 tr|B9WAA8|B9WAA8_CANDC
170 tr|F8MRU1|F8MRU1_NEUT8
171 tr|A1DJS0|A1DJS0_NEOFI
172 tr|B6HLM8|B6HLM8_PENRW
173 tr|A0A117DWS0|A0A117DWS0_ASPNG
174 tr|A0A124BXM1|A0A124BXM1_ASPNG
175 tr|J3NZQ6|J3NZQ6_GAGT3
176 tr|W0THH8|W0THH8_KLUMD


177 tr|B2AEF2|B2AEF2_PODAN

178 tr|Q96TX1|Q96TX1_NEUCS
179 tr|E9DYA6|E9DYA6_METAQ
180 tr|B9WAI2|B9WAI2_CANDC
181 tr|Q6C171|Q6C171_YARLI
182 tr|A6ZMV1|A6ZMV1_YEAS7
183 tr|B6K7X7|B6K7X7_SCHJY
184 tr|I1RSA2|I1RSA2_GIBZE
185 tr|F7VWZ9|F7VWZ9_SORMK
186 tr|A0A1B2J789|A0A1B2J789_PICPA
187 tr|G0V8N4|G0V8N4_NAUCC
188 tr|H8WXN8|H8WXN8_CANO9
189 tr|A6ZZS0|A6ZZS0_YEAS7
190 tr|G9MQD1|G9MQD1_HYPVG
191 tr|A0A1G4JVK5|A0A1G4JVK5_9SACH
192 tr|A0A0J5PNX6|A0A0J5PNX6_ASPFM
193 tr|A0A254UET1|A0A254UET1_ASPNG
194 tr|A1D587|A1D587_NEOFI
195 tr|Q1K7I4|Q1K7I4_NEUCR
196 tr|G8JS88|G8JS88_ERECY
197 tr|A0A136J7D3|A0A136J7D3_9PEZI
198 tr|E9E4X8|E9E4X8_METAQ
199 tr|Q6CP42|Q6CP42_KLULA
200 tr|G0WHL4|G0WHL4_NAUDC
201 tr|A0A0L8RFQ5|A0A0L8RFQ5_SACEU
202 tr|F9WYX6|F9WYX6_ZYMTI
203 tr|A0A0C4DZP6|A0A0C4DZP6_MAGP6
204 tr|J3P147|J3P147_GAGT3
205 tr|I2H842|I2H842_TETBL
206 tr|S6E3R3|S6E3R3_ZYGB2
207 tr|E5AD94|E5AD94_LEPMJ
208 tr|Q6FJM7|Q6FJM7_CANGA
209 tr|A5DUW0|A5DUW0_LODEL
210 tr|G8BF46|G8BF46_CANPC
211 tr|W3XAF6|W3XAF6_PESFW
212 tr|B6H7I2|B6H7I2_PENRW
213 tr|Q5ATF9|Q5ATF9_EMENI
214 tr|A0A167CDD3|A0A167CDD3_9ASCO
215 tr|G0RXS3|G0RXS3_CHATD
216 tr|B8NX26|B8NX26_ASPFN
217 tr|A0A0E1RZZ1|A0A0E1RZZ1_COCIM
218 tr|A0A0J5SN29|A0A0J5SN29_ASPFM
219 tr|A1CMM3|A1CMM3_ASPCL
220 tr|B6HU84|B6HU84_PENRW
221 tr|A0A0B0E7F0|A0A0B0E7F0_NEUCS
222 tr|G2WHY6|G2WHY6_YEASK
223 tr|G0W7G8|G0W7G8_NAUDC
224 tr|G4NGV8|G4NGV8_MAGO7
225 tr|W3X855|W3X855_PESFW
226 tr|A0A2N6NY19|A0A2N6NY19_BEABA
227 tr|J8Q2K7|J8Q2K7_SACAR
228 tr|A0A1E4RBW2|A0A1E4RBW2_9ASCO
229 tr|J6EGN8|J6EGN8_SACK1
230 tr|A0A100ISD9|A0A100ISD9_ASPNG
231 tr|G8YQC0|G8YQC0_PICSO
232 tr|F7VVP8|F7VVP8_SORMK
233 tr|A0A0L8VJB2|A0A0L8VJB2_9SACH
234 tr|F8MBP4|F8MBP4_NEUT8
235 tr|A0A254TXZ7|A0A254TXZ7_ASPNG
236 tr|A1CD27|A1CD27_ASPCL
237 tr|A5DHF3|A5DHF3_PICGU
238 tr|D4ATT7|D4ATT7_ARTBC
239 tr|E2PT42|E2PT42_ASPNC
240 tr|C7ZPE5|C7ZPE5_NECH7
241 tr|A0A0A2VM24|A0A0A2VM24_PARBA
242 tr|Q4WKP7|Q4WKP7_ASPFU
243 tr|F8MXT4|F8MXT4_NEUT8
244 tr|A0A177AJ74|A0A177AJ74_9PEZI
245 tr|J7RQR5|J7RQR5_KAZNA

246 tr|Q75CW7|Q75CW7_ASHGO
247 tr|G3AKK4|G3AKK4_SPAPN

248 tr|B2W762|B2W762_PYRTR
249 tr|A0A0L8VLX1|A0A0L8VLX1_9SACH
250 tr|C4XWK1|C4XWK1_CLAL4

1	2	3	4	5	6	7	8	9
Variable		Average			Conserved			

 - Insufficient data - the calculation for this site was performed on less than 10% of the sequences.