

Asymmetric Intramolecular C–H Aminations with Chiral-at-Ruthenium Complexes

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Publications and Poster Presentations

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- Zijun Zhou, Yuqi Tan, Tatsuya Yamahira, Sergei Ivlev, Xiulan Xie, Radostan Riedel, Marcel Hemming, Masanari Kimura, Eric Meggers, Catalytic Enantioselective Ring-Closing C(sp³)–H Amination of Urea Derivatives. *Chem* 2020, *6*, 2024. ("Free-featured Article") ("Most Read") (Highlighted by Bas de Bruin et al. *Chem* 2020, *6*, 1847.)
- Zijun Zhou, Shuming Chen, Yubiao Hong, Erik Winterling, Yuqi Tan, Klaus Harms, K. N. Houk, Eric Meggers, Non-C₂-Symmetric Chiral-at-Ruthenium Catalyst for Highly Efficient Enantioselective Intramolecular C(sp³)–H Amidation. *J. Am. Chem. Soc.* 2019, *141*, 19048.
- Zijun Zhou, Shuming Chen, Jie Qin, Xin Nie, Xingwen Zheng, Klaus Harms, Radostan Riedel, K. N. Houk, Eric Meggers, Catalytic Enantioselective Intramolecular C(sp³)–H Amination of 2-Azidoacetamides. *Angew. Chem. Int. Ed.* 2019, 58, 1088. ("Hot Paper")
- <u>Zijun Zhou</u>, Xin Nie, Klaus Harms, Radostan Riedel, Lilu Zhang, Eric Meggers, Enantioconvergent Photoredox Radical-Radical Coupling Catalyzed by a Chiral-at-Rhodium Complex. *Sci. China Chem.* 2019, *62*, 1512. ("Invited Submission")
- Jie Qin, <u>Zijun Zhou</u>, Tianjiao Cui, Marcel Hemming, Eric Meggers, Enantioselective Intramolecular C–H Amination of Aliphatic Azides by Dual Ruthenium and Phosphine Catalysis. *Chem. Sci.* 2019, 10, 3202.
- Guanghui Wang, <u>Zijun Zhou</u>, Xiang Shen, Sergei Ivlev, Eric Meggers, Asymmetric Catalysis with a Chiral-at-Osmium Complex. *Chem. Commun.* 2020, 56, 7714.

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Abstract

Chiral transition-metal catalysts in which the chirality only originates from a stereogenic metal center have attracted much attention in the past few years as their excellent catalytic performance has been illustrated through diverse applications in catalytic asymmetric reactions, especially enantioselective intramolecular C–H amination reactions. This thesis reports the synthesis of a series of newly modified chiral-at-metal ruthenium catalysts and their applications in challenging enantioselective intramolecular C–H amination reactions.

1) Synthesis of diverse ruthenium-based chiral catalysts with exclusive metal-centered chirality as a catalyst tool box was accomplished. Through modifications of the chelating carbene ligand, the catalyst's property were changed, thus, behaved differently in catalytic asymmetric transformations. The newly modified ruthenium catalysts were used in different catalytic asymmetric intramolecular C–H amination reactions which are reported on chapter 2.1-2.4 of this thesis.

2) An catalytic enantioselective ring-closing C–H amination of 2-azidoacetamides was developed. A chiral-at-metal ruthenium complex served as the catalyst and provided chiral imidazolidin-4-ones in 31-95% yields, with enantioselectivities up to 95% ee, and catalyst loadings down to 0.1 mol% (740 TON). Mechanistic experiments reveal the importance of the amide group presumably by enabling an initial bidentate coordination of the 2-azidoacetamides to the catalyst (Chapter 2.1).

3) An application of a new class of chiral-at-metal ruthenium catalysts for enantioselective C-H amindations was developed. In the catalyst scaffold, ruthenium is cyclometalated by two 7-methyl-1,7-phenanthrolinium heterocycles, resulting in chelating pyridylidene remote N-heterocyclic carbene ligands (rNHCs). The non- C_2 -symmetric chiral-at-ruthenium complexes displayed unprecedented catalytic activity for the intramolecular C–H amidation of 1,4,2-dioxazol-5-ones and provided chiral γ -lactams with up to 98% ee and catalyst loadings down to 0.005 mol% (up to 11200 TON), while the C_2 -symmetric diastereomer favored an undesired Curtius-type rearrangement. DFT calculations elucidated the origins of the superior C–H amidation reactivity displayed by the non- C_2 -symmetric catalysts compared to related C_2 -symmetric counterparts (Chapter 2.2).

4) An enantioselective intramolecular C–H amination of *N*-benzoyloxyureas using a chiral-at-metal ruthenium catalyst was reported, providing chiral 2-imidazolidinones in yields of up to

99% and with up to 99% ee. Catalyst loadings down to 0.05 mol% were feasible. Control experiments were performed which support a stepwise nitrene insertion mechanism through hydrogen atom transfer of a ruthenium nitrenoid intermediate followed by a radical recombination. Chiral imidazolidines are prevalent in bioactive compounds and can be converted to chiral vicinal diamines in a single step. The synthetic value of the new method was demonstrated for the synthesis of intermediates of the drugs levamisole and dexamisole, the bisindole alkaloids topsentine D and spongotine A, and a chiral organocatalyst. (Chapter 2.3).

5) Chiral β -amino alcohols are important building blocks for the synthesis of drugs, natural products, chiral auxiliaries, chiral ligands and chiral organocatalysts. The catalytic asymmetric β -amination of alcohols offers a direct strategy to access this class of synthetic intermediates. In this part, we report a general intramolecular C–H nitrene insertion method for the synthesis of chiral oxazolidin-2-ones as precursors of chiral β -amino alcohols was developed. Specifically, the ring-closing C–H amination of *N*-benzoyloxycarbamates with just 2 mol% of a chiral ruthenium catalyst provided cyclic carbamates in up to 99% yield and with up to 99% ee. The method is applicable to benzylic, allylic, and propargylic C–H bonds and can even be applied to completely non-activated C–H bonds, although with somewhat reduced yields and stereoselectivities. The obtained cyclic carbamates can subsequently be hydrolyzed to obtain chiral β -amino alcohols. The method is highly practical as the catalyst can be easily synthesized in a gram scale and can be recycled after the reaction for further use. The synthetic value of the new method was demonstrated with the asymmetric synthesis of chiral β -amino alcohols that are intermediates for the synthesis of chiral product (-)-aurantiolavine and chiral β -amino alcohols that are intermediates for the synthesis of chiral box-ligand and the natural products hamacanthin A and dragmacidin A (Chapter 2.4).

Zusammenfassung

Chirale Übergangsmetall-Katalysatoren, deren Chiralität exklusiv auf einem stereogenen Metallzentrum basiert, haben in den letzten Jahren viel Aufmerksamkeit bekommen, da ihre hervorragenden katalytischen Eigenschaften durch verschiedene Anwendungen in asymmetrischen Reaktionen, insbesondere enantioselektiven, intramolekularen C–H Aminierungen, gezeigt wurden. Diese Arbeit berichtet über die Synthese einer Reihe neuer *chiral-at-metal* Ruthenium-Katalysatoren sowie deren Anwendung bei herausfordernden, enantioselektiven, intramolekularen C–H Aminierungen.

1) Synthese verschiedener chiraler Ruthenium-Katalysatoren mit exklusiv metallzentrierter Chiralität, die entweder eine Λ - (linkshändiger Propeller) oder Δ - (rechtshändiger Propeller) Konfiguration aufweisen. Durch Modifikationen des chelatisierenden Carbenliganden wurden die elektronischen Eigenschaften des Katalysators verändert und deren Einfluss in asymmetrischen Transformationen untersucht. Die neu modifizierten Ruthenium-Katalysatoren wurden in verschiedenen katalytischen, intramolekularen C–H-Aminierungen verwendet, über die in Kapitel 2.1-2.4 berichtet wurde.

2) Eine enantioselektive, ringschließende C–H-Aminierung von 2-Azidoacetamiden wird durch einen *chiral-at-metal* Rutheniumkomplex katalysiert und liefert chirale Imidazolidin-4-one in 31 bis 95% Ausbeute mit Enantioselektivitäten von bis zu 95% *ee* und Katalysatorbeladungen von bis zu 0.1 mol% (740 TON). Mechanistische Experimente zeigen die Bedeutung der Amid-Gruppe, die vermutlich eine anfängliche bidentate Koordination der 2-Azidoacetamide an den Katalysator ermöglicht (Kapitel 2.1).

3) Eine neue Klasse von chiralen Ruthenium-Katalysatoren wird vorgestellt, bei der Ruthenium durch zwei 7-Methyl-1,7-phenanthrolinium–Heterocyclen cyclometalliert wird, was zu chelatisierenden Pyridyliden-Liganden (entfernte N–Heterocyclische Carben-Liganden (rNHCs)) führt. Diese Arbeit befasst sich mit der Bedeutung der relativen metallzentrierten Stereochemie. Nur die nicht C₂-symmetrischen chiralen Rutheniumkomplexe zeigen eine beispiellose katalytische Aktivität für die intramolekulare C–H-Amidierung von 1,4,2-Dioxazol-5-onen, um chirale γ-Lactame mit einem Enantiomerenverhältnis von bis zu 98% ee bei einer Katalysatorbeladung von bis zu 0.005 mol% (bis-

zu 11200 TON) zu erhalten. Das C₂-symmetrische Diastereomer führt dagegen zu einer unerwünschten Umlagerung vom CURTIUS-Typ. Für diesen Teil wurden zudem DFT-Berechnungen durchgeführt, die eine Erklärung für das überlegenere Reaktionsverhalten des nicht C₂-symmetrischen Katalysators gegenüber den verwandten C₂-symmetrischen Komplexen in der C–H-Amidierung liefern sollen (Kapitel 2.2).

4) Eine enantioselektive intramolekulare C-H-Aminierung von N-Benzoyloxyharnstoff unter Verwendung eines chiral-at-metal Ruthenium-Katalysators wird berichtet, die chirale 2-Imidazolidinone in Ausbeuten von bis zu 99% und mit bis zu 99% ee liefert. Katalysatorbeladungen bis **Z**11 0.05 mol%sind möglich. Kontrollexperimente unterstützen einen schrittweisen Nitren-Insertionsmechanismus durch Wasserstoffatomtransfer eines Rutheniumnitrenoid-Intermediats gefolgt von einer radikalischen Rekombination. Chirale Imidazolidine sind in bioaktiven Verbindungen weit verbreitet und können in einem einzigen Schritt in chirale, vicinale Diamine umgesetzt werden. Die synthetische Bedeutung dieser neuen Methode wird für die Darstellung von Zwischenprodukten der Wirkstoffe Levamisol und Dexamisol, der Bisindolalkaloide Topsentin D und Spongotin A sowie eines chiralen Organokatalysators demonstriert (Kapitel 2.3).

5) Chirale β -Aminoalkohole sind wichtige Bausteine für die Synthese von Arzneimitteln, Naturstoffen, chiralen Auxiliaren, chiralen Liganden und chiralen Organokatalysatoren. Die katalytische, asymmetrische β-Aminierung von Alkoholen bietet eine direkte Strategie für den Zugang zu dieser Klasse synthetischer Zwischenprodukte. In diesem Teil berichten wir über eine allgemeine intramolekulare C-H-Nitren-Insertionsmethode zur Synthese von chiralen Oxazolidin-2-onen als Vorläufer von chiralen β-Aminoalkoholen. Insbesondere liefert die ringschließende C(sp³)–H-Aminierung *N*-Benzoyloxycarbamaten mit nur 2 mol% von eines chiralen Ruthenium-Katalysators cyclische Carbamate mit bis zu 99% Ausbeute und 99% ee. Das Verfahren ist auf benzylische, allylische und propargylische C-H-Bindungen anwendbar und kann sogar bei vollständig unaktivierten C-H-Bindungen verwendet werden, wenn auch mit verringerten Ausbeuten und Stereoselektivitäten. Die erhaltenen cyclischen Carbamate können anschließend hydrolysiert werden, um chirale β-Aminoalkohole zu erhalten. Das Verfahren ist sehr praktisch, da der Katalysator ohne Probleme im Gramm-Maßstab synthetisiert und nach der Reaktion zur weiteren Verwendung wiedergewonnen werden kann. Der synthetische Wert der neuen Methode wird anhand der asymmetrischen Darstellung von chiralem Oxazolidin-2-on als Intermediat für die Synthese des

Naturstoffs (-)-Aurantiolavin und chiraler β -Aminoalkohole demonstriert, die Zwischenprodukte für die Synthese von chiralen Box-Liganden und den Naturstoffen Hamacanthin A und Dragmacidin A sind (Kapitel 2.4).

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Chapter 1: Theoretical Part

1.1 Introduction

The prevalence of chiral amines in pharmaceuticals, natural products, fine chemicals, agrochemicals, and as reagents or chiral ligands of asymmetric catalysts in organic synthesis has motivated the organic chemist for developing mild, efficient and convenient methodology for the construction of chiral C-N bonds. The typical conventional synthetic methods rely on functional group conversions, such as the cross-coupling reaction of aryl- or alkenyl (pseudo)halides with amines under optimal catalytic systems (**Figure 1a**).¹ Recently, the construction of C-N bonds by direct C–H amination, without the preinstallation of any reactive functional group, has attracted much attention. Among them, C–H amination via nitrene insertion is one of the most efficient method for building a chiral C-N bond (**Figure 1b**). With high motivation for making progress in this field, recently, different linear and cyclic chiral amines and amides have been synthesized through this methodology.²

Catalytic asymmetric amination via nitrene insertion:

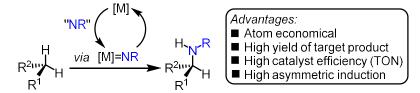


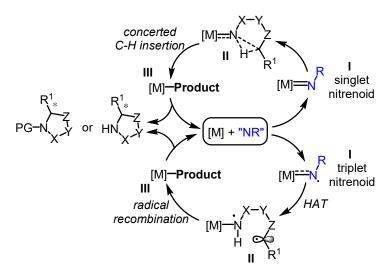
Figure 1. General concept of asymmetric amination via nitrene insertion.

1.2 Asymmetric Intramolecular C-H Aminations

The transition-metal catalyzed ring-closing C–H amination via nitrenoid intermediate is one of the most powerful tool for the synthesis of chiral nitrogen-containing heterocycles. In one mechanistic manifold, intermediate transition metal nitrenoid insert a nitrogen atom between C–H bonds to build a C–N bond in either a stepwise or concerted fashion. As shown in **Figure 2**, nitrene precursor undergo the activation of transition-metal catalyst and generate either singlet or triplet metal-nitrenoid intermediate. Singlet nitrenes undergo a concerted C–H insertion and triplet nitrenes undergo hydrogen atom transfer (HAT) followed by fast radical recombination, which leads to the formation of catalyst bound product. Product release and new substrate participation finishes the whole catalytic cycle. Different functional groups served as nitrene precursors have been developed in the past few decades¹ and the intramolecular version of the reaction renders directing groups obsolete, while exerting high control over the regioselectivity. This methodology has been applied to the catalytic

asymmetric synthesis of cyclic sulfamidates, sulfamides, sulfonamides, carbamates, lactams, ureas, Boc-protected pyrrolidines, and related Boc-protected heterocycles. The following section will briefly review the asymmetric intramolecular ring-closing C–H amination reactions.

Mechanism of intramolecular C-H amination



[M] = Transition-metal catalyst "NR" = Nitrene precursor PG = Protecting group Figure 1. Mechanism of intramolecular C–H amination via metal-nitrenoid intermediate.

1.2.1 Synthesis of Cyclic Sulfamidates

Chiral cyclic sulfamidates are very useful organic small moleculars as it can be used for further one step ring-open to the corresponding β - or γ -amino alcohols which are widely used as intermediates for the synthesis of durgs, natural products and as chiral ligands used in asymmetric catalysis.³



Figure 3. Ring-opening of chiral cyclic sulfamidates.

In 2002, Chi-Ming Che and co-workers demonstrated for the first time about highly enantioselective intramolecular ring-closing C–H aminations by detailed investigation of the ruthenium porphyrin catalyzed asymmetric synthesis of chiral cyclic sulfamidates (**Figure 4**).³ The ring-closing C–H aminations were of high diastereoselectivity and enantioselectivity which provided 5- or 6-membered chiral cyclic sulfamidates in up to 77% yield and 87% ee. Different functional groups on the phenyl moiety were well tolerated. Although the yields were only modest, the overall results were quite impressive.

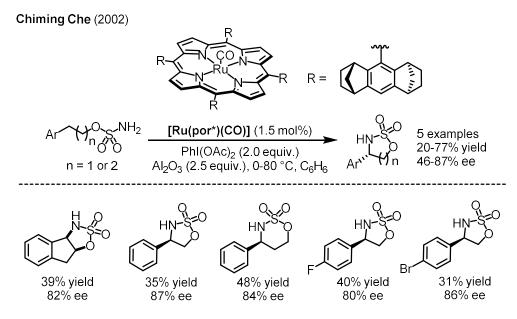


Figure 4. Research from the Chi-Ming Che group on the synthesis of chiral cyclic sulfamidates.

In 2008, the Blakey group subsequently reported a simplified ruthenium catalyst system for this reaction. Such ruthenium(II)–pybox complexes were readily prepared by using a method developed by Nishiyama's group.⁴ For the same transformations of linear sulfamidates to cyclic sulfamidates, this catalyst system provided much better results with up to 93% yield and 92% ee and in some cases with excellent diastereoselectivity for the formation of 6-membered cyclic sulfamidates as benzylic C–H bonds are more active than aliphatic C–H bonds at the stage of C–H insertion (**Figure 5**).

Simon Blakey (2008)

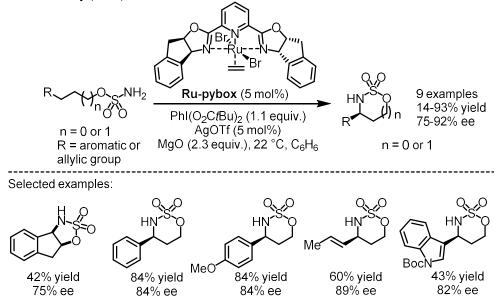


Figure 5. Research from the Simon Blakey group on the synthesis of chiral cyclic sulfamidates.

Almost at the same time, the Du Bois group reported a chiral rhodium carboxamidate catalyzed

enantioselective ring-closing C–H amination of sulfamidates.⁵ The newly designed dirhodium catalyst contains a strongly donating carboxamidate ligand instead of carboxylate which increases the capacity of the dirhodium centers for backbonding to the π -acidic nitrene ligand, thus affording a more stable and potentially more discriminating oxidant. As shown in **Figure 6**, **Rh**₂(*S*-**nap**)₄ displays unprecedented catalytic performance for the enantioselective intramolecular amination of benzylic C–H bonds with up to 98% yield and 99% ee. The design and development of this unique dirhodium complex further advance methods on enantioselective intramolecular C–H amination for the synthesis of chiral cyclic sulfamidates.

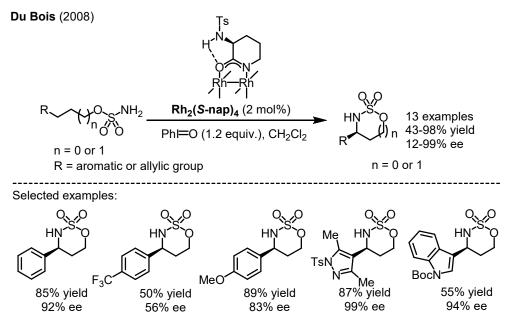
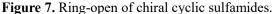


Figure 6. Research from the Du Bois group on the synthesis of chiral cyclic sulfamidates.

1.2.2 Synthesis of Cyclic Sulfamides



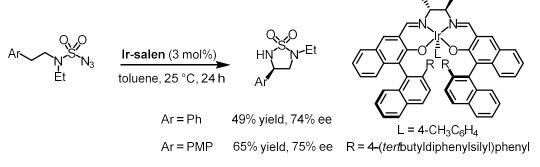


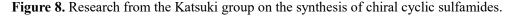
Although the structure of sulfamides are similar with sulfamidates which contain a nitrogen instead of the oxygen atom, they are different types of useful scaffolds as it can be manipulated for further one step ring-opening to the corresponding 1,2- or 1,3-diamines which are well-known synthetic intermediates for drugs, natural products and as chiral ligands used in asymmetric catalysis (**Figure 7**).⁶⁻⁸

In 2011, the Katsuki group firstly reported the synthesis of chiral cyclic sulfamides by efficient

intramolecular nitrene C–H insertion using sulfonyl azide substrates together with their newly invented **Ir-salen** catalyst (**Figure 8**).⁶ The C–H aminations proceeded smoothly and provided target 5-membered sulfamides in 49% yield and 74% ee for a phenyl substituent and 65% yield with 75% ee for the *para*-methoxy substituent. Although they only showed two examples, this pioneering work showcased that chiral cyclic sulfamides can be synthesized through efficient nitrene C–H insertion methodology.

Katsuki (2011)





In 2018, the Peter Zhang group explored the synthesis of 6-membered cyclic chiral sulfamides by using their unique chiral cobalt porphyrin catalysts (**Figure 9**).^{7a} The ring-closing C–H aminations were of high diastereoselectivity as the major products were always forming in 6-membered rings. This methodology was broadly applicable as they showed it can be applied to efficient benzylic, propargylic, allylic, electron-deficient and even non-activated aliphatic C–H aminations with 25 examples in up to 95% yield and 98% ee.

After systematic research work on modifications of their chiral porphyrin catalysts, they successfully invented a novel D_2 -symmetric chiral amidoporphyrin with alkylbridges across two chiral amide units on both sides of the porphyrin plane which is constructed in a modular fashion to permit variation of the bridge length. As showcased with enantioselective intramolecular C–H amination of sulfamoyl azides, the asymmetric 1,5-C–H amination is not easy for enantiocontrol (as shown in **Figure 8** with two examples in up to 75% ee).

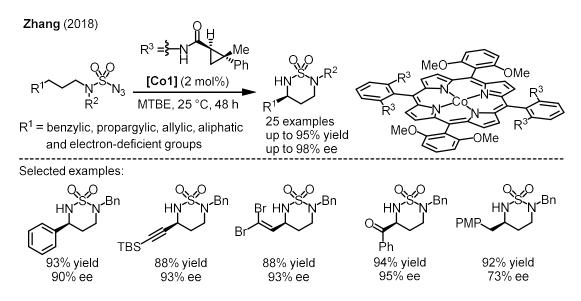


Figure 9. Research from the Peter Zhang group on the synthesis of 6-membered chiral cyclic sulfamides.

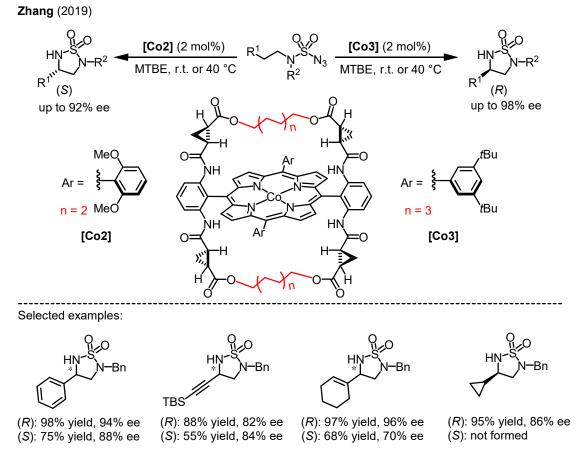


Figure 10. Research from the Peter Zhang group on the synthesis of 5-membered chiral cyclic sulfamides.

With the support of new catalysts, the Zhang group found that ring-closing C–H amination reactions could also be applied to the highly enantioselective synthesis of 5-membered chiral cyclic sulfamides. Interestingly, by changing the substituents of the chiral porphyrin ligand, the desired cyclic sulfamide products were formed with switched absolute configurations between (R) and (S) (**Figure**

10). As shown by the authors, the C-H aminations proceeded effectively at benzylic, propargylic, allylic and also aliphatic C-H bonds with up to 98% yield and 98% ee.7b

Subsequently, the Arnold's group invented an enzymatic platform for the asymmetric amination of primary, secondary and tertiary C-H bonds which lead to the formation of corresponding 5- and 6-membered chiral cyclic sulfamides in up to 51400 TTN and 99% ee (Figure 11)⁸. The concept "TTN" is a dimensionless number, defined as the ratio of moles of product generated divided by the moles of biocatalyst used in a reaction.

Arnold (2019)

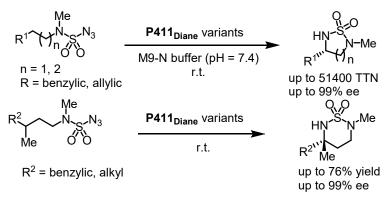


Figure 11. Research from the Arnold group on the synthesis of 5- and 6-membered chiral cyclic sulfamides.

Meggers (2020)

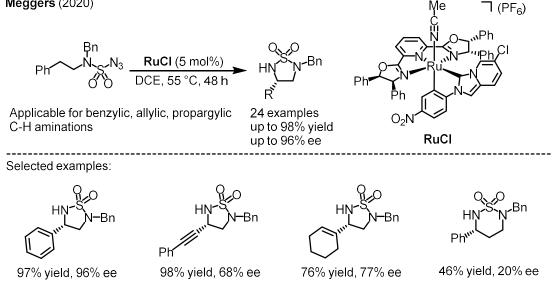


Figure 12. Research from the Meggers group on the synthesis of 5- and 6-membered chiral cyclic sulfamides.

In 2020, our group introduced a strategy to increase the utility of chiral pybox metal complexes for asymmetric catalysis by complementing the pybox ligand with an additional bidentate ligand. Such chiral ruthenium catalysts could be successfully applied in asymmetric C-H aminations of sulfamoyl azides. The C–H aminations could happen at benzylic, propargylic and allylic positions to provide cyclic sulfamides in up to 98% yield and 96% ee (Figure 12).⁹

1.2.3 Synthesis of Cyclic Sulfonamides

Since 2011, several groups have reported the enantioselective intramolecular C–H amination of sulfamoyl azides to chiral cyclic sulfonamides which is an important structure core in several biological active organic small molecules (**Figure 13**). The Katsuki group firstly invented this kind of transformation by using their chiral **Ir-salen** catalyst in the presence of sulfamoyl azide substrates providing desired 5-membered ring product in 77% yield and 92% ee which is quite impressive.⁶ Later, the Arnold group used their unique enzymatic catalyst system for a similar transformation which achieved up to 89% ee.¹⁰ The Fasan group used the same catalyst as Arnold but with some modification of the reaction conditions and the substrate's structure finally improved the results to up to 310 TTN and 91% ee.¹¹ Further progress was reported by the Hartwig group, they used an **Ir(Me)-PIX CYP**₁₁₉ catalyst system, which is a cytochrome P450 enzyme derived from a thermophilic organism and containing an iridium porphyrin cofactor (**Ir(Me)-PIX**) in place of the heme to improve the results to up to 98% yield and 90% ee.¹²

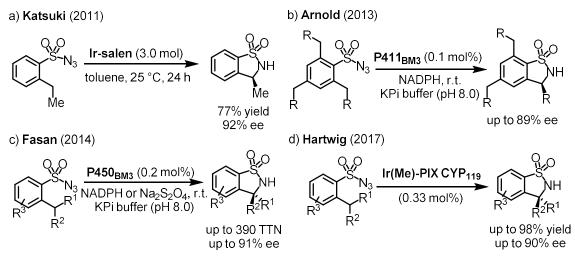


Figure 13. Synthesis of chiral sulfonamides via nitrene insertion.

1.2.4 Synthesis of Cyclic Carbamates

Compared with chiral sulfamidates, carbamates have the advantage of a more facile ring-opening which can proceed smoothly under mild reaction conditions. Besides, the catalytic intramolecular C–H amination of sulfamidates often lead to the formation of 6-membered cyclic sulfamidates which is not suitable for synthesizing chiral β -amino alcohols. Thus, it makes more sense to put efforts in finding a

suitable catalyst system for the synthesis of chiral cyclic carbamates via nitrene insertion.

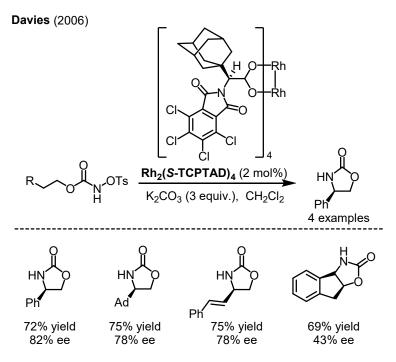


Figure 14. Research from the Davies group on the synthesis of chiral cyclic carbamates.

In 2008, the Davies group reported preliminary results on enantioselective intramolecular C–H amination for the synthesis of chiral cyclic carbamates.¹³ In their report, they provided only four examples using *N*-tosyloxycarbamates as nitrene precursor under chiral dirhodium catalysis. Unfortunately, yields (62-75%) and enantioselectivities (43-82% ee) were only very modest and not of practical value (**Figure 14**).

1.2.5 Synthesis of Lactams

Different with the acylnitrenoid intermediate involved in the synthesis of cyclic carbamates, intramolecular insertion of metal nitrenes into C–H bonds to form γ -lactams has traditionally been hindered by competing isocyanate formation (Curtius rearrangement).¹⁴ In 2018, the Chang's group solved this problem by optimizing a class of pentamethylcyclopentadienyl iridium(III) catalysts for suppression of the competing Curtius decomposition pathway. Modulation of the stereoelectronic properties of the bidentate ligands to be more electron-donating was suggested by density functional theory calculations to lower the C–H insertion barrier favoring the desired C–H amination.¹⁴

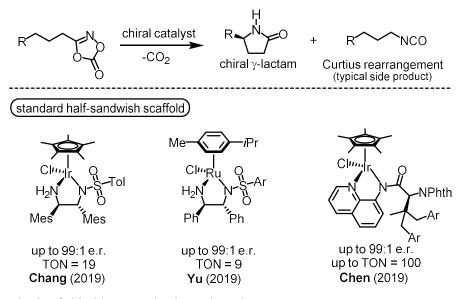


Figure 15. Synthesis of chiral lactams via nitrene insertion.

Subsequent research progress of this reaction was reported by the same group as an asymmetric version.¹⁵ By introducing a chiral bidentate ligand, the desired chiral lactam products were formed in up to 99% yield and 99:1 e.r., despite the TON of this transformation was quite low as only 19. Further progresses of this transformation were reported by the Yu group and Chen group,^{16,17} the results were further improved to 100 TON. Both of these three reports, the catalysts were similar half-sandwish structures (**Figure 15**).

1.2.6 Synthesis of Cyclic Ureas

The asymmetric intramolecular C–H aminations were already reported for several times and have been illustrated as a powerful tool for the synthesis of chiral nitrogen-containing heterocycles. This methodology has been applied to the catalytic asymmetric synthesis of cyclic sulfamidates, sulfamides, sulfonamides, carbamates, and lactams. However, interestingly, urea derivatives as nitrene precursors leading to the catalytic enantioselective formation of chiral cyclic ureas, specifically 2-imidazolidinones, were not reported yet when the work for this thesis was initiated. The only reports were from Du Bois on racemic rhodium-catalzyed ring-closing amination of urea derivatives using PhI(OAc)₂ as the oxidant.¹⁸ The absence of catalytic enantioselective versions to obtain chiral imidazolidin-2-ones is unfortunate considering the prevalence of this motif in bioactive compounds and their use as chiral auxiliaries.

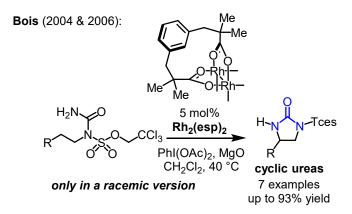


Figure 16. Synthesis of cyclic ureas in a racemic version from the Bois group.

Furthermore, chiral 2-imidazolidinones can be converted in a single step to chiral vicinal diamines which are valuable building blocks for the synthesis of medicinal agents, natural products, chiral ligands, and chiral catalysts.

1.2.7 Synthesis of Boc-Protected Cyclic Amines

Nitrenes with different substituents

EWG	EDG
[M]=N	[M] = N
more electrophilic	less electrophilic
(<i>higher reactivity</i>)	(lower reactivity)

Figure 17. Nitrenes with different substituents.

Because of metal-nitrenoid intermediate's electrophilic nature, such kind of nitrenoid intermediates with electron withdrawing groups lead to the formation of more electrophilic nitrenes which exhibit higher reactivity for the C–H amination reactions (**Figure 17**). In contrast, electron donating groups as substituents decrease the reactivity of corresponding nitrene intermediates, thus, the related C–H amination reactions become more difficult.

In 2013, the Betley group firstly reported an iron-catalyzed intramolecular C–H amination of aliphatic organic azide for the synthesis of Boc-protected cyclic amines namely pyrrolidines and the de Bruin group introduced a chiral cobalt porphyrin catalyst system which could get a preliminary result in an enantioselective version as showcased with only one example with 22% yield and 46% ee (**Figure 18**).¹⁹

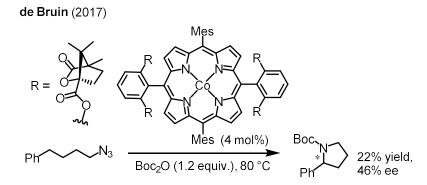


Figure 18. Research progress on the synthesis of the chiral Boc-protected pyrrolidine.

1.3 Asymmetric Intermolecular C-H Aminations

As compared with intramolecular C–H aminations, the intermolecular version has its particular advantages for the synthesis of chiral amines. As discussed in the above chapter 1.2, intramolecular C–H aminations are more efficient, with high asymmetric induction, but they often need several steps of additional transformations to achieve the synthesis of final chiral amines. Due to this further ring-opening step, some functional groups are not well tolerated (**Figure 19a**). The asymmetric intermolecular C–H amination via nitrene insertion to the C–H bonds can build a chiral C–N bond directly, which is much more convenient than the corresponding intramolecular version. The intermolecular C–H amination reactions also face several challenges, such as low efficiency and site-selectivity in outer-sphere C–H aminations and the enantioselectivity of the desired amination products are not as good as related intramolecular C–H aminations (**Figure 19b**).

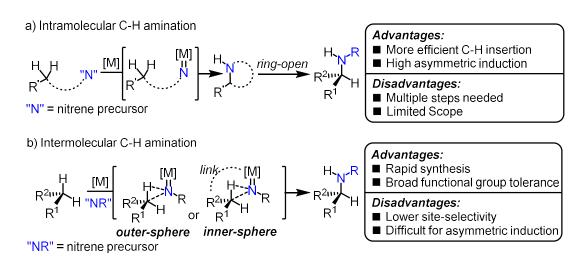


Figure 19. Comparison of intra- and intermolecular C-H amination for the synthesis of chiral amines.

Although, the asymmetric intermolecular C–H amination reactions are very challenging, several successful examples have already been reported in the past few decades. The following section will

briefly review the asymmetric intermolecular C–H amination reactions and category them by different transition-metal catalyst systems.

1.3.1 Rhodium Catalyst Systems

Among the reported examples of asymmetric intermolecular C–H aminations, rhodium-centered catalysts were the most frequently used transition metal catalysts. Such as dirhodium paddlewheel complexes, which feature significant structural rigidity, ease of ligand exchange, facile diaxial coordination sites, and relatively low oxidation potential, are widely utilized in asymmetric C–H aminations. Pioneering work by the Müller group for direct C–H amination of an indane substrate with dirhodium catalyst provided the desired C–H amination product in 71% yield and 30% ee as a groundbreaking result.²⁰ The result has been further improved by the Hashimoto group to 82% yield and 70% ee by modification of the dirhodium catalyst's bidentate chiral ligand (**Figure 20**).²¹

In 2006, the Davies group disclosed an intermolecular benzylic C–H amination system using a dirhodium(II) complex as the catalyst, which led to the formation of desired intermolecular C–H amination products in up to 95% yield and 94% ee (Figure 21).¹³

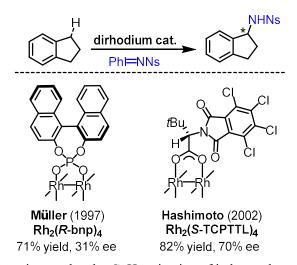


Figure 20. Pioneering work on intermolecular C-H amination of indane substrate.

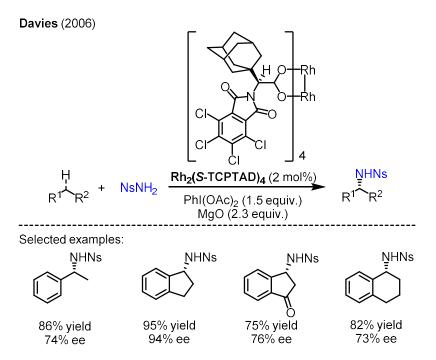


Figure 21. Intermolecular C–H amination work from the Davies group.

In 2019, the Dauban group made further modifications of the dirhodium catalysts, combining the pentafluorobenzyl sulfamate with the chiral rhodium complex $Rh_2(S-tfptad)_4$ which has led to the development of a catalytic asymmetric intermolecular C–H amination reaction with a broad scope in excellent yields and enantioselectivities. The reaction could scale up to gram-scale with equal efficiency (Figure 22).²²

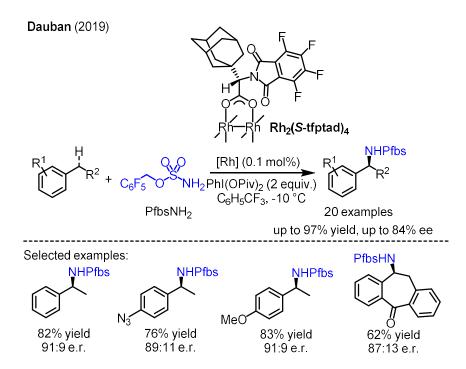


Figure 22. Intermolecular C-H amination work from the Dauban group.

In 2013, the Bach group developed a dual catalysis system, in which the rhodium center is responsible for activating PftbsNH₂ substrate to generate rhodium-nitrenoid intermediate, and the ligand contains a chiral amide functional group which can form hydrogen bond activation with another substrate, thus, providing high asymmetric induction with up to 74% ee (**Figure 23**).²³

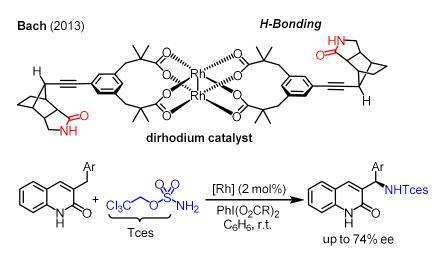


Figure 23. Intermolecular C–H amination work from the Bach group.

Besides, chiral rhodium cyclopentadiene complexes have been reported as efficient catalysts used in asymmetric C–H functionalization reactions. In 2019, the Matsunaga group reported a chiral carboxylic acid assisted enantioselective intermolecular benzylic C–H amination with 46-99% yield and up to 88% ee. From their proposed mechanism, the binaphthyl-based chiral carboxylic acid assisted the enantioselective cleavage of methylene C–H bonds in building a chiral C-N bond (**Figure 24**).²⁴

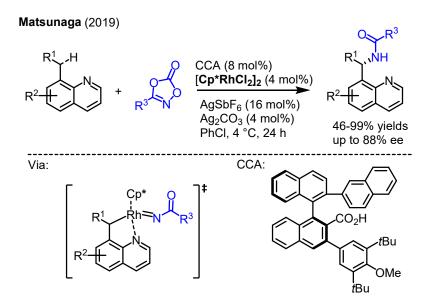


Figure 24. Intermolecular C-H amination work from the Matsunaga group.

Recently, the Blakey group reported an asymmetric inner-sphere C–H amination reaction (**Figure 25**). They developed a new planar chiral rhodium indenyl complex for regio- and enantioselective amination of allylic C–H bonds. The reaction exhibited broad functional group compatibility, provided an array of enantioenriched allylic amide products from readily available alkene starting materials. Crystallographic experiments supported the electronic transmission of asymmetry from the planar chiral indenyl ligand to the π -allyl ligand as a critical intermediate in the catalytic cycle. The overall results are quite impressive as 24 examples in up to 90% yield and 99:1 e.r.²⁵

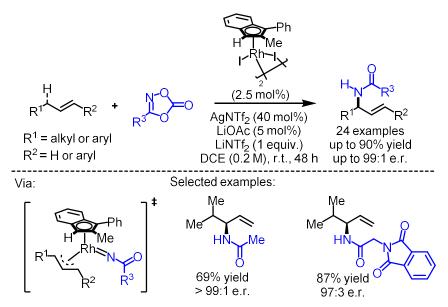


Figure 25. Intermolecular C-H amination work from the Blakey group.

1.3.2 Ruthenium Catalyst Systems

In 1999, the Chi-Ming Che group reported the first example of a ruthenium-centered chiral catalyst for asymmetric intermolecular C–H aminations (**Figure 26**). The enantioselective intermolecular amination at benzylic C–H bonds provided target chiral sulfonylamides in up to 58% yield as preliminary results.²⁶

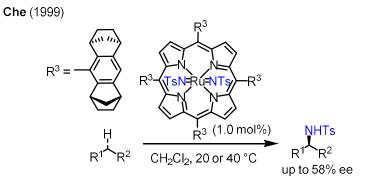


Figure 26. Intermolecular C-H amination work from the Chi-Ming Che group.

In 2003, the Katsuki group invented the **Ru(salen)(CO)** catalyst and successfully applied it in the asymmetric intermolecular C–H amination of allylic C–H bonds in up to 80% ee. The yield of the desired product was relatively low. The same group reported an improved result in 2011. They used the modified substrate **SESN**₃ and catalyst. Finally, the results were improved to 99% ee with broad substrate scope as shown for 22 examples (**Figure 27**).²⁷

Katsuki (2003 & 2011)

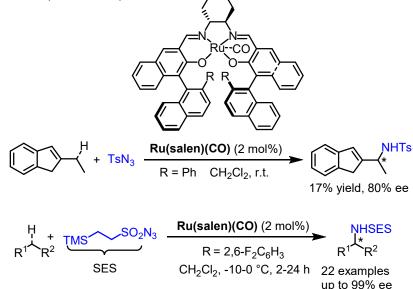


Figure 27. Intermolecular C-H amination work from the Katsuki group.

1.3.3 Silver, Manganese, and Iron Catalyst Systems

Transition metals such as silver, manganese, and iron were also reported to be used as catalysts in this kind of transformations. In 2001, the Katsuki group reported a chiral manganese salen complex catalyzed intermolecular C–H aminations of benzylic C–H bonds in up to 71% yield and 89% ee (**Figure 28a**).²⁸ Later, in 2017, the Arnold group successfully applied their chiral iron-porphyrin based enzymatic catalyst system in asymmetric intermolecular C–H amination of benzylic C–H bonds. The desired chiral amides were obtained in up to >99% ee and up to 1300 TON (**Figure 28b**).²⁹ Subsequently, in 2020, the same group reported the asymmetric primary C–H aminations at both benzylic and allylic C–H bonds provided desired chiral amines in up to 3930 TTN and 96% ee (**Figure 28c**).³⁰ Recently, the Bach group reported a dual catalytic system in which the chiral ligand forming hydrogen bonds with substrates and led to high asymmetric induction for the synthesis of desired amination products in up to 88% yield and 97% ee (**Figure 28d**).³¹

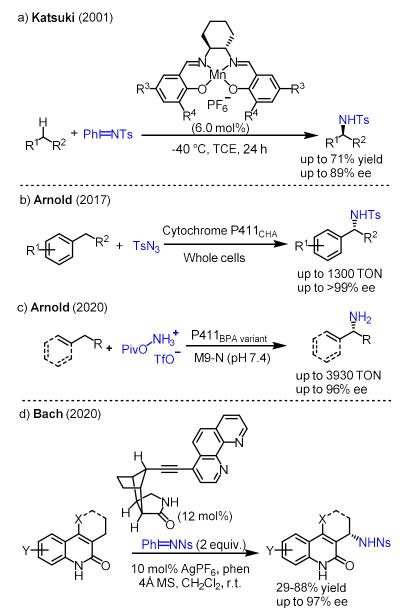


Figure 28. Other transition-metals catalyzed asymmetric intermolecular C-H aminations.

1.4 Aim of the Thesis

Chiral-at-metal ruthenium complexes developed in our group are powerful catalysts and can be applied in efficient asymmetric catalysis as demonstrated for enantioselective alkynylations of trifluoromethyl ketones.³² Since ruthenium-centered transition-metal complexes were already reported as effective catalysts for asymmetric C–H amination reactions, we envisioned that our chiral-at-ruthenium complexes could also catalyze asymmetric C–H amination reactions.

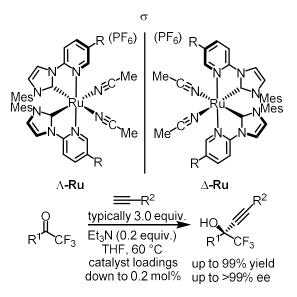


Figure 29. Chiral-at-metal ruthenium catalysts for asymmetric alkynylation reactions.

1) Building a chiral-at-ruthenium catalyst toolbox

In previous investigations on the catalytic asymmetric alkynylation of trifluoromethyl ketones, only two different chiral-at-metal ruthenium complexes were investigated. In this thesis, we aimed to make further modifications of such chiral-at-metal ruthenium complexes to build a chiral catalyst toolbox for detailed investigations of asymmetric intramolecular C–H amination reactions.

2) Apply chiral-at-ruthenium catalysts in asymmetric intramolecular C–H aminations

Transition-metal catalyzed asymmetric C–H aminations are a very attractive topic in current organic synthesis. Despite much efforts were already being put into this area, the intramolecular C–H amination remained some unsolved problems. To find an efficient asymmetric catalyst system is always an appealing project in this area. In this thesis, we aimed at achieving efficient asymmetric intramolecular C–H aminations with judiciously modified chiral-at-ruthenium catalysts and suitable substrates.

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Chapter 2. Results and Discussion

2.1 Catalytic Enantioselective Intramolecular C-H Amination of 2-Azidoacetamides

2.1.1 Research Background and Reaction Design

Ruthenium catalyzed enantioselective intramolecular C–H amination using aliphatic organic azides¹ as nitrene precursors led to the formation of target cyclic Boc-protected pyrrolidine products as initial ground-breaking results obtained by Jie Qin, who is a former member of the Meggers group (**Figure 30a**). The same transformations were reported for several times as racemic versions^{2,3}. Thus, the highly enantioselective intramolecular C–H aminations using aliphatic organic azides as nitrene precursors is an unsolved problem. The author of this thesis started to pursue his doctoral degree in the Meggers group when Jie Qin needed a partner to help him work together to improve the initial results.

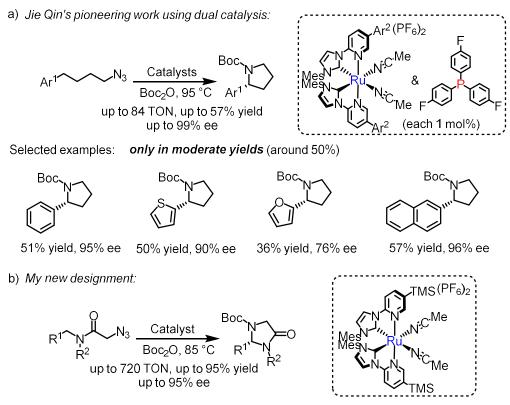


Figure 30. a) Pioneering work on ruthenium catalyzed intramolecular C–H aminations using aliphatic organic azides as nitrene precursors by the former group member Jie Qin. b) My new reaction designment.

The author of this thesis helped Jie Qin to improve the results by synthesizing a series of newly modified chiral ruthenium catalysts and improved the results to 95% ee. As shown in the above figure, the target chiral pyrrolidine products were always forming in only modest yields of around 50%,⁵ thus, the author of this thesis decided to explore a new class of aliphatic organic azides as nitrene precursors for intramolecular C–H aminations with the purpose of achieving higher yields of the target $\frac{22}{22}$

ring-closing C–H amination products in combination with modified chiral-at-ruthenium catalysts (Figure 30b).⁶

2.1.2 Synthesis of Modified Chiral-at-Ruthenium Catalysts

The general synthetic route of chiral-at-ruthenium catalysts was established by our previous group member Yu Zheng. I directly followed his method for the attempts on the synthesis of different racemic ruthenium catalysts. To our delight, the methodology was generally applicable for modifications of various substituents on the pyridine moiety (**Figure 31**). The desired racemic ruthenium catalysts were obtained in 51-89% yield. It's noteworthy that the TMS-substituted ruthenium catalyst was obtained in a low yield of 51% under standard complexation conditions. Further optimization of the reaction conditions by reducing the temperature from 200 °C to 185 °C successfully improved the yield to above 70%.

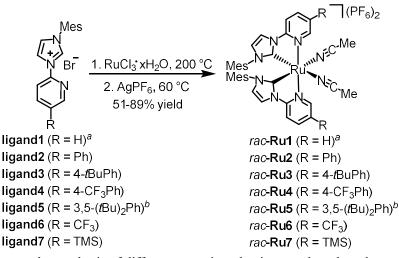


Figure 21. Attempts on the synthesis of different racemic ruthenium catalysts based on modifications of the pyridine moiety. [a] Yu Zheng's reported results. [b] Jie Qin's reported results.

With isolated analytical pure racemic ruthenium catalysts in hand, next, we performed the chiral-auxiliary-mediated synthesis of enantiomerically pure Λ -RuAux1-7 via Yu Zheng's established synthetic method. Under the standard 2-steps procedure, the desired enantiomerically pure Λ -RuAux1-7 were obtained (Figure 32).

The absolute configuration of standard ruthenium *bis*-NHC catalyst Λ -**Ru1** was comfirmed via single crystal diffraction. As similar structures, we identified the absolute configurations of newly modified ruthenium catalysts by comparing their CD spectra. As shown in **Figure 33**, the CD spectra of the representative ruthenium catalyst Λ -**Ru7** contains a similar trend with the reported Λ -**Ru1**.

Synthetic route of Λ -Ru1-7:

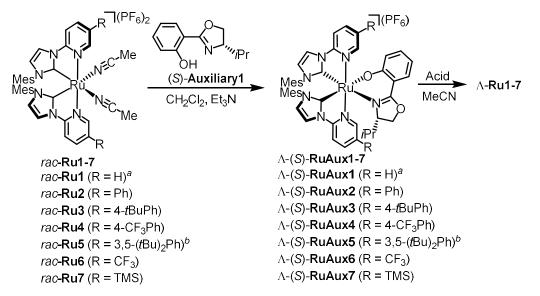


Figure 3. Chiral-auxiliary-mediated synthesis of enantiomerically pure Λ-**RuAux1-7**. [a] Yu Zheng's reported results. [b] Jie Qin's reported results.

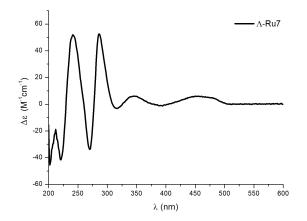


Figure 33. CD spectra of Λ -Ru7 recorded in CH₃OH (0.2 mM).

2.1.3 Initial Experiments and Reaction Development

During an initial screening of suitable substrates for a ring-closing C–H amination, I followed exactly the same conditions which Jie Qin used in his system (without phosphine additive instead) and found that 2-azido-*N*,*N*-dibenzylacetamide (**1a**) was converted to Boc-protected 3-benzyl-2-phenylimidazolidin-4-one (*S*)-**2a** in 28% NMR yield and 52% ee using ruthenium catalyst Λ -**Ru1** (1 mol%) in the presence of Boc₂O (1.2 equiv) in 1,2-dichlorobenzene (1 M) at 95 °C (Table 1, entry 1). This kind of product is a new structure motif in ring-closing C–H amination area, nobody have achieved it through nitrene insertion. Thus, it was promising to further optimize the reaction conditions to improve the preliminary result we obtained. Since I already had a series of newly

modified ruthenium catalysts in the fridge, I just tested all of them and hoped to get better results.

Ph N Ph	N ₃ Boc 1,2-dic H he	ol% Ru Cat 20 (1.2 eq) hlorobenzene eating, 48 h lard conditions	Mes ^N Mes-N		u1 : R = H u2 : R = Ph u3 : R = 4- <i>t</i> BuPh u4 : R = 4-CF ₃ Ph u5 : R = 3,5-(<i>t</i> Bu) ₂ Ph u6 : R = CF ₃ u7 : R = SiMe ₃
Entry	Catalyst	Conditions ^b	T (°C)	NMR yield ($(\%)^c$ ee $(\%)^d$
1	Λ- Ru1	standard	95	28	52 (<i>S</i>)
2	Λ- Ru2	standard	85	68	60 (<i>S</i>)
3	Λ- Ru3	standard	85	72	73 (<i>S</i>)
4	Λ-Ru4	standard	85	59	76 (<i>S</i>)
5	Λ- Ru5	standard	85	83	56 (<i>S</i>)
6	Λ- Ru6	standard	85	37	86 (<i>S</i>)
7	Λ -Ru7	standard	85	$84 (80)^e$	91 (<i>S</i>)
8	Δ -Ru7	standard	85	83	91 (<i>R</i>)
9	Λ -Ru7	$0.2 \text{ mol}\% \text{ cat}^f$	90	71 ^g	91 (<i>S</i>)
10	Λ -Ru7	$0.1 \text{ mol}\% \text{ cat}^f$	95	74^h	90 (<i>S</i>)
11	Λ -Ru7	under air	85	58	90 (<i>S</i>)
12	Λ -Ru7	no Boc ₂ O	85	$<1^i$	-
13	Λ-IrS	standard	85	9	n.d. ^j
14	Λ- RhS	standard	85	<1	-

 Table 1. Reaction Conditions Optimization^a

[a] Standard conditions: **1a** (0.2 mmol), Boc₂O (0.24 mmol), Ru catalyst (0.002 mmol) in 1,2-dichlorobenzene (0.2 mL) stirred at the indicated temperature for 48 h under N₂ unless noted otherwise. [b] Deviations from standard conditions are shown. [c] Determined by ¹H NMR of the crude products using Cl₂CHCHCl₂ as internal standard. [d] Enantiomeric excess determined by HPLC analysis of the crude main product on a chiral stationary phase. [e] Isolated yield in brackets. [f] Reaction time increased to 96 h. [g] 19% starting material left. [h] 14% starting material left. [i] Product refers to the unprotected amine. [j] Not determined.

As expected, the ruthenium catalyst played an important role in getting different results. A phenyl-modified catalyst Λ -**Ru2** provided an improved yield of 68% at 85 °C but only 60% ee (entry 2). Different substituents within the phenyl moiety (Λ -**Ru3**-**Ru5**) did not provide satisfactory enantioselectivities (entries 3–5). Interestingly, the CF₃-functionalized catalyst Λ -**Ru6** afforded 86% ee but only 37% yield (entry 6). The best results were obtained with the bulky trimethylsilyl (TMS)-functionalized ruthenium complex Λ -**Ru7** which provided 84% NMR yield (80% isolated yield) and 91% ee (entry 7). In addition, the catalyst Δ -**Ru7** featuring a metal-centered Δ -configuration provided the product **2a** in the same yield and enantioselectivity, but with opposite absolute configuration (entry 8). The catalyst loading can be reduced to 0.2 mol% (entry 9) and even 0.1 mol%

(entry 10) without affecting the enantioselectivity. The reduced reaction rate needs to be compensated by increasing the reaction temperature and time. Importantly, air atmosphere reduces the yield significantly (entry 10) while Boc₂O is required for this conversion (entry 12). Finally, it is noteworthy that our previously reported bis-cyclometalated chiral-at-metal catalysts Λ -**IrS** and Λ -**RhS** are almost or completely inactive for this transformation (entries 13-14).

2.1.4 Substrate Scope

With the optimized catalyst and reaction conditions in hand, we started to investigate the substrate scope with respect of 2-azido-N,N-dibenzylacetamides (Figure 34). Methyl groups in paraor meta-position of the phenyl moiety were well tolerated (imidazolidinones 2b and 2c), but the sterically hindering ortho-methyl group (2d) led to a reduced yield of 52% and 87% ee. Electron-donating groups appeared to be beneficial cause transition-metal nitrenoid intermediate prefers electron-rich C-H bonds. For example, a para-methoxy group provided the Boc-protected imidazolidinone 2e with a higher yield of 95% and 90% ee. Chlorines on the phenyl moiety (2f, 2g) did not affect the outcome of the reaction whereas strongly electron withdrawing CF₃ and fluorine substituents provided lower yields. Substrates with the heteroaromatic furan (2j) and thiophene (2k) moieties afforded slightly reduced yields and enantioselectivities. Replacing the aryl moiety with an ester group provided the desired cyclic product (21) with 32% ee. We were not satisfied with the result of only 32% ee and luckily found that the it can be improved to 70% ee by using Λ -Ru5 catalyst instead. Finally, 2-azidoacetamides with two different substituents could also be employed and provided the imidazolidinones (2m-2o). However, a bulky *tert*-butyl group at the amide nitrogen lead to a vastly diminished enantioselectivity of just 23% ee (20). No desired product was obtained for 2-azido-*N*,*N*-di(phenethyl)acetamide or 2-azido-*N*,*N*-di(allyl)acetamide as the substrates.

Next, we investigated azidoacetamides derived from isoindolines. We were delighted to find isoindolyl-*N*-(2-azidoacetamide) provided the desired tricyclic ring-closing C–H amination product **2p** smoothly in 74% yield and with 94% ee. An extended aromatic system (**2q**), electron withdrawing (**2r**), and electron donating (**2s**) moieties were well tolerated. Furthermore, azidoacetamides derived from 1,2,3,4-tetrahydroisoquinoline and 1,2,3,4-tetrahydro- β -carboline provided the desired cyclization products **2t** and **2u**, respectively, upon increasing the reaction temperature and reaction time, albeit with reduced yields (**Figure 35**).

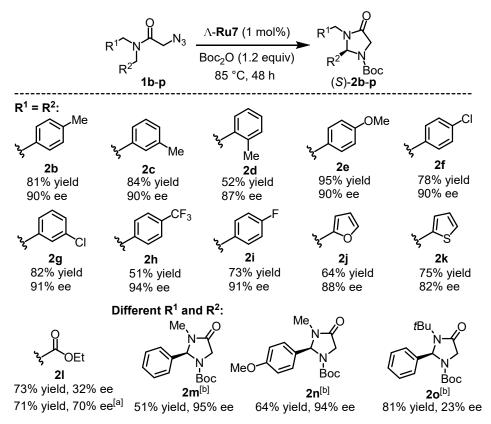


Figure 34. Substrate scope. ^[a]Λ-Ru5 as the catalyst instead. ^[b]Reaction temperature of 95 °C instead.

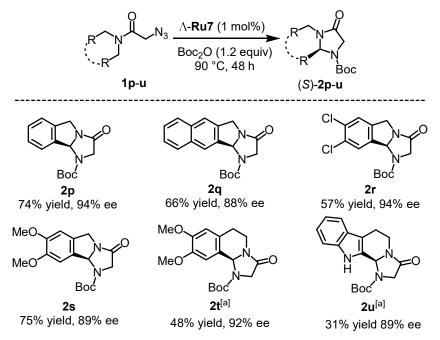


Figure 35. Synthesis of tri- and tetracyclic structures. ^[a]Reaction temperature of 105 °C for 60 h instead.

2.1.5 Mechanistic Study

The proposed mechanism is shown on **Figure 36**. The reaction is initiated by ruthenium coordination to the 2-azidoacetamide in a bidentate fashion through the amide carbonyl group and the α -nitrogen of the azide (intermediate I). Such coordination modes of organic azides have been

reported.⁷ A subsequent release of N_2 generates the chelated ruthenium-imido intermediate II, followed by dissociation of the amide (III), and a subsequent stereocontrolled insertion of the nitrene moiety into the C–H bond to provide a ruthenium-coordinated imidazolidinone (IV). Finally, the released imidazolidinone product is Boc-protected to suppress product inhibition. The Houk group coordinated to our research by performing Density Functional Theory (DFT) calculations to probe the validity of this mechanistic proposal. The maximum free energy barrier of this proposed reaction pathway was calculated to be 22.0 kcal/mol for the major enantiomer (*vide infra*), with irreversible C–H insertion and overall highly exergonic formation of the aminated product.

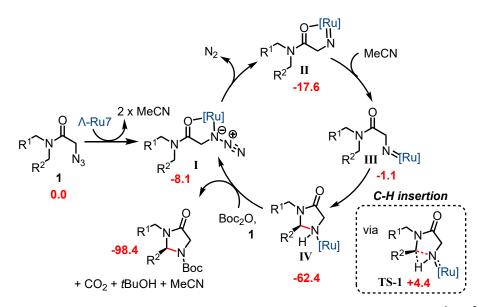


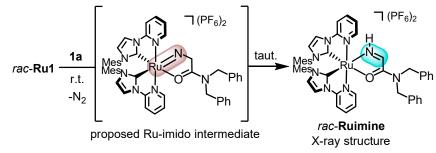
Figure 36. Proposed mechanism and DFT free energies (in kcal/mol) calculated with $R^1 = R^2 = Ph$ at the M06-D3/6-311++G(d,p)-SDD, SMD (1,2-dichlorobenzene)//B3LYP-D3/6-31G(d)-LANL2DZ level of theory. These DFT calculations were performed by Dr. Shuming Chen (Houk group, UCLA).

Our mechanistic experiments support the proposed mechanism. Reaction of the 2-azidoacetamide **1a** with equimolar amounts of a racemic ruthenium catalyst (*rac*-**Ru1**) at room temperature provided quantitatively the chelated imine complex *rac*-**Ruimine** which was characterized by X-ray crystallography (**Figure 37a**).

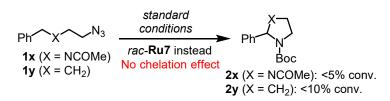
The generation of *rac*-**Ruimine** is a strong indication of the intermediate formation of the proposed chelated azide (**I**) and subsequent ruthenium-imido complex **II** (**Figure 36**), which tautomerizes to its corresponding imine upon 1,2–H shift. Such tautomerizations of ruthenium-imido complexes to their imines are well established.⁸ DFT calculations revealed that in this system the tautomerization is strongly exergonic, with a free energy change of -46.2 kcal/mol. However, at high temperatures the 1,2–H shift apparently cannot compete with the C–H amination step.

Changing the position of the amide functionality (1x) or removing it altogether (1y) suppresses the reaction or leads to very low conversions, respectively, further supporting the important role of the amide group in the ring-closing C-H amination of 2-azidoacetamides (Figure 37b). In another important mechanistic control experiment, we found that the bulky substrate 1aMe₂, bearing two additional methyl groups in α -position to the azido group, furnished the ring-closing C-H amination product 2aMe₂ in 11% yield in the absence of Boc₂O (Figure 37c). From this we conclude that Boc₂O is not required for the C-H amination step but important for protecting the secondary amine and preventing its coordination to the catalyst. In 2aMe₂, the steric hindrance of the two methyl groups makes this protection unnecessary. The observed kinetic isotope effects (KIE) are low and indicate singlet nitrene insertion with concerted N-C and N-H formation rather than stepwise radical reaction through a triplet nitrene (Figure 37d) as compared with reported literatures.⁹ An internal competition experiment with a 2-azido-N,N-dibenzylacetamide (1z) which contains one electron-rich and one electron-deficient benzyl group reveals a preference for electron rich C-H groups and suggests an electrophilic character of the ruthenium nitrene intermediate (Figure 37e). To elucidate the origins of the asymmetric induction in this reaction, DFT calculations were used to locate the C-H insertion transition states (Figure 38). TS-1-major, which leads to the observed major enantiomer of the product, is favored by 2.0 kcal/mol, in excellent agreement with the experimental value of 1.7–1.8 kcal/mol. A variety of functionals were tested, and all gave similar results to those reported in the text. In **TS-1-major**, a favorable π - π stacking interaction is observed between the upright Ph ring on the substrate and the PyNHC ligand. This stabilizing interaction is absent in TS-1-minor. In addition, a steric clash exists in TS-1-minor between the C-H bond undergoing insertion and the N-Mes group on the ligand, with a H...H distance of 2.23 Å. The steric pocket created by the ligand framework has a sterically congested bottom half due to the presence of TMS and N-Mes groups. As a result, the longer and more flexible N-Bn moiety is better accommodated in this part of the steric pocket (TS-1-major) than the shorter and more rigid Ph moiety (TS-1-minor). The substrate and the catalyst framework are calculated to be 1.0 and 0.7 kcal/mol less distorted in **TS-1-major** than in **TS-1-minor**, respectively, which also indicates a superior steric fit between the substrate and the catalyst framework in TS-1-major.

a) Ru imine complex formation under mild conditions:



b) Probing presence and position of amide group:



c) Isolated C-H amination product without Boc₂O:

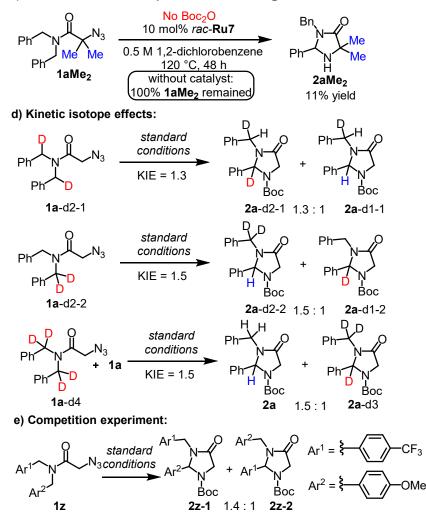


Figure 37. Mechanistic experiments.

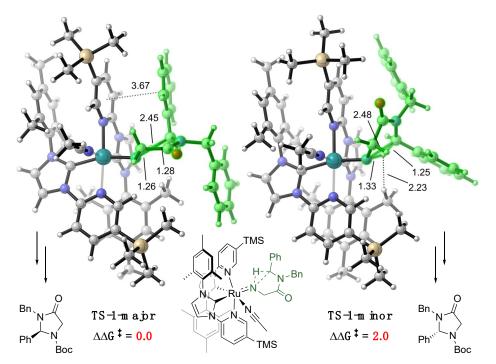


Figure 38. Calculated transition state structures for the C–H insertion step at the M06-D3/6-311++G(d,p)-SDD, SMD (1,2-dichlorobenzene)//B3LYP-D3/6-31G(d)-LANL2DZ level of theory. Energies are shown in kcal/mol. Interatomic distances are denoted in Ångströms. Calculations performed by Dr. Shuming Chen (Houk group, UCLA).

2.1.6 Conclusions

In conclusion, we here presented a ruthenium-catalyzed enantioselective ring-closing C–H amination of 2-azidoacetamides giving rise to chiral imidazolidin-4-ones, which find applications as auxiliaries for the synthesis of unnatural amino acids,¹⁰ as building blocks for the synthesis of natural products,¹¹ and as asymmetric organocatalysts¹². This work demonstrates the capability of chiral-at-metal bis(pyridyl-NHC) ruthenium complexes for highly efficient asymmetric intramolecular C–H aminations.

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2.2 Enantioselective Synthesis of y-Lactams by Intramolecular C-H Amidation

2.2.1 Research Background and Reaction Design

The group member Yubiao Hong firstly explored the synthetic route of the new ruthenium complexes Λ -**Ru8** and Λ -**Ru9** starting from heterocyclic 7-methyl-3-phenyl-1,7-phenanthrolinium hexafluorophosphate as the ligand of choice (**Figure 39**). It's no doubt that his pioneering exploration provided us the initial inspiration. But there were several specific issues that remained unsolved, which were very important for making further progress. The synthetic route needed some further modifications for getting higher yields, and the enantioselectivity of the final catalyst was not sufficient for using it in asymmetric catalysis. During that time, Yubiao Hong was busy making progress on his very promising chiral-at-iron catalyst project and decided not to pursue this catalyst anymore. Therefore, the author of this thesis decided to investigate further this new class of chiral-at-ruthenium complexes and apply them to new enantioselective C-H amination reactions.

Yubiao Hong's initial pioneering exploration of new ruthenium catalysts

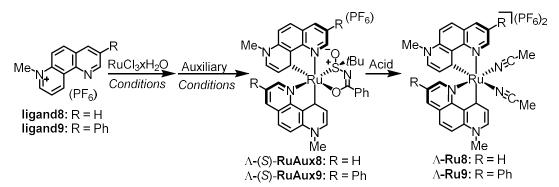


Figure 39. Yubiao Hong's pioneering exploration of new ruthenium complexes.

As shown in **Figure 39**, the main difference between the previous ruthenium scaffold and the new one are the different NHC ligands. The previous ruthenium scaffold contains normal NHC ligands, which are less electron-donating compared with the new scaffold, which contains remote NHC ligands.¹ Thus, the ruthenium center of the new platform is more electron-rich than the previous ruthenium scaffold. After understanding the most significant differences between these ruthenium catalysts, the author of this thesis had a feeling that we should explore reactions which need an especially electron-rich ruthenium metal center for catalysis.

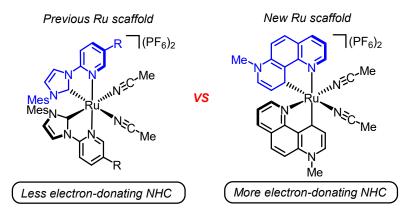


Figure 39. Main difference between previous ruthenium scaffold and the new one.

In 2018, the Chang group reported a streamlined synthetic methodology for γ -lactams via nitrene insertion catalyzed by electron-rich iridium catalysts (**Figure 40**). Modulation of the stereoelectronic properties of the auxiliary bidentate ligands to be more electron-donating was suggested by density functional theory calculations to lower the C–H insertion barrier favoring the desired reaction.²

Chang (2018): Electron rich iridium complex catalyzed intramolecular C-H amidation

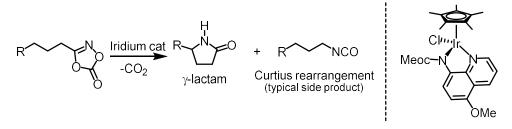
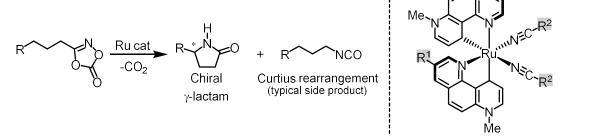


Figure 40. Intramolecular C-H amidation catalyzed by electron rich iridium complex from Chang's group.

After checking their supplementary files, the author of this thesis found that some ruthenium complexes (such as ruthenium porphyrin catalysts) could also catalyze this transformation and provide Curtius rearrangement products as major products. Thus, the author of this thesis thought the chiral-at-ruthenium catalysts could also catalyze the same transformation to afford the desired C–H insertion products (**Figure 41**).





 $(BF_4)_2$

Figure 41. Idea of applying our new ruthenium catalyst scaffold in asymmetric intramolecular C–H amidation reactions.

2.2.2. Synthesis of Chiral-at-Ruthenium Catalysts

According to Yubiao Hong's synthetic route with slight modifications, we chose the heterocycle 7-methyl-3-phenyl-1,7-phenanthrolinium hexafluorophosphate (**ligand9**) as our ligand of choice. Reaction of RuCl₃ hydrate with **ligand9** in 2-ethoxyethanol:water (4:1) at 125 °C afforded the racemic chloro-bridged dimer complex *rac*-Rudimer2 (88% yield). Each ruthenium is cyclometalated by two 1,7-phenanthroline ligands, which are electronically best described as chelating pyridyl pyridylidene ligands. The racemic mixture was next reacted with (*R*)- or (*S*)-*N*-benzoyl-*tert*-butanesulfinamide in the presence of K₂CO₃ to provide the complexes Λ -(*S*)-**RuAux9** or Δ -(*R*)-**RuAux9** as single stereoisomers in 46% and 44% yield, respectively. Interestingly, in the course of the formation of the *N*-sulfinylcarboximidate complexes, an unexpected but important isomerization of the chelating pyridylidene ligands occurred (this step is the crucial step for getting non-*C*₂-symmetric catalyst's structure, namely, auxiliary induced isomerization). The non-*C*₂-symmetric structure and corresponding absolute configuration of this ruthenium auxiliary complex are confirmed by the X-ray diffraction study of a simplified auxiliary complex Λ -(*S*)-**RuAux8** conducted by Yubiao Hong.

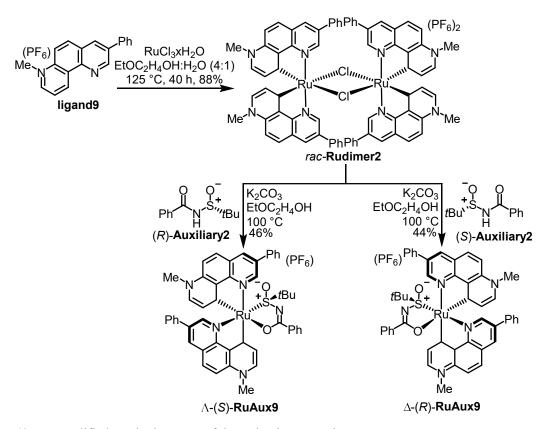


Figure 42. My modified synthetic routes of the ruthenium complexes.

After having the auxiliary complexes, we started the investigation of how to remove the chiral

auxiliaries. At the beginning, the author of this thesis directly used Yubiao Hong's standard reaction conditions with overnight as reaction time instead. Accidently, the author of this thesis found the final catalyst completely racemized. To judge the catalyst's enantio-purity, the author of this thesis performed an indirect way as using the result of the intramolecular C-H amidation reaction (it was due to the dicationic complex that couldn't be separated well by HPLC conditions). As shown in **Figure 43**, Λ -(*S*)-**RuAux9** under acid promoted acetonitrile substitution of the bidentate ligand led to the formation of the final ruthenium catalyst. When the author of this thesis used Yubiao Hong's synthesized ruthenium catalyst, we got 45% ee of the target product, but with my newly synthesized one, 0% ee was obtained. It was quite unfortunate to get this result, but it also meant that the removal of auxiliary played an important role in getting highly enantiomeric pure final ruthenium catalysts.

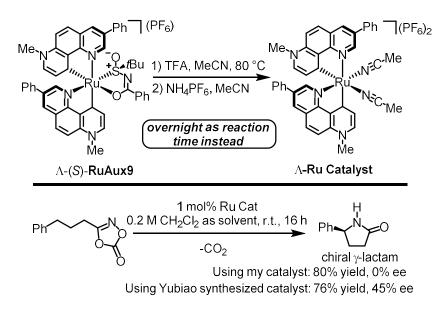


Figure 43. Comparison of catalytic results by using different ruthenium catalysts.

The author of this thesis thought maybe TFA is too acidic and led to the racemization of the ruthenium catalyst. The longer the reaction time, the more racemization occurred. So the author of this thesis decided to perform the auxiliary removal step in a shorter time as 30 mins. To our delight, the result further improved to 60% ee. The author of this thesis also tried different reaction solvents and finally found 1,2-dichlorobenzene was the solvent of choice, which led to 66% ee by using the same batch of ruthenium catalysts. Next, using NH_4PF_6 instead of TFA for the same transformation was performed in order to get higher ee values. Surprisingly, the result further improved to 84% ee. Due to NH_4PF_6 is a weak acid, the reaction needed 100 °C as the reaction temperature (**Figure 44**).

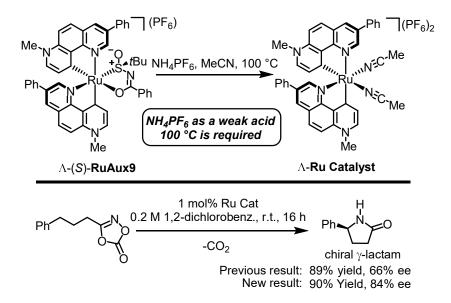


Figure 44. New results by performing the reaction using NH₄PF₆ as source of acid in 100 °C.

In order to further improve the result to around 90% ee, the author of this thesis tested some other acids and different reaction temperatures and finally found by performing the reaction with NH_4BF_4 as acid of choice in 75 °C could provide the best result (**Figure 45**). However, the removal of auxiliary step couldn't achieve full conversion.

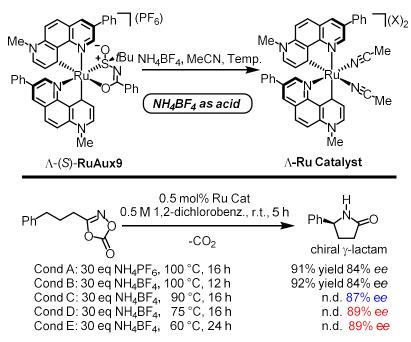


Figure 45. New results by performing the reaction using NH_4BF_4 as source of acid in different conditions. $X = PF_6$ or BF_4 .

Since 30 equiv. of NH_4BF_4 didn't dissolve in acetonitrile very well, the author of this thesis decided to use a solvent mixture (acetonitrile: $H_2O = 3:1$) instead, in order to achieve higher reactivity for the removal of auxiliary. To our delight, the reaction proceeded smoothly and got full conversion,

the desired ruthenium catalyst was isolated in 95% yield. By performing further modification of coordinated acetonitrile ligands to isobutylnitrile we got the final catalyst. The absolute configuration of this catalyst was confirmed by the X-ray diffraction study of a simplified ruthenium catalyst as Λ -configuration.

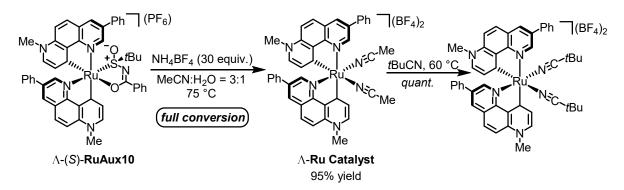


Figure 46. New results by performing the reaction using NH_4BF_4 as source of acid in different conditions. $X = PF_6$ or BF_4 . water acetonitrile solvent mixture and further modification of catalyst.

2.2.3 Initial Experiments and Reaction Development

We firstly investigated the catalytic properties of the new chiral-at-ruthenium complexes and found that Λ -**Ru10** displayed excellent catalytic activity for the intramolecular C–H amidation of 1,4,2-dioxazol-5-ones. For example, using just 0.5 mol% of Λ -**Ru10** at room temperature catalyzed the conversion of **3a** to the γ -lactam (*S*)-**4a** in 92% yield and with 90% ee (Table 1, entry 1). The related catalysts Λ -**Ru11** (acetonitriles instead of pivalonitriles) and Λ -**Ru12** (devoid of phenyl groups and acetonitriles instead of pivalonitriles) provided lower yields and enantioselectivities for this conversion and higher yields of side product isocyanate formation (entries 2 and 3). Importantly, as a comparison, our previous chiral-at-ruthenium complexes **Ru1** and **Ru6-7**, which were demonstrated to be very suitable catalysts for enantioselective C–H aminations of aliphatic azides, did not afford any detectable amounts of the C–H amidation product but instead quantitatively provided the isocyanate compound **5** (entries 4-6). Thus, Λ -**Ru10** has very distinct catalytic properties. The yield and enantioselectivity of Λ -**Ru10**-catalyzed conversion **3a** \rightarrow (*S*)-**4a** could be further improved to 95% yield and 92% ee by performing the reaction at 4 °C. To demonstrate the exceptional catalytic activity of Λ -**Ru10**, catalyst loading was further reduced (entries 7-9). At a catalyst loading of just 0.05 mol% the intramolecular C–H amidation still occured with 93% yield and 90% ee.

	-	,			,		
		$\begin{array}{c} \begin{array}{c} \mathbb{R}^{1} & (BF_{4})_{2} \\ \mathbb{R}^{1} & \mathbb{R}^{2} \\ \mathbb{R}^{1} & \mathbb{R}^{2} \\ \mathbb{R}^{1} & \mathbb{R}^{2} \\ \mathbb{R}^{3} \\ \mathbb{R}^{$					
		Λ - Ru12 : R ¹ = H, R ² = Me Λ - Ru7 : R ³ = TMS					
		$Ph \xrightarrow{N} CO_2$	PhC (S)-4a	+ Ph 5 yield (%) ^b	NCO		
entry	catalyst	(mol %)	T (°C)	<u>Jiela (70)</u> 3a	5	ee (%) ^c	
1	Λ- Ru10	0.5	r.t.	93 (92) ^d	6	90	
2	Λ- Ru11	0.5	r.t.	92 $(91)^d$	7	89	
3	Λ- Ru12	0.5	r.t.	$84(82)^d$	15	84	
ŀ	Λ-Ru1	0.5	r.t.	-	>99	-	
5	Λ -Ru6	0.5	r.t.	-	>99	-	
5	Λ- Ru7	0.5	r.t.	-	>99	-	
		o -	4	95 $(95)^d$	5	92	
7	Λ -Ru10	0.5	4	95 (95)	5	14	
7 8 ^e	Λ- Ru10 Λ- Ru10	0.5 0.1	4	$95(95)^d$ 95(95) ^d	5	92 92	

 Table 2. Comparison of different ruthenium catalysts^a

"Standard conditions: 3a (0.2 mmol) and Ru catalyst (0.05-0.5 mol%) in 1,2-dichlorobenzene (0.4 mL) stirred at the indicated temperature for 8 h under an atmosphere of nitrogen. ^bYields based on ¹H NMR analysis. 'Enantiomeric ratio of the crude product determined by HPLC analysis on a chiral stationary phase. ^dIsolated yields in brackets. ^eReaction time of 30 h instead. ^{*f*}Reaction time of 48 h instead.

2.2.4 Substrate Scope

With the optimized catalyst and reaction conditions in hand, we performed a substrate scope (Figure 47). A methyl group in para-position of the phenyl moiety was well tolerated and provided 96% yield with 90% ee (4b). An electron-donating methoxy group in the para-position of the phenyl moiety provided an almost quantitative yield but with a reduced enantioselectivity (4c). An electron-withdrawing nitro substituent at the para-position of the phenyl moiety gave 84% yield with 90% ee (4d). Halogen substituents at the para-position of the phenyl moiety were also well tolerated and provided high yields and good enantioselectivities (4e, 4f). Of note, both the brominated (4e) and iodinated (4f) γ -lactam products were improved to 98% ee after a single recrystallization in ethyl acetate. Replacing the phenyl moiety with a sterically more demanding naphthyl moiety provided the desired lactam product (4g) with 95% yield and 84% ee. A substrate with the heteroaromatic thiophene (4h) afforded a lower yield with 88% ee.

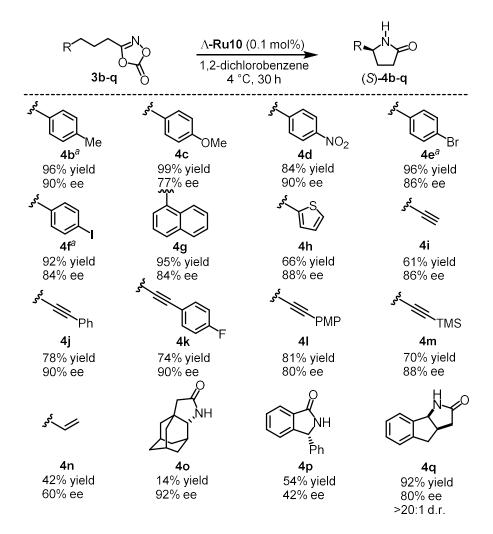


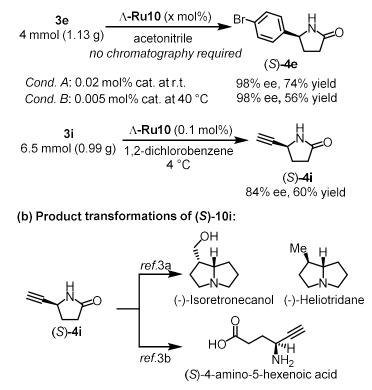
Figure 4. Substrate scope.

Next, we performed reactions of 1,4,2-dioxazol-5-ones with alkynyl substituents adjacent to the γ -C–H group which afforded chiral γ -alkynyl lactams in 61-81% yield and with up to 90% ee (**4i-m**). It is noteworthy that a terminal alkyne (γ -lactam **4i**) and TMS-functionalized alkyne (γ -lactam **4m**) were not included in previous reports on the direct enantioselective C–H amidation using 1,4,2-dioxazol-5-ones. Finally, with respect to substrate scope, a vinyl group next to the γ -C–H afforded the γ -vinyl lactam **4n** with modest 42% yield and 60% ee. If the γ -C–H is not activated by a π -system, the yields are low as shown for the adamantyl product **4o**. The synthesis of a chiral

isoindolinone (4p) and a desymmetrization generating two stereocenters (4q) are also shown in the substrate scope.

2.2.5 Synthetic Applications

The practical value of our developed C-H amidation catalyst was demonstrated in a gram-scale synthesis of (S)-4e with a catalyst loading of 0.02 mol% (Figure 48a). Interestingly, the isolation of the desired C-H amidation product was performed without column chromatography by only precipitation and crystallization in 74% yield and with 98% ee. It is noteworthy that the catalyst loading can be further reduced to merely 0.005 mol% to provide (S)-4e with 56% yield and 98% ee (TON number = 11200) at the same reaction scale. We believe this is the highest TON number reported for asymmetric transition-metal complex catalyzed ring-closing C-H nitrogenations. A gram-scale synthesis was also demonstrated for the terminal alkyne-functionalized γ -lactam (S)-4i and obtained in 60% yield with 84% ee. Chiral y-lactams are useful intermediates for the synthesis of bioactive molecules such as natural products and drugs. For example, the γ -lactam (R)-4e has been reported as an intermediate for the synthesis of a compound for the treatment of inflammatory disorders and a hydroxamate-based inhibitor of deacetylases B. Chiral γ -lactam (S)-4i, containing a terminal alkyne, was reported as a synthetic intermediate used in the total syntheses of the natural products (-)-isoretronecanol, (-)-Heliotridane,^{3a} and the GABA aminotransferase and glutamic acid decarboxylase inhibitor (S)-(+)-4-amino-5-Hexynoic acid^{3b} (Figure 48b). (S)-4i was previously synthesized in multiple steps starting from chiral amino acids or amino ester.



(a) Gram-scale synthesis of (S)-10e and (S)-10i:

Figure 48. Gram-scale reaction with low catalyst loading and further transformations. (a) Gram-scale synthesis of (S)-4e and (S)-4i. (b) Synthetic applications of (S)-4i.

2.2.6 Mechanistic Study

The proposed mechanism is shown in **Figure 49**. The reaction is initiated by ruthenium coordination to the 1,4,2-dioxazol-5-one (intermediate II). A subsequent fragmentation and release of CO_2 gas generates the ruthenium-imido intermediate III, followed by stereocontrolled insertion of the nitrene moiety into the C–H bonds (IV) and subsequent release of the lactam product finish the whole catalytic cycle.

We started our mechanistic study by investigating the influence of the relative stereochemistry of the new ruthenium catalyst scaffold on promoting C–H amidation reactivity. The complex **Ru10** features non- C_2 -symmetry, whereas all catalysts previously developed in our group for asymmetric C–H aminations (**Ru1** and **Ru6-7**) display C_2 -symmetry, but only provide the undesired Curtius-type rearrangement products (Table 2, entries 4-6). To evaluate the importance of the relative stereochemistry around the metal center, and the potential electronic role of the rNHC ligands, we synthesized the C_2 -symmetric diastereomer of **Ru10** (**C**₂-**rNHCRu**) and subjected it to standard reaction conditions with dioxazolone **Ru10**. Revealingly, the Curtius-type rearrangement product **11** was formed as the major reaction product in 60% yield with only 38% C–H amidation, demonstrating that the relative stereochemistry of **Ru10** has a significant effect on the reaction outcome (**Figrue** 50a).

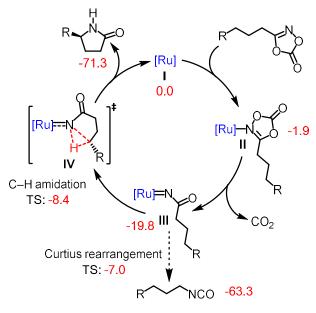


Figure 5. Proposed mechanism. Dr. Shuming Chen from the Houk group at UCLA performed the DFT calculations.

An intermolecular KIE value of 1.2 was determined by measuring initial C–H amidation rates of non-deuterated and bis-deuterated substrates (Figure 50b). The C–H amidations with substrate bearing a benzylic-CHD stereocenter were tested by using different Ru10 catalysts. Both Λ - or Δ -Ru10 gave a KIE value of 1.1, while C₂-rNHCRu, the C₂-symmetric diastereomer of Ru10, afforded a KIE value of 1.3. Low KIE values were also obtained in related work by Chang, Yu, and Chen using dioxazolones as nitrene precursors.^{4,5,6} These results indicate a singlet nitrene insertion with a concerted N-C and N–H formation pathway,⁷ although a stepwise radical reaction through a triplet nitrene cannot be totally excluded. We also tested the stereochemistry of the reaction by subjecting the non-racemic (98% ee) chiral substrate (*R*)-**3a**-Me to the C–H amidation conditions. As a result, (*S*)-**4a**-Me with retention of configuration at the reacted carbon was obtained as the major product with both Λ - and Δ -Ru10, but the Λ -catalyst reacts significantly faster with higher yield and affords the γ -lactam with a higher enantiomeric excess, thus revealing a high stereochemical discrimination between the two enantiotopic C–H bonds (Figure 50c).

Transition metal nitrenoids have been reported to transfer the nitrene fragment to phosphines to furnish iminophosphoranes.⁸⁻¹⁰ To gain experimental evidence for the formation of an intermediate ruthenium nitrenoid in our catalytic system, we performed a trapping experiment with PPh₃ using a dioxazolone substrate (**3t**) that is not capable of undergoing an intramolecular δ -C–H insertion (**Figure**

50d). As a result, the expected iminophosphorane 4t was formed in 81% yield under our standard reaction conditions which is consistent with our assumption that the ruthenium-catalyzed reaction proceeds through an intermediate ruthenium nitrenoid.

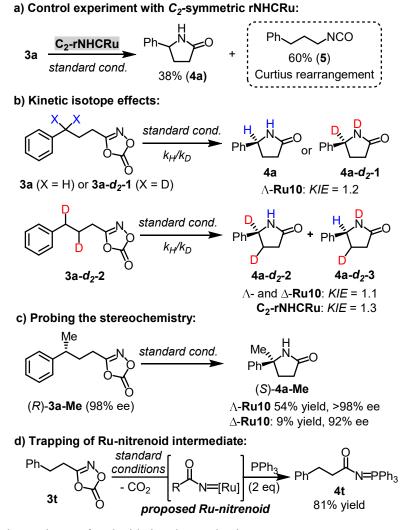


Figure 50. Control experiments for elucidating the mechanism.

Calculated structures of ruthenium *N*-acylnitrenes (**a**–**c**) formed with the three model catalysts **Ru12**, its *C*₂-symmetric diastereomer, and **Ru1**, are shown in **Figure 51**. Both **Ru12** and **C**₂-**Ru12** bear rNHC ligands, while **Ru1** has normal NHC (nNHC) ligands. Hirshfeld charge analysis shows that rNHC-bearing **a** (from **Ru12**) and **b** (from **C**₂-**Ru12**) have more electron-rich nitrene fragments (highlighted in green) than **c**.¹¹ This is consistent with the expected higher σ -donating ability of rNHC over nNHC ligands.¹² The calculated structures also reveal another factor that influences the electron-richness of the nitrenes, namely NCH…O=C hydrogen bonds between the carbonyl group of the acylnitrene and the polarized C–H bond in 2-position of the closeby pyridyl ligand. This NCH…O=C hydrogen bond is significantly stronger in **b** (2.17 Å) and **c** (2.05 Å), which are both

derived from C_2 -symmetric catalysts, than in **a** (2.30 Å) derived from a non- C_2 -symmetric catalyst. In **a**, the metal's coordination bond to one of the pyridine nitrogens is lengthened (2.24 Å, versus 2.15 Å in **b** and **c**) due to being positioned *trans* to the strongly σ -donating rNHC carbene carbon.¹³ The longer Ru–N(pyridine) bond leads to a more electron-rich pyridine nitrogen and a less acidic α -CH bond which consequently weakens the NCH···O=C hydrogen bond in **a** over **b** and **c**. Both Hirshfeld charges and orbital energies (see **Figure 51**) show that the overall nucleophilicity of the nitrene fragment decreases in the order of **a**>**b**>**c**, predicting that rNHC-bearing, non- C_2 -symmetric catalysts should be the most selective for C–H amidation, in agreement with the experimental results (**Table 2**, entries 1–3)

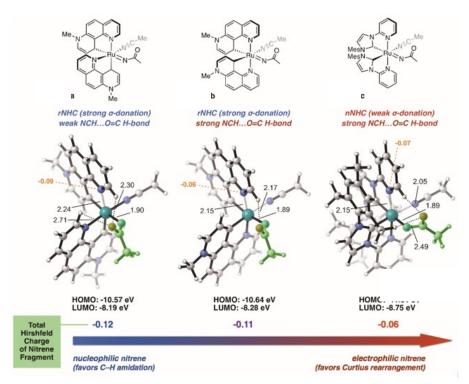


Figure 51. Calculated structures of ruthenium nitrene complexes at the B3LYP-D3/6-31G(d)–LANL2DZ (Ru) level of theory. Interatomic distances are in Ångströms (Å). Bolded numbers in orange indicate hirshfeld charges on individual atoms. Calculations performed by Dr. Shuming Chen (Houk group, UCLA).

Figure 52 shows the calculated transition states for the C–H amidation and Curtius rearrangement with substrate 3a and the catalysts Ru11 (non- C_2 -symmetry) and Ru1 (C_2 -symmetry). For Ru11, C–H amidation proceeds with a barrier of 11.4 kcal/mol, 1.4 kcal/mol lower than for the Curtius rearrangement pathway. This result is in excellent agreement with the experimentally observed product ratio (4a:5 = 92:7, $\Delta\Delta G^{\ddagger}$ = 1.6 kcal/mol). For Ru1, Curtius rearrangement is more facile with a barrier of 10.5 kcal/mol, while the C–H amidation barrier increases to 15.1 kcal/mol. The large $\Delta\Delta G^{\ddagger}$ of 4.6 kcal/mol is consistent with the experimental observation that Ru1 leads exclusively to the

Curtius rearrangement product.

Aside from higher nitrene electrophilicity, we hypothesized that steric factors may also play a role. In **Ru1** two mesityl groups are in close vicinity to the active site and may disfavor C–H amidation because of the more sterically demanding transition state, in which the nitrene alkyl chain must fold into a particular conformation in order to cyclize.

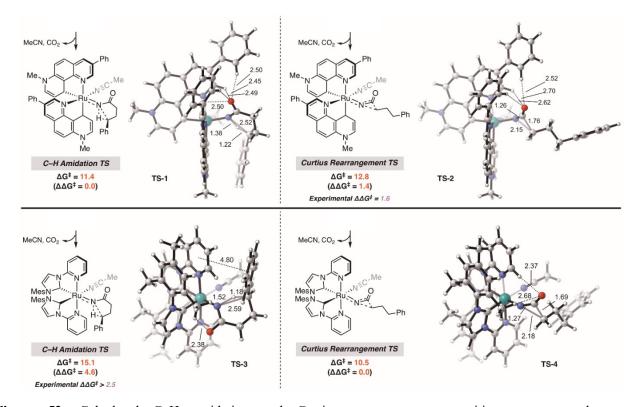


Figure 52. Calculated C–H amidation and Curtius rearrangement transition states at the M06-D3/6-311++G(d,p)–SDD (Ru), SMD (1,2-dichlorobenzene) // B3LYP-D3/6-31G(d)–LANL2DZ (Ru) level of theory. Interatomic distances are in ångströms (Å). Energies are in kcal/mol. Activation barriers are calculated with respect to the lowest-energy conformers of Ru nitrene intermediate III. Calculations performed by Dr. Shuming Chen (Houk group, UCLA).

Finally, the Houk group helped us for calculating C–H amidation transition states for the reaction $3a \rightarrow 4a$ catalyzed by Λ -Ru11, leading to different enantiomers of the lactam product. TS-1, which leads to the major enantiomer (*S*)-4a, is favored by 1.9 kcal/mol, in good agreement with the observed e.r. of 94:6 ($\Delta\Delta G^{\ddagger} = 1.6$ kcal/mol). A stabilizing π - π stacking interaction exists between the nitrene phenyl group and the rNHC ligand. In TS-1-ent, the cyclizing nitrene fragment is oriented quite differently, with two weaker π - π stacking interactions between the nitrene phenyl group and the rNHC ligands (Figure 53).

The calculations also established the energetic feasibility of the proposed catalytic cycle (Figure 3). After coordination of the substrate to the Ru center, extrusion of CO₂ to furnish **III** is exergonic

(-17.9 kcal/mol for **Ru11** and **3a**). Both C–H amidation and Curtius rearrangement pathways have low barriers, consistent with the observation that the reactions generally proceed at room temperature or below.

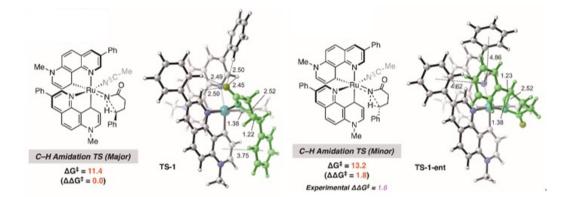


Figure 53. Calculated C–H amidation transition states leading to major and minor lactam enantiomers. at the M06-D3/6-311++G(d,p)–SDD (Ru), SMD (1,2-dichlorobenzene) // B3LYP-D3/6-31G(d)–LANL2DZ (Ru) level of theory. Interatomic distances are in ångströms (Å). Energies are in kcal/mol. Activation barriers are calculated with respect to the lowest-energy conformer of Ru nitrene intermediate III. Calculations performed by Dr. Shuming Chen (Houk group, UCLA).

2.2.7 Conclusions

In conclusion, we explored the applications of a new class of chiral ruthenium catalysts **Ru10**. The new ruthenium catalysts exhibited an exceptional catalytic activity for the enantioselective C–H amidation of 1,4,2-dioxazol-5-ones to chiral γ -lactams, reaching TON of up to 11200 (0.005 mol% catalyst loading). We believe that such a low catalyst loading are unprecedented for enantioselective C–H nitrogenations with non-enzymatic catalysts. Interestingly, the *C*₂-symmetric diastereomer of **Ru10**, as well as previously reported *C*₂-symmetric ruthenium catalysts suitable for enantioselective C–H aminations, provided instead an undesired Curtius-type rearrangement as the main product. In this catalyst architecture, the relative metal-centered stereochemistry (non-*C*₂ vs. *C*₂-symmetry) is therefore crucial for the reactivity and fate of the catalyzed reaction, while the absolute metal-centered stereochemistry (Λ vs. Δ) determines the absolute configuration of the C–H amidation product. Thus, whereas *C*₂-symmetric chiral transition metal catalysts are typically desirable for reducing the number of competing processes and transition states, in this catalyst architecture only the diastereomer with lower symmetry provides the desired catalytic activity.

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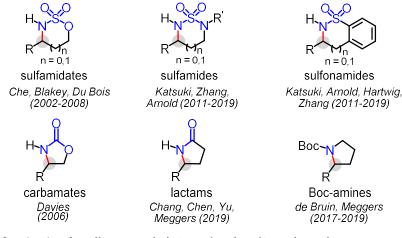
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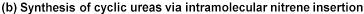
2.3 Enantioselective Synthesis of 2-Imidazolidinones by Intramolecular C-H Amidations

2.3.1 Research Background and Reaction Design

As has been described in the previous chapters, the direct catalytic asymmetric conversion of prochiral C(sp³)–H into C-N bonds offers an efficient synthetic route to non-racemic chiral nitrogen-containing molecules. This has been applied to the catalytic asymmetric synthesis of cyclic sulfamidates,¹⁻³ sulfamides,⁴⁻⁶ sulfonamides,⁷⁻¹⁰ carbamates,¹¹⁻¹² lactams,¹³⁻¹⁶ Boc-protected pyrrolidines,^{17,18} and related Boc-protected heterocycles (**Figure 54**).¹⁹ However, interestingly, urea derivatives as nitrene precursors leading to chiral cyclic urea, specifically 2-imidazolidinones, are absent from this list. This is unfortunate considering the prevalence of chiral 2-imidazolidinones in bioactive compounds and their use as chiral auxiliaries.^{20,21} Furthermore, chiral 2-imidazolidinones can be converted in a single step to chiral vicinal diamines which are valuable building blocks for the synthesis of medicinal agents, natural products, chiral ligands, and chiral catalysts.²²⁻²³ It was thus the goal to fill this gap and develop an enantioselective intramolecular C-H amination of urea derivatives.

(a) Chiral azacycles via intramolecular nitrene insertion





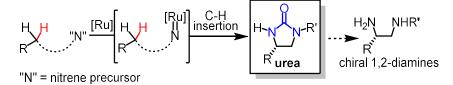
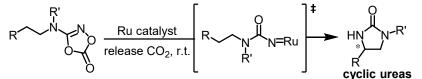


Figure 54. Enantioselective intramolecular amination of prochiral C(sp³)–H bonds. (a) Chiral heterocycles accessible by this strategy. (b) Reaction design.

As described in chapter 2.3, our ruthenium complexes were very efficient catalysts for activating 1,4,2-dioxazolone substrates for intramolecular C–H aminations providing chiral cyclic

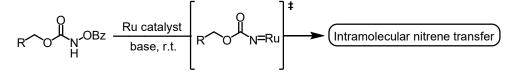
nitrogen-containing heterocycles. Thus, we envisioned to use similar substrates for the synthesis of cyclic ureas but no synthetic methods are currently available for their synthesis. During that time, the previous group member Yuqi Tan found that the *N*-benzoyloxycarbamates can serve as nitrene precursors together with chiral-at-ruthenium catalysts to afford cyclic carbamates. The author of this thesis therefore decided to investigate related *N*-benzoyloxyureas as nitrene precursors of intramolecular C–H aminations to synthesize chiral cyclic ureas (**Figure 55**).

(a) Initial idea on the synthesis of cyclic ureas via intramolecular nitrene insertion



Reason for not persue of the reaction: currently no synthetic method available for substrate synthesis

(b) Yuqi Tan's exploration on using N-benzoyloxycarbamate as nitrene precursor



(c) Idea of using N-benzoyloxyurea as nitrene precursor for intramolecular amination

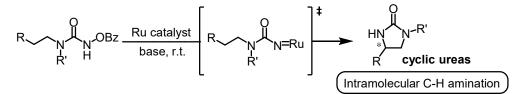


Figure 55. (a) Initial idea on the synthesis of cyclic ureas via intramolecular C–H amination. (b) Yuqi Tan's pioneering exploration on using *N*-benzoyloxycarbamates as nitrene precursors for intramolecular nitrene transfer reactions. (c) My idea of reaction design for the synthesis of chiral cyclic ureas.

2.3.2 Initial Experiments and Reaction Development

In chapters 2.1 and 2.2 we disclosed enantioselective nitrene insertion chemistry of organic azides and 1,4,2-dioxazol-5-ones using "chiral-at-metal" ruthenium catalysts with exclusive metal-centered chirality (only achiral ligands). We anticipated that this novel class of catalysts would allow us to address the challenge of accessing chiral 2-imidazolidinones by enantioselective $C(sp^3)$ –H amination from judiciously chosen urea derivatives. We initiated our study with the *N*-benzoyloxyurea **6aa** and envisioned that after release of the benzoate leaving group, an intermediate ruthenium nitrenoid would form and engage in an intramolecular C–H amination. Indeed, in the presence of 1 mol% **Ru1** and K_2CO_3 (3 equiv.) in CH₂Cl₂ at room temperature for 16 hours, the 2-imidazolidinone **7a** was formed in quantitative yield and with 86% ee (Table 3, entry 1).

Table 3. Evaluation of the C–H amination reaction^a

$R = 1(PF_6)_2$ $Ru1: R = H$ $Ru4: R = 4-CF_3Ph$ $Ru5: R = 3,5-tBu_2Ph$ $Ru6: R = CF_3$ $Ru7: R = SiMe_3$							
	PhN Mi starting mat		Me 6aa-ad	Ru cat (1 mol K ₂ CO ₃ (3 equ CH ₂ Cl ₂ , r.t., 1 standard condit	iv.) 6 h H−N N−Me	1	
	X = 2 664		Gab	to the second se	CF ₃	u	
entry	catalysts	Х	conditions ^t		yield $(\%)^c$	$ee (\%)^d$	
1	Λ -Ru1	6aa	standard		quant.	86	
2	Λ- Ru4	6aa	standard		quant.	94	
3	Λ- Ru5	6aa	standard		quant.	95	
4	Λ -Ru6	6aa	standard		quant.	94	
5	Λ- Ru7	6aa	standard		quant. (99)	95	
6	Λ- Ru7	6ab	standard		quant.	94	
7	Λ- Ru7	6ac	standard		quant	94	
8	Λ- Ru7	6ad	standard		27	91	
9	Λ- Ru7	6aa	0.5 mol% c	eat.	quant.	94	
10	Λ- Ru 7	6aa	0.1 mol% c	cat. ^g	quant.	94	
11	Λ- Ru7	6aa	0.05 mol%	cat. ^g	66	93	
12	Λ- Ru7	6aa	no base ^h		quant.	94	

^aStandard conditions: Substrate **6aa-6ad** (0.2 mmol), K₂CO₃ (0.6 mmol), Ru catalyst (0.002 mmol) in CH₂Cl₂ (2 mL) stirred at the indicated temperature for 16 h under N₂ unless noted otherwise. ^bDeviations from standard conditions are shown. ^cDetermined by ¹H NMR of the crude products using Cl₂CHCHCl₂ as internal standard. ^dEnantiomeric excess determined by HPLC analysis of the crude main product on a chiral stationary phase. ^fIsolated yield in brackets. ^gReaction executed at 40 °C for 24 h. ^hIncreased reaction time of 48 h.

Optimization of the chiral-at-metal ruthenium catalyst (Ru4-Ru7, entries 2-5) improved the enantioselectivity to 95% ee using the trimethylsilyl-modified ruthenium catalyst Ru7.

Functionalization of the benzoate leaving group with an electron-donating methoxy (**6ab**) (entry 6) or electron-withdrawing CF₃-group (**6ac**) (entry 7) slightly affected the enantioselectivity. A pivaloate leaving group (**6ad**) only provided the 2-imidazolidinone in 27% yield with 91% ee (entry 8). Interestingly, the catalyst loading can be reduced down to 0.05 mol% upon increasing the reaction time to 24 hours and the temperature to 40 °C. (entries 9-11). The addition of a base is not required but increases the rate of reaction (entry 12).

2.3.3 Substrate scope

To explore the scope of this new method, we applied the reaction conditions to a variety of N-benzoyloxyurea as shown in Figure 56. Benzylic C(sp³)-H aminations to chiral 2-imidazolidinones occured with high yields and high enantioselectivities. For example, a para-, meta-, and even a sterically very hindering ortho-methyl group in the phenyl moiety are well tolerated (products 7d-f, 92-99% yield, 94-97% ee), as well as an electron-withdrawing fluorine (7g) and chlorine substituent (7h), and an electron-donating methoxy group (7i). A 1-naphthyl group provided the cyclic urea 7j with 99% yield and 99% ee, while a 2-naphthyl group afforded the cyclic urea 7k with 97% yield and 90% ee. The smaller 2-thiophene moiety provided the cyclic urea 71 with 99% yield but a somewhat reduced 88% ee. 2-Imidazolidinone 7m with stereocenters in the 4- and 5-position was obtained in sluggish 29% yield but 89% ee by desymmetrization of an indane substrate using the ruthenium catalyst Λ -Ru6 instead of Λ -Ru7. However, the desymmetrization of a N-benzoyloxyurea derived from 1,3-diphenyl-2-propanamine provided the 4,5-difunctionalized 2-imidazolidinones 7n with two adjacent stereocenters as a single diastereomer in 93% yield with 94% ee. Besides C(sp³)-H aminations at benzylic and allylic positions, ring-closing C(sp³)-H amination is also possible at a propargylic position in 89% yield and with 87% ee (70). The ring-closing $C(sp^3)$ -H amination to 2-imidazolidinones tolerates different N-alkyl substituents as shown in Figure 57. Ethyl, n-butyl, isobutyl, and phenethyl substituents are well tolerated providing the corresponding N-alkylated 2-imidazolidinones 7p-s in 68-99% yield and with 92-95% ee. However, a benzyl substituents results in reduced yields of 37% yield for 7t (93% ee). Importantly, the ring-closing C(sp³)-H amination is applicable to non-alkylated substrates as demonstrated with the product 7u containing two N-H groups which was afforded in 91% yield and 91% ee. It's worth noting that a substrate bearing a *N*-phenyl substituent (6v) only provided the corresponding $C(sp^2)$ -H amination product (7v), which is not desired in this context but by itself a useful transformation.

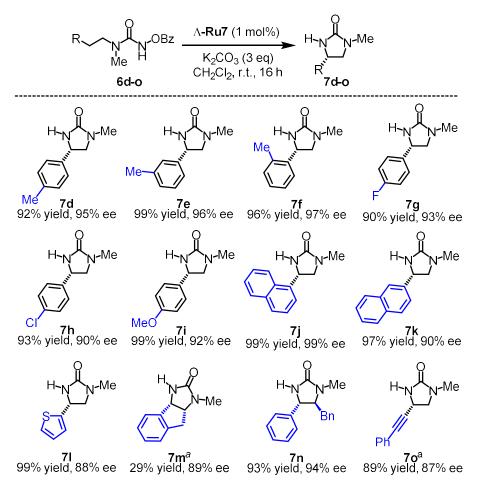


Figure 56. Scope with respect to synthesis of *N*-methyl cyclic ureas. ${}^{a}\Lambda$ -Ru6 was used as the catalyst instead.

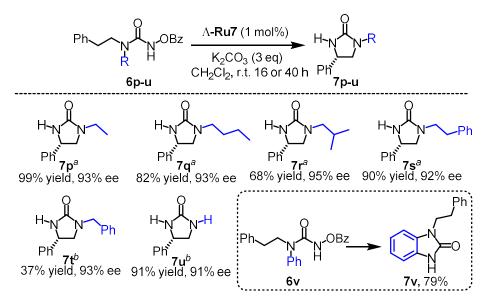


Figure 57. Substrate scope. Products 7p-r formed with regioisomeric ratios of more than 20:1. a40 h reaction time. b16 hour reaction time.

2.3.4 Mechanistic Study

The proposed mechanism is shown in Figure 58. Upon release of benzoic acid from the

N-benzoyloxyurea, the ruthenium catalyst forms a ruthenium nitrenoid intermediate (**I**). The ruthenium nitrenoid from its triplet state subsequently performs a 1,5–Hydrogen atom transfer (HAT) to provide the radical intermediate **II**. This is followed by C-N bond formation through radical-radical recombination to provide the ruthenium-coordinated product (**III**), which is released to regenerate the active catalyst for a new catalytic cycle.

The radical mechanism is supported by a significant kinetic isotope effect (KIE) of 4.35, which was determined with an intramolecular competition experiment using the deuterated substrate 6aa' to provide the cyclized products 7a' and 7a'' with a ratio of 4.35:1 (Figure 59a). The high KIE value obtained is a strong indication for the formation of a triplet nitrene intermediate which then engages in radical chemistry. This is in contrast to our previous work on nitrene insertion of 2-azidoacetamides in which a KIE value of 1.5 was determined by an analogous intramolecular competition experiment. That the mechanism proceeds through intermediate radicals is further indicated by an experiment performed with the diastereomeric substrates (Z)-6b and (E)-6b (Figure 59b). While (E)-6b formed (E)-7b under complete retention of the alkene configuration (80% yield, 76% ee), (Z)-6b (Z/E ratio > 20:1) was converted to (Z/E)-7b with an eroded Z/E diastereometic ratio of just 4.4:1 (73% yield, 91%) ee). This can be rationalized with an isomerization from the thermodynamically less stable Z-isomer to the preferred *E*-isomer in the course of the C–H amination at the stage of the allyl radical intermediate. However, the radical is apparently short-lived so that no complete isomerization can occur. As expected, the Z/E-ratio is temperature dependent with a higher Z/E-ratio at lower temperature (5.1:1 at 4 °C) and a lower Z/E-ratio at higher temperature (2.9:1 at 40 °C). Finally, a radical mechanism is also supported by the ring-closing C–H amination of the chiral non-racemic substrate (S)-6c (89% ee) to provide (S)-7c under retention of configuration for both catalyst enantiomers (Figure 59c). However, while the Λ -catalyst provides (S)-7c with only slightly decreased enantioselectivity (85% ee), the mismatched Δ -catalyst leads to a strong erosion of the enantiomeric excess of (S)-7c (40% ee). This erosion in enantiomeric excess is not consistent with a concerted C-H insertion mechanism but rather indicates a radical pathway in which the A-catalyst matches the S-configuration of the chiral substrate while the Δ -catalyst constitutes a mismatch, thus resulting in a slower radical recombination and subsequent partial racemization.

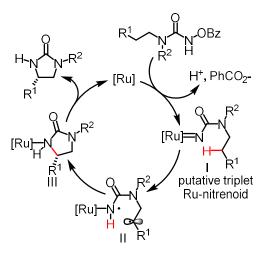


Figure 58. Proposed mechanism through an intermediate triplet ruthenium nitrenoid.

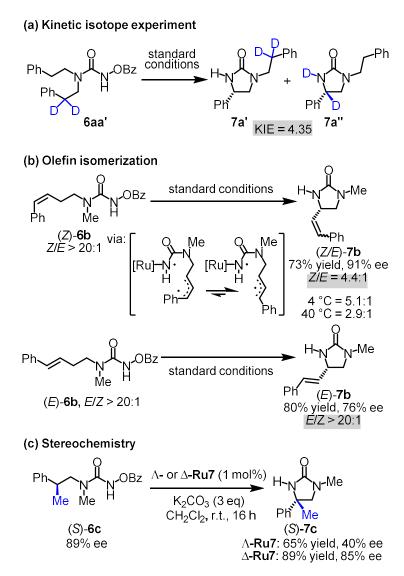


Figure 59. Control experiments to probe the proposed radical mechanism.

2.3.5 Synthetic Applications

Chiral 2-imidazolidinones are highly valuable building blocks for the synthesis of bioactive compounds such as medicinal agents and natural products. For example, (S)-4-phenyl-2-imidazolidinone ((S)-7**u**) can be obtained from the N-benzoyloxyurea **6u** and just 0.2 mol% Λ -Ru7 in a yield of 74% with almost perfect enantioselectivity of 99.6% ee after a single recrystallization step (Figure 60a). According to reported procedures, this enantiomerically pure 2-imidazolidine (S)-7**u** can be converted to the drug levamisole²⁴ by first thiation of the urea with Lawesson's reagent²⁵ followed by cycloalkylation with 1,2-dibromoethane.^{26,27} Analogously, the drug dexamisole, which is the enantiomer of levamisole,²⁸ can be synthesized from (R)-7u which was obtained from 6u using Δ -Ru7 instead of Λ -Ru7.²⁹⁻³¹

A second example provides a concise route to the bisindole alkaloids topsentine D and spongotine A (**Figure 60b**). Accordingly, ring-closing C(sp³)–H amination of the indole-containing substrates **6w** and **6x** provided under standard conditions the 4-indolyl-2-imidazolidinones **7w** and **7x**, respectively, in high yields and with high ee. These 2-imidazolidinones can be hydrolyzed with concentrated HCl in AcOH at 85 °C under microwave conditions³² for 10 min to the respective vicinal diamines **8a** and **8b** with almost unchanged enantiomeric excess, and constitute intermediates of the natural products topsentine D³³ and spongotine A,³⁴ respectively. A third example provides access to the mono-*N*-methylated 1,2-diamine **8c** which was reported as an intermediate for the synthesis of a chiral organocatalyst (**Figure 60c**).³⁵

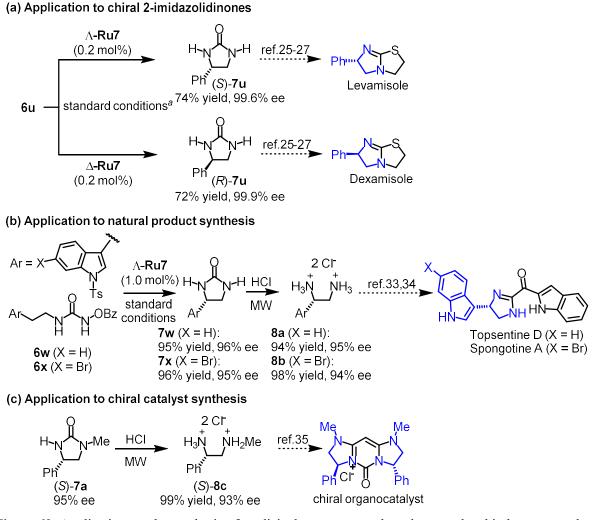


Figure 60. Applications to the synthesis of medicinal agents, natural products, and a chiral organocatalyst. ^{*a*}Standard conditions followed by recrystallization in EtOAc/n-Hexane. MW = microwave.

2.3.6 Conclusions

To conclude, we here reported the first example of a ring-closing C(sp³)–H amination of urea substrates to chiral 2-imidazolidinones in a catalytic enantioselective fashion. Starting from abundant primary or secondary amines, *N*-benzoyloxyurea can be synthesized in just 2 steps and enantioselectively cyclized to cyclic urea under mild reaction conditions and using low loadings of a chiral-at-metal ruthenium catalyst. This method will be of significant synthetic interest for future researches since the furnished chiral 2-imidazolidinones and their corresponding chiral 1,2-diamines, obtained through efficient hydrolysis with HCl, are highly valuable chiral building blocks.

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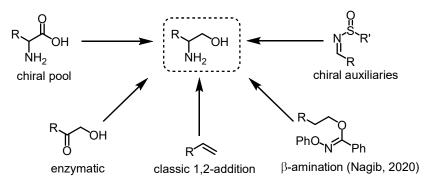
2.4 Enantioselective Synthesis of Cyclic Carbamates by Nitrene Insertion

2.4.1 Research Background and Reaction Design

Chiral vicinal amino alcohols are indispensable synthetic building blocks. For example, they are frequently used for the synthesis of bioactive compounds¹⁻³ and are substrates for the synthesis of chiral oxazolines, which represent one of the most frequently used class of chiral ligands in asymmetric transition metal catalysis.⁴ Enantiomerically pure chiral β -amino alcohols are typically synthesized by accessing the chiral pool, using chiral auxiliaries, or by applying catalytic asymmetric methods (**Figure 61a**). However, all these standard methods face significant limitations and drawbacks. The reduction of α -amino acids is only attractive for naturally occurring amino acid side chains and the L-stereochemistry.⁵ Chiral auxiliaries, such as Ellman's sulfinamide,⁶ have to be employed in uneconomic equimolar amounts, and catalytic asymmetric aminooxygenations of alkenes often lack a high degree of regioselectivity, and entail other drawbacks such as the use of toxic osmium in the case of the Sharpless aminohydroxylation.⁷ Biocatalytic approaches have also been reported but rely on substrates and reaction conditions that are compatible with enzymes.⁸

An appealing strategy for the asymmetric synthesis of chiral β -amino alcohols starts from ubiquitous alcohols and builds a $C(sp^3)$ -N bond including a stereocenter in a single step through regioand enantioselective ring-closing C-H amination chemistry. The Nagib group recently introduced an impressive method to accomplish this task through a photoredox radical protocol.⁹ However, the method requires a complicated cocktail out of iridium photocatalyst and chiral copper catalyst, the use of expensive BARF counterions, in addition to 25 mol% camphoric acid. We envisioned to provide a stereocontrolled and regioselective β -C(sp³)–H amination of alcohols by exploiting catalytic asymmetric nitrene insertion (Figure 61b). Great progress has been made for the catalytic asymmetric synthesis of chiral amines using transition metal nitrenoid chemistry under typically very mild reaction conditions.¹⁰⁻¹⁵ However, a general access to chiral β-amino alcohols in high yields and with high enantioselectivity through enantioselective nitrene insertion remains an unsolved problem. Sulfamate esters have been demonstrated to undergo intramolecular ring closing C-H aminations in the presence of hypervalent iodine reagents to provide cyclic sulfamidates,¹⁶⁻¹⁸ but are difficult to hydrolyze and would preferably provide γ -amino alcohols. The formation of cyclic carbamates through ring-closing $C(sp^3)$ -H amination would be advantageous. Davies provided four examples using N-tosyloxycarbamates under chiral dirhodium catalysis.¹⁹ Unfortunately, yields (62-75%) and enantioselectivities (43-82% ee) were only very modest and not of practical value.

a) Overview of representative methods for the synthesis of chiral β -amino alcohols



b) Strategy to chiral β-amino alcohols via transition-metal nitrenoid intermediates

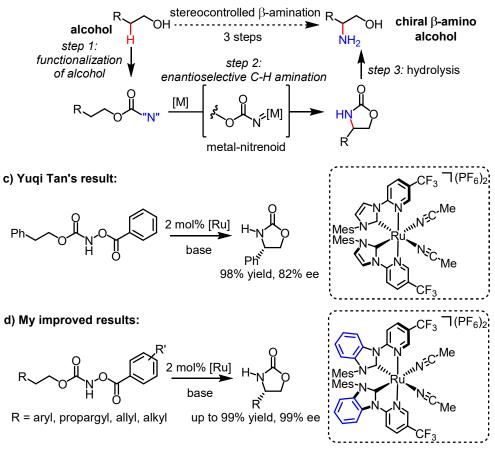


Figure 61. (a) Overview of representative methods for the synthesis of chiral β -amino alcohol. (b) Strategy to chiral b-amino alcohols via transition-metal nitrenoid intermediates. (c) Preliminary results obtained by the previous group member Y. Tan. (d) My improved results.

The previous group member Yuqi Tan obtained the preliminary result of the ring-closing C-H amination reaction, which lead to the formation of carbamate product in 98% yield and 82% ee using our standard chiral-at-Ru catalyst (**Figure 61c**). We were not satisfied with the obtained enantioselectivity. The author of this thesis thought if we modified the catalyst and also the leaving group of the substrate, maybe we could improve the enantioselectivity to above 90% ee (**Figure 61d**).

Although, the reaction has been reported with preliminary results, the synthetic utility of this reaction has not been illustrated. There is still no report for using ring-closing C–H amination methodology to synthesize chiral 1,2-amino alcohol via nitrene insertion.

2.4.2 Synthesis of New Chiral-at-Ruthenium Catalyst

As the previous group member Yuqi Tan already screened different reaction conditions in the purpose of getting above 90% ee of the desired carbamate product but often failed. The author of this thesis decided to start to modify the ruthenium catalyst's structure. As shown in **Figure 62**, starting from **ligand10** under standard complexation conditions, the desired *rac*-**Ru13** was obtained in 81% yield (**Figure 62**). The catalyst structure of the newly modified *rac*-**Ru13** has been proved by the single crystal X-ray diffraction study.

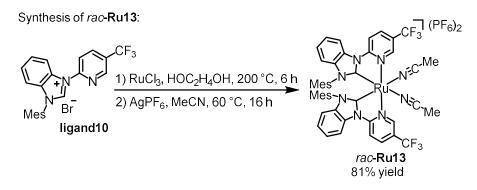


Figure 62. Attempts on the synthesis of *rac*-Ru13 based on modification of the NHC moiety.

 Λ -RuAux10 was obtained through the chiral-auxiliary-mediated synthesis. It's noteworthy that Λ -RuAux10 couldn't be synthesized through the standard auxiliary-mediated synthetic route (the intermediate auxiliary complexes couldn't be isolated well by flash column as the two diastereomers were of the same R_f value), thus, we explored a slightly modified synthetic route which usesd a new (*S*)-Auxiliary3 instead of the old one for getting enantiomeric pure Λ -Ru13 (Figure 63).

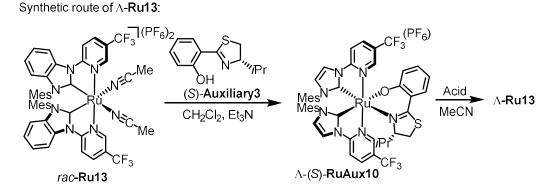


Figure 63. Chiral-auxiliary-mediated synthesis of enantiomerically pure Λ -Ru13.

2.4.3 Initial Experiments and Reaction Development

Azidoformates are well-established precursors of alkoxylcarbonylnitrenes upon release of dinitrogen. However, no reaction occured with phenethyl azidoformate 9aa at room temperature in the presence of chiral-at-Ru catalyst A-Ru6 (2.0 mol%) (entry 1). Likewise, sulfonyloxycarbamates did not provide satisfactory results. For example, the N-toluenesulfonyloxycarbamate 9ab afforded (S)-10ya in 69% NMR yield with just 68% ee (entry 2). The related N-methylsulfonyloxycarbamate **9ac** afforded (S)-10a in a higher NMR yield of 94% but with a reduced enantioselectivity of just 62% ee (entry 3). We next turned our attention to N-benzoyloxycarbamates. Previous work from our laboratory on intramolecular C-H oxygenations and amino-oxgenations revealed that our chiral-at-Ru catalysts efficiently generate ruthenium nitrenoid intermediates from N-benzoyloxycarbamates. In fact, in recent preliminary results we were able to convert the N-benzoyloxycarbamate 9ad to (S)-10a in 88% yield and with 78% ee (entry 4). Using carbamate 9ad we next screened solvents and found that 1,2-dichlorobenzene is the solvent of choice and provided almost quantitative NMR yields (98%) with improved 82% ee (entry 5). Since we were not able to further improve the enantioselectivity by modifying the reaction conditions we turned our attention to use modified ruthnium catalyst and discovered that the newly modified ruthenium catalyst (Ru13) provided an improved enantiomeric excess of 86% ee under otherwise identical reaction conditions (entry 6). Final improvements were accomplished by functionalizing the benzoate leaving group. Best results were obtained with 3,4,5-trimethoxybenzoate (9ae) (entry 7) and 2,4-difluorobenzoate (9af) (entry 8) which both afforded the cyclic carbamate (S)-10a with 90% ee. The 2,4-difluorobenzoyloxycarbamate appears slightly more suitable due to a somewhat higher reactivity. Finally, without base the reaction proceeded very sluggish (entry 9) or performing the reaction under air resulted in a slightly reduced yield (entry 10).

Ph	if 0	2 mol% [Ru] 1,2-dichlorobenz. K_2CO_3 (3 equiv) .05 M, 30 °C, 20 andard conditions	h (S)-10a	Mes N Ru N	$CF_3^{(PF_6)_2}$
N3 0 9aa		R =	9ac 9ad 9ae OMe	Me 9af	F
Entry	Substrate	Catalyst	Conditions ^b	NMR yield (%	$)^d$ ee (%) ^e
1	9aa	Λ -Ru6	CH ₂ Cl ₂ at 25 °C without base	0	n.d.
2	9ab	Λ -Ru6	CH ₂ Cl ₂ at 25 °C	69	68
3	9ac	Λ -Ru6	CH ₂ Cl ₂ at 25 °C	94	62
4 ^f	9ad	Λ -Ru6	CHCl ₃ at 25 °C	88	78
5	9ad	Λ-Ru6	standard	98	82
6	9ad	Λ-Ru13	standard	95	86
7	9ae	Λ-Ru13	40 °C instead	97	90
8	9af	Λ- Ru13	standard	quant. (99) ^g	90
9	9af	Λ-Ru13	no base	< 5	n.d.
10	9af	Λ- Ru13	under air	92	90
^a Standard	condition	s: 9aa (0.2	mmol), K ₂ CO ₃ (0.6 mmol), Ru	catalyst (0.002	mmol) in

Table 4. Initial experiments and optimization.^a

^{*a*}Standard conditions: **9aa** (0.2 mmol), K₂CO₃ (0.6 mmol), Ru catalyst (0.002 mmol) in 1,2-dichlorobenzene (4 mL) stirred at the 30 °C for 20 h under N₂ unless noted otherwise. ^{*b*}Deviations from standard conditions are shown. ^{*c*}Conversion. ^{*d*}Determined by ¹H NMR of the crude products using Cl₂CHCHCl₂ as internal standard. ^{*e*}Enantiomeric excess determined by HPLC analysis of the crude main product on a chiral stationary phase. ^{*f*}Taken from ref. 27. ^{*g*}Isolated yield in brackets.

2.4.4 Substrate Scope

With the optimized reaction conditons in hand we investigated the substrate scope. *N*-Benzoyloxycarbamates bearing different aryl substituents at the β -position were tested first. As shown in **Figure 64**, oxazolidin-2-ones with electron donating methoxy substituents in *para-* (**10b**), *meta-* (**10c**) or *ortho-* (**10d**) positions of the phenyl moiety were obtained in almost quantitative yields and with excellent enantioselectivities (90-92% ee). A substrate bearing a sterically very hindering *ortho-*methyl substituent (**10e**) was also well tolerated and provided 99% yield and 92% ee. A 4-phenyl substituent on the phenyl moiety (**10f**) provided almost quantitative yield together with an excellent enantioselectivity of 98% ee. Different electron-withdrawing groups are also accommodated in *para*-position as demonstrated for an electron-withdrawing fluorine (**10g**, 99% yield, 91% ee), chlorine (**10h**, 92% yield, 88% ee), or bromine (**10i**, 99% yield, 90% ee). A 1-naphthyl group provided the cyclic carbamate **10j** with 99% yield and 93% ee, while a 2-naphthyl group afforded the cyclic carbamate **10k** with 97% yield and 92% ee. A tryptophol-derived substrate provided oxazolidin-2-one in 66% yield and 85% ee (**10l**). The smaller 3-thiophene moiety provided the cyclic carabamate **10m** with 77% yield and 93% ee. Oxazolidin-2-one **10n** with stereocenters in the 4- and 5-positions was obtained in 99% yield and 95% ee by desymmetrization of an indane substrate. A *N*-benzoyloxycarbamate derived from 1,3-diphenyl-2-propanol provided the 4,5-difunctionalized oxazolidin-2-one **10o** with two adjacent stereocenters as a 1.8:1 *trans/cis* mixture with respective 92% and 97% ee in overall 83% yield.

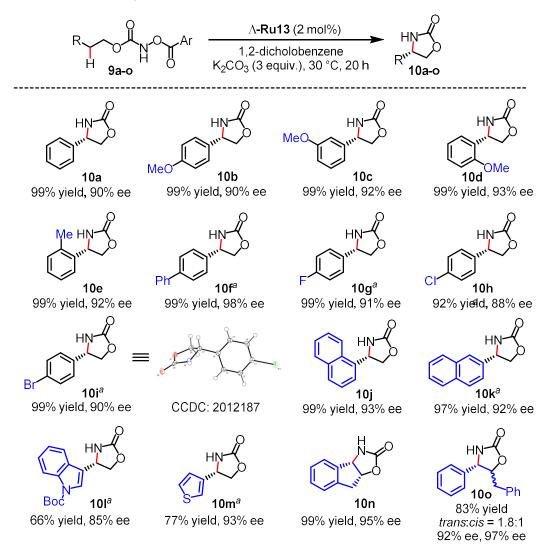


Figure 64. Substrate scope with respect to benzylic C–H aminations. Ar = 2,4-difluorophenyl unless noted otherwise. ^{*a*}Modified substrate and reaction conditions: Ar = 3,4,5-trimethoxyphenyl and reacted at 40 °C.

Non-benzylic C-H amination reactions are of particular interest since they are more difficult to

achieve in high yields and with high enantioselectivities. In fact, previous reported ruthenium catalyzed enantioselective intramolecular C-H aminations at non-benzylic positions from our group often failed to get satisfactory results. As shown in Figure 64, we investigated propargylic C-H aminations first and encouragingly found the product 10p was formed in 91% yield and 91% ee under standard reaction conditions. An electron-donating methoxy substituent on the phenyl moiety provided cyclic carbamate 10q with 91% yield and 94% ee while an electron-withdrawing fluorine substituent provided cyclic carbamate 10r with slightly reduced 85% yield and 90% ee. Replacement of the phenyl moiety with an alkyl group is also accommodated as shown by the alkyne product 10s (84%) yield and 90% ee). The aminated C(sp³)-H bond can also be flanked by an alkenyl group. While (E)-9t converted to (E)-10t under complete retention of the alkene configuration (88% yield, 63% ee), (Z)-9t (Z/E ratio > 20:1) was converted to (Z/E)- 10t with an eroded Z/E diastereometric ratio of 10.3:1 (56% yield, 45% ee). This can be explained with an isomerization from the thermodynamically less stable Z-isomer to the preferred E-isomer in the course of the C-H amination through an intermediate allyl radical. However, the radical is apparently not long-lived enough to furnish complete isomerization. We were delighted to find that a cyclohexene substitutent provided cyclic carbamate 10u in 96% yield and 92% ee. It is noteworthy that the late-stage functionalization of (R)-nopol provided cyclic carbamate 10v in 87% yield with 96:4 d.r.. In this example, the stereoselectivity of the C-N bond formation was controlled only by chiral ruthenium catalyst since the racemic ruthenium catalyst lead to the formation of 10v with only 1:1 d.r.. In addition, C-H amination next to a small isopropenyl substituent, provided the product 10w in 79% yield with 61% ee. Beside C(sp³)-H aminations at propargylic and allylic positions, ring-closing C-H amination was also possible at aliphatic methylene groups without any adjacent activating group. The adamantyl substituted cyclic carbamate 10x was formed in 99% yield with 99% ee. The *n*-butyl substituted oxazolidin-2-one 10y was formed in 53% yield with 80% ee. The late-stage functionalization of 10-undecen-1-ol provided 10z in 41% yield and 79% ee. Finally, a substrate bearing an adjacent chiral center was also employed, which is the late-stage functionalization of (S)-catronnellol, providing 10za in 18% yield and > 20:1d.r. as determined by ¹H NMR.

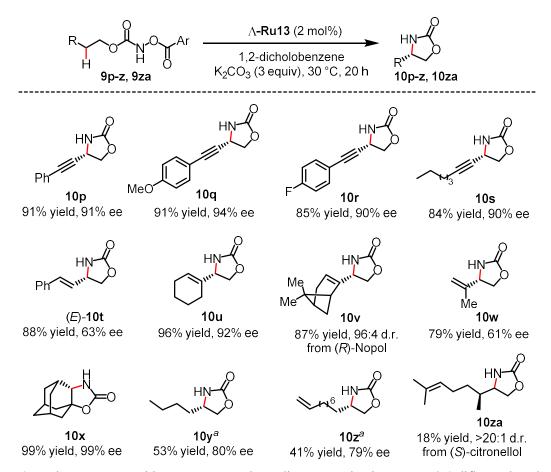
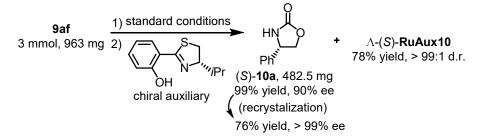


Figure 65. Substrate scope with respect to non-benzylic C–H aminations. Ar = 2,4-difluorophenyl unless noted otherwise. ${}^{a}Ar = Ph$.

2.4.5 Synthetic Applications

For practical purposes it is important to note that the catalytic intramolecular C–H amination of substrate **9af** was tested on a gram scale and proceeded smoothly under standard reaction conditions to provide (*S*)-**10a** in 99% yield with 90% ee (**Figure 66a**). Upon a following simple recrystallization step, (*S*)-**10a** could even be obtained with > 99% ee. Furthermore, upon addition of the auxiliary after the reaction, the chiral-at-Ru catalyst was recycled in 78% yield and with >99:1 d.r. as the auxiliary complex Λ -(*S*)-**RuAux10**. We also investigated follow-up conversions of the carbamate products. Accordingly, the oxazolidin-2-one (*S*)-**10a** (recrystallized with > 99% ee) was converted to the Boc-protected β -amino alcohol (*S*)-**11** without any loss of enantiomeric excess (**Figure 66b**, method A). The aminoalcohol (*S*)-**11** was reported as a valuable synthetic intermediate for the synthesis of chiral 1,2-diamine²⁰ and also the chiral α -amino acid²¹. In another follow-up chemistry using bis(2-aminoethyl)amine as the ring-opening reagent, the oxazolidin-2-one **10n**, bearing two vicinal stereochenters, was converted to the corresponding chiral amino alcohol (*IS*,2*R*)-**12** which is a building block for the frequently used chiral Box-ligand (**Figure 66b**, method B).²²

(a) Gram-scale synthesis of (*S*)-**16a** and catalyst recovery:



(b) Synthesis of chiral β -amino alcohol and further transformations:

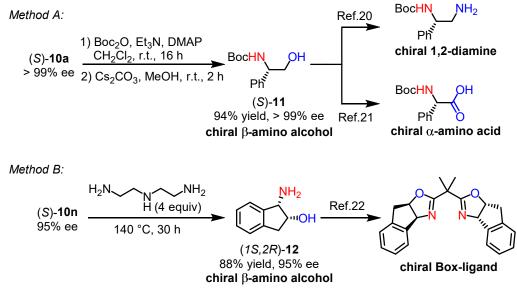


Figure 66. Gram-scale reaction and ring opening of chiral cyclic carbamates.

The utility of our new method for the straightforward synthesis of natural products is shown in **Figure 67**. Substrate **9zb** undergoes an intramolecular enantioselective cyclization to provide in 92% yield and with 91% ee (*S*)-**10zb** which was reported as an intermediate for the synthesis of the natural product (-)-aurantioclavine (**Figure 67a**).²³ A second example provides a concise route to the bisindole alkaloids hamacanthin A and dragmacidin A. Accordingly, ring-closing C–H amination of the indole containing substrate **9zc** provided under standard conditions the cyclic carbamate intermediate (*S*)-**10zc** in 74% yield and 94% ee. Further ring-opening provided the chiral β -amino alcohol **13** in 88% yield with 94% ee which was reported as an intermediate for the synthesis of natural products hamacanthin A and dragmacidin A (**Figure 67b**).²⁴

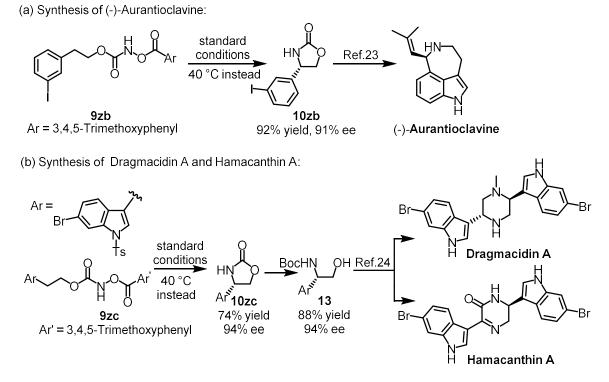


Figure 67. Synthetic application to natural products.

2.4.6 Conclusions

In summary, we here reported an economic and practical method to chiral oxazolidin-2-ones and corresponding β -amino alcohols, both of which are highly valuable chiral building blocks. The method is based on a ring-closing C(sp³)-H amination of N-benzoyloxycarbamates using a new benzimidazol-2-ylidene chiral-at-ruthenium catalyst. 2,4-Difluorobenzoate carbene and 3,4,5-trimethoxy benzoate leaving groups afford for most substrates the best results. The intramolecular C-H amination provides cyclic carbamates in up to 99% yield and with up to 99% ee for benzylic, allylic, and propargylic C-H bonds. Completely non-activated C(sp³)-H bonds provide somewhat reduced yields and enantioselectivities. We demonstrated the synthetic value of this new method with the catalytic enantioselective synthesis of chiral oxazolidin-2-ones as intermediates of the natural products aurantiolavine, hamacanthin A and dragmacidin A, a chiral amino acids, and a chiral Box ligand.

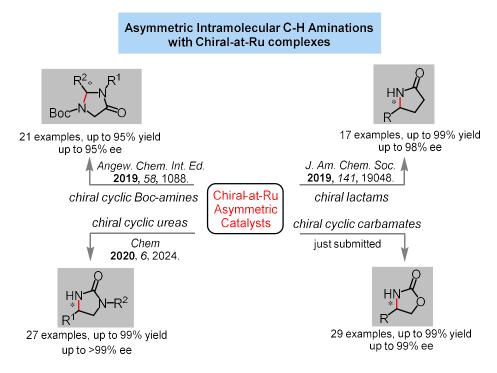
Reference

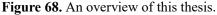
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Chapter 3: Summary and Outlook

3.1 Summary

The synthetic access to a variety of newly modified chiral-at-ruthenium asymmetric catalysts has been accomplished through a chiral-auxiliary-mediated synthetic strategy. These ruthenium catalysts are composed of two chelating inert NHC ligands, which generate metal-centered chirality, and two nitrile ligands. The dicationic complexes are complemented by two hexafluorophosphate or tetrafluoroborate anions.





The excellent catalytic activity of these chiral ruthenium catalysts has been demonstrated through diverse asymmetric intramolecular C–H amination reactions. Accordingly, the efficient asymmetric intramolecular C–H aminations have been applied into the construction of chiral cyclic Boc-amines, lactams, cyclic ureas, and carbamates in high yields and excellent enantioselectivities. Further manipulation of these chiral nitrogen-containing heterocycles by performing the one-step ring-open transformations given rise to the formation of intermediates for the synthesis of drugs, natural products, chiral ligands, and chiral catalysts.

1) Catalytic enantioselective intramolecular C-H amination of 2-azidoacetamides

Asymmetric intramolecular C-H amination of 2-azidoacetamides was catalyzed by a newly modified ruthenium catalyst **Ru7**. The desired imidazolidin-4-one products were formed in excellent yields and enantioselectivities with up to 95% yield and 95% ee (错误!未找到引用源。). This work demonstrated our chiral-at-ruthenium asymmetric catalysts could be applied in challenging intramolecular C-H amination reactions.

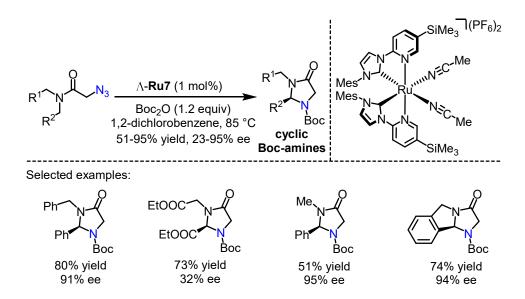


Figure 69. Catalytic enantioselective intramolecular C-H amination of 2-azidoacetamides.

2) Enantioselective synthesis of γ -lactams by intramolecular C–H amidation

The exploration of the catalytic property of the non- C_2 -symmetric ruthenium catalyst **Ru10**, which was first synthesized by the group member Yubiao Hong, has been accomplished. The non- C_2 -symmetric chiral-at-ruthenium catalyst was applied in highly efficient enantioselective intramolecular C(sp³)–H amidation for the synthesis of chiral γ -lactams. Due to the strong electron-donating effect of the rNHC ligands, the metal center of the new ruthenium catalyst is more electron-rich compared with our previous chiral-at-ruthenium catalysts, which instead exhibited Curtius decomposition via transition-metal acyl-nitrenoid intermediates (**Figure 70**).

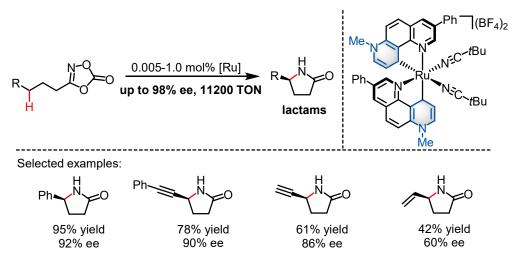


Figure 70. Enantioselective synthesis of γ -lactams by intramolecular C–H amidation.

3) Enantioselective synthesis of 2-imidazolidinones by intramolecular C-H amidation

The enantioselective synthesis of 2-imidazolidinones was accomplished by a chiral-at-metal ruthenium catalyst **Ru7**. It has been demonstrated for the first time on the synthesis of chiral cyclic ureas via ring-closing C-H aminations. The reaction proceeded smoothly and the target chiral cyclic ureas were formed in up to 99% yield and 99% ee (错误!未找到引用源。). Further ring-opening of cyclic ureas provided different chiral 1,2-diamines which were already been reported as intermediates for the synthesis of natural products, and a chiral organo-catalyst.

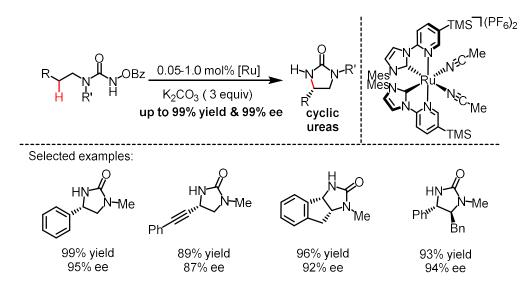


Figure 71. Enantioselective synthesis of cyclic ureas by intramolecular C-H amidation.

4) Enantioselective synthesis of β-amino alcohols by nitrene insertion

The enantioselective synthesis of chiral cyclic carbamates was accomplished by a newly modified ruthenium catalyst **Ru13** catalyzed intramolecular C–H amination of *N*-benzoyloxycarbamates. The

desired chiral cyclic carbamates were obtained in up to 99% yield and 99% ee with broad substrate scope. It can be applied in the amination of benzylic, propargylic, allylic and even non-activated aliphatic C–H bonds. Further ring-opening of cyclic carbamates provided chiral β -amino alcohols as intermediates for the synthesis of natural products and chiral ligand used in asymmetric catalysis (**Figure 72**).

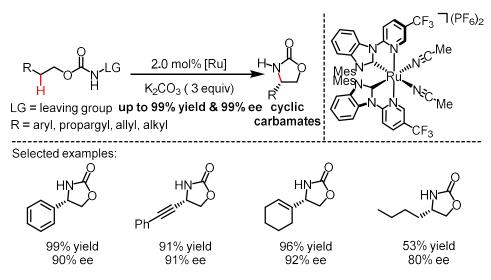


Figure 72. Enantioselective synthesis of cyclic carbamates by intramolecular C-H amidation.

3.2 Outlook

Considering the diverse synthetic applications of C–H amination products to drugs, natural products, chiral ligands, and chiral catalysts. Therefore, it should be of high synthetic value for further investigations on new types of C–H amination reactions.

1) Synthesis of new chiral *N*-heterocycles by asymmetric intramolecular C–H aminations: This thesis showed the synthesis of four different chiral *N*-heterocycles by asymmetric intramolecular C–H aminations, there are still some untapped structure motifs¹ which could be constructed through ruthenium catalyzed asymmetric intramolecular C–H amiantions, such as the structures shown in Figure 73.

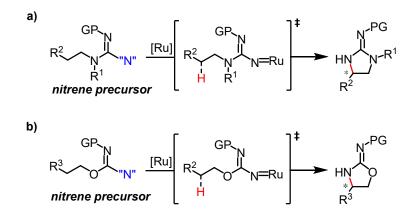


Figure 73. Synthesis of new chiral *N*-heterocycles via asymmetric intramolecular C-H amiantions.

2) Exploit asymmetric intermolecular C–H amination reactions: Compared with intramolecular C–H amination reactions, the intermolecular version is more direct for the synthesis of chiral amines. Cyclic *N*–heterocycles synthesized via asymmetric intramolecular C–H aminations need further ring-opening to get final amine products. Thus, some functional groups are not well tolerated. The intermolecular version has it's special advantages on the direct and rapid synthesis of chiral amines. Our ruthenium catalysts have been proved for asymmetric intramolecular C–H aminations, and the next stage should be asymmetric intermolecular C–H aminations²⁻⁵.

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Chapter 4: Experimental Part

4.1 Materials and Methods

All reactions were carried out under an atmosphere of nitrogen with magnetic stirring. The catalysis reactions were performed using standard Schlenk glassware techniques.

Solvents and Reagents

Solvents were distilled under nitrogen from calcium hydride (CH₂Cl₂, CH₃CN) or sodium/benzophenone (Et₂O, THF and toluene). Super-dry solvents, such as 1,2-dichlorobenzene (from Acros) and DMF (from Sigma Aldrich) were perchased from commercial available source and used directly without further drying. All reagents purchased from Acros, Alfa Aesar, Sigma Aldrich, TCI, ChemPur, Merck and Fluorochem were used without any further purifications.

Chromatographic Methods

The course of the reactions and the column chromatographic elution were monitored by thin layer chromatography (TLC) [Macherey-Nagel (ALUGRAM®Xtra Sil G/UV254)]. Flash column chromatography was performed with silica gel from Merck (particle size 0.040-0.063 mm).

Nuclear Magnetic Resonance Spectroscopy (NMR)

¹H NMR, proton decoupled ¹³C NMR, and proton coupled ¹⁹F NMR spectra were recorded on Bruker Avance 300 system (¹H NMR: 300 MHz and 500 MHz, ¹³C NMR: 75 MHz and 125 MHz, ¹⁹F NMR: 282 MHz) spectrometers at ambient temperature. Chemical shifts are given in ppm on the δ scale, and were determined after calibration to the residual signals of the solvents, which were used as an internal standard. NMR standards were used are as follows: ¹H NMR spectroscopy: $\delta = 7.26$ ppm (CDCl₃), $\delta =$ 5.32 ppm (CD₂Cl₂), $\delta = 2.50$ ppm (DMSO-*d*6), $\delta = 3.31$ ppm (CD₃OD); ¹³C-NMR spectroscopy: $\delta =$ 77.0 ppm (CDCl₃), $\delta = 53.8$ ppm (CD₂Cl₂), $\delta = 118.26$, 1.32 ppm (CD₃CN), $\delta = 206.26$, $\delta = 39.52$ ppm (DMSO-*d*6), $\delta = 49.0$ ppm (CD₃OD). ¹⁹F NMR spectroscopy: $\delta = 0$ ppm (CFCl₃). The characteristic signals were specified from the low field to high field with the chemical shifts (δ in ppm). ¹H NMR spectra peak multiplicities indicated as singlet (s), doublet (d), doublet of doublet (dd), doublet of doublet (ddd), triplet (t), doublet of triplet (dt), quartet (q), multiplet (m). The coupling constant J indicated in hertz (Hz).

High-Performance Liquid Chromatography (HPLC)

Chiral HPLC was performed with an Agilent 1200 Series, Agilent 1260 Series HPLC System or Shimadzu Lc-2030c. All the HPLC conditions were detailed in the individual procedures. The type of the columns, mobile phase and the flow rate were specified in the individual procedures.

Infrared Spectroscopy (IR)

IR measurements were recorded on a Bruker Alpha-P FT-IR spectrometer. The absorption bands were indicated a wave numbers v (cm⁻¹). All substances were measured as films or solids.

Mass Spectrometry (MS)

High-resolution mass spectra were recorded on a Bruker En Apex Ultra 7.0 TFT-MS instrument using ESI or APCI or FD technique. Ionic masses are given in units of m/z for the isotopes with the highest natural abundance.

Circular Dichroism Spectroscopy (CD)

CD spectra were recorded on a JASCO J-810 CD spectropolarimeter. The parameters we used as follows: from 600 nm to 200 nm; data pitch (0.5 nm); band with (1 nm); response (1 second); sensitivity (standard); scanning speed (50 nm/min); accumulation (5 times). The concentration of the compounds for the measurements was 0.2 mM. The formula for converting θ to ε is shown as below.

$$\Delta \varepsilon = \frac{\theta[m \deg]}{32980 \times c(mol/L) \times L(cm)}$$

C = concentration of the sample; L = thickness of the measurement vessel

Crystal Structure Analysis

Crystal X-ray measurements and the crystal structure analysis were carried out by Dr. Klaus Harms (Chemistry Department, Philipps University of Marburg) and Sergei Ivlev (Chemistry Department, Philipps University of Marburg). X-ray data were collected with a Bruker 3 circuit D8 Quest diffractometer with MoKa radiation (microfocus tube with multilayer optics) and Photon 100 CMOS detector. Scaling and absorption correction was performed by using the SADABS software package of Bruker. Structures were solved using direct methods in SHELXS and refined using the full matrix least squares procedure in SHELXL-2013 or SHELXL-2014. The Flack parameter is a factor used to estimate the absolute configuration of the coumounds. Disorder of PF_6 ions, solvent molecules or methylene groups was refined using restraints for both the geometry and the anisotropic displacement factors.

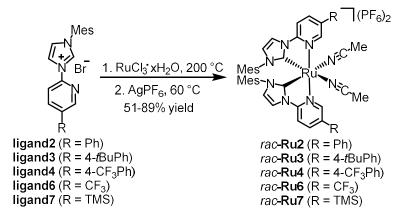
Optical Rotation Polarimeter

Optical rotations were measured on a Krüss P8000-T or Perkin-Elmer 241 polarimeter with $[\alpha]_D^{20}$ or $[\alpha]_D^{25}$ values reported in degrees with concentrations reported in g/100 mL.

4.2 Catalytic Enantioselective Intramolecular C-H Amination of 2-Azidoacetamides

4.2.1 Synthetic of the Ruthenium Catalysts

1) Synthesis of Racemic Ruthenium Catalysts:



General procedure of racemic ruthenium catalysts synthesis: A solution of RuCl₃•xH₂O (1 equiv, 0.24 mmol) and ligand (2 equiv, 0.48 mmol) in ethylene glycol (2.4 mL) was heated at 200 °C for 6 h. The reaction mixture was treated with saturated aqueous NH₄PF₆ after cooling down to room temperature. A yellow precipitate was formed, which was dissolved and extracted with CH₂Cl₂ for three times. The combined organic layers were washed with water and concentrated under reduced pressure to obtain an orange solid, which was dissolved in CH₃CN (3 mL) followed by adding AgPF₆ (76 mg, 0.3 mmol). The mixture was stirred at 60 °C overnight. After cooling to room temperature, the mixture was filtered, and the filtrate was collected, evaporated to dryness and purified by column chromatography on silica gel (CH₂Cl₂:CH₃CN = 200:1 to 20:1) to give pure racemic ruthenium catalyst.

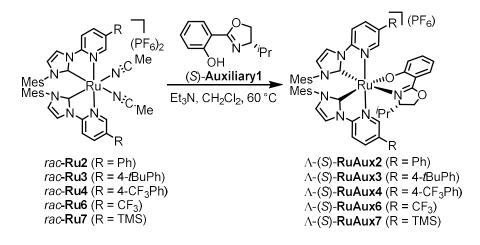
rac-**Ru2**: pale yellow solid (204 mg, 74% yield) was obtained. ¹H NMR (300 MHz, CD₂Cl₂) δ 8.53 (d, J = 2.0 Hz, 2H), 8.08 (dd, J = 8.6, 2.1 Hz, 2H), 8.02 (d, J = 2.3 Hz, 2H), 7.65-7.52 (m, 12H), 6.93 (d, J = 2.3 Hz, 2H), 6.60 (d, J = 6.3 Hz, 4H), 2.31 (s, 6H), 2.01 (s, 6H), 1.97 (s, 6H), 1.52 (s, 6H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 189.6, 152.4, 149.2, 140.6, 137.1, 135.6, 135.3, 135.1, 134.3, 134.2, 130.2, 130.2, 129.8, 129.6, 127.0, 126.0, 125.0, 118.0, 111.8, 21.1, 17.7, 17.5, 4.0. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -71.32, -73.84. IR (film): v (cm⁻¹) 2922, 2852, 1511, 1483, 1453, 1422, 1290, 1247, 1142, 1081, 1036, 952, 930, 835, 764, 745, 696, 678, 620, 591, 556, 542, 488, 457, 434, 388.

rac-Ru3: pale yellow solid (212 mg, 0.17 mmol, 56% yield) was obtained from ligand 3 (0.6 mmol). ¹H NMR (300 MHz, CD₂Cl₂) δ 8.51 (d, J = 1.9 Hz, 2H), 8.07 (dd, J = 8.6, 2.1 Hz, 2H), 8.02 (d, J = 2.3 Hz, 2H), 7.68-7.52 (m, 10H), 6.93 (d, J = 2.3 Hz, 2H), 6.60 (s, 4H), 2.31 (s, 6H), 2.03 (s, 6H), 1.95 (s, 6H), 1.51 (s, 6H), 1.41 (s, 18H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 189.3, 153.0, 151.9, 148.7, 140.3, 136.5, 135.2, 135.0, 134.0, 134.0, 131.9, 130.0, 129.4, 127.0, 126.4, 125.6, 124.7, 117.8, 111.7, 35.1, 31.4, 20.8, 17.5, 17.2, 3.7. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -72.21, -74.73. IR (film): v (cm⁻¹) 3138, 2959, 1611, 1490, 1425, 1373, 1309, 1258, 1103, 1033, 932, 834, 738, 629, 588, 554, 434.

rac-**Ru4**: pale yellow solid (348 mg, 89% yield) was obtained from **ligand 4** (0.6 mmol). ¹**H NMR** (300 MHz, CD₂Cl₂) δ 8.57 (d, J = 1.9 Hz, 2H), 8.10 (dd, J = 8.7, 2.1 Hz, 2H), 8.03 (d, J = 2.3 Hz, 2H), 7.89 (d, J = 8.3 Hz, 4H), 7.78 (d, J = 8.2 Hz, 4H), 7.67 (d, J = 8.6 Hz, 2H), 6.93 (d, J = 2.3 Hz, 2H), 6.64 (s, 2H), 6.57 (s, 2H), 2.31 (s, 6H), 2.00 (s, 12H), 1.52 (s, 6H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 189.7, 152.9, 149.4, 140.4, 138.6, 137.3, 135.2, 134.2, 134.1, 134.0, 131.6, 131.1, 130.2, 129.3, 127.5, 127.1, 127.0, 127.0, 126.9, 126.3, 126.0, 125.2, 122.7, 117.9, 111.9, 21.0, 17.5, 17.3, 3.7. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -62.90, -71.13, -73.65. IR (film): v (cm⁻¹) 1612, 1492, 1426, 1322, 1255, 1166, 1117, 1069, 823, 689, 552.

rac-**Ru6**: pale yellow solid (246 mg, 72% yield) was obtained from **ligand 6** (0.6 mmol). ¹**H NMR** (**300 MHz, CD₂Cl₂)** δ 8.55 (s, 2H), 8.21-7.93 (m, 4H), 7.79 (d, *J* = 8.7 Hz, 2H), 6.94 (d, *J* = 2.2 Hz, 2H), 6.76 (s, 2H), 6.64 (s, 2H), 2.32 (s, 6H), 2.19 (s, 6H), 2.00 (s, 6H), 1.47 (s, 6H). ¹³C **NMR (75 MHz, CD₂Cl₂)** δ 189.9, 155.8, 148.6, 141.0, 136.6, 135.1, 134.0, 133.6, 130.2, 126.5, 125.8, 118.6, 112.5, 20.8, 17.3, 17.3, 3.9. ¹⁹F **NMR** (235 MHz, CD₂Cl₂) δ -62.73, -71.10, -74.12. **IR (film):** *v* (cm⁻¹) 3144, 2926, 2174, 1620, 1505, 1427, 1382, 1325, 1256, 1174, 1139, 1073, 1039, 929, 830, 736, 708, 626, 583, 554, 506, 463, 435.

rac-**Ru7**: pale yellow solid (175 mg, 51% yield) was obtained from **ligand** 7 (0.6 mmol). ¹**H NMR** (**300 MHz, CD₂Cl₂)** δ 8.43 (s, 2H), 7.97 (s, 2H), 7.88 (d, *J* = 8.1 Hz, 2H), 7.48 (d, *J* = 8.1 Hz, 2H), 6.86 (d, *J* = 1.8 Hz, 2H), 6.63 (d, *J* = 6.5 Hz, 4H), 2.31 (s, 6H), 2.17 (s, 6H), 1.98 (s, 6H), 1.49 (s, 6H), 0.40 (s, 18H). ¹³**C NMR (75 MHz, CD₂Cl₂)** δ 190.0, 154.7, 153.6, 143.8, 140.1, 135.0, 134.2, 134.0, 129.9, 129.7, 125.8, 124.6, 117.7, 111.0, 21.3, 17.6, 17.5, 3.6, -1.4. ¹⁹F NMR (235 MHz, CD₂Cl₂) δ -71.14, -74.16. **IR (film):** *v* (cm⁻¹) 2955, 2156, 2090, 2054, 2010, 1982, 1948, 1685, 1634, 1595, 1491, 1421, 1327, 1304, 1252, 1194, 1143, 1089, 1035, 931, 833, 760, 694, 627, 590, 553, 429.



2) Intermediate Ruthenium Auxiliary Complexes:

General procedure of chiral ruthenium catalysts synthesis: A mixture of racemic ruthenium catalyst (1 equiv, 0.11 mmol), chiral auxiliary (S)-Auxiliary1 (2 equiv, 62.2 mg, 0.22 mmol) and triethylamine (3 equiv, 50 μ L, 0.33 mmol) in CH₂Cl₂ (3.6 mL) was heated at 60 °C for 18 h. The reaction mixture was cooled to room temperature and concentrated to dryness. The residue was subjected to flash silica gel chromatography (CH₃CN:CH₂Cl₂= 1:600 to 1:50) to isolate the first diastereomer which was assigned as Λ -(S)-RuAux2-4,6-7. The absolute configuration of ruthenium auxiliary complexes were assigned as Λ -(S)-configurataion, due to the auxiliary complexes after removal of auxiliary and got the final ruthenium catalyst's absolute configuration on chapter 2.1.2, the catalyst's configuration was assigned by comparing the CD spectra with previous reported one), thus we assigned here the ruthenium auxiliary complexes as Λ -(S)-configurataion.

Λ-(*S*)-**RuAux2**: orange solid (33.8 mg, 28% yield). ¹**H NMR (500 MHz, CD₂Cl₂)** δ 8.74 (d, *J* = 2.0 Hz, 1H), 8.19 (d, *J* = 2.0 Hz, 1H), 7.95 (d, *J* = 2.3 Hz, 1H), 7.92 (dd, *J* = 8.6, 2.2 Hz, 1H), 7.87 (d, *J* = 2.3 Hz, 1H), 7.74 (dd, *J* = 8.6, 2.2 Hz, 1H), 7.54-7.43 (m, 7H), 7.42-7.38 (m, 3H), 7.37-7.31 (m, 3H), 7.00 (ddd, *J* = 8.7, 6.9, 1.9 Hz, 1H), 6.89 (d, *J* = 2.3 Hz, 1H), 6.84 (d, *J* = 2.2 Hz, 1H), 6.56 (s, 1H), 6.51 (s, 1H), 6.50-6.45 (m, 3H), 6.24-6.18 (m, 1H), 4.29 (dd, *J* = 9.3, 3.2 Hz, 1H), 4.12 (t, *J* = 9.1 Hz, 1H), 3.91 (dt, *J* = 8.9, 3.1 Hz, 1H), 2.28 (s, 3H), 2.05 (s, 3H), 2.00 (s, 3H), 1.95 (s, 3H), 1.61 (s, 3H), 1.42 (s, 3H), 0.55 (d, *J* = 7.0 Hz, 3H), 0.30-0.19 (m, 1H), -0.03 (d, *J* = 6.8 Hz, 3H). ¹³C **NMR (126 MHz, CD₂Cl₂)** δ 197.5, 196.2, 172.0, 165.2, 153.0, 152.9, 148.5, 148.0, 139.5, 137.3, 135.8, 135.4, 135.1, 135.0, 134.7, 134.4, 134.2, 133.9, 133.9, 133.6, 133.4, 130.2, 129.8, 129.6, 129.4, 129.4, 129.1, 129.0, 126.4, 125.6, 125.1, 123.8, 116.6, 115.9, 112.7, 110.8, 110.5, 110.0, 74.9, 66.6, 30.1, 30.0, 20.9,

20.9, 18.8, 18.5, 17.9, 17.9, 17.4, 13.5. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -71.73, -74.25. HRMS (ESI, *m/z*) calcd. for C₅₈H₅₆RuN₇O₂ [M-PF₆]⁺: 984.3533, found: 984.3565. IR (film): *v* (cm⁻¹) 1605, 1537, 1507, 1471, 1413, 1355, 1322, 1250, 1222, 1067, 969, 921, 836, 756, 685, 580, 553.

A-(*S*)-**RuAux3**: orange solid (45.9 mg, yield: 34%). ¹**H NMR (300 MHz, CD₂Cl₂)** δ 8.73 (d, J = 2.0 Hz, 1H), 8.20 (d, J = 1.9 Hz, 1H), 7.95-7.82 (m, 3H), 7.75 (dd, J = 8.6, 2.1 Hz, 1H), 7.49 (dt, J = 20.9, 9.3 Hz, 6H), 7.40-7.22 (m, 5H), 6.99 (ddd, J = 8.7, 6.9, 1.9 Hz, 1H), 6.87 (dd, J = 13.1, 2.3 Hz, 2H), 6.61-6.37 (m, 5H), 6.22 (t, J = 7.1 Hz, 1H), 4.29 (dd, J = 9.2, 3.2 Hz, 1H), 4.15 (t, J = 9.0 Hz, 1H), 3.96-3.87 (m, 1H), 2.28 (s, 3H), 2.06 (s, 3H), 1.99 (s, 3H), 1.93 (s, 3H), 1.60 (s, 3H), 1.42 (s, 3H), 1.39 (s, 9H), 1.35 (s, 9H), 0.55 (d, J = 6.9 Hz, 3H), 0.33-0.16 (m, 1H), -0.03 (d, J = 6.8 Hz, 3H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 197.1, 195.9, 171.7, 165.0, 152.7, 152.7, 152.4, 152.3, 148.1, 147.7, 139.5, 139.5, 137.2, 135.0, 134.9, 134.7, 134.3, 134.1, 133.9, 133.8, 133.4, 133.3, 132.6, 132.4, 130.1, 129.7, 129.4, 129.4, 129.0, 126.8, 126.5, 126.0, 126.0, 125.5, 125.0, 123.8, 116.6, 115.8, 112.6, 110.8, 110.5, 109.9, 74.9, 66.5, 35.0, 34.9, 31.4, 31.3, 30.0, 20.9, 20.8, 18.8, 18.5, 17.9, 17.9, 17.4, 13.5. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -71.68, -74.19. IR (film): v (cm⁻¹) 2958, 1607, 1486, 1420, 1372, 1320, 1251, 1151, 1068, 1032, 926, 836, 747, 688, 553. HRMS (ESI, *m/z*) calcd. for C₆₆H₇₂RuN₇O₂ [M-PF₆]⁺: 1096.4786, found: 1096.4824.

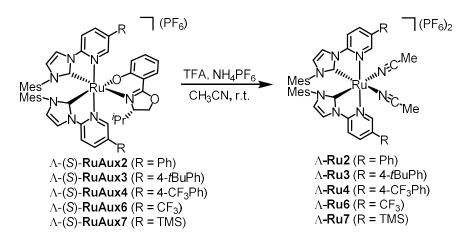
Λ-(*S*)-**RuAux4**: orange solid (56.7 mg, yield: 41%). ¹**H NMR (300 MHz, CD₂Cl₂)** δ 8.78 (d, *J* = 2.2 Hz, 1H), 8.22-8.17 (m, 1H), 8.02-7.99 (m, 1H), 7.97 (dd, *J* = 8.6, 1.9 Hz, 1H), 7.93-7.89 (m, 1H), 7.82-7.76 (m, 3H), 7.73 (s, 1H), 7.71 (s, 1H), 7.58 (d, *J* = 8.7 Hz, 1H), 7.53-7.41 (m, 6H), 7.05-6.97 (m, 1H), 6.91 (d, *J* = 2.3 Hz, 1H), 6.86 (d, *J* = 2.3 Hz, 1H), 6.58 (s, 1H), 6.54 (s, 1H), 6.51-6.44 (m, 3H), 6.28-6.22 (m, 1H), 4.33-4.27 (m, 1H), 4.10 (t, *J* = 9.1 Hz, 1H), 3.92-3.85 (m, 1H), 2.27 (s, 3H), 2.04 (s, 3H), 2.02 (s, 3H), 1.96 (s, 3H), 1.61 (s, 3H), 1.43 (s, 3H), 0.55 (d, *J* = 7.0 Hz, 3H), 0.27-0.15 (m, 1H), -0.04 (d, *J* = 6.9 Hz, 3H). ¹³**C NMR (75 MHz, CD₂Cl₂)** δ 197.5, 196.12 172.0, 165.3, 153.8, 153.6, 148.7, 148.2, 139.6, 139.5, 139.0, 137.2, 135.2, 134.9, 134.7, 134.5, 134.4, 134.1, 134.0, 132.1, 131.8, 130.9, 130.8, 130.7, 130.5, 130.1, 129.9, 129.6, 129.2, 128.8, 126.9, 126.8, 126.8, 126.6, 126.6, 126.5, 125.75, 125.56, 125.54, 125.34, 123.79, 123.40, 123.38, 116.84, 116.13, 112.87, 110.82, 110.32, 74.8, 66.6, 30.1, 20.9, 20.9, 18.8, 18.5, 17.9, 17.8, 17.4, 13.5. ¹⁹**F NMR (282 MHz, CD₂Cl₂)** δ -63.02, -63.04, -71.64, -74.16. **IR (film):** ν (cm⁻¹) 2925, 1608, 1531, 1489, 1422, 1379, 1322, 1285, 1251, 1166, 1117, 1068, 1018, 925, 836, 757, 686, 596, 553, 507, 458, 429. **HRMS (ESI**,

m/z) calcd. for C₆₀H₅₄RuF₆N₇O₂ [M-PF₆]⁺: 1120.3281, found: 1120.3317.

Λ-(*S*)-**RuAux6**: orange solid (39.2 mg, yield: 32%). ¹**H NMR (300 MHz, CD₂Cl₂)** δ 8.93 (s, 1H), 8.15 (s, 1H), 8.06 (d, J = 2.4 Hz, 1H), 7.91 (dt, J = 5.3, 2.4 Hz, 2H), 7.73 (dd, J = 8.6, 1.7 Hz, 1H), 7.66 (d, J = 8.7 Hz, 1H), 7.45 (d, J = 8.7 Hz, 1H), 7.38 (dd, J = 8.1, 1.8 Hz, 1H), 6.93 (ddd, J = 8.7, 6.3, 2.6 Hz, 2H), 6.86 (d, J = 2.3 Hz, 1H), 6.73 (d, J = 5.3 Hz, 2H), 6.62 (s, 1H), 6.50 (s, 1H), 6.33 (dd, J = 8.6, 0.9 Hz, 1H), 6.23 (ddd, J = 8.0, 6.9, 1.1 Hz, 1H), 4.29 (dd, J = 9.4, 3.3 Hz, 1H), 4.14 (t, J = 9.1Hz, 1H), 3.67 (dt, J = 8.7, 3.1 Hz, 1H), 2.25 (s, 3H), 2.19 (s, 3H), 2.09 (s, 3H), 1.57 (s, 3H), 1.36 (s, 3H), 0.50 (d, J = 7.0 Hz, 3H), 0.06 (d, J = 6.8 Hz, 3H), -0.11-0.31 (m, 1H).¹³**C NMR (75 MHz, CD₂Cl₂**) δ 172.2, 171.9, 165.6, 156.2, 155.7, 147.8, 147.4, 139.8, 137.0, 136.8, 134.5, 134.2, 134.1, 133.9, 133.9, 133.6, 133.5, 133.4, 130.0, 129.7, 129.4, 129.2, 125.9, 125.6, 122.9, 116.6, 116.2, 114.6, 113.0, 110.5, 110.3, 110.0, 74.1, 66.4, 30.0, 20.4, 20.3, 18.4, 17.8, 17.4, 17.2, 17.0, 16.9, 13.1. ¹⁹F NMR (235 MHz, CD₂Cl₂) δ -62.73, -71.10, -74.12. **IR (film):** *v* (cm⁻¹) 3141, 2965, 2924, 2868, 2120, 2083, 2050, 1976, 1928, 1608, 1536, 1500, 1472, 1421, 1381, 1321, 1253, 1225, 1170, 1134, 1066, 1035, 925, 834, 757, 706, 688, 627, 582, 555, 531, 504, 463, 435, 391. **HRMS (ESI,** *m/z***)** calcd. for C₄₈H₄₆F₆RuN₇O₂ [M-PF₆]⁺: 968.2655, found: 968.2665.

A-(*S*)-**RuAux7**: orange solid (44.4 mg, yield: 36%). ¹**H NMR (300 MHz, CD₂Cl₂)** δ 8.53 (s, 1H), 8.05 (s, 1H), 7.90 (d, J = 2.3 Hz, 1H), 7.82 (d, J = 2.2 Hz, 1H), 7.70 (dd, J = 8.1, 1.3 Hz, 1H), 7.60 (dd, J = 8.1, 1.3 Hz, 1H), 7.45 (dd, J = 8.0, 1.6 Hz, 1H), 7.35 (d, J = 8.1 Hz, 1H), 7.26 (d, J = 8.1 Hz, 1H), 7.02-6.91 (m, 1H), 6.85 (d, J = 2.3 Hz, 1H), 6.78 (d, J = 2.2 Hz, 1H), 6.57 (t, J = 9.2 Hz, 4H), 6.36 (d, J = 8.4 Hz, 1H), 6.17 (d, J = 6.8 Hz, 1H), 4.26 (dd, J = 9.2, 3.2 Hz, 1H), 4.11 (dd, J = 11.3, 6.8 Hz, 1H), 3.88 (d, J = 8.5 Hz, 1H), 2.24 (s, 3H), 1.60 (s, 3H), 1.41 (s, 3H), 0.50 (d, J = 6.8 Hz, 3H), 0.22 (s, 9H), 0.17 (s, 9H), 0.02 (dd, J = 14.3, 7.9 Hz, 1H), -0.09 (d, J = 6.6 Hz, 3H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 154.5, 154.4, 154.3, 141.3, 141.1, 139.3, 139.2, 137.4, 135.2, 135.0, 134.9, 134.4, 134.0, 133.7, 132.5, 131.9, 130.1, 129.9, 129.6, 129.4, 129.2, 125.4, 125.0, 123.8, 116.6, 115.8, 110.0, 109.4, 74.9, 66.4, 30.0, 21.2, 21.1, 18.9, 18.7, 18.2, 17.9, 17.6, 13.7, -1.3, -1.4. HRMS (ESI, *m/z*) calcd. for C₅₂H₆₄RuN₇O₂Si₂ [M-PF₆]⁺: 976.3698, found: 978.3719. **IR (film):** *v* (cm⁻¹) 2953, 2158, 2117, 2024, 1605, 1537, 1487, 1415, 1379, 1353, 1321, 1249, 1142, 1064, 1035, 960, 924, 836, 754, 687, 622, 584, 554, 505, 413.

3) Enantiopure Ruthenium Catalysts:



General Procedure: To a mixture of single ruthenium auxiliary complex (0.044 mmol) in CH₃CN (4.4 mL) was added CF₃COOH (0.44 mmol). The reaction mixture was stirred at room temperature for 5 min, then concentrated under reduced pressure. The residue was subjected to flash silica gel chromatography (CH₃CN:CH₂Cl₂ = 1:100 to 1:10) and added 2 g NH₄PF₆ above the seasand band (to change the counter anion on the column) to afford chiral ruthenium catalyst as pale yellow solid. The counter anion exchange will lead to two yellow bands on the column but they are the same compound. It should be purified within 30 min (It should be eluted fast!).

 Λ -**Ru2**: pale yellow solid (44.8 mg, 88% yield) All other spectroscopic data of enantiopure ruthenium catalyst Λ -**Ru2** were in agreement with the racemic catalyst *rac*-**Ru2**.

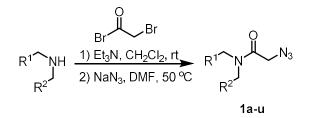
 Λ -Ru3: pale yellow solid (46 mg, yield: 82%). All other spectroscopic data of enantiopure ruthenium catalyst Λ -Ru3 were in agreement with the racemic catalyst *rac*-Ru3.

 Λ -Ru4: pale yellow solid (54 mg, yield: 95%). All other spectroscopic data of enantiopure ruthenium catalyst Λ -Ru4 were in agreement with the one of racemic catalyst *rac*-Ru4.

 Λ -**Ru6**: pale yellow solid (43.8 mg, 87% yield). All other spectroscopic data of enantiopure ruthenium catalyst Λ -**Ru6** were in agreement with the racemic catalyst *rac*-**Ru6**.

 Λ -Ru7: pale yellow solid (41.6 mg, 82% yield). All other spectroscopic data of enantiopure ruthenium catalyst Λ -Ru7 were in agreement with the racemic catalyst *rac*-Ru7.

4.2.2 Synthesis of Substrates



General procedure for the preparation 2-azidoacetamides. To a solution of amines (1.0 equiv, 5 mmol) and Et_3N (1.0 equiv, 5 mmol) in fresh distilled CH_2Cl_2 (10 mL) was added bromoacetylbromide (1.05 equiv, 5.25 mmol) in CH_2Cl_2 (2 mL) dropwise at 0 °C. The resulting mixture was then reacted at room temperature for 4 h. After that, the reaction was quenched by saturated aqueous NaHCO₃ (10 mL) at 0 °C. The aqueous phase was extracted three times with diethyl ether. The combined organic phases were washed with brine, dried over Na₂SO₄. After removal of the solvent under reduced pressure, the crude mixture was used without any further purification.

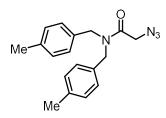
To a solution of the above 2-bromoacetamide in DMF (10 mL, 0.5 M) was added sodium azide (650.1 mg, 2.0 equiv). The resulting solution was stirred at 50 °C overnight (14 h to 16 h, monitored by TLC). After that, a 1:1 mixture of H₂O with diethyl ether were added to the reaction mixture, and the aqueous phase was extracted three times with 10 mL diethyl ether. The combined organic phases were washed several times (5-8 times) with 10 mL H₂O (deionized) to remove DMF. Then it was washed with brine, and dried over anhydrous Na₂SO₄. After removal of the solvent under reduced pressure, the crude mixture was purified by flash column chromatography on a silica gel column (*n*-hexane:EtOAc = 1:5 to 1:3, R_f value = 0.4 to 0.6) which resulted in the analytical pure azides.

Compound 1a

1a: white solid. Yield: 95%. ¹H NMR (300 MHz, CDCl₃) δ 7.34 (m, 8H), 7.17 (m, 2H), 4.68 (s, 2H),
4.41 (s, 2H), 4.01 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 168.1, 136.6, 135.6, 129.3, 128.9, 128.6,
128.1, 127.9, 126.4, 50.8, 49.6, 49.1. IR (film): v (cm⁻¹) 3028, 2907, 2229, 2169, 2110, 1649, 1492,
1423, 1359, 1290, 1271, 1208, 1168, 1077, 1026, 993, 949, 816, 746, 697, 622, 570, 509, 456.

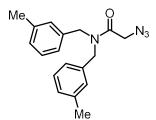
85

Compound 1b



1b: white solid. Yield: 88%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.31-7.17 (m, 2H), 7.07 (dd, *J* = 17.3, 11.3 Hz, 4H), 6.93 (d, *J* = 6.6 Hz, 2H), 4.62 (s, 2H), 4.34 (s, 2H), 3.97 (s, 2H), 2.35 (s, 6H). ¹³**C NMR (75 MHz, CDCl₃)** δ 168.1, 139.2, 138.7, 136.6, 135.7, 129.4, 129.2, 128.9, 128.7, 127.0, 125.7, 123.5, 50.9, 49.6, 49.2, 21.5. **IR (film):** *v* (cm⁻¹) 3308, 2944, 2099, 1923, 1654, 1619, 1545, 1510, 1474, 1416, 1364, 1321, 1221, 1162, 1115, 1060, 1014, 974, 937, 816, 755, 711, 662, 632, 586, 549, 525, 490, 432, 392.

Compound 1c



1c: white solid. Yield: 79%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.23-7.09 (m, 6H), 7.01 (d, *J* = 7.7 Hz, 2H), 4.59 (s, 2H), 4.31 (s, 2H), 3.96 (s, 2H), 2.35 (s, 6H). ¹³**C NMR (75 MHz, CDCl₃)** δ 170.0, 137.0, 137.7, 133.7, 132.6, 130.0, 129.6, 128.7, 126.4, 50.1, 49.2, 48.7, 21.2. **IR (film):** *v* (cm⁻¹) 2926, 2201, 2098, 1889, 1659, 1485, 1446, 1404, 1343, 1282, 1211, 1163, 1085, 1012, 981, 930, 833, 800, 717, 662, 613, 569, 547, 477, 445, 409.

Compound 1d

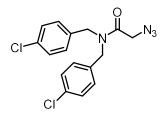
Me

1d: white solid. Yield: 93%. ¹H NMR (300 MHz, CDCl₃) δ 7.16 (t, J = 26.0 Hz, 8H), 4.72 (s, 2H), 4.29 (s, 2H), 3.95 (s, 2H), 2.17 (d, J = 28.2 Hz, 6H). ¹³C NMR (75 MHz, CDCl₃) δ 168.3, 136.9, 135.5, 134.0, 133.3, 130.8, 128.4, 127.9, 126.9, 126.4, 124.5, 50.8, 46.9, 19.0. IR (film): v (cm⁻¹) 3024, 2919, 2100, 1656, 1608, 1420, 1349, 1277, 1210, 1175, 1093, 1038, 985, 949, 889, 822, 777, 741, 696, 644, 583, 550, 518, 476, 429.

Compound 1e

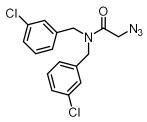
1e: white solid. Yield: 90%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.17 (d, *J* = 8.0 Hz, 2H), 7.04 (d, *J* = 8.0 Hz, 2H), 6.96-6.73 (m, 4H), 4.54 (s, 2H), 4.28 (s, 2H), 3.96 (s, 2H), 3.80 (s, 6H). ¹³**C NMR (75 MHz, CDCl₃)** δ 167.8, 159.4, 159.3, 130.0, 128.7, 127.7, 127.4, 114.6, 114.2, 55.4, 50.8, 48.8, 48.1. **IR** (film): *v* (cm⁻¹) 2998, 2925, 2835, 2170, 2098, 1646, 1610, 1583, 1505, 1457, 1418, 1357, 1294, 1237, 1170, 1107, 1026, 926, 835, 808, 755, 709, 631, 598,515, 474, 439, 395.

Compound 1f



1f: white solid. Yield: 91%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.32 (dd, *J* = 15.9, 8.0 Hz, 4H), 7.16 (d, *J* = 7.9 Hz, 2H), 7.06 (d, *J* = 7.8 Hz, 2H), 4.56 (s, 2H), 4.33 (s, 2H), 3.96 (s, 2H). ¹³**C NMR (75 MHz, CDCl₃)** δ 168.0, 134.9, 134.2, 133.9, 130.0, 129.6, 129.1, 127.9, 50.9, 49.2, 48.4. **IR (film):** *v* (cm⁻¹) 3119, 2923, 2101, 1657, 1503, 1450, 1421, 1387, 1348, 1282, 1249, 1188, 1148, 1075, 1010, 940, 884, 813, 733, 636, 596, 552, 508, 416.

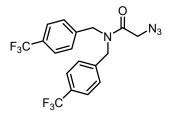
Compound 1g



1g: white solid. Yield: 90%. ¹H NMR (300 MHz, CDCl₃) δ 7.35-6.84 (m, 8H), 4.51 (s, 2H), 4.28 (s,

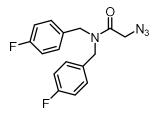
2H), 3.90 (s, 2H). ¹³C NMR (**75 MHz, CD₂Cl₂**) δ 168.4, 139.2, 138.2, 135.4, 134.9, 130.9, 130.5, 128.7, 128.6, 128.3, 127.0, 125.1, 51.0, 49.8, 49.2. **IR (film):** *v* (cm⁻¹) 3066, 3042, 2923, 2217, 2098, 1647, 1603, 1505, 1460, 1417, 1353, 1279, 1210, 1154, 1097, 1022, 985, 928, 849, 816, 768, 709, 631, 583, 549,519, 489, 423, 400.

Compound 1h



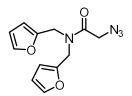
1h: white solid. Yield: 79%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.62 (dd, *J* = 17.3, 7.9 Hz, 4H), 7.34 (d, *J* = 7.7 Hz, 2H), 7.26 (d, *J* = 7.6 Hz, 2H), 4.68 (s, 2H), 4.46 (s, 2H), 4.00 (s, 2H). ¹³**C NMR (75 MHz, CDCl₃)** δ 168.2, 140.3, 139.4, 128.8, 126.8, 126.4, 125.9, 50.9, 49.8, 48.9. ¹⁹**F NMR** (235 MHz, CD₂Cl₂) δ -62.90, -62.97. **IR (film):** *v* (cm⁻¹) 3013, 2921, 2860, 2227, 2193, 2097, 1653, 1510, 1446, 1414, 1353, 1269, 1201, 1167, 1108, 1032, 983, 926, 798, 749, 697, 617, 591, 554, 527, 477, 417, 391.

Compound 1i



1i: white solid. Yield: 93%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.32-6.87 (m, 8H), 4.56 (s, 2H), 4.34 (s, 2H), 3.97 (s, 2H). ¹³**C NMR (75 MHz, CDCl₃)** δ 167.9, 164.1, 160.8, 132.3, 131.1, 130.4, 130.3, 128.2, 128.1, 116.4, 116.1, 115.9, 115.6, 50.8, 49.0, 48.2. ¹⁹**F NMR** (235 MHz, CD₂Cl₂) δ -115.06, -115.44. **IR (film):** *v* (cm⁻¹) 3023, 2944, 2912, 2183, 2099, 1649, 1606, 1450, 1352, 1284, 1215, 1105, 1049, 1024, 990, 948, 934, 875, 838, 811, 747, 713, 639, 583, 547, 512, 483, 435, 408.



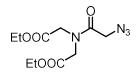


1j: colorless oil. Yield: 88%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.33 (d, *J* = 6.7 Hz, 2H), 6.29 (s, 2H), 6.23 (d, *J* = 19.5 Hz, 2H), 4.56 (s, 2H), 4.32 (s, 2H), 4.09 (s, 2H). ¹³**C NMR (75 MHz, CDCl₃)** δ 167.3, 150.0, 149.0, 143.0, 142.5, 110.5, 109.2, 108.8, 50.4, 43.0, 41.3. **IR (film):** *v* (cm⁻¹) 3103, 2921, 2100, 1653, 1536, 1446, 1421, 1367, 1331, 1280, 1239, 1203, 1152, 1077, 1041, 983, 934, 844, 825, 697, 586, 551, 514, 463, 408.

Compound 1k

1k: colorless oil. Yield: 82%. ¹H NMR (300 MHz, CDCl₃) δ 7.28-6.88 (m, 8H), 4.56 (s, 2H), 4.34 (s, 2H), 3.97 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 167.1, 138.6, 127.7, 127.3, 126.8, 126.3, 126.1, 125.9, 44.9, 43.3. IR (film): v (cm⁻¹) 2986, 2105, 1738, 1669, 1459, 1412, 1375, 1347, 1292, 1184, 1100, 1057, 1022, 966, 930, 867, 828, 736, 632, 556, 522, 436, 400.

Compound 11



11: colorless oil. Yield: 64%. ¹H NMR (300 MHz, CDCl₃) δ 4.18 (m, 6H), 4.05 (s, 2H), 3.93 (s, 2H),
1.25 (q, J = 7.0 Hz, 6H). ¹³C NMR (75 MHz, CDCl₃) δ 168.7, 168.4, 168.1, 62.1, 61.6, 50.5, 49.7,
48.4, 14.2. IR (film): v (cm⁻¹) 3061, 3027, 2923, 2853, 2196, 2150, 2098, 1951, 1700, 1644, 1485,
1449, 1419, 1369, 1263, 1223, 1180, 1150, 1080, 1030, 989, 920, 826, 792, 747, 699, 627, 598, 550,
496, 421.

Compound 1m

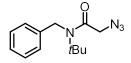
1m: colorless oil. Yield: 86%. **¹H NMR (300 MHz, CDCl₃)** δ 7.33 (m, 5H), 4.63 (s, 2H), 3.99 (s, 2H), 2.88 (s, 3H). with rotation ratio: 1.5:1. ¹³C NMR (75 MHz, CDCl₃) δ 167.8, 167.4, 136.6, 135.7, 129.3, 128.9, 128.4, 128.2, 127.9, 126.4, 53.0, 51.4, 50.8, 50.7, 34.5, 34.0. **IR (film):** *v* (cm⁻¹) 3047, 2956, 2919, 2860, 2283, 2196, 2152, 2099, 1727, 1648, 1418, 1353, 1320, 1272, 1227, 1172, 1147, 1087, 1010, 943, 912, 835, 797, 743, 649, 591, 559, 514, 419.

Compound 1n

Мe

1n: colorless oil. Yield: 95%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.17 (d, *J* = 8.6 Hz, 2H), 6.84 (d, *J* = 8.6 Hz, 2H), 4.51 (s, 2H), 3.93 (s, 2H), 3.77 (s, 3H), 2.82 (s, 3H). With rotation ratio = 1.6:1. ¹³**C NMR (75 MHz, CDCl₃)** δ 167.6, 167.2, 159.4, 159.2, 129.7, 128.7, 127.7, 127.5, 114.6, 114.1, 55.4, 55.3, 52.3, 50.7, 50.6, 34.1, 33.7. **IR (film):** *v* (cm⁻¹) 3069, 3002, 2939, 2862, 2192, 2160, 2106, 2026, 1666, 1614, 1509, 1445, 1418, 1341, 1300, 1262, 1223, 1190, 1165, 1100, 1031, 994, 916, 864, 843, 774, 731, 646, 557, 505, 481, 391.

Compound 1o

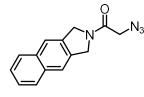


10: white solid. Yield: 94%. ¹**H NMR (300 MHz, CDCl₃)** δ 7.30 (dd, *J* = 10.0, 4.6 Hz, 2H), 7.20 (dd, *J* = 9.2, 5.3 Hz, 1H), 7.11 (d, *J* = 7.1 Hz, 2H), 4.44 (s, 2H), 3.73 (s, 2H), 1.40 (s, 9H). ¹³**C NMR (75 MHz, CDCl₃)** δ 168.6, 138.3, 129.2, 127.6, 125.4, 58.9, 52.6, 48.0, 28.6. **IR (film):** *v* (cm⁻¹) 3099, 3027, 2928, 2902, 2854, 2225, 2176, 2147, 2103, 1744, 1659, 1573, 1439, 1416, 1339, 1253, 1183, 1153, 1114, 994, 932, 908, 867, 841, 816, 749, 693, 663, 634, 560, 488, 429.

Compound 1p

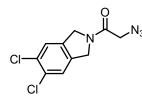
1p: white solid. Yield: 74%. **¹H NMR (300 MHz, CD₂Cl₂)** δ 7.31 (m, 4H), 4.81 (s, 2H), 4.73 (s, 2H), 3.94 (s, 2H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 166.3, 136.4, 136.6, 128.4, 128.0, 123.4, 123.1, 52.8, 52.1, 51.2. **IR (film):** *v* (cm⁻¹) 3062, 3014, 2959, 2920, 2857, 2262, 2185, 2104, 1765, 1652, 1502, 1431, 1339, 1312, 1270, 1235, 1171, 993, 911, 873, 836, 775, 747, 639, 576, 541, 478, 454, 395.

Compound 1q



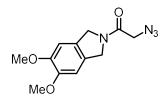
1q: white solid. Yield: 61%. **¹H NMR** (300 MHz, CD₂Cl₂) δ 7.90-7.81 (m, 2H), 7.78 (s, 1H), 7.74 (s, 1H), 7.53-7.43 (m, 2H), 4.96 (s, 2H), 4.87 (s, 2H), 3.99 (s, 2H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 166.4, 135.2, 135.0, 133.8, 133.5, 128.2, 128.1, 126.5, 126.4, 121.9, 121.7, 52.3, 51.5, 51.3. **IR (film)**: *v* (cm⁻¹) 3062, 3030, 2925, 2100, 1652, 1486, 1447, 1406, 1355, 1265, 1216, 1111, 1025, 990, 948, 920, 817, 736, 699, 619, 583, 550, 514, 458, 398.

Compound 1r



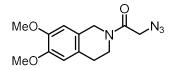
1r: white solid. Yield: 81%. ¹H NMR (300 MHz, CDCl₃) δ 7.42 (s, 1H), 7.35 (s, 1H), 4.80 (s, 2H),
4.73 (s, 2H), 3.95 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 166.2, 136.1, 135.8, 132.7, 132.3, 125.2,
124.8, 52.1, 51.2, 51.1. IR (film): v (cm⁻¹) 3002, 2931, 2838, 2101, 1652, 1611, 1509, 1461, 1407,
1354, 1242, 1176, 1112, 1030, 989, 928, 812, 756, 693, 587, 550, 516, 444, 397.

Compound 1s



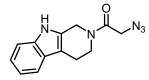
1s: white solid. Yield: 82%. ¹**H NMR (300 MHz, CDCl₃)** δ 6.79 (s, 1H), 6.73 (s, 1H), 4.77 (s, 2H), 4.69 (s, 2H), 3.95 (s, 2H), 3.88 (s, 3H), 3.86 (s, 3H). ¹³**C NMR (75 MHz, CDCl₃)** δ 166.1, 149.8, 149.5, 127.7, 127.0, 105.9, 105.6, 56.3, 56.3, 52.7, 52.0, 51.0. **IR (film):** *v* (cm⁻¹) 2987, 2947, 2900, 2838, 2149, 2103, 1650, 1607, 1515, 1454, 1363, 1335, 1281, 1250, 1201, 1107, 1060, 1014, 965, 912, 850, 821, 787, 747, 683, 606, 550, 484, 444, 402.

Compound 1t



1t: white solid. Yield: 92%. ¹**H NMR (300 MHz, CDCl₃)** δ 6.61 (m, 2H), 4.65 (s, 2H), 3.99 (s, 2H), 3.87-3.77 (m, 6H), 3.56 (t, J = 5.9 Hz, 2H), 2.86-2.70 (m, 2H). With rotation ratio = 4:3. ¹³**C NMR (75 MHz, CDCl₃)** δ 166.2, 148.3, 148.2, 126.9, 125.7, 124.9, 123.5, 112.0, 111.5, 109.7, 109.2, 56.2, 51.2, 46.6, 44.4, 43.1, 40.4, 29.0, 28.1. **IR (film):** *v* (cm⁻¹) 2985, 2949, 2900, 2838, 2205, 2162, 2105, 1651, 1607, 1515, 1454, 1362, 1337, 1282, 1251, 1201, 1106, 1060, 1015, 966, 913, 851, 821, 787, 747, 684, 606, 550, 486, 443, 401.

Compound 1u

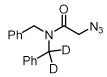


1u: white solid. Yield: 73%. ¹**H NMR** (300 MHz, CD₂Cl₂) δ 8.08 (d, *J* = 61.5 Hz, 1H), 7.48 (t, *J* = 7.5 Hz, 1H), 7.36 (d, *J* = 7.8 Hz, 1H), 7.12 (ddd, *J* = 14.8, 13.8, 6.9 Hz, 2H), 4.71 (d, *J* = 78.7 Hz, 2H), 4.15-3.99 (m, 2H), 3.98-3.57 (m, 2H), 2.86 (dd, *J* = 11.5, 5.9 Hz, 2H). ¹³**C NMR** (75 MHz, CD₂Cl₂) δ 166.8, 136.7, 130.3, 127.2, 122.2, 120.0, 118.2, 111.4, 108.2, 51.3, 43.8, 41.2, 22.3. **IR (film):** *v* (cm⁻¹) 3292, 3058, 2914, 2846, 2099, 1642, 1446, 1375, 1277, 1215, 1163, 1051, 985, 950, 919, 894, 848, 787, 735, 682, 656, 613, 577, 550, 499, 470, 429.

Compound 1a-d2-1

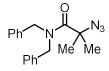
1a-d2-1: colorless oil. Yield: 87%. ¹**H NMR** (300 MHz, CD₂Cl₂) δ 7.45-7.29 (m, 6H), 7.25 (d, *J* = 7.2 Hz, 2H), 7.16 (d, *J* = 7.1 Hz, 2H), 4.61 (s, 1H), 4.36 (s, 1H), 3.97 (s, 2H). ¹³**C NMR** (75 MHz, CD₂Cl₂) δ 168.3, 137.2, 136.1, 129.5, 129.1, 128.7, 128.3, 128.0, 126.9, 51.0, 50.0, 49.7, 49.4, 49.1, 48.9. **IR** (film): *v* (cm⁻¹) 3060, 3031, 2915, 2099, 1653, 1492, 1443, 1415, 1358, 1266, 1211, 1079, 1020, 981, 920, 837, 814, 726, 699, 612, 553, 502, 454, 396.

Compound 1a-d2-2



1a-d2-2: colorless oil. Yield: 91%. **¹H NMR** (300 MHz, CDCl₃) δ 7.34-7.20 (m, 6H), 7.16 (d, *J* = 5.2 Hz, 2H), 7.05 (d, *J* = 6.9 Hz, 2H), 4.56 (s, 1H), 4.28 (s, 1H), 3.89 (s, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 168.0, 136.6, 135.6, 135.5, 129.3, 128.9, 128.6, 128.1, 127.9, 126.4, 50.8, 49.5, 49.0. **IR (film):** *v* (cm⁻¹) 3061, 3029, 2917, 2101, 1653, 1493, 1441, 1414, 1357, 1267, 1211, 1077, 1018, 982, 921, 834, 811, 730, 696, 609, 551, 500, 452, 399.

Compound 1aMe₂



1aMe₂: white solid. Yield: 87% ¹**H NMR** (300 MHz, CDCl₃) δ 7.57-6.96 (m, 10H), 4.88 (s, 2H), 4.54 (s, 2H), 1.64 (s, 6H). ¹³**C NMR** (75 MHz, CDCl₃) δ 172.1, 136.9, 128.9, 128.2, 127.6, 127.1, 64.3, 50.8, 48.9, 25.7. **IR (film):** *v* (cm⁻¹) 3062, 3033 2978, 2925, 2857, 2469, 2100, 1629, 1494, 1444, 1416, 1361, 1315, 1245, 1172, 1144, 1077, 1025, 944, 896, 849, 816, 739, 696, 615, 589, 555, 508, 470, 429, 413.

4.2.3 Catalytic Asymmetric Intramolecular C-H Aminations

General procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with azides 1a-v (0.2 mmol) and Λ -Ru7 (2.3 mg, 0.002 mmol, 1 mol%) under N₂ atmosphere. Boc₂O (55 μ L, 0.24 mmol) and super dried 1,2-dichlorobenzene (0.2 mL, 1.0 M) bought from "Acros" was added via syringe in sequence. The reaction mixture was stirred at the indicated temperature for the indicated time under N₂ atmosphere. Afterwards, the mixture was directly transferred to a column and purified by flash chromatography on silica gel (*n*-Hexane/EtOAc= 10:1 to 3:1) to afford the analytically pure products 2a-l, 2n-v, and 2a'. Enantiomeric excess was determined by HPLC analysis on chiral stationary phase. The absolute configuration of the product 2f was measured by single-crystal X-ray analysis as *S*-configuration.

Additional experimental informations:

Is the solvent 1,2-dichlorobenzene special? It can be replaced by other solvents with little bit drop of the catalytic performance.

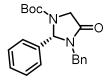
Ph Ph	H $N_3 \frac{1}{1,2}$	1 mol% Ru Cat Boc ₂ O (1.2 eq) -dichlorobenzene heating, 48 h andard conditions 7a	Mes Mes-	N N N Me	+ 2 PF ₆ -
entry	catalyst	conditions ^b	T (°C)	NMR yield (%) ^c	ee (%) ^d
1	∆-Ru7	standard	85	84 (80) ^e	91
2	∆-Ru7	Dioxane as Sol.	85	82	90
3	Λ- Ru7	Chlorobenzene as Sol.	85	79	91

^aStandard reaction conditions: **6a** (0.2 mmol), Boc₂O (0.24 mmol), Ruthenium catalyst (0.002 mmol) in 1,2-dichlorobenzene (0.2 mL) stirred at 85 °C for 48 h under N₂ unless noted otherwise. ^bDeviations from standard conditions are shown. ^cDetermined by ¹H NMR of the crude products using Cl₂CHCHCl₂ as internal standard. ^dEnantiomeric excess determined by HPLC analysis of the crude main product on a chiral stationary phase. ^eIsolated yield in the brackets.

How was the reaction executed? All catalytic reactions were carried with a 10 mL Schlenk tube made by "Synthware" and the vial was sealed under N₂ atmosphere (see following picture on the left).

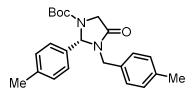
TLC of the reaction involved with standard substrate **1a** was monitored after 24 h (*n*-hexane:EtOAc = 3:1, Product $R_f = 0.45$, see following picture on the right). When the reaction time was 48 h, the starting material was fully consumed (>99% conversion).

Compound 2a



Starting from 1a (56.0 mg, 0.20 mmol) according to the general procedure to provide 2a as a white solid (56.2 mg, 80% yield). Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak IG column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 85:15, flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 19.7 min, t_r (minor) = 15.3 min). $[\alpha]_{D}^{22} = +21.6^{\circ}$ (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.44-7.36 (m, *J* = 6.2, 3.6 Hz, 3H), 7.34-7.19 (m, 5H), 7.18-7.09 (m, 2H), 5.62 (br, m, 1H), 5.00 (br, m, 1H), 4.17 (s, 2H), 3.39 (br, m, 1H), 1.42-0.96 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.4, 148.2, 138.3, 135.6, 129.3, 128.7, 128.4, 127.8, 127.4, 80.8, 74.7, 48.9, 48.4, 43.2, 27.9, 27.7. HRMS (ESI, *m*/*z*) calcd. for C₂₁H₂₄N₂O₃Na[M+Na]⁺: 375.1679, found: 375.1687.

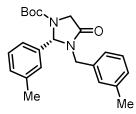
Compound 2b



Starting from **1b** (61.7 mg, 0.20 mmol) according to the general procedure to provide **2b** as a colorless oil (61.4 mg, 81% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 14.9 min, t_r (minor) = 36.4 min). [α] $_{D}^{22}$ = +78.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.31-7.15 (m, 3H), 7.13-6.97 (m, 3H), 6.93 (d, J = 6.2 Hz, 2H), 5.57 (br, m, 1H), 4.92 (br, m, 1H), 4.16 (s, 2H), 3.36 (br, m, 1H), 2.35 (s, 3H), 2.32 (s, 3H), 1.27 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.7, 152.7, 139.0, 135.9,

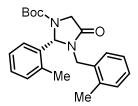
130.4, 129.5, 128.9, 128.4, 125.9, 125.1, 81.2, 75.3, 49.5, 48.8, 43.7, 28.4, 28.2, 21.5. **HRMS (ESI,** *m/z*) calcd. for C₂₃H₂₈N₂O₃Na[M+Na]⁺: 403.1992, found: 403.2004.

Compound 2c



Starting from 1c (61.7 mg, 0.20 mmol) according to the general procedure to provide 2c as a white solid (63.8 mg, 84% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 8.0 min, t_r (minor) = 22.0 min). [α] $_{D}^{22}$ = +86.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.12 (d, *J* = 7.8 Hz, 2H), 7.05 (d, *J* = 7.9 Hz, 4H), 6.94 (d, *J* = 8.0 Hz, 2H), 5.47 (br, m, 1H), 4.90 (br, m, 1H), 4.02 (s, 2H), 3.21 (br, m, 1H), 2.28 (s, 3H), 2.25 (s, 3H), 1.33-0.97 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.7, 152.7, 139.7, 138.1, 135.6, 135.3, 133.0, 129.8, 128.8, 127.8, 81.1, 74.9, 74.7, 49.4, 48.9, 43.3, 28.2, 21.4, 21.3. HRMS (ESI, *m*/z) calcd. for C₂₃H₂₈N₂O₃Na[M+Na]⁺: 403.1992, found: 403.2006.

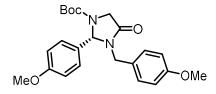
Compound 2d



Starting from 1d (61.7 mg, 0.20 mmol) according to the general procedure to provide 2d as a white solid (39.6 mg, 52% yield). Enantiomeric excess was established by HPLC analysis as 87% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 85:15, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 13.9 min, t_r (minor) = 20.9 min). [α] $_{D}^{22}$ = +88.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.27-7.08 (m, 7H), 6.88 (m, 1H), 5.83 (s, 1H), 5.09-4.88 (br, m, 1H), 4.20 (s, 2H), 3.38 (br, m, 1H), 2.10-1.92 (m, 6H), 1.48-0.96 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 168.0, 153.0, 137.2,

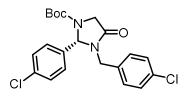
136.1, 133.3, 131.3, 131.0, 130.7, 129.4, 128.3, 127.8, 126.9, 126.5, 81.3, 72.2, 49.5, 49.3, 41.6, 28.4,
28.3, 19.2, 18.6. HRMS (ESI, *m/z*) calcd. for C₂₃H₂₈N₂O₃Na[M+Na]⁺: 403.1992, found: 403.2003.

Compound 2e



Starting from **1e** (68.1 mg, 0.20 mmol) according to the general procedure to provide **2e** as a colorless oil (78.3 mg, 95% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 50:50, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: tr (major) = 10.0 min, tr (minor) = 14.0 min). [α] $_{0}^{22}$ = +22.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300 MHz**, **CD₂Cl₂**) δ 7.16 (s, 2H), 7.09-7.00 (m, 2H), 6.91 (d, *J* = 8.6 Hz, 2H), 6.86 (dd, *J* = 6.7, 4.8 Hz, 2H), 5.55 (br, m, 1H), 4.93 (br, m, 1H), 4.13 (s, 2H), 3.83 (s, 3H), 3.78 (s, 3H), 3.32 (br, m, 1H), 1.44-0.99 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.5, 160.8, 159.8, 152.8, 130.2, 129.2, 128.0, 114.5, 81.1, 74.7, 55.7, 55.7, 49.4, 48.9, 43.0, 28.3. HRMS (ESI, *m*/*z*) calcd. for C₂₃H₂₈N₂O₅Na[M+Na]⁺: 435.1890, found: 435.1901.

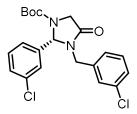
Compound 2f



Starting from **1f** (69.6 mg, 0.20 mmol) according to the general procedure to provide **2f** as a white solid (65.5 mg, 78% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 8.9 min, t_r (minor) = 12.9 min). [α]_D²² = +4.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.43-7.34 (m, 2H), 7.30 (d, *J* = 8.4 Hz, 2H), 7.17 (s, 2H), 7.07 (d, *J* = 8.4 Hz, 2H), 5.74-5.38 (br, m, 1H), 4.97-4.81 (br, m, 1H), 4.16 (s, 2H), 3.56-3.20 (br, m, 1H), 1.47-1.04 (br, m, 9H).

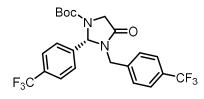
¹³C NMR (75 MHz, CD₂Cl₂) δ 167.8, 152.9, 146.5, 137.1, 135.5, 134.4, 134.1, 130.3, 129.4, 129.3, 81.6, 74.5, 48.7, 43.2, 28.2. HRMS (ESI, *m/z*) calcd. for C₂₁H₂₂N₂O₃Cl₂[M+Na]⁺: 443.0900, found: 443.0911.

Compound 2g



Starting from 1g (69.6 mg, 0.20 mmol) according to the general procedure to provide 2g as a white solid (68.8 mg, 82% yield). Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 75:25, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: tr (major) = 10.1 min, tr (minor) = 12.2 min). $[\alpha]_D^{22} = +6.2^\circ$ (c = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.43-7.07 (m, 7H), 7.03 (d, J = 3.9 Hz, 1H), 5.74-5.46 (br, m, 1H), 4.98-4.74 (br, m, 1H), 4.18 (s, 2H), 3.73-3.34 (br, m, 1H), 1.56-1.05 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.8, 153.0, 152.4, 140.7, 137.9, 134.9, 130.5, 130.0, 128.8, 128.5, 128.1, 127.0, 126.2, 81.7, 74.8, 49.2, 48.6, 43.5, 28.2. HRMS (ESI, *m*/*z*) calcd. for C₂₁H₂₂N₂O₃Cl₂[M+Na]⁺: 443.0900, found: 443.0910.

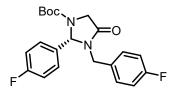
Compound 2h



Starting from **1h** (83.2 mg, 0.20 mmol) according to the general procedure to provide **2h** as a white solid (49.8 mg, 51% yield). Enantiomeric excess was established by HPLC analysis as 94% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 4.9 min, t_r (minor) = 6.4 min). [α]_D²² = +18.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.64 (d, *J* = 8.1 Hz, 2H), 7.56 (d, *J* = 8.1 Hz, 2H), 7.36 (s, 2H), 7.24 (d, *J* = 8.0 Hz, 2H), 5.82-5.51 (br, m, 1H), 5.01-4.78 (br, m, 1H), 4.22 (s, 2H), 3.81-3.55 (br, m, 1H), 1.50-0.97 (br, m, 9H).

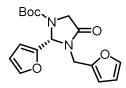
¹³C NMR (75 MHz, CD₂Cl₂) δ 168.0, 153.0, 152.4, 142.7, 142.2, 140.1, 132.05, 131.3, 130.6, 130.4-129.6 (m), 129.2, 128.4, 126.7-125.6 (m), 122.7 (d, J = 13.4 Hz), 81.9, 74.9, 60.7, 49.1, 48.6, 43.7, 28.2. ¹⁹F NMR (235 MHz, CD₂Cl₂) δ -63.01, -63.08. HRMS (ESI, *m/z*) calcd. for $C_{23}H_{22}N_2O_3F_6Na[M+Na]^+$: 511.1427, found: 511.1441.

Compound 2i



Starting from **1i** (63.2 mg, 0.20 mmol) according to the general procedure to provide **2i** as a white solid (56.5 mg, 73% yield). Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 8.4 min, t_r (minor) = 11.4 min). [α] $_{D}^{22}$ = +29.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300 MHz**, **CD₂Cl₂**) δ 7.38-6.82 (m, 8H), 5.79-5.43 (br, m, 1H), 5.02-4.78 (br, m, 1H), 4.16 (s, 2H), 3.59-3.28 (br, m, 1H), 1.43-1.00 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.8, 165.3, 164.5, 162.0, 161.2, 152.6, 134.5, 131.8, 130.7, 130.6, 129.9, 129.8, 116.2, 116.1, 115.9, 115.8, 81.5, 74.6, 49.2, 48.7, 41.6, 43.1, 28.3. ¹⁹F NMR (235 MHz, CD₂Cl₂) δ -112.77, -115.06. HRMS (ESI, *m/z*) calcd. for C₂₁H₂₂F₂N₂O₃Na[M+Na]⁺: 411.1491, found: 411.1491.

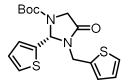
Compound 2j



Starting from **1j** (52.0 mg, 0.20 mmol) according to the general procedure to provide **2j** as a colorless oil (42.5 mg, 64% yield). Enantiomeric excess was established by HPLC analysis as 88% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 8.6 min, t_r (minor) = 18.3 min). [α]_D²² = +18.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.41 (dd, *J* = 12.8, 1.0 Hz, 2H), 6.49 (d, *J* = 15.0 Hz, 1H), 6.37 (ddd, *J* = 20.3, 3.1, 1.8 Hz,

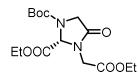
2H), 6.21 (d, J = 3.2 Hz, 1H), 5.98-5.69 (br, m, 1H), 4.99-4.75 (br, m, 1H), 4.17-3.94 (br, m, 2H), 3.76-3.56 (br, m, 1H), 1.45-1.20 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.8, 150.0, 149.5, 143.7, 143.2, 110.8, 110.8, 109.4, 81.4, 68.6, 68.4, 48.8, 48.1, 36.6, 28.3. HRMS (ESI, *m/z*) calcd. for C₁₇H₂₀N₂O₅Na[M+Na]⁺: 355.1264, found: 355.1274.

Compound 2k



Starting from 1k (58.4 mg, 0.20 mmol) according to the general procedure to provide 2k as a white solid (54.6 mg, 75% yield). Enantiomeric excess was established by HPLC analysis as 82% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: tr (major) = 6.4 min, tr (minor) = 15.1 min). [α] $_{D}^{22}$ = +71.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.40 (d, *J* = 5.0 Hz, 1H), 7.29 (dd, *J* = 5.1, 1.0 Hz, 1H), 7.17 (s, 1H), 7.03 (dd, *J* = 5.0, 3.5 Hz, 1H), 6.97 (dd, *J* = 5.1, 3.5 Hz, 1H), 6.92 (d, *J* = 3.4 Hz, 1H), 6.18-5.90 (br, m, 1H), 5.16-5.00 (br, m, 1H), 4.15-3.98 (br, m, 2H), 3.93-3.75 (br, m, 1H), 1.45-1.19 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.1, 152.6, 142.6, 138.1, 128.4, 127.9, 127.4, 127.2, 126.4, 81.6, 70.3, 48.6, 48.1, 38.1, 28.3. HRMS (ESI, *m*/*z*) calcd. for C₁₇H₂₀N₂O₃S₂Na[M+Na]⁺: 387.0808, found: 387.0818.

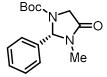
Compound 21



Starting from **11** (54.4 mg, 0.20 mmol) according to the general procedure to provide **21** as a colorless oil (50.3 mg, 73% yield, 32% ee). It's noteworthy that when treated with Λ -**Ru5** instead under general procedure provided **21** (49.0 mg, 71% yield, 70% ee). Enantiomeric excess was established by HPLC analysis (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 9.76 min, t_r (minor) = 15.78 min). [α]_D²² = +7.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.43-5.31 (m, 1H), 4.43-4.32 (m, 1H), 4.32-4.15 (m, 4H), 4.12-4.00 (m, 2H), 3.95-3.76 (m,

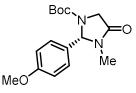
1H), 1.52-1.38 (m, 9H), 1.35-1.21 (m, 6H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 169.2, 168.9, 167.9, 82.1, 72.6, 62.7, 62.2, 62.1, 61.8, 50.0, 49.1, 42.3, 30.1, 28.4, 28.3, 14.3. HRMS (ESI, *m/z*) calcd. for C₁₅H₂₄N₂O₇Na[M+Na]⁺: 367.1476, found: 367.1485.

Compound 2m



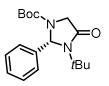
Starting from **1m** (40.8 mg, 0.20 mmol) according to the general procedure to provide **2m** as a colorless oil (28.2 mg, 51% yield). Enantiomeric excess was established by HPLC as 95% ee (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 9.3 min, t_r (minor) = 10.9 min). [α]_D²² = +46.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.50-7.17 (m, 5H), 5.90-5.55 (br, m, 1H), 4.09 (s, 2H), 2.60 (s, 3H), 1.55-1.06 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.7, 152.9, 138.8, 129.6, 129.1, 127.6, 81.3, 77.4, 49.2, 48.7, 28.3, 26.7. HRMS (ESI, *m/z*) calcd. for C₁₅H₂₀N₂O₃Na[M+Na]⁺: 299.1366, found: 299.1366.

Compound 2n



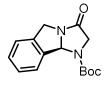
Starting from **1n** (46.8 mg, 0.20 mmol) according to the general procedure to provide **2n** as a colorless oil (39.1 mg, 64% yield). Enantiomeric excess was established by HPLC analysis as 94% ee (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 10.0 min, t_r (minor) = 13.0 min). [α]_D²² = +52° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.23 (s, 2H), 6.91 (d, *J* = 8.7 Hz, 2H), 5.81-5.53 (br, m, 1H), 4.06 (s, 2H), 3.81 (s, 3H), 2.59 (s, 3H), 1.51-1.10 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.6, 160.8, 152.9, 130.6, 128.9, 114.4, 81.1, 77.0, 55.7, 49.1, 48.6, 28.4, 26.7. HRMS (ESI, *m*/*z*) calcd. for C₁₆H₂₂N₂O₄Na[M+Na]⁺: 329.1472, found: 329.1481.

Compound 2o



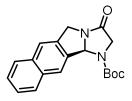
Starting from **10** (49.2 mg, 0.20 mmol) according to the general procedure to provide **20** as a white solid (51.5 mg, 81% yield). Enantiomeric excess was established by HPLC analysis as 23% ee (column: Daicel Chiralpak ADH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 98:2 flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 17.0 min, t_r (minor) = 20.8 min). [α]_D²² = +11.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300 MHz**, **CD₂Cl₂**) δ 7.38 (d, *J* = 11.8 Hz, 5H), 6.26-5.90 (br, m, 1H), 4.12-3.84 (br, m, 2H), 1.42-1.32 (br, m, 9H), 1.30 (s, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 169.3, 169.0, 152.4, 152.0, 141.3, 129.1, 128.8, 127.7, 127.5, 81.3, 81.0, 75.3, 74.9, 55.7, 49.6, 49.1, 28.4, 28.1. HRMS (ESI, *m/z*) calcd. for C₁₈H₂₆N₂O₃Na[M+Na]⁺: 341.1836, found: 341.1836.

Compound 2p



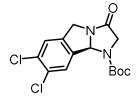
Starting from **1p** (40.4 mg, 0.20 mmol) according to the general procedure to provide **2p** as a white solid (40.5 mg, 74% yield). Enantiomeric excess was established by HPLC analysis as 94% ee (column: Daicel Chiralpak OJH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: tr (major) = 16.9 min, tr (minor) = 19.2 min). [α]_D²² = +123.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300 MHz**, **CD₂Cl₂)** δ 7.77 (s, 1H), 7.42-7.11 (m, 3H), 6.41 (s, 1H), 5.03-4.93 (br, m, 1H), 4.32-4.12 (br, m, 2H), 3.89-3.59 (br, m, 1H), 1.70-1.44 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 174.1, 154.7, 140.3, 139.6, 129.4, 128.4, 128.0, 123.3, 79.6, 50.7, 49.5, 28.6, 28.4, 28.3. HRMS (ESI, *m/z*) calcd. for C₁₅H₁₈N₂O₃Na[M+Na]⁺: 297.1210, found: 297.1221.

Compound 2q



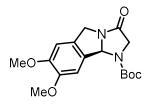
Starting from 1q (50.4 mg, 0.20 mmol) according to the general procedure to provide 2q as a white solid (42.5 mg, 66% yield). Enantiomeric excess was established by HPLC analysis as 88% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: tr (major) = 7.3 min, tr (minor) = 5.7 min). [α] $_{D}^{22}$ = +193.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 8.43-8.03 (m, 1H), 7.98-7.80 (m, 2H), 7.73 (s, 1H), 7.62 -7.38 (m, 2H), 6.55 (br, s, 1H), 5.25-5.11 (br, m, 1H), 4.45-4.14 (br, m, 2H), 3.89-3.65 (br, m, 1H), 1.78-1.47 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 173.7, 138.6, 137.5, 134.2, 133.7, 128.9, 128.2, 127.0, 126.5, 124.3, 121.9, 81.7, 79.0, 50.8, 48.8, 28.6. HRMS (ESI, *m*/z) calcd. for C₁₉H₂₀N₂O₃Na[M+Na]⁺: 347.1366, found: 347.1379.

Compound 2r



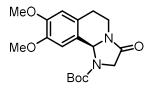
Starting from **1r** (54.0 mg, 0.20 mmol) according to the general procedure to provide **2r** as a white solid (39.0 mg, 57% yield). Enantiomeric excess was established by HPLC analysis as 94% ee (column: Daicel Chiralpak IB column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 10.6 min, t_r (minor) = 8.6 min). [α] $_{D}^{22}$ = +62.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300 MHz**, **CD₂Cl₂**) δ 7.99-7.66 (m, 1H), 7.39 (s, 1H), 6.44-6.20 (br, m, 1H), 5.03-4.86 (br, m, 1H), 4.35-4.04 (br, m, 2H), 3.89-3.60 (br, m, 1H), 1.70-1.47 (br, m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 173.8, 154.8, 140.5, 139.6, 133.5, 132.4, 127.3, 126.4, 125.2, 100.5, 82.1, 78.8, 50.6, 48.9, 28.5, 28.4, 28.3. HRMS (ESI, *m/z*) calcd. for C₁₅H₁₇N₂O₃Cl₂[M+H]⁺: 343.0611, found: 343.0624.

Compound 2s



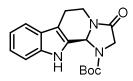
Starting from **1s** (52.4 mg, 0.20 mmol) according to the general procedure to provide **2s** as a white solid (50.4 mg, 75% yield). Enantiomeric excess was established by HPLC analysis as 89% ee (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: t_r (major) = 11.7 min, t_r (minor) = 10.7 min). [α]_D²² = +199.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (**300** MHz, **CD₂Cl₂**) δ 7.25 (d, *J* = 49.2 Hz, 1H), 6.76 (s, 1H), 6.33 (s, 1H), 5.04-4.83 (br, m, 1H), 4.31-4.01 (br, m, 2H), 3.91-3.57 (m, 7H), 1.69-1.40 (br, m, 9H). ¹³C NMR (**75** MHz, **CD₂Cl₂**) δ 174.2, 150.9, 149.9, 132.0, 131.4, 108.3, 107.1, 106.1, 81.6, 79.8, 56.4, 56.4, 50.7, 50.4, 49.6, 28.7. HRMS (**ESI**, *m/z*) calcd. for C₁₇H₂₃N₂O₅[M+H]⁺: 335.1601, found: 335.1613.

Compound 2t



Starting from **1t** (55.2 mg, 0.20 mmol) according to the general procedure to provide **2t** as a white powder (33.4 mg, 48% yield). Enantiomeric excess was established by HPLC analysis as 92% ee (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 40 °C, retention times: t_r (major) = 11.9 min, t_r (minor) = 15.3 min). [α]_D²² = +56.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.36-6.84 (m, 1H), 6.67-6.55 (m, 1H), 6.21 (s, 1H), 4.33-4.04 (m, 2H), 3.80 (s, 7H), 3.72-3.51 (m, 1H), 3.39-3.13 (m, 1H), 3.08-2.89 (m, 1H), 2.73-2.57 (m, 1H), 1.54 (s, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 168.9, 149.5, 148.5, 128.8, 126.9, 111.8, 110.1, 109.5, 81.6, 71.1, 56.2, 56.1, 49.4, 38.3, 28.5, 27.0. HRMS (ESI, *m*/*z*) calcd. for C₁₈H₂₄N₂O₅Na[M+Na]⁺: 371.1577, found: 371.1588

Compound 2u



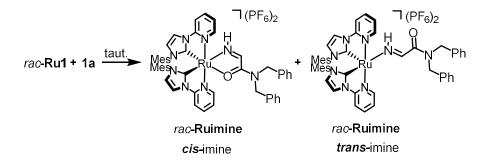
Starting from **1u** (51.0 mg, 0.20 mmol) according to the general procedure to provide **2u** as a white solid (20.2 mg, 31% yield). Enantiomeric excess was established by HPLC analysis as 89% ee (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: tr (major) = 7.7 min, tr (minor) = 8.7 min). [α]₀²² = +43.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, **CDCl₃**) δ 9.36 (s, 1H), 7.50 (d, *J* = 7.7 Hz, 1H), 7.39 (d, *J* = 8.1 Hz, 1H), 7.25-7.17 (m, 1H), 7.12 (t, *J* = 7.0 Hz, 1H), 6.36 (s, 1H), 4.62 (dd, *J* = 13.2, 5.3 Hz, 1H), 4.19-4.05 (m, 1H), 3.92 (dd, *J* = 15.8, 2.0 Hz, 1H), 3.19 (td, *J* = 12.2, 4.6 Hz, 1H), 3.08-2.90 (m, 1H), 2.80 (dd, *J* = 15.3, 4.8 Hz, 1H), 1.55 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 167.6, 155.2, 136.1, 126.1, 123.1, 119.9, 118.9, 111.7, 109.6, 82.4, 68.6, 49.3, 38.7, 28.4, 20.8. HRMS (ESI, *m*/z) calcd. for C₁₈H₂₁N₃O₃Na[M+Na]⁺: 350.1475, found: 350.1475.

Compound 2a'

Starting from **1a** (56.0 mg, 0.20 mmol) according to the general procedure using Cbz₂O instead of Boc₂O to provide **2a**' as a white solid (45.6 mg, 59% yield). Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak ASH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 40:60, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 11.6 min, t_r (minor) = 40.8 min). [α]_D²² = +30.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.53-7.08 (m, 14H), 6.88 (s, 1H), 5.89-5.57 (m, 1H), 5.19-4.79 (m, 3H), 4.26 (s, 2H), 3.52-3.27 (m, 1H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 167.5, 153.5, 137.7, 136.7, 135.8, 131.0, 129.9, 129.3, 129.2, 128.8, 128.6, 128.6, 128.3, 128.0, 75.1, 67.7, 49.0, 43.7. HRMS (ESI, *m/z*) calcd. for C₂₄H₂₂N₂O₃Na[M+Na]⁺: 409.1523, found: 409.1519.

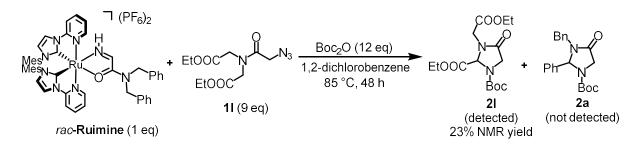
4.2.4 Mechanistic Studies

a) Ru-imine complex formation under mild conditions



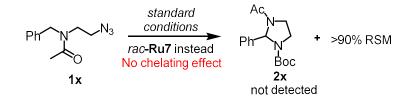
Reaction at room temperature: To a solution of rac-Ru1 (50.1 mg, 0.05 mmol) in 0.5 mL mixed solvent (CH₂Cl₂:1,2-dichlorobenzene = 1:1) was added 1a (28.0 mg, 0.05 mmol). After stirring at room temperature for 24 h, the above solution was concentrated under reduced pressure and subsequently purified by fast flash column (CH₂Cl₂:EtOAc = 2:1) to afford *rac*-Ruimine (58.7 mg, quantitative yield). The ¹H NMR analysis of the crude reaction solution showed only *cis*-imine formation. Spectrum analysis of *rac*-Ruimine: ¹H NMR (300 MHz, CD₂Cl₂) δ 12.09 (d, J = 11.7 Hz, 1H), 8.99 (d, J = 11.7 Hz, 1H), 8.00 (d, J = 2.3 Hz, 1H), 7.90 (d, J = 2.3 Hz, 1H), 7.86-7.70 (m, 2H), 7.54 (dd, *J* = 18.1, 5.0 Hz, 3H), 7.44 -7.31 (m, 4H), 7.18-7.02 (m, 5H), 6.92 (td, *J* = 5.5, 2.9 Hz, 4H), 6.75 (d, J = 7.3 Hz, 2H), 6.65 (d, J = 5.3 Hz, 2H), 6.52 (d, J = 7.0 Hz, 2H), 5.00-4.77 (m, 3H), 4.43 (d, J = 14.9 Hz, 1H), 2.16 (d, J = 5.9 Hz, 6H), 2.02 (s, 3H), 1.89 (s, 3H), 1.51 (s, 3H), 1.47 (s, 3H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 192.2, 190.8, 169.5, 163.8, 153.4, 153.1, 151.6, 149.6, 140.3, 138.5, 138.1, 135.8, 135.6, 134.6, 134.4, 134.2, 134.1, 134.0, 133.7, 132.8, 130.2, 130.1, 129.6, 129.6, 128.7, 127.7, 127.6, 125.9, 125.7, 123.2, 122.7, 117.5, 117.1, 111.4, 110.7, 52.2, 21.0, 20.9, 17.8, 17.5, 17.2, 17.1. ¹⁹F NMR (282 MHz, CD₂Cl₂) δ -71.97, -74.50. HRMS (ESI, *m/z*) calcd. for C₅₀H₅₀N₈OP₁F₆[M-PF₆]⁺: 1025.2787, found: 1025.2823. **IR (film):** v (cm⁻¹) 3670, 3294, 2921, 2249, 2178, 2096, 2012, 1650, 1609, 1484, 1451, 1421, 1376, 1330, 1301, 1255, 1158, 1130, 1081, 1030, 931, 831, 766, 741, 698, 587, 554, 500, 453, 411.

Reaction at 85 °C: To a solution of *rac*-Ru1 (50.1 mg, 0.05 mmol) in 0.5 mL 1,2-dichlorobenzene was added 1a (28.0 mg, 0.1 mmol). After stirring at 85 °C for 1 h the reaction was complete. ¹H NMR analysis showed a ratio of *cis*-imine to *trans*-imine at 1:1.5 (*trans*-imine is not stable enough to be purified). Thus, this indicates that the amide group dissociates at high temperature which is in consistent with our mechanistic proposal.



To a solution of *rac*-**Ruimine** (11.7 mg, 0.01 mmol) in 0.5 mL 1,2-dichlorobenzene was added 11 (24.5 mg, 0.18 mmol). After stirring at 85 °C for 48 h only 21 was detected from ¹HNMR. It was noteworthy that 2a was not detected which indicates that the Ru-nitrene \rightarrow imine tautomerization is irreversible.

b) Probing presence and position of the amide group

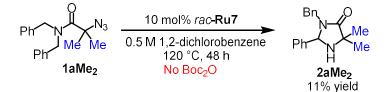


1x (43.6 mg, 0.2 mmol) and *rac*-Ru7 (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 24 h. To the resulting solution was added $Cl_2CHCHCl_2$ as internal standard. The ¹H NMR analysis of the above solution showed >90% remaining starting material.



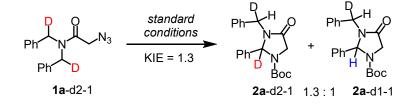
1y (35.1 mg, 0.2 mmol) and *rac*-**Ru7** (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 24 h. To the resulting solution was added Cl₂CHCHCl₂ as internal standard. The ¹H NMR analysis of the above solution showed <5% formation of **2y** and >90% remained starting material.

c) Isolated C-H amination product without Boc₂O



1aMe₂ (30.8 mg, 0.1 mmol) and *rac*-**Ru7** (11.5 mg, 0.01 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 120 °C under N₂ atmosphere for 48 h. The above solution was transformed to a flash silica gel column and purified (*n*-hexane:EtOAc:Et₃N = 1:1:0.001) to afford **2aMe**₂ (3.1 mg, 11% yield). **2aMe**₂ : ¹**H NMR** (300 MHz, CDCl₃) δ 7.44-7.37 (m, 3H), 7.30-7.26 (m, 1H), 7.25-7.20 (m, 4H), 7.06-6.94 (m, 2H), 5.05 (s, 1H), 5.01 (d, *J* = 14.5 Hz, 1H), 3.53 (d, *J* = 14.5 Hz, 1H), 1.82 (s, br, 1H), 1.46 (s, 3H), 1.28 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 178.2, 138.2, 136.0, 129.5, 129.1, 128.6, 128.4, 127.7, 127.4, 73.2, 59.5, 44.5, 25.7, 24.5. **HRMS (ESI,** *m/z***)** calcd. for C₁₈H₂₁N₂O [M+H]⁺: 281.1648, found: 281.1657.

d) Kinetic isotope effects



1a-d2-1 (56.0 mg, 0.2 mmol) and *rac*-**Ru7** (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 48 h (standard conditions). The ¹H NMR analysis of the above solution showed a ratio of **2a**-d2-1:**2a**-d1-1 = 1.3:1. The peaks from 5.5 to 5.7 ppm belong to **2a**-d2-1 (blue H), and the peaks from 4.1 to 4.2 belong to **2a**-d2-1 and **2a**-d1-1 (green dot). Thus, we calculated the ratio of **2a**-d2-1 to **2a**-d1-1 as 1.3 to 1.

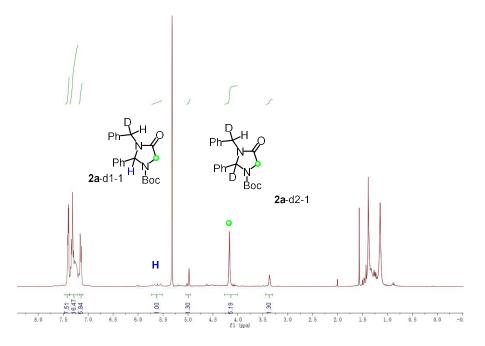
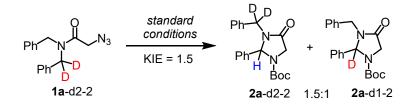


Figure 74. ¹H NMR of 2a-d2-1 reaction analysis.



1a-d2-2 (56.0 mg, 0.2 mmol) and *rac*-**Ru7** (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 48 h. The ¹H NMR analysis of the above solution showed a ratio of **2a**-d2-2:**2a**-d1-2 =1.5:1. The peaks from 4.1 to 4.2 ppm belong to **2a**-d2-2 and **2a**-d1-2 (green dot), and the peak from 3.25 to 3.35 ppm belongs to **2a**-d1-2 (blue H). Thus, we calculated the ratio of **2a**-d2-2 to **2a**-d1-2 as 1.5 to 1.

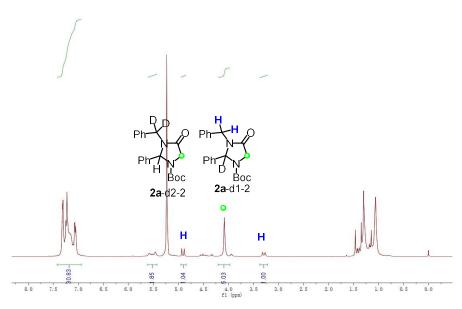
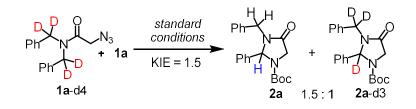


Figure 75. ¹H NMR of 2a-d2-2 reaction analysis.



1a-d4 (56.0 mg, 0.2 mmol) and *rac*-**Ru7** (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 16 h. The ¹H NMR analysis of the above solution showed a ratio of **2a**:**2a**-d3 = 1.5:1. The peak from 4.1 to 4.2 ppm belongs to **2a** and **2a**-d3 (green dot), and the peak from 3.25 to 3.35 ppm belongs to **2a** (blue H). Thus, we calculated the ratio of **2a** to **2a**-d3 as 1.5 to 1.

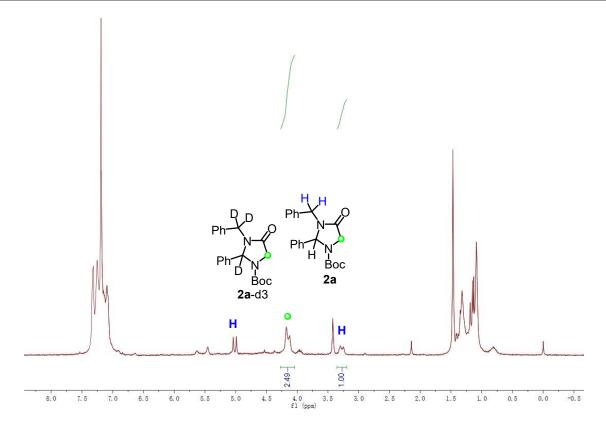
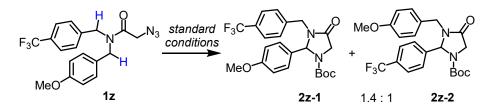


Figure 76. ¹H NMR of 2a and 2a-d4 reaction analysis.

e) Competition experiment



1z (75.6 mg, 0.2 mmol) and *rac*-Ru7 (2.3 mg, 0.002 mmol) in 0.2 mL 1,2-dichlorobenzene were stirred at 85 °C under N₂ atmosphere for 48 h. The resulting solution was concentrated under vacuo and purified by flash silical gel column (*n*-hexane:EtOAc = 3:1) to afford a mixture of 2z-1 and 2z-2 (79% total yield). ¹H NMR analysis showed a ratio of the products of 1.4:1.

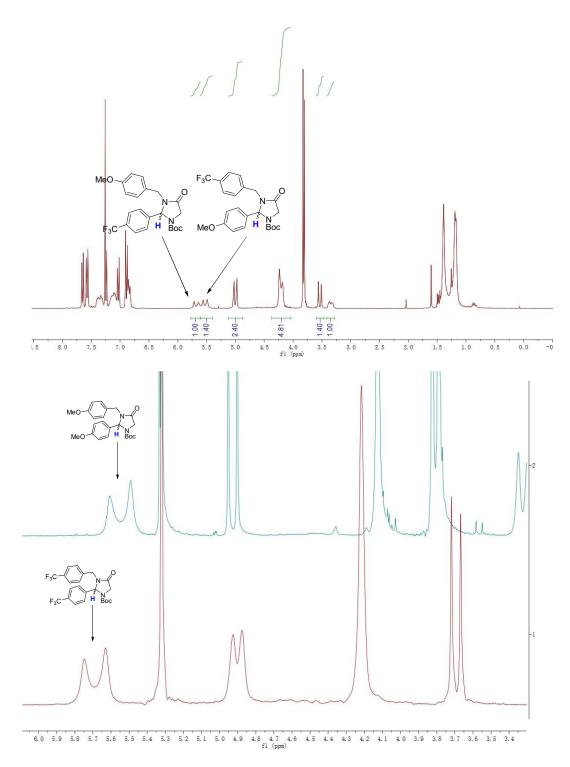


Figure 77. ¹H NMR of competition experiment analysis.

4.2.5 Single Crystal X-Ray Diffraction Studies

Single crystals of (S)-2e were obtained by slow diffusion from the solution in EtOAc layered with n-hexane at room temperature.

Single crystals of *rac*-**Ruimine** were obtained by slow volatilization from the solution in CDCl₃ and CH₂Cl₂ (ratio = 1:1) at room temperature.

Data was collected with an STOE STADIVARI diffractometer equipped with with CuK_a radiation, a graded multilayer mirror monochromator (l = 1.54178 Å) and a DECTRIS PILATUS 300K detector using an oil-coated shock-cooled crystal at 100(2) K. Absorption effects were corrected semi-empirical using multiscanned reflexions (STOE LANA, absorption correction by scaling of reflection intensities.). Cell constants were refined using 72887 of observed reflections of the data collection. The structure was solved by direct methods by using the program XT V2014/1 (Bruker AXS Inc., 2014) and refined by full matrix least squares procedures on F² using SHELXL-2018/3 (Sheldrick, 2018). The non-Hydrogen atoms have been refined anisotropically, carbon bonded hydrogen atoms were included at calculated positions and refined using the 'riding model' with isotropic temperature factors at 1.2 times (for CH₃ groups 1.5 times) that of the preceding carbon atom. CH₃ groups were allowed to rotate about the bond to their next atom to fit the electron density. Nitrogen bonded hydrogen atoms were located and allowed to refine isotropically. Disordered solvent contribution to the calculated structure factors were calculated by using the PLATON/SQUEEZE procedure. Crystallographic data for rac-Ruimine has been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 1857105. Absolute configuration could be established: The Flack parameter refined to -0.005(14). Crystallographic data for (S)-2e has been deposited with the Cambridge Crystallographic Data Centre as supplementary publication number CCDC 1857104.

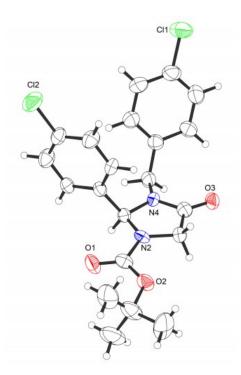


Figure 78. Crystal structure of compound (*S*)-2e.

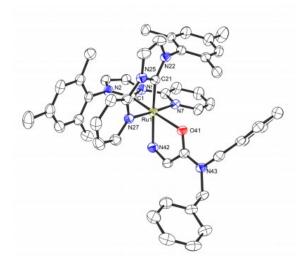


Figure 79. Crystal structure of compound *rac*-Ruimine.

 Table 5. Crystal data and structure refinement for (S)-2e.

Habitus, colour	needle, colourless		
Crystal size	$0.21 \text{ x} 0.06 \text{ x} 0.04 \text{ mm}^3$		
Crystal system Space group Unit cell dimensions	Monoclinic $P2_1$ $Z = 4$ $a = 12.8650(8)$ Å $= 90^{\circ}$. $b = 6.1505(3)$ Å $= 101.330(5)^{\circ}$. $c = 27.0802(15)$ Å $= 90^{\circ}$.		
Volume	2101.0(2) Å ¹		
Cell determination Empirical formula Moiety formula Formula weight	6303 peaks with Theta 3.5 to 67.1°. C_{21} H_{22} Cl_2 N_2 O_3 C_{21} H_{22} Cl_2 N_2 O_3 421.30		
Density (calculated)	1.332 Mg/m^3		
Absorption coefficient	2.978 mm ⁻¹		
F(000) Diffractometer type Wavelength Temperature Theta range for data collection Index ranges Data collection software Cell refinement software Data reduction software	880 STOE STADIVARI 1.54186 Å 230(2) K 3.504 to 68.472°. -14<=h<=14, -7<=k<=7, -31<=1<=32 X-Area Pilatus3_SV 1.31.127.0 (STOE, 2016) ² X-Area Recipe 1.33.0.0 (STOE, 2015) ³ X-Area Integrate 1.71.0.0 (STOE, 2016) ⁴ X-Area LANA 1.68.2.0 (STOE, 2016) ⁵ MERGEHKLF5 (Lutz, 2016) ⁶		
Reflections collected Independent reflections Completeness to theta = 67.686° Observed reflections Reflections used for refinement Extinction coefficient Absorption correction Max. and min. transmission Flack parameter (absolute struct.)	38035 $8441 [R(int) = 0.0686]$ 98.2% $5312[I > 2\sigma(I)]$ 8441 $X = 0.0033(3)$ Semi-empirical from equivalents ⁷ $0.7032 \text{ and } 0.1984$ $-0.005(14)^{8}$		
Largest diff. peak and hole	$0.279 \text{ and } -0.252 \text{ e.Å}^{-3}$		
Solution	dual space ⁹		
Refinement	Full-matrix least-squares on F ²		

Treatment of hydrogen atoms Programs used	Calculated positions, constr. ref. XT V2014/1 (Bruker AXS Inc., 2014) ¹⁰ SHELXL-2018/1 (Sheldrick, 2018) DIAMOND (Crystal Impact) ShelXle (Hübschle, Sheldrick, Dittrich, 2011)
Data / restraints / parameters	8441 / 1 / 513
Goodness-of-fit on F^2	0.899
R index (all data) R index conventional [I>2sigma(I)]	wR2 = 0.1031 R1 = 0.0449

 Table 6. Crystal data and structure refinement for rac-Ruimine.

Habitus, colour Crystal size Crystal system Space group Unit cell dimensions	prism, red $0.17 \ge 0.14 \ge 0.06 \text{ mm}$ Monoclinic $P2_{1/c} \qquad Z = 4$ $a = 19.4935(2) \text{ Å} \qquad = 90^{\circ}.$ $b = 25 \le 0.022(2) \text{ Å} \qquad = 0.75020(10)^{\circ}$
Volume Cell determination Empirical formula Moiety formula Cl ₂)	b = 25.6083(3) Å = 97.5930(10)°. c = 23.9361(3) Å = 90°. 11844.0(2) Å 72887 peaks with Theta 2.5 to 76.2°. C ₁₀₃ H _{104.50} Cl _{7.50} F ₂₄ N ₁₆ O ₂ P ₄ Ru ₂ 2(C ₅₀ H ₄₉ N ₈ O Ru), 4(F ₆ P), 1.5(C H Cl ₃), 1.5(C H ₂
Formula weight	2646.41
Density (calculated)	1.484 Mg/m^3
Absorption coefficient	4.930 mm ⁻¹
F(000) Diffractometer type Wavelength Temperature Theta range for data collection Index ranges Data collection software Cell refinement software Data reduction software Data reduction software Reflections collected Independent reflections Completeness to theta = 67.679° Observed reflections Reflections used for refinement Absorption correction Max. and min. transmission	5368 STOE STADIVARI 1.54178 Å 100(2) K 2.539 to 75.866°. -24 <=h <= 23, -30 <=k <= 32, -18 <=l <= 29 X-Area Pilatus3_SV 1.31.127.0 (STOE, 2016) ² X-Area Recipe 1.33.0.0 (STOE, 2015) ³ X-Area Integrate 1.71.0.0 (STOE, 2016) ⁴ X-Area LANA 1.68.2.0 (STOE, 2016) ⁵ PLATON/SQUEEZE (Spek, 2015) ⁶ 130276 24295 [R(int) = 0.0654] 99.8 % 18362[I > 2 σ (I)] 24295 Semi-empirical from equivalents ⁷ 0.3444 and 0.0969
Largest diff. peak and hole	1.721 and -0.919 e.Å ⁻³
Solution	intrinsic phases ⁸
Refinement	Full-matrix least-squares on F ²⁹
Treatment of hydrogen atoms Programs used	CH calculated, "riding", NH located, isotr. ref. XT V2014/1 (Bruker AXS Inc., 2014) ¹⁰ SHELXL-2018/3 (Sheldrick, 2018) 116

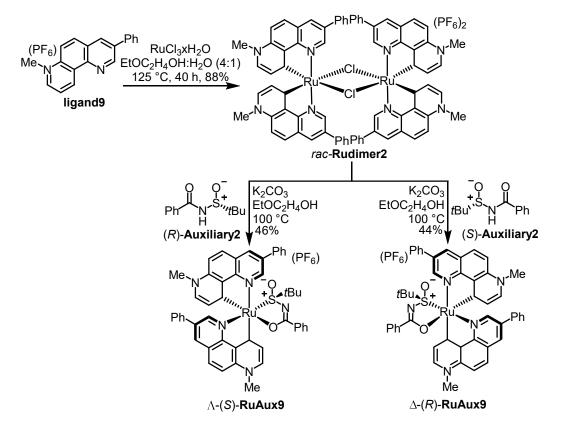
	DIAMOND (Crystal Impact)	
	ShelXle (Hübschle, Sheldrick, Dittrich, 2011)	
Data / restraints / parameters	24295 / 32 / 1524	
Goodness-of-fit on F^2	1.052	
R index (all data)	wR2 = 0.1293	
R index conventional [I>2sigma(I)]	R1 = 0.0470	

References

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4.3 Enantioselective Synthesis of γ-Lactams by Intramolecular C-H Amidation

4.3.1 Synthesis of the Ruthenium Catalysts

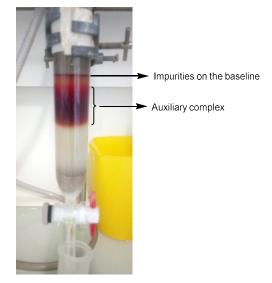


Synthesis of *rac*-Rudimer2: To a 10 mL Schlenk tube was added ligand9 (125 mg, 0.3 mmol), RuCl₃xH₂O (31 mg, 0.15 mmol) and 3.0 mL solvent mixture (EtOC₂H₄OH:H₂O = 4:1). The resulting solution was heated at 125 °C for 40 h. After that, 50 mg NH₄PF₆ dissolved in 0.5 mL deionized H₂O was added to the reaction solution. Ultrasonic was used for 3 min to achieve counteranion exchange. The liquid phase was removed by filtration and the resulting residue was washed with water (3 x 3 mL) and Et₂O (3 x 5 mL). The resulting residue was dried under high *vacuo* to provide crude *rac*-Rudimer2 (108 mg, 88% yield) as dark purple solid for the next step without further purification.

Synthesis of Λ -(*S*)-RuAux9: To a 10 mL Schlenk tube was added *rac*-Rudimer2 (164 mg, 0.1 mmol), (*R*)-*N*-(*tert*-butylsulfinyl)benzamide (34 mg, 0.15 mmol), K₂CO₃ (27.6 mg, 0.2 mmol) and EtOC₂H₄OH (3 mL) in sequence. The resulting solution was heated at 100 °C for 16 h. After that, solvent was removed under high *vacuo*. After cooling to room temperature, NH₄PF₆ (50 mg) in MeOH (3 mL) was added and stirred at room temperature for 30 min. Organic solvent was removed under *vacuo*. The resulting residue was diluted with CH₂Cl₂ and purified by flash chromatography (see

details below) to provide analytical pure Λ -(S)-RuAux9 (93 mg, 46% yield) as dark red solid.

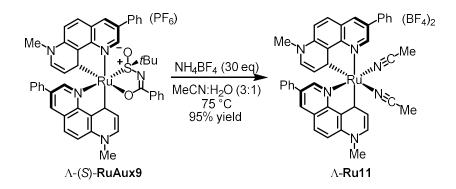
Procedures for the purification of Λ -(*S*)-**RuAux9**: First, 400 mL CH₂Cl₂:MeOH = 50:1 was used as eluent. After that, 300 mL CH₂Cl₂:MeOH = 20:1 was used. Next, 300 mL CH₂Cl₂:MeOH = 10:1 was used to provide the desired compound Λ -(*S*)-**RuAux9** with 95% ¹H NMR purity. A second short flash column was performed to remove some ruthenium complex decomposition products during the first column. For this second column, 600 mL CH₂Cl₂:MeOH = 15:1 was used to remove impurities on the baseline (see below).



A-(*S*)-**RuAux9:** ¹**H NMR** (500 MHz, CD₃CN) δ 10.11 (d, J = 6.9 Hz, 1H), 9.50 (d, J = 2.0 Hz, 1H), 9.00 (d, J = 2.0 Hz, 1H), 8.49 (d, J = 9.3 Hz, 1H), 8.43 (d, J = 1.9 Hz, 1H), 8.22-8.12 (m, 3H), 8.09 (d, J = 9.3 Hz, 1H), 7.99 (dd, J = 13.3, 8.1 Hz, 2H), 7.73-7.64 (m, 2H), 7.56 (d, J = 1.9 Hz, 1H), 7.48 (ddd, J = 22.7, 10.4, 5.4 Hz, 5H), 7.34 (dddd, J = 11.8, 8.9, 6.1, 4.7 Hz, 6H), 7.19-7.07 (m, 2H), 4.13 (s, 3H), 3.97 (s, 3H), 0.56 (s, 9H). ¹³C **NMR** (126 MHz, CD₃CN) δ 177.6, 151.8, 151.5, 150.7, 150.3, 142.2, 141.8, 138.1, 137.1, 136.8, 136.2, 136.1, 136.0, 135.7, 135.6, 135.6, 135.3, 135.3, 134.3, 133.4, 132.1, 131.9, 130.5, 130.3, 130.2, 130.1, 129.7, 129.0, 128.2, 127.6, 127.2, 125.9, 120.7, 120.4, 68.0, 55.3, 43.1, 42.9, 28.4, 23.7. ¹⁹F **NMR** (283 MHz, CD₃CN) δ -71.02, -73.52. **IR** (film): ν (cm⁻¹) 1612, 1531, 1504, 1455, 1330, 1173, 830, 779, 718, 669, 586, 556, 491, 428, 398. **HRMS** (ESI, *m/z*) calcd. for C₄₉H₄₂N₅O₂Ru₁S₁ [M-PF₆]⁺: 866.2110, found: 866.2117.

Synthesis of Δ -(*R*)-RuAux9: Identical synthetic procedures as for Λ -(*S*)-RuAux10 but with slightly reduced yield (89 mg, 44% yield).

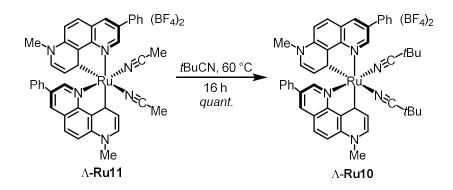
 Δ -(*R*)-**RuAux9**: Dark red solid. Analytical data of Δ -(*R*)-**RuAux10** identical with Λ -(*S*)-**RuAux10**.



Synthesis of Λ -Ru11: To a 10 mL Schlenk tube was added Λ -(*S*)-RuAux9 (50.6 mg, 0.05 mmol) and NH₄BF₄ (157 mg, 1.5 mmol) in MeCN:H₂O = 3:1 (3 mL). The resulting solution was heated at 75 °C for 16 h. After that, the solution was directly transferred to a column and purified by flash chromatography (see details below) to provide analytical pure Λ -Ru11 (42.7 mg, 95% yield) as dark red solid.

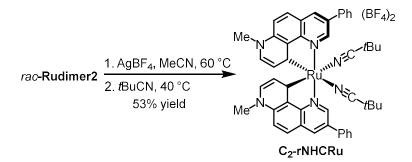
Procedures for the purification of Λ -**Ru11**: First, CH₂Cl₂:MeCN = 20:1 (300 mL) was used as eluent to remove the recovered auxiliary. Secondly, pure MeCN (200 mL) was used as eluent to provide the desired compound Λ -**Ru11** mixed with NH₄BF₄. Finally, the excess NH₄BF₄ was removed by running a short celite column using CH₂Cl₂:MeCN = 20:1 (60 mL). The solvent was removed under *vacuo* to provide analytical pure Λ -**Ru11** as red solid.

A-**Ru11:** ¹**H** NMR (500 MHz, CD₃CN) δ 9.98 (d, J = 2.0 Hz, 1H), 9.04 (d, J = 2.0 Hz, 1H), 8.68 (d, J = 6.5 Hz, 1H), 8.53 (dd, J = 16.6, 5.6 Hz, 2H), 8.35 (d, J = 9.3 Hz, 1H), 8.20 (d, J = 9.3 Hz, 1H), 8.13-8.04 (m, 4H), 8.02 (d, J = 1.9 Hz, 1H), 7.74-7.66 (m, 2H), 7.61 (ddd, J = 7.4, 4.0, 1.2 Hz, 1H), 7.41-7.23 (m, 6H), 7.04 (d, J = 6.5 Hz, 1H), 4.30 (s, 3H), 4.04 (s, 3H), 2.17 (s, 3H), 1.96 (s, 3H). ¹³C NMR (126 MHz, CD₃CN) δ 154.1, 153.8, 151.9, 150.6, 142.7, 142.1, 138.2, 137.6, 137.6, 137.1, 136.7, 136.0, 135.8, 135.1, 134.9, 134.6, 133.9, 133.7, 132.9, 132.4, 130.6, 130.2, 130.0, 129.7, 128.7, 128.0, 127.0, 126.6, 126.1, 125.4, 120.3, 119.9, 43.3, 43.1, 4.2. ¹⁹F NMR (283 MHz, CD₃CN) δ -151.08, -151.13.



Synthesis of Λ -Ru10: Λ -Ru11 (20 mg) was dissolved in 3 mL pivalonitrile and heated at 60 °C for 16 h. After that, the solvent was removed under *vacuo* to provide Λ -Ru10 as a red solid in quantitative yield.

A-**Ru10**: ¹**H NMR** (300 MHz, CD₃CN) δ 9.97 (d, J = 2.0 Hz, 1H), 9.04 (d, J = 2.0 Hz, 1H), 8.59-8.50 (m, 3H), 8.35 (d, J = 9.3 Hz, 1H), 8.20 (d, J = 9.3 Hz, 1H), 8.14-7.99 (m, 5H), 7.70 (t, J = 7.4 Hz, 2H), 7.62 (d, J = 7.3 Hz, 1H), 7.38-7.26 (m, 6H), 7.02 (d, J = 6.4 Hz, 1H), 4.30 (s, 3H), 4.04 (s, 3H), 1.34 (s, 9H), 1.17 (s, 9H). ¹³C NMR (126 MHz, CD₃CN) δ 154.0, 153.8, 151.8, 150.5, 142.6, 142.1, 138.3, 137.8, 137.6, 137.1, 136.7, 136.1, 135.8, 135.2, 134.9, 134.4, 133.9, 133.8, 133.4, 133.0, 132.0, 130.6, 130.2, 130.1, 129.7, 128.7, 128.7, 128.0, 127.0, 126.2, 120.3, 119.9, 79.2, 78.9, 78.6, 43.4, 43.2, 28.4, 28.2. ¹⁹F NMR (235 MHz, CD₃CN) δ -151.82, -151.87. **IR** (film): v (cm⁻¹) 1624, 1584, 1535, 1460, 1393, 1353, 1304, 1229, 1170, 1022, 931, 871, 814, 761, 726, 696, 651, 569, 518, 446.

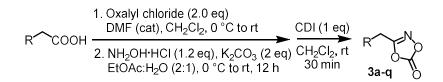


Synthesis of C₂-rNHCRu: To a 10 mL Schlenk tube were added *rac*-Rudimer2 (33 mg, 0.02 mmol) and AgBF₄ (9.7 mg, 0.05 mmol) in 3 mL MeCN. The resulting solution was heated at 60 °C for 16 h. After that, the above solution was directly transferred to a column and purified by flash chromatography (CH₂Cl₂:MeCN = 5:1) to provide analytical pure ruthenium intermediate. The intermediate was dissolved in 2 mL pivalonitrile. The resulting solution was heated at 40 °C for 10

min. After that, organic solvent was removed under *vacuo* to provide analytical pure C₂-rNHCRu (20.9 mg, 53% yield) as a brown solid.

C₂-rNHCRu: ¹**H NMR** (300 MHz, CD₃CN) δ 9.86 (d, J = 1.8 Hz, 1H), 8.93 (d, J = 1.8 Hz, 1H), 8.55 (d, J = 9.3 Hz, 1H), 8.15 (d, J = 9.3 Hz, 1H), 8.09-8.03 (m, 2H), 7.73 (t, J = 7.4 Hz, 2H), 7.65 (d, J = 7.3 Hz, 1H), 7.50 (d, J = 6.5 Hz, 1H), 6.81 (d, J = 6.4 Hz, 1H), 4.08 (s, 3H), 1.38 (s, 9H). ¹³C **NMR** (75 MHz, CD₃CN) δ 154.6, 154.4, 142.2, 138.5, 137.9, 137.5, 134.9, 133.7, 133.4, 131.0, 130.6, 130.3, 130.1, 128.8, 128.2, 127.1, 112.0, 43.3, 28.5. ¹⁹F **NMR** (235 MHz, CD₃CN) δ -152.37, -152.42. **IR** (film): 1624, 1584, 1535, 1460, 1422, 1393, 1353, 1304, 1229, 1170, 1022, 931, 871, 814, 761, 726, 696, 651, 569, 518, 446.

4.3.2 Synthesis of the Substrates



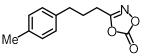
All substrates were synthesized according to the published procedures.¹⁻³

General procedure: Oxalyl chloride (4.0 mmol) and DMF (2 drops) were added to a solution of the carboxylic acid (2.0 mmol) in dichloromethane (30 mL) at 0 °C. The mixture was allowed to stir at room temperature for 2.5-4 h. Then, the reaction mixture was concentrated, and the crude product was used directly in the next reaction. Hydroxylamine hydrochloride (1.2 equiv) was added to a biphasic mixture of K_2CO_3 (2.0 equiv) in a 2:1 mixture of EtOAc (16 mL) and H_2O (8 mL). The resulting solution was cooled to 0 °C followed by dropwise addition of the unpurified acid chloride dissolved in a minimum amount of EtOAc under air. The flask containing the acid chloride was then rinsed with additional EtOAc. The reaction was warmed to room temperature for stirring additional 12 h. The phases were separated and the aqueous phase was extracted twice with EtOAc. The combined organic layers were dried over MgSO₄, filtered, and evaporated under reduced pressure. The resulting solid was dissolved in CH₂Cl₂ (10 mL), CDI (1 equiv) was added to the resulting solution. The organic layer was collected and dried over MgSO₄. The resulting solid was purified by flash chromatography (CH₂Cl₂, R_f value = 0.95) to provide the desired analytical pure 1,4,2-dioxazol-5-ones.

Compound 3a

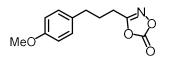
3a, 72% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 7.28-7.20 (m, 2H), 7.19-7.07 (m, 3H), 2.67 (t, *J* = 7.4 Hz, 2H), 2.54 (t, *J* = 7.5 Hz, 2H), 1.99 (p, *J* = 7.5 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.6, 154.2, 139.9, 128.8, 128.6, 126.7, 34.7, 26.0, 24.2.

Compound 3b



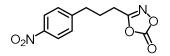
3b, 78% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 7.13 (d, *J* = 8.0 Hz, 2H), 7.07 (d, *J* = 8.1 Hz, 2H), 2.71 (t, *J* = 7.3 Hz, 2H), 2.61 (t, *J* = 7.5 Hz, 2H), 2.34 (s, 3H), 2.05 (p, *J* = 7.5 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.6, 154.2, 136.8, 136.2, 129.5, 128.5, 34.2, 26.1, 24.1, 21.1.

Compound 3c



3c, 65% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 7.09 (d, *J* = 8.6 Hz, 2H), 6.85 (d, *J* = 8.7 Hz, 2H), 3.80 (s, 3H), 2.69 (t, *J* = 7.3 Hz, 2H), 2.60 (t, *J* = 7.5 Hz, 2H), 2.03 (p, *J* = 7.4 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.6, 158.5, 154.2, 131.9, 129.5, 114.3, 55.4, 33.8, 26.3, 24.1.

Compound 3d

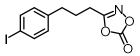


3d, 80% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.17 (d, *J* = 8.7 Hz, 2H), 7.36 (d, *J* = 8.7 Hz, 2H), 2.93-2.80 (m, 2H), 2.68 (t, *J* = 7.4 Hz, 2H), 2.19-2.03 (m, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.1, 154.0, 147.7, 147.0, 129.4, 124.1, 34.5, 25.5, 24.2.

Compound 3e

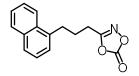
3e, 82% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 7.44 (d, *J* = 8.4 Hz, 2H), 7.06 (d, *J* = 8.4 Hz, 2H), 2.70 (t, *J* = 7.4 Hz, 2H), 2.62 (t, *J* = 7.5 Hz, 2H), 2.04 (p, *J* = 7.5 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.4, 154.1, 138.9, 131.9, 130.3, 120.5, 34.1, 25.9, 24.1.

Compound 3f



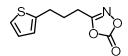
3f, 58% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 7.63 (d, *J* = 8.3 Hz, 2H), 6.94 (d, *J* = 8.3 Hz, 2H), 2.69 (t, *J* = 7.5 Hz, 2H), 2.61 (t, *J* = 7.5 Hz, 2H), 2.04 (p, *J* = 7.5 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.4, 154.1, 139.6, 137.9, 130.6, 91.8, 34.2, 25.8, 24.1.

Compound 3g



3g, 74% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 7.99 (d, *J* = 8.0 Hz, 1H), 7.93-7.86 (m, 1H), 7.77 (d, *J* = 8.2 Hz, 1H), 7.53 (pd, *J* = 6.8, 1.5 Hz, 2H), 7.46-7.38 (m, 1H), 7.32 (d, *J* = 6.8 Hz, 1H), 3.21 (t, *J* = 7.4 Hz, 2H), 2.68 (t, *J* = 7.5 Hz, 2H), 2.21 (p, *J* = 7.5 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 166.6, 154.2, 136.0, 134.2, 131.7, 129.2, 127.6, 126.6, 126.4, 125.9, 125.6, 123.4, 31.9, 25.2, 24.5.

Compound 3h

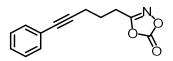


3h, 62% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 7.17 (dd, *J* = 5.1, 1.1 Hz, 1H), 6.94 (dd, *J* = 5.1, 3.4 Hz, 1H), 6.83 (dd, *J* = 3.4, 0.9 Hz, 1H), 2.98 (t, *J* = 7.2 Hz, 2H), 2.67 (t, *J* = 7.5 Hz, 2H), 2.11 (p, *J* = 7.3 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.4, 154.2, 142.4, 127.1, 125.3, 124.0, 28.8, 26.4, 4.0.

Compound 3i

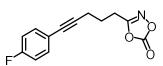
3i, 62% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 2.80 (t, *J* = 7.5 Hz, 2H), 2.37 (td, *J* = 6.7, 2.6 Hz, 2H), 2.04 (t, *J* = 2.7 Hz, 1H), 1.96 (dt, *J* = 18.0, 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.2, 154.2, 81.8, 70.5, 23.8, 23.3, 17.8.

Compound 3j



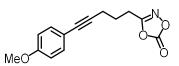
3j, 70% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 7.44-7.35 (m, 2H), 7.33-7.27 (m, 3H), 2.85 (t, *J* = 7.5 Hz, 2H), 2.59 (t, *J* = 6.7 Hz, 2H), 2.04 (dt, *J* = 18.1, 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.3, 154.2, 131.7, 128.5, 128.2, 123.3, 87.2, 82.7, 24.0, 23.6, 18.8.

Compound 3k



3k, 73% yield, colorless oil. ¹**H NMR** (250 MHz, CDCl₃) δ 7.37 (dd, *J* = 8.8, 5.4 Hz, 2H), 6.99 (t, *J* = 8.7 Hz, 2H), 2.84 (t, *J* = 7.5 Hz, 2H), 2.58 (t, *J* = 6.7 Hz, 2H), 2.10-1.96 (m, 2H).

Compound 31

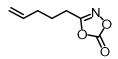


31, 77% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 7.32 (d, *J* = 8.8 Hz, 2H), 6.82 (d, *J* = 8.8 Hz, 2H), 3.80 (s, 3H), 2.84 (t, *J* = 7.5 Hz, 2H), 2.57 (t, *J* = 6.7 Hz, 2H), 2.09-1.95 (m, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.4, 159.6, 154.2, 133.1, 115.4, 114.1, 85.6, 82.5, 55.4, 24.0, 23.7, 18.8.

Compound 3m

3m, 62% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 2.77 (t, *J* = 7.5 Hz, 2H), 2.39 (t, *J* = 6.7 Hz, 2H), 2.01-1.85 (m, 2H), 0.15 (s, 9H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.3, 154.2, 104.2, 87.1, 23.9, 23.4, 19.2, 0.1.

Compound 3n



3n, 71% yield, colorless oil. ¹**H NMR** (300 MHz, CD₂Cl₂) δ 5.79 (ddt, *J* = 17.0, 10.2, 6.7 Hz, 1H), 5.18-4.94 (m, 2H), 2.64 (t, *J* = 7.5 Hz, 2H), 2.18 (q, *J* = 7.0 Hz, 2H), 1.82 (p, *J* = 7.4 Hz, 2H). ¹³**C NMR** (75 MHz, CD₂Cl₂) δ 167.3, 154.7, 137.0, 116.5, 32.9, 24.4, 24.0.

Compound 3o



30, 80% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 2.37 (s, 2H), 2.03 (s, 3H), 1.83-1.53 (m, 12H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.1, 154.5, 42.3, 39.0, 36.5, 33.3, 28.5.

Compound 3p



3p, 58% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 7.79 (dd, *J* = 7.8, 1.2 Hz, 1H), 7.56 (td, *J* = 7.6, 1.4 Hz, 1H), 7.42 (td, *J* = 7.7, 1.0 Hz, 1H), 7.35-7.17 (m, 4H), 7.14 (dd, *J* = 6.6, 5.2 Hz, 2H), 4.35 (s, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 163.9, 153.7, 141.9, 139.0, 133.4, 132.1, 129.5, 129.2, 128.8, 127.2, 126.7, 119.5, 40.0.

Compound 3q

3q, 84% yield, white solid. ¹**H NMR** (300 MHz, CDCl3) δ 7.25-7.14 (m, 4H), 3.22 (dd, J = 15.3, 7.5 Hz, 2H), 2.91 (tt, J = 13.4, 6.9 Hz, 1H), 2.83-2.75 (m, 3H), 2.72 (d, J = 6.3 Hz, 1H). ¹³**C NMR** (75 MHz, CDCl3) δ 166.1, 154.2, 141.5, 126.9, 124.8, 38.8, 35.7, 30.4.

4.3.3 Ruthenium Catalyzed Enantioselective Intramolecular C-H Amidations

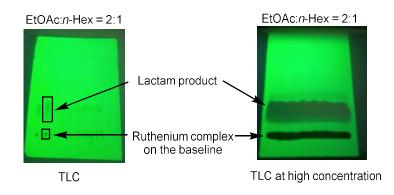
General Procedures: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrates **3a-q** (0.2 mmol) and Λ -**Ru10** (0.2 mg, 0.0002 mmol, 0.1 mol%) under an atmosphere of N₂. Pre-cooled (using an ice water bath) "super dried" (Acros) 1,2-dichlorobenzene (0.4 mL, 0.5 M) from "Acros" was added via syringe in sequence. The reaction mixture was stirred at the indicated temperature for the indicated time under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc with 5% MeOH/*n*–Hexane = 1:2 to 2:1) to afford the analytical pure products **4a-q**. Enantiomeric ratios were determined by HPLC analysis on chiral stationary phase. The absolute configuration of the product **4e** was confirmed by single crystallography and X-ray structure analysis as *S*-configuration.

Additional experimental informations:

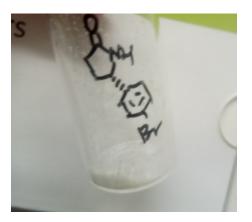
- a) Control of reaction temperature. The catalytic reactions were set up in a cold room at 4 °C.
- b) Execution of the reactions. All catalytic reactions were carried out in 10 mL Schlenk tubes from "Synthware" under N₂ atmosphere (see image below).



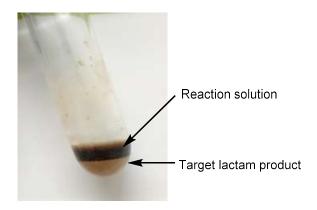
c) TLC monitoring of the final reaction solution. See images below.



- d) Isolation of the final product.
- The C-H amidation products can be isolated by flash chromatography without any trace contaminations from the ruthenium catalyst as evident by the white color of the products (see image below).



2) For the catalytic reactions of 3b, 3e, 3f, 3g, and 3q. The lactam products can also be isolated without any chromatography by only filtration (see image below). This method was used for the gram-scale synthesis of 4e.



Compound 5 (side product)

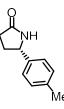
5: The Curtius-type decomposition pathway provide the formation of isocyanate 5 which can be isolated by flash chromatography on silica gel (pure CH₂Cl₂, R_f value = 0.9-0.95). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.41-7.26 (m, 2H), 7.25-7.08 (m, 3H), 3.32 (t, *J* = 6.6 Hz, 2H), 2.79-2.65 (m, 2H), 2.01-1.85 (m, 2H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 141.3, 128.87, 128.85, 126.5, 42.7, 33.1, 33.0.

Compound (S)-4a



Starting from **3a** (41.0 mg, 0.20 mmol) according to the general procedure to provide **4a** as a white solid (30.6 mg, 95% yield) and with 96:4 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 18.6 min, t_r (minor) = 21.6 min). [α]_D²² = -24.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.44-7.22 (m, 5H), 6.27 (s, 1H), 4.75 (t, *J* = 7.1 Hz, 1H), 2.69-2.31 (m, 3H), 1.97 (tdd, *J* = 9.1, 8.4, 4.4 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 178.6, 142.6, 129.1, 128.1, 125.8, 58.2, 31.5, 30.4. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₁N₁O₁Na₁[M+Na]⁺: 184.0733, found: 184.0733.

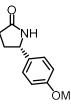
Compound (S)-4b



Starting from **3b** (43.8 mg, 0.20 mmol) according to the general procedure to provide **4b** as a white solid (33.6 mg, 96% yield) and with 95:5 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 20.1 min, t_r (minor) = 21.2 min). [α]_D²² = -25.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.21-7.10 (m, 4H), 6.14 (s,

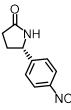
1H), 4.71 (t, J = 7.1 Hz, 1H), 2.64-2.48 (m, 1H), 2.47-2.37 (m, 2H), 2.34 (s, 3H), 2.05-1.83 (m, 1H).
¹³C NMR (75 MHz, CDCl₃) δ 178.7, 139.4, 137.9, 129.7, 125.8, 58.2, 31.5, 30.5, 21.2. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₃N₁O₁Na₁[M+Na]⁺: 198.0889, found: 198.0889.

Compound (S)-4c



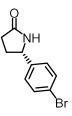
Starting from **3c** (47.0 mg, 0.20 mmol) according to the general procedure to provide **4c** as a white solid (37.9 mg, 99% yield) and with 89:11 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 31.8 min, t_r (minor) = 21.2 min). [α]_D²² = -28.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.20 (d, *J* = 8.6 Hz, 2H), 6.88 (d, *J* = 8.7 Hz, 2H), 6.45 (s, 1H), 4.70 (t, *J* = 7.0 Hz, 1H), 3.79 (s, 3H), 2.62-2.31 (m, 3H), 2.03-1.84 (m,12 1H). ¹³C NMR (75 MHz, CDCl₃) δ 178.6, 159.5, 134.5, 127.0, 114.4, 57.9, 55.5, 31.6, 30.5. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₃N₁O₂Na₁[M+Na]⁺: 214.0838, found: 214.0838.

Compound (S)-4d



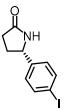
Starting from **3d** (50.0 mg, 0.20 mmol) according to the general procedure to provide **4d** as a white solid (34.5 mg, 84% yield) and with 95:5 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 38.9 min, t_r (minor) = 46.1 min). [α]_D²² = -47.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 8.21 (d, *J* = 8.8 Hz, 2H), 7.50 (d, *J* = 8.6 Hz, 2H), 6.86 (s, 1H), 2.70-2.55 (m, 1H), 2.45-2.35 (m, 2H), 2.03-1.82 (m, 1H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 178.8, 150.9, 148.0, 127.0, 124.5, 57.8, 31.4, 30.4. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₀N₂O₃Na₁[M+Na]⁺: 229.0584, found: 229.0584.

Compound (S)-4e



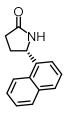
Starting from **3e** (56.6 mg, 0.20 mmol) according to the general procedure to provide **4e** as a white solid (45.8 mg, 96% yield) and with 93:7 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 26.8 min, t_r (minor) = 24.0 min). [α]_D²² = -30.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.49 (d, *J* = 8.4 Hz, 2H), 7.17 (d, *J* = 8.4 Hz, 2H), 6.41 (s, 1H), 4.73 (t, *J* = 7.1 Hz, 1H), 2.68-2.34 (m, 3H), 2.05-1.81 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 178.7, 141.5, 132.2, 127.5, 122.0, 57.8, 31.4, 30.3. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₀Br₁N₁O₁Na₁[M+Na]⁺: 261.9838, found: 261.9838.

Compound (S)-4f



Starting from **3f** (66.2 mg, 0.20 mmol) according to the general procedure to provide **4f** as a white solid (52.6 mg, 92% yield) and with 92:8 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 29.7 min, t_r (minor) = 26.4 min). [α]_p²² = -11.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.70 (d, *J* = 8.3 Hz, 2H), 7.05 (d, *J* = 8.3 Hz, 2H), 5.93 (s, 1H), 4.70 (t, *J* = 7.0 Hz, 1H), 2.69-2.30 (m, 3H), 2.02-1.83 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 178.4, 142.4, 138.2, 127.7, 93.4, 57.7, 31.5, 30.2. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₀I₁N₁O₁Na₁[M+Na]⁺: 309.9699, found: 309.9698.

Compound (S)-4g



Starting from **3g** (51.0 mg, 0.20 mmol) according to the general procedure to provide **4g** as a white solid (40.0 mg, 95% yield) and with 92:8 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 32.3 min, t_r (minor) = 37.1 min). [α]_D²² = -142.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.01-7.86 (m, 2H), 7.80 (d, *J* = 7.9 Hz, 1H), 7.61-7.42 (m, 4H), 6.60 (s, 1H), 5.54 (dd, *J* = 8.0, 5.1 Hz, 1H), 2.81 (ddd, *J* = 15.7, 12.7, 8.2 Hz, 1H), 2.54-2.34 (m, 2H), 2.14-1.98 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 179.0, 138.2, 134.1, 130.2, 129.2, 128.3, 126.6, 126.0, 125.6, 122.5, 121.4, 54.6, 30.1, 29.8. HRMS (ESI, *m/z*) calcd. for C₁₄H₁₃N₁O₁Na₁[M+Na]⁺: 234.0889, found: 234.0888.

Compound (S)-4h

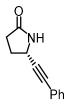
Starting from **3h** (42.2 mg, 0.20 mmol) according to the general procedure to provide **4h** as a white solid (22.1 mg, 66% yield) and with 94:6 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol =95:5, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 23.3 min, t_r (minor) = 27.9 min). [α]_p²² = -6.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR δ 7.26 (dd, *J* = 6.2, 1.3 Hz, 1H), 7.02-6.93 (m, 2H), 6.51 (s, 1H), 5.03 (t, *J* = 6.8 Hz, 1H), 2.68-2.33 (m, 3H), 2.22-2.05 (m, 1H). ¹³C NMR δ 177.9, 146.5, 127.1, 125.0, 124.3, 54.1, 31.8, 30.2. HRMS (ESI, *m/z*) calcd. for C₈H₉N₁O₁S₁ [M+Na]⁺: 190.0297, found: 190.0296.

Compound (S)-4i



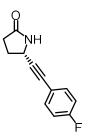
Starting from **3i** (30.6 mg, 0.20 mmol) according to the general procedure to provide **4i** as a white solid (13.3 mg, 61% yield). and with 93:7 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 27.8 min, t_r (minor) = 19.2 min). [α]_D²² = -24.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 6.58 (s, 1H), 4.37 (ddd, *J* = 7.2, 5.4, 2.1 Hz, 1H), 2.53-2.07 (m, 5H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 177.8, 83.6, 72.1, 44.9, 29.7, 29.3.

Compound (S)-4j



Starting from **3j** (45.8 mg, 0.20 mmol) according to the general procedure to provide **4j** as a white solid (28.7 mg, 78% yield) and with 95:5 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 70:30, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 16.0 min, t_r (minor) = 11.3 min). [α]_D²² = -5.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 7.49-7.38 (m, 2H), 7.37-7.25 (m, 3H), 6.52 (s, 1H), 4.60 (dd, *J* = 7.2, 5.2 Hz, 1H), 2.59-2.38 (m, 2H), 2.38-2.17 (m, 2H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 177.8, 132.0, 129.0, 128.8, 122.8, 88.8, 83.9, 45.7, 29.9, 29.6.

Compound (S)-4k

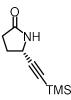


Starting from **3k** (49.4 mg, 0.20 mmol) according to the general procedure to provide **4k** as a white solid (29.9 mg, 74% yield) and with 95:5 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 23.2 min, t_r (minor) = 14.7 min). [α] $_{p^{22}}$ = -26.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.47-7.30 (m, 2H), 7.10-6.89 (m, 2H), 6.14 (s, 1H), 4.65-4.51 (m, 1H), 2.62-2.44 (m, 2H), 2.43-2.20 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 177.5, 164.5, 161.2, 133.8, 133.7, 118.5, 118.4, 116.0, 115.7, 87.8, 83.2, 45.4, 29.6, 29.3. ¹⁹F NMR (235 MHz, CDCl₃) δ -110.29.

Compound (S)-41

Starting from **31** (51.8 mg, 0.20 mmol) according to the general procedure to provide **41** as a white solid (34.6 mg, 82% yield) and with 90:10 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 92 : 8, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 21.1 min, t_r (minor) = 19.5 min). [α]_D²² = -25.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.34 (d, *J* = 8.9 Hz, 2H), 6.83 (d, *J* = 8.9 Hz, 2H), 6.05 (s, 1H), 4.59 (dd, *J* = 7.4, 5.1 Hz, 1H), 3.79 (d, *J* = 7.6 Hz, 3H), 2.58-2.17 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 177.6, 160.0, 133.3, 114.4, 114.1, 86.6, 84.1, 55.4, 45.5, 29.6, 29.4.

Compound (S)-4m



Starting from **3m** (45.0 mg, 0.20 mmol) according to the general procedure to provide **4m** as a white solid (25.3 mg, 70% yield) and with 94:6 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IC column 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 31.2 min, t_r (minor) = 17.2 min). [α]_D²² = -9.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 6.07 (s, 1H), 4.41-4.28 (m, 1H), 2.48-2.06 (m, 4H), 0.21-0.10 (m, 9H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 177.5, 105.3, 88.7, 45.6, 29.7, 29.5, -0.1.

Compound (S)-4n



Starting from **3n** (32.0 mg, 0.20 mmol) according to the general procedure to provide **4n** as a white solid (9.3 mg, 42% yield) and with 80:20 e.r. as determined by HPLC analysis (column: Daicel Chiralpak ADH column 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 96:4, flow rate: 0.8 mL/min, column temperature: 30 °C, retention times: t_r (major) = 17.4 min, t_r (minor) = 20.4 min). [α]_D²² = -4.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 6.57 (s, 1H), 5.82 (ddd, *J* = 17.0, 10.2, 6.7 Hz, 1H), 5.15 (dd, *J* = 32.3, 13.6 Hz, 2H), 4.19-4.00 (m, 1H), 2.36-2.16 (m, 3H), 1.88-1.72 (m, 1H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 178.5, 139.6, 115.4, 57.0, 30.2, 28.5. HRMS (ESI, *m/z*) calcd. for C₆H₉N₁O₁Na₁[M+Na]⁺: 134.0576, found: 134.0577.

Compound (R)-40

Starting from 30 (47.0 mg, 0.20 mmol) according to the general procedure to provide 40 as a white

solid (5.3 mg, 14% yield) and with 96:4 e.r. as determined by HPLC analysis (column: Daicel Chiralpak ODH column 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 0.7 mL/min, column temperature: 25 °C, retention times: t_r (major) = 14.5 min, t_r (minor) = 16.7 min). [α]_D²² = -9.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₂Cl₂) δ 5.59 (s, 1H), 3.45 (d, *J* = 2.2 Hz, 1H), 2.10-1.95 (m, 3H), 1.91-1.80 (m, 5H), 1.79-1.55 (m, 6H), 1.42 (dd, *J* = 10.1, 2.3 Hz, 1H). ¹³C NMR (75 MHz, CD₂Cl₂) δ 179.0, 64.3, 46.6, 40.4, 38.9, 37.6, 37.5, 37.2, 30.1, 29.9, 29.6, 28.0. HRMS (ESI, *m/z*) calcd. for C₁₂H₁₇N₁O₁Na₁[M+Na]⁺: 214.1202, found: 214.1203.

Compound (R)-4p



Starting from **3p** (50.6 mg, 0.20 mmol) according to the general procedure to provide **4p** as a white solid (22.6 mg, 54% yield) and with 71.29 e.r. as determined by HPLC analysis (column: Daicel Chiralpak ODH column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 14.5 min, t_r (minor) = 16.6 min). [α]_p²² = -14.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.89 (dd, *J* = 6.2, 1.9 Hz, 1H), 7.57-7.42 (m, 2H), 7.40-7.30 (m, 3H), 7.30-7.20 (m, 3H), 6.71 (s, 1H), 5.63 (s, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 171.1, 148.1, 138.6, 132.5, 130.9, 129.2, 128.7, 128.5, 127.0, 124.0, 123.5, 60.9. HRMS (ESI, *m/z*) calcd. for C₁₄H₁₁N₁O₁H[M+H]⁺: 210.0913, found: 210.0913.

Compound (3*R*, 8*S*)-4q

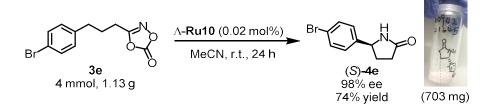


Starting from **3q** (43.4 mg, 0.20 mmol) according to the general procedure to provide **4q** as a white solid (31.8 mg, 92% yield) and with 90:10 e.r. as determined by HPLC analysis (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 6.9 min, t_r (minor) = 12.1 min). [α]_D²² = +14.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.28-7.10 (m, 4H), 7.02 (s, 1H), 4.95 (d, *J* = 7.0 Hz, 1H), 3.32-3.13 (m, 2H), 2.84-2.70 (m, 1H), 2.69-2.56 (m, 1H),

2.20-2.07 (m, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 177.8, 142.6, 141.7, 128.8, 127.4, 125.5, 124.9, 63.6, 38.6, 37.7, 37.7. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₁N₁O₁Na[M+Na]⁺: 196.0733, found: 196.0732.

4.3.4 Gram-Scale Reactions and Mechanistic Study

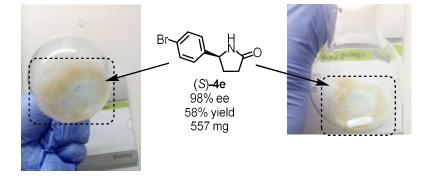
1) Gram-Scale Synthesis of (S)-4e, (R)-4e and (S)-4i



Procedure: To a 100 mL round bottom Schlenk flask was added **3e** (1.13 g, 4 mmol) and Λ -**Ru10** (0.8 mg, 0.0008 mmol, 0.02 mol% catalyst loading). Freshly distilled MeCN (8 mL) was added to the flask and the resulting reaction solution was stirred at room temperature for 24 h under an atmosphere of N₂. After complete consumption of **3e** monitored by TLC, (*S*)-**4e** was isolated without column chromatography by precipitation and crystallization.

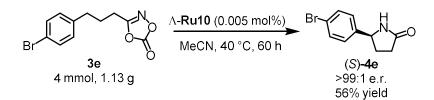
Work-up of the above reaction:

Enantiopure (S)-4e directly precipitated (557 mg, 58% yield) during the reaction and was isolated easily by removal of the remaining solution via syringe since the product was sticking to the wall of the flask (see image below). Enantiomeric ratio was established by HPLC analysis as 98% ee

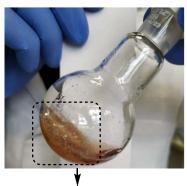


The remaining solution was cooled with an ice-water bath and (S)-4e crystals were formed and collected (146 mg).

The combined two batches of (S)-4e provided a total yield of 74% (703 mg). Enantiomeric ratio was established by HPLC analysis as 98% ee

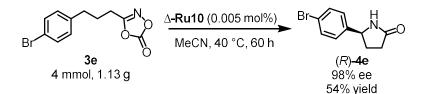


Procedure: To a 25 mL round bottom flask was added **3e** (1.13 g, 4 mmol) and Λ -**Ru10** (0.2 mg, 0.0002 mmol, 0.005 mol% catalyst loading). Freshly distilled MeCN (4 mL) was added to the flask and the resulting reaction solution was stirred at 40 °C for 60 h under an atmosphere of N₂. Afterwards, (*S*)-**4e** was isolated without column chromatography by crystallization upon cooling with an ice-water bath (Note: No precipitation of (*S*)-**4e** occurred at the elevated temperature of 40 °C).

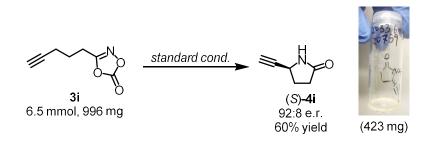


(S)-4e crystal formed

Work up of the above reaction: The reaction solution was cooled with an ice-water bath and (*S*)-4e crystals were formed and collected by filtration. The resulting filtrate was washed with MeCN (pre-cooled to $0 \,^{\circ}$ C) (3 x 2 mL) to provide analytical pure (*S*)-4e (535 mg, 56% yield) with 98% ee.

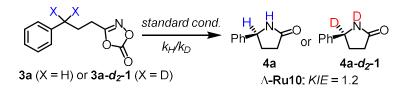


Procedure: The same transformation of **3e** to **4e** by using 0.005 mol% Δ -configuration **Ru10** catalyst provided lactam product **4e** in 54% yield with 98% ee as (*R*)-configuration.



Procedure: To a 50 mL round bottom Schlenk flask (pre-dried for 3 times by using heating gun) was added **3i** (996 mg, 6.5 mmol) and Λ -**Ru10** (6.5 mg, 0.0065 mmol, 0.1 mol% catalyst loading). 1,2-Dichlorobenzene (13 mL) was added to the flask and the resulting reaction solution was stirred in a cold room for 30 h under an atmosphere of N₂. After that, the reaction solution was directly transferred to flash column chromatography (*n*-Hexane:EtOAc = 1:1 R_f = 0.3-0.4) to provide analytical pure (*S*)-**4i** as white solid (423 mg, 60% yield) with 84% ee.

2) Kinetic Isotope Effects



Procedure: 3a (21.0 mg, 0.2 mmol) and **3a**- d_2 -**1** (21.0 mg, 0.2 mmol) were seperated as individual reactions. **3a** or **3a**- d_2 -**1** was added to a solution of Λ -**Ru10** (0.2 mg, 0.0002 mmol) in pre-cooled (using an ice water bath) super dried 1,2-dichlorobenzene (0.4 mL, 0.5 M). Then 10.6 µL Cl₂CHCHCl₂ was added as the internal standard. The reaction mixture was stirred at 0 °C under an atmosphere of nitrogen. Aliquots were taken at time intervals as indicated in the figure below. The aliquot was analyzed by ¹H NMR spectroscopy for the formation of product (blue square for **4a**, red square for **4a**- d_2 -1).

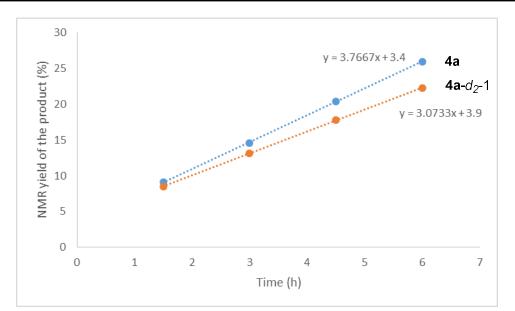
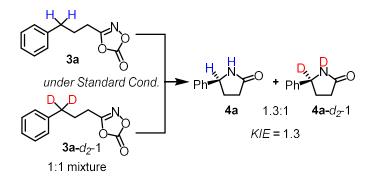


Figure 80. Initial rate of 4a and $4a-d_2-1$ formation under the above reaction conditions.



Procedure: 3a (21.0 mg, 0.1 mmol) and **3a**- d_2 -**1** (21.0 mg, 0.1 mmol) were added to a solution of Λ -**Ru10** (0.2 mg, 0.0002 mmol) in pre-cooled (using an ice water bath) super dried 1,2-dichlorobenzene (0.4 mL, 0.5 M). The reaction solution was set up in a cold room at 4 °C. After 6 h reaction time (<50% conv., monitored by crude ¹H NMR), the C–H amidation product was isolated. ¹H NMR analysis showed a ratio **4a**:**4a**- d_2 -1 of 1.3:1. (Both **4a** and **4a**- d_2 -1 have the same signal at 5.96 ppm but only **4a** has the signal at 4.76 ppm)

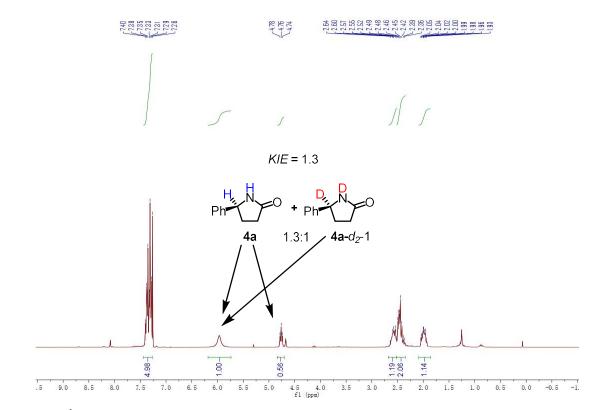
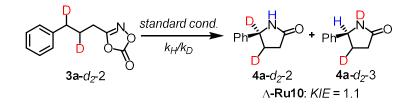


Figure 81. ¹H NMR spectra for the analysis of KIE value.



Procedure: 3a- d_2 -2 (21.0 mg, 0.1 mmol) was added to a solution of Λ -**Ru10** (0.2 mg, 0.0002 mmol) in pre-cooled (using an ice water bath) super dried 1,2-dichlorobenzene (0.4 mL, 0.5 M). The reaction solution was setup in a cold room at 4 °C room temperature. After 30 h reaction time, the C–H amidation product was isolated. ¹H NMR analysis showed a ratio **4a**- d_2 -2:**4a**- d_2 -3 of 1.1:1.

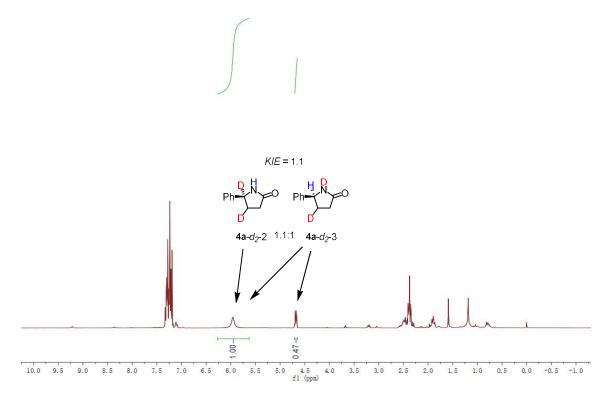
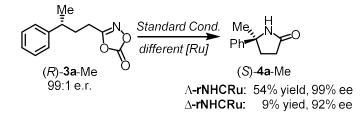


Figure 82. ¹H NMR spectra for the analysis of KIE value.

3) Probing the Stereochemistry



Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrate (*R*)-**3a**-Me (0.2 mmol) and Λ or Δ -**Ru10** (0.2 mg, 0.0002 mmol, 0.1 mol%) under an atmosphere of N₂. Pre-cooled (using an ice water bath) "super dried" (Acros) 1,2-dichlorobenzene (0.4 mL, 0.5 M) from "Acros" was added via syringe in sequence. The reaction mixture was stirred at 4 °C for 30 h under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc with 5% MeOH/*n*-*H*exane = 1:2 to 2:1) to afford the analytical pure products (*S*)-**4a**-Me. Enantiomeric ratios were determined by HPLC analysis on chiral stationary phase. (column: Daicel Chiralpak IA column 250 x 4.6 mm, absorption: λ = 220 nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate 1.0

mL/min, column temperature: 30 °C, retention times: t_r (major) = 16.9 min, t_r (minor) = 19.3 min). The HPLC spectrums were shown below. The observed results indicated a singlet nitrene was formed in our system rather than a triplet nitrene. Although, a triplet nitrene pathway cannot be totally excluded.

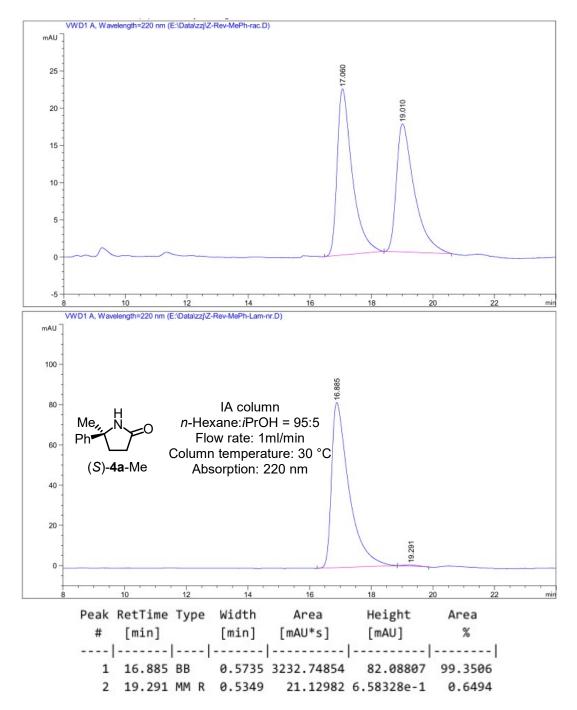


Figure 83. HPLC spectrums of *rac*-4a-Me and (S)-4a-Me obtained by using Λ -Ru10.

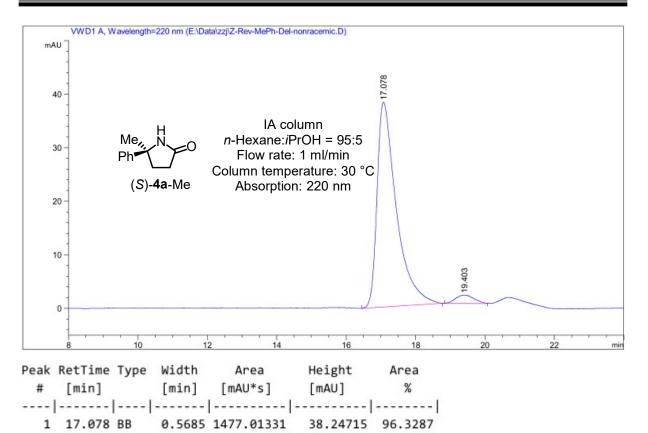


Figure 84. HPLC spectrum of (S)-4a-Me obtained by using Δ -Ru10.

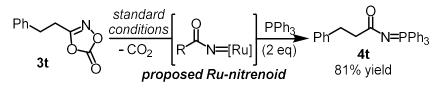
56.29272

0.6075

4) Trapping of Ru-nitrenoid Intermediate

19.403 MM R

2



1.54431

3.6713

Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrate **3t** (38.2 mg, 0.2 mmol), PPh₃ (52.4 mg, 0.4 mmol) and Λ -**Ru10** (0.2 mg, 0.0002 mmol, 0.1 mol%) under an atmosphere of N₂. Pre-cooled (using an ice water bath) "super dried" (Acros) 1,2-dichlorobenzene (0.4 mL, 0.5 M) from "Acros" was added via syringe in sequence. The reaction mixture was stirred at 4 °C for 30 h under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc:*n*-Hexane = 1:3 R_f value = 0.4-0.45) to afford the analytical pure products **4t**.

4t: 81% yield, color less oil. ¹**H NMR** (300 MHz, CD₃CN) δ 7.78-7.58 (m, 9H), 7.50 (ddd, *J* = 7.2, 5.4, 2.4 Hz, 6H), 7.31-7.13 (m, 5H), 3.01 (t, *J* = 7.4 Hz, 2H), 2.68 (td, *J* = 7.4, 1.5 Hz, 2H). ¹³**C NMR** (75

MHz, CD₃CN) δ 184.0, 183.8, 143.7, 133.9, 133.8, 133.3, 133.3, 130.2, 129.7, 129.6, 129.5, 129.1, 128.9, 126.6, 42.6, 42.3, 33.5, 33.4. ³¹**P** NMR (122 MHz, CD₃CN) δ 19.22. **HRMS (ESI**, *m/z*) calcd. for C₂₇H₂₄N₁Na₁O₁P₁ [M+Na]⁺: 432.1488, found: 432.1484.

4.3.5 Single Crystal X-Ray Diffraction Studies

Crystallography of compound (*S*)-**4e**: Single crystals of **4e** were obtained by recrystalization on EtOAc at 80 °C. Data was collected with an STOE STADIVARI diffractometer equipped with with CuK radiation, a graded multilayer mirror monochromator (1.54186 Å) and a DECTRIS PILATUS 300K detector using an oil-coated shock-cooled crystal at 100(2) K. Absorption effects were corrected semi-empirical using multiscanned reflections (STOE LANA, absorption correction by scaling of reflection intensities.). Cell constants were refined using 15678 of observed reflections of the data collection. The structure was solved by direct methods by using the program XT V2014/1 (Bruker AXS Inc., 2014) and refined by full matrix least squares procedures on F² using SHELXL-2018/3 (Sheldrick, 2018). The non–Hydrogen atoms have been refined anisotropically, carbon bonded hydrogen atoms were included at calculated positions and refined using the 'riding model' with isotropic temperature factors at 1.2 times (for CH₃ groups 1.5 times) that of the preceding carbon atom. CH₃ groups were allowed to rotate about the bond to their next atom to fit the electron density. Nitrogen or oxygen bonded hydrogen atoms were located and allowed to refine isotropically. The absolute configuration has been determined: The "Flack parameter" (Parsons, Flack, *et al.*, 2013) refined to -0.005(14). CCDC number: 1910815.

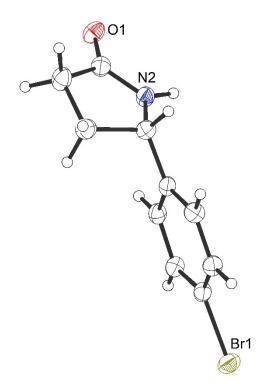


Figure 85. Crystal structure of (*S*)-4e.

Table 7. Crystal data and structure refinement for (S)-4e.

Crystal data

Identification code	(<i>S</i>)-4e	
Habitus, colour	plate, colourless	
Crystal size	0.18 x 0.17 x 0.05 mm ³	
Crystal system	Monoclinic	
Space group	P21	Z = 2
Unit cell dimensions	a = 4.8070(2) Å	= 90°.
	b = 7.0174(2) Å	= 96.901(3)°.
	c = 14.3984(6) Å	= 90°.
Volume	482.18(3) Å ³	
Cell determination	15678 peaks with Theta 3.1 to 76.1°.	
Empirical formula	C10 H10 Br N O	
Moiety formula	C10 H10 Br N O	
Formula weight	240.10	
Density (calculated)	1.654 Mg/m ³	

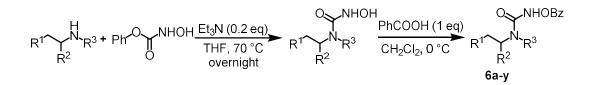
Absorption coefficient	5.468 mm ⁻¹
-	
F(000)	240
Data collection:	
Diffractometer type	STOE STADIVARI
Wavelength	1.54186 Å
Temperature	100(2) K
Theta range for data collection	3.092 to 75.455°.
Index ranges	-5<=h<=6, -6<=k<=8, -17<=l<=17
Data collection software	X-Area Pilatus3_SV 1.31.127.0 (STOE, 2016) ⁴
Cell refinement software	X-Area Recipe 1.33.0.0 (STOE, 2015) ⁵
Data reduction software	X-Area Integrate 1.71.0.0 (STOE, 2016) ⁶
	X-Area LANA 1.68.2.0 (STOE, 2016) ⁷
Solution and refinement:	
Reflections collected	8380
Independent reflections	1728 [R(int) = 0.0289]
Completeness to theta = 67.686°	100.0 %
Observed reflections	$1725[I > 2\sigma(I)]$
Reflections used for refinement	1728
Absorption correction	Semi-empirical from equivalents ⁸
Max. and min. transmission	0.0516 and 0.0138
Flack parameter (absolute struct.)	$-0.005(14)^9$
Largest diff. peak and hole	0.324 and -0.590 e.Å ⁻³
Solution	intrinsic phases ¹⁰
Refinement	Full-matrix least-squares on F ^{2¹¹}
Treatment of hydrogen atoms Programs used	CH calculated positions, constr., NH located, isotr. ref. XT V2014/1 (Bruker AXS Inc., 2014) ¹²
Tograms used	SHELXL-2018/3 (Sheldrick, 2018) ¹³
	DIAMOND (Crystal Impact) ¹⁴
	ShelXle (Hübschle, Sheldrick, Dittrich, 2011) ¹⁵
Data / restraints / parameters	1728 / 1 / 122
-	17207 17 122
Goodness-of-fit on F ²	1.098
R index (all data)	wR2 = 0.0625
R index conventional [I>2sigma(I)]	R1 = 0.0235

References

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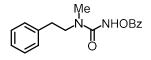
4.4 Enantioselective Synthesis of 2-Imidazolidinones by Intramolecular C-H Amidation

4.4.1 Synthesis of Substrates



General procedure: Amine (3.0 mmol, 1 equiv) phenyl hydroxycarbamate (3.0 mmol, 1 equiv) and Et_3N (0.6 mmol, 0.2 equiv) dissolved in THF (10 mL) were heated at 70 °C overnight (16-18 h, monitored by TLC) under N₂ atmosphere. After cooling to room temperature, the organic solvent was removed under reduced pressure. The resulting solid was dissolved in 20 mL EtOAc, and washed with brine (3 x 10 mL). The organic layer was collected and dried under anhydrous MgSO₄. The organic solvent was removed and the residue was subjected to flash silica gel chromatography to provide crude intermediate (EtOAc, R_f value = 0.3-0.4) as a white solid. The above intermediate (3.0 mmol, 1 equiv) together with benzoic acid (3.0 mmol, 1 equiv) were dissolved in 30 mL CH₂Cl₂. The resulting solution was cooled to 0 °C (ice-water bath). After that, DCC (3.0 mmol, 1 equiv) dissolved in 10 mL CH₂Cl₂ was added dropwisely to the solution. After stirring at 0 °C for 3 h, 20 mL Et₂O was added to the solution in one portion. The resulting solution was filtered and the filtrate was collected. The organic solvent was removed under reduced pressure. The residue was subjected to a flash silica gel chromatography to provide analytical pure *N*-benzoyloxyurea substrates.

Compound 6aa

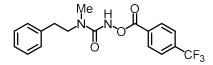


6aa, 74% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) 8.39 (s, 1H), 8.22-8.12 (m, 2H), 7.66 (t, *J* = 7.4 Hz, 1H), 7.52 (t, *J* = 7.7 Hz, 2H), 7.37 (dd, *J* = 10.2, 4.3 Hz, 2H), 7.32-7.23 (m, 3H), 3.62 (t, *J* = 7.4 Hz, 2H), 3.04-2.89 (m, 5H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.9, 157.7, 138.7, 134.1, 130.0, 128.9, 128.9, 128.7, 127.3, 126.7, 51.4, 34.8, 34.3. **HRMS (ESI,** *m/z*) calcd. for C₁₇H₁₈N₂O₃Na₁[M+Na]⁺: 321.1210, found: 321.1207.

Compound 6ab

6ab, 78% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.34 (s, 1H), 8.18-8.07 (m, 2H), 7.33 (qd, *J* = 7.6, 2.2 Hz, 5H), 7.05-6.93 (m, 2H), 3.93 (s, 3H), 3.67-3.57 (m, 2H), 3.02-2.91 (m, 5H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.7, 164.4, 157.9, 138.8, 132.2, 129.0, 128.9, 126.8, 119.5, 114.1, 55.7, 51.5, 34.9, 34.4. **HRMS (ESI**, *m/z*) calcd. for C₁₈H₂₀N₂O₄Na₁[M+Na]⁺: 351.1315, found: 351.1313.

Compound 6ac

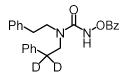


6ac, 75% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.14 (d, *J* = 8.0 Hz, 3H), 7.67 (d, *J* = 8.3 Hz, 2H), 7.35-7.21 (m, 2H), 7.17 (dd, *J* = 8.0, 4.6 Hz, 3H), 3.50 (t, *J* = 7.3 Hz, 2H), 2.90-2.79 (m, 5H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.7, 157.4, 138.6, 135.7, 135.3, 130.7, 130.5, 128.97, 128.95, 126.9, 125.87, 125.82, 125.8, 125.7, 125.4, 121.8, 51.5, 34.8, 34.3. **HRMS** (**ESI**, *m/z*) calcd. for C₁₈H₁₇F₃N₂O₃Na₁[M+Na]⁺: 389.1083, found: 389.1086.

Compound 6ad

6ad, 52% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.38 (s, 1H), 7.41-7.16 (m, 5H), 3.55-3.39 (m, 2H), 2.93-2.77 (m, 5H), 1.28 (s, 9H). ¹³**C NMR** (75 MHz, CD₃CN) δ 178.7, 158.5, 140.2, 129.8, 129.5, 127.3, 51.2, 38.8, 34.7, 34.5, 27.3. **HRMS (ESI,** *m/z*) calcd. for C₁₅H₂₂N₂O₃Na₁[M+Na]⁺: 301.1523, found: 301.1520.

Compound 6aa'



6aa', 57% yield, white solid. ¹H NMR (300 MHz, CD₃CN) δ 8.74 (s, 1H), 8.16-8.04 (m, 2H), 7.73 (t,

J = 7.4 Hz, 1H), 7.58 (t, J = 7.7 Hz, 2H), 7.41-7.17 (m, 10H), 3.41 (dd, J = 9.5, 5.5 Hz, 4H), 2.95-2.80 (m, 2H). ¹³C NMR (75 MHz, CD₃CN) δ 167.3, 158.2, 140.2, 140.1, 135.0, 130.4, 129.9, 129.9, 129.5, 128.8, 127.3, 55.3, 50.1, 50.0, 35.0. HRMS (ESI, *m/z*) calcd. for C₂₄H₂₂D₂N₂O₃Na₁[M+Na]⁺: 413.1805, found: 413.1805.

Compound (Z)-6b

(Z)-6b, 77% yield, white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.38 (s, 1H), 8.09-7.97 (m, 2H), 7.61-7.49 (m, 1H), 7.40 (t, J = 7.6 Hz, 2H), 7.34-7.09 (m, 5H), 6.52 (d, J = 11.6 Hz, 1H), 5.60 (dt, J = 11.6, 7.4 Hz, 1H), 3.38 (t, J = 7.2 Hz, 2H), 2.85 (s, 3H), 2.56 (qd, J = 7.4, 1.6 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 167.0, 157.8, 137.2, 134.1, 132.0, 130.1, 128.8, 128.5, 128.0, 127.3, 127.1, 49.0, 34.1, 26.9. HRMS (ESI, *m*/*z*) calcd. for C₁₉H₂₀N₂O₃Na₁[M+Na]⁺: 347.1366, found: 347.1363.

Compound (E)-6b

(*E*)-6b, 63% yield, white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.38 (s, 1H), 8.09-7.98 (m, 2H), 7.54 (t, J = 7.4 Hz, 1H), 7.40 (t, J = 7.7 Hz, 2H), 7.33-7.09 (m, 6H), 6.43 (d, J = 15.8 Hz, 1H), 6.20-6.04 (m, 1H), 3.42 (t, J = 7.2 Hz, 2H), 2.97 (s, 3H), 2.47 (q, J = 7.2 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 167.0, 157.8, 137.3, 134.1, 132.9, 130.1, 128.8, 128.7, 127.5, 127.3, 126.3, 49.3, 34.7, 31.6. HRMS
(ESI, *m/z*) calcd. for C₁₉H₂₀N₂O₃Na₁[M+Na]⁺: 347.1366, found: 347.1363.

Compound (S)-6c

Me NHOBz

6c, 64% yield, colorless oil, and with 89% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 9.8 min, t_r (minor) = 8.5 min). ¹H NMR (300 MHz, CDCl₃) δ 8.26 (s, 1H), 8.20-8.09 (m, 2H), 7.64 (t, *J* = 7.4 Hz, 1H), 7.50

(t, J = 7.6 Hz, 2H), 7.42-7.20 (m, 5H), 3.71 (dd, J = 13.7, 6.6 Hz, 1H), 3.25 (ddd, J = 21.6, 14.3, 8.0 Hz, 2H), 2.81 (s, 3H), 1.35 (d, J = 6.8 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 166.9, 158.0, 144.1, 134.1, 130.1, 128.9, 128.7, 127.4, 127.3, 127.0, 57.2, 38.8, 35.2, 18.7. HRMS (ESI, *m/z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1366, found: 335.1363.

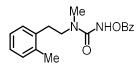
Compound 6d

6d, 72% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.27 (s, 1H), 8.11 (d, *J* = 7.9 Hz, 2H), 7.62 (t, *J* = 7.4 Hz, 1H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.21 (t, *J* = 7.7 Hz, 1H), 7.07 (t, *J* = 15.6 Hz, 3H), 3.56 (t, *J* = 7.4 Hz, 2H), 2.99-2.83 (m, 5H), 2.33 (d, *J* = 5.4 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.9, 157.7, 138.6, 134.1, 130.1, 129.8, 128.8, 128.8, 127.5, 127.3, 125.9, 51.5, 34.9, 34.3, 21.5. **HRMS** (**ESI**, *m/z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1366, found: 335.1363.

Compound 6e

6e, 68% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.28 (s, 1H), 8.11 (dd, *J* = 8.3, 1.2 Hz, 2H), 7.61 (dd, *J* = 10.6, 4.3 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 2H), 7.25-7.00 (m, 4H), 3.65-3.44 (m, 2H), 2.99-2.81 (m, 5H), 2.33 (d, *J* = 5.5 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.9, 157.7, 136.3, 135.6, 134.1, 130.1, 129.6, 128.8, 128.7, 127.3, 77.6, 77.2, 76.7, 51.5, 34.8, 33.9, 21.2. **HRMS (ESI,** *m/z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1366, found: 335.1363.

Compound 6f

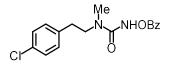


6f, 71% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.37 (s, 1H), 8.12 (dd, *J* = 8.4, 1.3 Hz, 2H), 7.65-7.58 (m, 1H), 7.46 (dd, *J* = 10.6, 4.7 Hz, 2H), 7.16 (s, 4H), 3.52 (dd, *J* = 8.4, 6.8 Hz, 2H), 3.04-2.85 (m, 5H), 2.38 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.8, 157.7, 136.8, 136.3, 134.1, 130.6, 130.0, 129.7, 128.7, 127.3, 126.9, 126.5, 50.3, 34.8, 31.6, 19.4. **HRMS (ESI,** *m/z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1366, found: 335.1363.

Compound 6g

6g, 72% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.71 (s, 1H), 8.07 (dd, *J* = 8.3, 1.2 Hz, 2H), 7.78-7.67 (m, 1H), 7.57 (t, *J* = 7.7 Hz, 2H), 7.37-7.24 (m, 2H), 7.13-7.00 (m, 2H), 3.61-3.41 (m, 2H), 2.98-2.79 (m, 5H). ¹³**C NMR** (75 MHz, CD₃CN) δ 167.2, 164.1, 160.9, 158.5, 136.2, 136.2, 135.0, 131.6, 131.5, 130.4, 129.9, 128.7, 116.2, 115.9, 51.2, 34.8, 33.6. **HRMS (ESI,** *m/z*) calcd. for C₁₇H₁₇F₁N₂O₃Na₁[M+Na]⁺: 339.1115, found: 339.1113.

Compound 6h



6h, 73% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.28 (s, 1H), 8.11-7.95 (m, 2H), 7.64-7.49 (m, 1H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.29-7.16 (m, 2H), 7.09 (d, *J* = 8.4 Hz, 2H), 3.47 (t, *J* = 7.3 Hz, 2H), 2.90-2.76 (m, 5H). ¹³**C NMR** (75 MHz, CDCl₃) δ 167.0, 157.7, 137.2, 134.2, 132.6, 130.3, 130.1, 129.0, 128.8, 127.2, 51.3, 34.9, 33.6. **HRMS (ESI,** *m/z*) calcd. for C₁₇H₁₇Cl₁N₂O₃Na₁[M+Na]⁺: 355.0820, found: 355.0818.

Compound 6i

6i, 70% yield, white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.26 (s, 1H), 8.15-8.07 (m, 2H), 7.61 (dd, J = 10.6, 4.3 Hz, 1H), 7.47 (t, J = 7.7 Hz, 2H), 7.14 (t, J = 5.7 Hz, 2H), 6.87 (dd, J = 6.7, 4.7 Hz, 2H), 3.79 (s, 3H), 3.53 (t, J = 7.3 Hz, 2H), 2.91 (s, 3H), 2.86 (t, J = 7.3 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 166.9, 158.6, 157.7, 134.1, 130.7, 130.1, 129.9, 128.7, 127.3, 114.4, 55.4, 51.2, 34.9, 33.4. HRMS
(ESI, *m/z*) calcd. for C₁₈H₂₀N₂O₄Na₁[M+Na]⁺: 351.1315, found: 351.1313.

Compound 6j

6j, 63% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.39 (s, 1H), 8.20-8.10 (m, 3H), 7.87 (d, *J* = 7.8 Hz, 1H), 7.76 (d, *J* = 7.5 Hz, 1H), 7.69-7.35 (m, 7H), 3.69 (dd, *J* = 8.5, 6.5 Hz, 2H), 3.46-3.34 (m, 2H), 2.90 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 167.0, 157.7, 134.9, 134.2, 134.1, 132.0, 130.1, 129.0, 128.8, 127.6, 127.3, 127.2, 126.6, 125.9, 125.8, 123.6, 51.1, 35.1, 31.5. **HRMS (ESI,** *m/z***)** calcd. for C₂₁H₂₀N₂O₃Na₁[M+Na]⁺: 371.1366, found: 371.1367.

Compound 6k

6k, 71% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.38 (s, 1H), 8.15-8.04 (m, 2H), 7.81 (dd, *J* = 4.6, 3.3 Hz, 3H), 7.69 (s, 1H), 7.62 (t, *J* = 7.5 Hz, 1H), 7.54-7.41 (m, 4H), 7.38 (dd, *J* = 8.4, 1.5 Hz, 1H), 3.66 (t, *J* = 7.4 Hz, 2H), 3.08 (t, *J* = 7.4 Hz, 2H), 2.91 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.9, 157.8, 136.2, 134.1, 133.8, 132.4, 130.1, 128.7, 128.5, 127.8, 127.7, 127.5, 127.3, 126.3, 125.7, 51.4, 35.0, 34.5. **HRMS (ESI,** *m/z***)** calcd. for C₂₁H₂₀N₂O₃Na₁[M+Na]⁺: 371.1366, found: 371.1364.

Compound 61

61, 69% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.25 (s, 1H), 8.01 (dd, *J* = 8.2, 1.0 Hz, 2H), 7.60-7.47 (m, 1H), 7.37 (t, *J* = 7.7 Hz, 2H), 7.07 (dd, *J* = 5.1, 1.1 Hz, 1H), 6.85 (dd, *J* = 5.1, 3.5 Hz, 1H), 6.78 (d, *J* = 3.3 Hz, 1H), 3.51 (t, *J* = 7.1 Hz, 2H), 3.04 (t, *J* = 7.1 Hz, 2H), 2.85 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.9, 157.8, 140.7, 134.1, 130.1, 128.8, 127.4, 127.3, 125.8, 124.3, 51.5, 34.9, 28.3. **HRMS (ESI**, *m/z*) calcd. for C₁₅H₁₆N₂O₃S₁Na₁[M+Na]⁺: 327.0774, found: 327.0771. **Compound 6m**

6m, 73% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.83 (s, 1H), 8.18-8.02 (m, 2H), 7.72 (t, *J* = 7.5 Hz, 1H), 7.57 (t, *J* = 7.6 Hz, 2H), 7.21 (ddd, *J* = 8.8, 7.0, 4.2 Hz, 4H), 5.14-4.99 (m, 1H), 3.19 (dd, *J* = 16.3, 8.4 Hz, 2H), 3.05 (dd, *J* = 16.3, 7.0 Hz, 2H), 2.81 (s, 3H). ¹³**C NMR** (75 MHz, CD₃CN) δ 167.2, 158.7, 142.2, 135.0, 130.4, 129.9, 128.8, 127.6, 125.3, 56.3, 36.7, 29.2. **HRMS (ESI,** *m/z*) calcd. for C₁₈H₁₈N₂O₃Na₁[M+Na]⁺: 333.1210, found: 333.1207.

Compound 6n

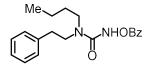
6n, 58% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 8.16-8.05 (m, 2H), 8.00 (s, 1H), 7.63 (dd, *J* = 10.6, 4.3 Hz, 1H), 7.48 (t, *J* = 7.6 Hz, 2H), 7.39-7.19 (m, 10H), 4.51 (s, 1H), 3.21-2.86 (m, 4H), 2.75 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.5, 158.0, 138.4, 134.0, 130.0, 129.0, 128.9, 128.7, 127.4, 126.8, 38.5. **HRMS (ESI**, *m/z*) calcd. for C₂₄H₂₄N₂O₃Na₁[M+Na]⁺: 411.1679, found: 411.1690.

Compound 60

60, 75% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.59 (s, 1H), 8.06 (dd, *J* = 8.1, 1.0 Hz, 2H), 7.57 (ddd, *J* = 6.9, 2.5, 1.2 Hz, 1H), 7.47-7.33 (m, 4H), 7.28-7.20 (m, 3H), 3.57 (t, *J* = 6.7 Hz, 2H), 3.09 (s, 3H), 2.70 (t, *J* = 6.7 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.7, 157.8, 134.1, 131.8, 130.0, 128.7, 128.4, 128.1, 127.3, 123.4, 86.8, 82.7, 48.6, 35.1, 19.1. **HRMS (ESI,** *m/z*) calcd. for C₁₉H₁₈N₂O₃Na₁[M+Na]⁺: 345.1221, found: 345.1210. **Compound 6p**

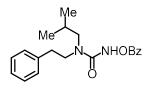
6p, 77% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.29 (s, 1H), 8.04 (dd, *J* = 5.2, 3.3 Hz, 2H), 7.62-7.49 (m, 1H), 7.39 (dd, *J* = 10.6, 4.7 Hz, 2H), 7.31-7.21 (m, 2H), 7.20-7.10 (m, 3H), 3.53-3.38 (m, 2H), 3.20 (q, *J* = 7.2 Hz, 2H), 2.93-2.81 (m, 2H), 1.12 (t, *J* = 7.2 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 167.1, 157.4, 138.8, 134.1, 130.1, 129.0, 128.9, 128.7, 127.3, 126.8, 49.2, 42.6, 34.9, 13.4. **HRMS (ESI,** *m*/*z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1377, found: 335.1363.

Compound 6q



6q, 71% yield, colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 8.30 (s, 1H), 8.12-7.99 (m, 2H), 7.53 (dd, *J* = 10.6, 4.3 Hz, 1H), 7.40 (t, *J* = 7.6 Hz, 2H), 7.31-7.10 (m, 5H), 3.53-3.38 (m, 2H), 3.19-3.05 (m, 2H), 2.96-2.80 (m, 2H), 1.51 (dt, *J* = 12.2, 7.2 Hz, 2H), 1.27 (dt, *J* = 15.0, 7.3 Hz, 2H), 0.86 (t, *J* = 7.3 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 167.1, 157.6, 138.8, 134.1, 130.1, 129.0, 128.9, 128.7, 127.3, 126.8, 49.6, 47.7, 34.7, 30.4, 20.2, 13.9. **HRMS (ESI,** *m/z*) calcd. for C₂₀H₂₄N₂O₃Na₁[M+Na]⁺: 363.1690, found: 363.1680.

Compound 6r



6r, 62% yield, colorless oil. ¹**H** NMR (300 MHz, CDCl₃) δ 8.50 (s, 1H), 8.16 (dd, *J* = 8.4, 1.3 Hz, 2H), 7.65 (ddd, *J* = 6.8, 2.5, 1.3 Hz, 1H), 7.50 (t, *J* = 7.7 Hz, 2H), 7.44-7.22 (m, 5H), 3.64-3.48 (m, 2H), 3.07 (d, *J* = 7.6 Hz, 2H), 3.02-2.92 (m, 2H), 2.13-1.95 (m, 1H), 0.98 (d, *J* = 6.7 Hz, 6H). ¹³**C** NMR (75 MHz, CDCl₃) δ 167.1, 157.8, 138.8, 134.1, 130.1, 128.9, 128.9, 128.7, 127.3, 126.7, 55.2, 50.1, 34.4, 27.7, 20.3. **HRMS (ESI**, *m/z*) calcd. for C₂₀H₂₄N₂O₃Na₁[M+Na]⁺: 363.1690, found: 363.1679. **Compound 6s**

6s, 61% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.14 (dd, *J* = 10.0, 2.9 Hz, 3H), 7.65 (t, *J* = 7.5 Hz, 1H), 7.51 (t, *J* = 7.7 Hz, 2H), 7.35 (t, *J* = 7.1 Hz, 4H), 7.30-7.18 (m, 7H), 3.47 (t, *J* = 7.4 Hz, 4H), 2.92 (t, *J* = 7.4 Hz, 4H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.8, 157.6, 138.7, 134.1, 130.1, 128.9, 128.7, 127.3, 126.8, 50.1, 34.7. **HRMS (ESI,** *m/z*) calcd. for C₂₀H₂₄N₂O₃Na₁[M+Na]⁺: 363.1690, found: 363.1680. **HRMS (ESI,** *m/z*) calcd. for C₂₄H₂₄N₂O₃Na₁[M+Na]⁺: 411.1690, found: 411.1680.

Compound 6t

6t, 68% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.81 (s, 1H), 8.08 (dd, *J* = 5.2, 3.3 Hz, 2H), 7.80-7.65 (m, 1H), 7.57 (dd, *J* = 10.6, 4.7 Hz, 2H), 7.44-7.20 (m, 10H), 4.46 (s, 2H), 3.58-3.41 (m, 2H), 3.01-2.83 (m, 2H). ¹³**C NMR** (75 MHz, CD₃CN) δ 167.3, 158.7, 140.0, 138.6, 135.0, 130.4, 129.9, 129.8, 129.6, 129.5, 128.7, 128.4, 127.4, 51.0, 49.3, 34.8. **HRMS (ESI,** *m/z*) calcd. for C₂₃H₂₂N₂O₃Na₁[M+Na]⁺: 397.1534, found: 397.1522.

Compound 6u

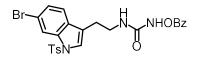
6u, 82% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.56 (s, 1H), 8.01-7.90 (m, 2H), 7.66 (ddd, *J* = 7.0, 2.5, 1.3 Hz, 1H), 7.57-7.44 (m, 2H), 7.25-7.08 (m, 5H), 5.39 (s, 1H), 3.56 (dd, J = 12.8, 6.6 Hz, 2H), 2.84 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 164.9, 158.3, 138.6, 134.5, 129.8, 129.0, 128.9, 128.8, 126.8, 126.7, 41.1, 35.9. **HRMS (ESI,** *m/z*) calcd. for C₁₆H₁₆N₂O₃Na₁[M+Na]⁺: 307.1053, found: 307.1050. **Compound 6v**

6v, 57% yield, colorless oil. ¹H NMR (300 MHz, CDCl₃) δ 8.15-8.08 (m, 2H), 8.02 (s, 1H), 7.63 (t, J = 7.4 Hz, 1H), 7.55-7.39 (m, 5H), 7.37-7.17 (m, 7H), 4.04-3.89 (m, 2H), 2.98 (dd, J = 8.9, 6.9 Hz, 2H).
¹³C NMR (75 MHz, CDCl₃) δ 166.9, 156.8, 140.1, 138.6, 134.1, 130.4, 130.1, 129.0, 128.8, 128.7, 128.6, 128.3, 127.2, 126.5, 52.2, 34.5.

Compound 6w

6w, 48% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.40 (s, 1H), 8.17-8.01 (m, 2H), 7.93 (d, J = 8.1 Hz, 1H), 7.78 (d, J = 8.4 Hz, 2H), 7.71 (t, J = 7.5 Hz, 1H), 7.64-7.46 (m, 4H), 7.41-7.20 (m, 4H), 6.16 (s, 1H), 3.49 (q, J = 6.6 Hz, 2H), 2.89 (t, J = 6.7 Hz, 2H), 2.32 (s, 3H). ¹³C NMR (75 MHz, CD₃CN) δ 166.3, 159.4, 146.5, 135.9, 135.6, 135.0, 132.1, 130.9, 130.6, 129.7, 128.3, 127.6, 125.6, 124.9, 124.2, 121.4, 120.6, 114.4, 39.8, 25.8, 21.4. **HRMS (ESI,** *m/z*) calcd. for C₂₅H₂₃N₃O₅S₁Na₁[M+Na]⁺: 500.1251, found: 500.1252.

Compound 6x



6x, 52% yield, white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.47 (s, 1H), 8.04 (d, J = 1.2 Hz, 1H), 7.89-7.79 (m, 2H), 7.65 (d, J = 8.4 Hz, 2H), 7.58 (t, J = 7.5 Hz, 1H), 7.41 (t, J = 7.7 Hz, 2H), 7.25 (dd, J = 13.8, 5.9 Hz, 3H), 7.21-7.11 (m, 3H), 5.38 (t, J = 5.7 Hz, 1H), 3.50 (q, J = 6.7 Hz, 2H), 2.83 (t, J = 6.8 Hz, 2H), 2.26 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 164.9, 158.3, 145.5, 136.0, 135.1, 134.6, 130.2, 129.7, 129.6, 129.1, 126.9, 126.8, 126.5, 124.1, 120.6, 119.4, 118.8, 117.0, 39.9, 25.4, 21.7. **HRMS (ESI**, *m/z*) calcd. for C₂₅H₂₂N₃O₅S₁Br₁Na₁[M+Na]⁺: 578.0356, 580.0338, found: 578.0357, 580.0337.

Compound 6y

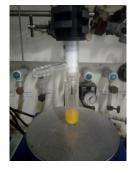
6y, 63% yield, white solid. ¹**H NMR** (300 MHz, CD₃CN) δ 8.72 (s, 1H), 8.07 (d, *J* = 7.3 Hz, 2H), 7.72 (t, *J* = 7.4 Hz, 1H), 7.56 (t, *J* = 7.7 Hz, 2H), 7.37-7.16 (m, 5H), 3.41-3.24 (m, 2H), 2.94 (s, 3H), 2.74-2.57 (m, 2H), 1.88 (dd, *J* = 15.2, 7.5 Hz, 2H). ¹³**C NMR** (75 MHz, CD₃CN) δ 167.2, 158.5, 142.9, 134.8, 130.3, 129.8, 129.3, 129.2, 128.7, 126.7, 49.0, 34.1, 33.4, 30.0. **HRMS (ESI**, *m/z*) calcd. for C₁₈H₂₀N₂O₃Na₁[M+Na]⁺: 335.1377, found: 335.1364.

4.4.2 Ruthenium Catalyzed Intramolecular C-H Amidations

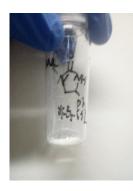
General procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrates **6a-y** (0.2 mmol), chiral ruthenium catalyst (0.002 mmol, 1 mol%) and K_2CO_3 (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (2.0 mL) was added via syringe. The reaction mixture was stirred at the indicated temperature for the indicated time under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc/*n*–*H*exane = 2:1 to EtOAc/MeOH 95:5) to afford the analytical pure products **7a-y**. Enantiomeric ratios were determined by HPLC analysis on chiral stationary phase. The absolute configuration of the product **7h** was confirmed by X-ray analysis (CCDC number: 1972573) as *S*-configuration. Racemic samples were obtained by the same catalytic reactions using racemic ruthenium catalyst instead of chiral ruthenium catalyst.

Additional experimental informations:

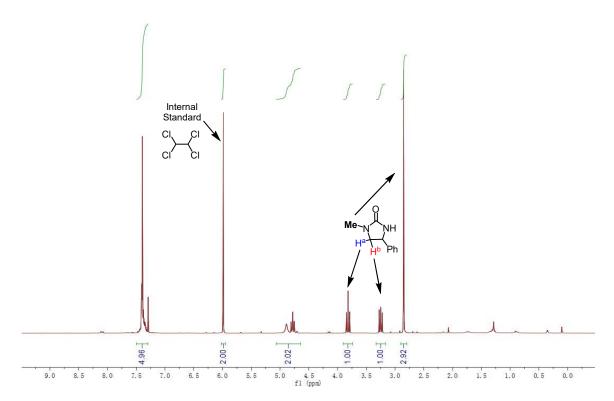
a) Execution of the reactions. All catalytic reactions were carried out in 10 mL Schlenk tubes from "Synthware" under N₂ atmosphere (see image below).



b) Isolation of the final product. The C-H amination products can be isolated by flash chromatography without any trace contaminations from the ruthenium catalyst as evident by the white color of the products (see image below).



c) Crude ¹H NMR spectra analysis of the standard reaction solution. See image below.



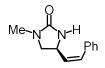
The above crude ¹H NMR analysis shows quantitative yield of the target cyclic urea product **7a** (**Table 3**, entry 5).

Compound (S)-7a



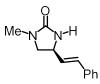
Starting from **6aa** (59.6 mg, 0.20 mmol) according to the general procedure to provide **7a** as a white solid (34.9 mg, 99% yield) and with 95% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 13.2 min, t_r (minor) = 11.4 min). [α] $_{D}^{22}$ = 10.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₃CN) δ 7.46-7.28 (m, 5H), 5.54 (s, 1H), 4.82-4.63 (m, 1H), 3.77 (t, *J* = 8.8 Hz, 1H), 3.12 (dd, *J* = 8.7, 7.5 Hz, 1H), 2.71 (s, 3H). ¹³C NMR (75 MHz, CD₃CN) δ 163.2, 143.6, 129.6, 128.7, 127.1, 56.5, 54.1, 30.8. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₂N₂O₁Na₁[M+Na]⁺: 199.0842, found: 199.0842.

Compound (S,Z)-7b



Starting from (*Z*)-**6b** (62.8 mg, 0.20 mmol) according to the general procedure to provide (*S*,*Z*)-**7b** as a white solid (29.5 mg, 73% yield) and with 91% ee as determined by HPLC analysis (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 15.5 min, t_r (minor) = 14.6 min). [α] $_{D}^{22}$ = 47.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₃CN) δ 7.44-7.36 (m, 2H), 7.35-7.30 (m, 1H), 7.30-7.23 (m, 2H), 6.65 (d, *J* = 11.6 Hz, 1H), 5.72 (dd, *J* = 11.5, 9.3 Hz, 1H), 5.14 (s, 1H), 4.57 (dd, *J* = 16.4, 8.7 Hz, 1H), 3.63 (t, *J* = 8.6 Hz, 1H), 3.14 (dd, *J* = 8.7, 7.4 Hz, 1H), 2.69 (s, 3H). ¹³C NMR (75 MHz, CD₃CN) δ 162.9, 137.2, 133.3, 132.4, 129.6, 129.4, 128.4, 54.3, 48.1, 30.8. HRMS (ESI, *m*/*z*) calcd. for C₁₂H₁₄N₂O₁Na₁[M+Na]⁺: 225.0998, found: 225.0997.

Compound (S,E)-7b

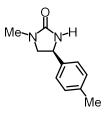


Starting from (*E*)-**6b** (62.8 mg, 0.20 mmol) according to the general procedure to provide (*S*,*E*)-**7b** as a white solid (32.2 mg, 80% yield) and with 76% ee as determined by HPLC analysis (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 15.5 min, t_r (minor) = 17.8 min). [α]_D²² = -15.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₃CN) δ 7.49-7.42 (m, 2H), 7.41-7.24 (m, 3H), 6.61 (d, *J* = 15.9 Hz, 1H), 6.28 (dd, *J* = 15.9, 7.3 Hz, 1H), 5.15 (s, 1H), 4.29 (q, *J* = 7.3 Hz, 1H), 3.60 (t, *J* = 8.7 Hz, 1H), 3.14 (dd, *J* = 8.8, 7.1 Hz, 1H), 2.70 (s, 3H). ¹³C NMR (75 MHz, CD₃CN) δ 162. 9, 137.6, 132.0, 130.7, 129.7, 128.8, 127.4, 54.0, 52.8, 30.8. HRMS (ESI, *m*/*z*) calcd. for C₁₂H₁₄N₂O₁Na₁[M+Na]⁺: 225.0998, found: 225.0998.

Compound (R)-7c

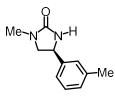
Starting from (*S*)-**6c** (62.4 mg, 0.20 mmol) according to the general procedure to provide (*R*)-**7c** as a white solid (33.7 mg, 89% yield) and with 85% ee (Δ -**Ru5** was applied), enantioselectivity was determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 215$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 11.6 min, t_r (minor) = 8.9 min). ¹H NMR (300 MHz, CDCl₃) δ 7.43-7.32 (m, 4H), 7.29 (dd, *J* = 5.7, 2.9 Hz, 1H), 4.83 (s, 1H), 3.45 (dd, *J* = 20.1, 8.6 Hz, 2H), 2.80 (s, 3H), 1.68 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.3, 145.9, 128.9, 127.5, 124.9, 62.1, 57.1, 30.6, 28.7. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₄N₂O₁Na₁[M+Na]⁺: 213.0998, found: 213.0999.

Compound (S)-7d



Starting from **6d** (62.4 mg, 0.20 mmol) according to the general procedure to provide **7d** as a white solid (35.0 mg, 92% yield) and with 95% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 95:5, flow rate: 1.0 mL/min, column temperature: 40 °C, retention times: t_r (major) = 21.1 min, t_r (minor) = 16.9 min). [α]_D²² = 18.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.26-6.97 (m, 4H), 4.76-4.47 (m, 2H), 3.70 (t, *J* = 8.9 Hz, 1H), 3.14 (dd, *J* = 8.6, 7.5 Hz, 1H), 2.75 (s, 3H), 2.29 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.3, 141.7, 138.9, 129.7, 129.1, 129.0, 126.8, 126.2, 123.3, 56.2, 53.8, 30.7, 21.5. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₄N₂O₁Na₁[M+Na]⁺: 213.0998, found: 213.0999.

Compound (S)-7e



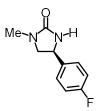
Starting from **6e** (62.4 mg, 0.20 mmol) according to the general procedure to provide **7e** as a white solid (37.7 mg, 99% yield) and with 96% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 11.4 min, t_r (minor) = 12.6 min). [α] $_{D}^{22}$ = 5.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CD₃CN) δ 7.27 (d, *J* = 8.1 Hz, 2H), 7.23-7.12 (m, 2H), 5.43 (s, 1H), 4.67 (t, *J* = 8.1 Hz, 1H), 3.74 (dd, *J* = 10.9, 6.5 Hz, 1H), 3.10 (dd, *J* = 8.7, 7.6 Hz, 1H), 2.71 (s, 3H), 2.35 (s, 3H). ¹³C NMR (75 MHz, CD₃CN) δ 163.1, 140.5, 138.5, 130.2, 127.1, 56.6, 53.9, 30.8, 21.1. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₄N₂O₁Na₁[M+Na]⁺: 213.0998, found: 213.0999.

Compound (S)-7f



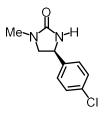
Starting from **6f** (62.4 mg, 0.20 mmol) according to the general procedure to provide **7f** as a white solid (36.6 mg, 96% yield) and with 97% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 85:15, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 8.8 min, t_r (minor) = 7.6 min). [α] $_{p^{22}} = 151.0^{\circ}$ (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.50-7.37 (m, 1H), 7.24 -7.04 (m, 3H), 4.92 (dd, *J* = 8.8, 7.4 Hz, 2H), 3.78 (t, *J* = 8.8 Hz, 1H), 3.05 (dd, *J* = 8.5, 7.3 Hz, 1H), 2.72 (s, 3H), 2.24 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.4, 139.7, 134.7, 130.8, 127.8, 126.8, 125.1, 54.9, 50.5, 30.6, 19.1. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₄N₂O₁Na₁[M+Na]⁺: 213.0998, found: 213.0998.

Compound (S)-7g



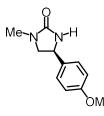
Starting from **6g** (63.2 mg, 0.20 mmol) according to the general procedure to provide **7g** as a white solid (34.8 mg, 90% yield) and with 93% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 40 °C, retention times: t_r (major) = 8.9 min, t_r (minor) = 9.8 min). [α]_D²² = 63.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.39-7.28 (m, 2H), 7.12-6.95 (m, 2H), 5.36 (s, 1H), 4.73 (t, *J* = 8.1 Hz, 1H), 3.75 (t, *J* = 8.8 Hz, 1H), 3.16 (dd, *J* = 8.7, 7.4 Hz, 1H), 2.78 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 164.3, 162.4, 161.0, 137.5, 137.5, 127.9, 127.8, 116.0, 115.7, 56.2, 53.2, 30.6. ¹⁹F NMR (235 MHz, CDCl₃) δ -114.16. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₁F₁N₂O₁Na₁[M+Na]⁺: 217.0748, found: 217.0748.

Compound (S)-7h



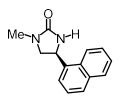
Starting from **6h** (66.4 mg, 0.20 mmol) according to the general procedure to provide **7h** as a white solid (39.1 mg, 93% yield) and with 90% ee as determined by HPLC analysis (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 85:15, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 12.1 min, t_r (minor) = 14.1 min). [α]_D²² = 23.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.31-7.17 (m, 4H), 5.22 (s, 1H), 4.72-4.59 (m, 1H), 3.70 (t, *J* = 8.9 Hz, 1H), 3.09 (dd, *J* = 8.5, 7.5 Hz, 1H), 2.72 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.4, 140.3, 134.1, 129.2, 127.6, 56.1, 53.2, 30.6. HRMS (ESI, *m/z*) calcd. for C₁₀H₁₁Cl₁N₂O₁Na₁[M+Na]⁺: 233.0452, found: 233.0451.

Compound (S)-7i



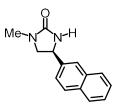
Starting from **6i** (65.6 mg, 0.20 mmol) according to the general procedure to provide **7i** as a white solid (41.0 mg, 99% yield) and with 92% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 19.4 min, t_r (minor) = 16.9 min). [α]_D²² = 56.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.34-7.18 (m, 2H), 6.88 (d, *J* = 8.7 Hz, 2H), 4.97 (s, 1H), 4.68 (t, *J* = 8.1 Hz, 1H), 3.79 (s, 3H), 3.73 (t, *J* = 8.8 Hz, 1H), 3.17 (dd, *J* = 8.6, 7.6 Hz, 1H), 2.79 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.3, 159.6, 133.7, 127.4, 114.4, 56.4, 55.4, 53.3, 30.7. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₄N₂O₂Na₁[M+Na]⁺: 229.0947, found: 229.0948.

Compound (S)-7j



Starting from **6j** (69.6 mg, 0.20 mmol) according to the general procedure to provide **7j** as a white solid (45.0 mg, 99% yield) and with 99% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 85:15, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 10.9 min, t_r (minor) = 9.5 min). [α]_D²² = 106.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.90 (dd, *J* = 11.5, 6.2 Hz, 2H), 7.82 (d, *J* = 8.2 Hz, 1H), 7.70 (d, *J* = 7.1 Hz, 1H), 7.60-7.44 (m, 3H), 5.53 (t, *J* = 7.9 Hz, 1H), 5.04 (s, 1H), 4.07 (t, *J* = 9.0 Hz, 1H), 3.26 (dd, *J* = 8.7, 6.9 Hz, 1H), 2.82 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.2, 137.1, 134.1, 130.4, 129.4, 128.5, 126.7, 126.0, 125.8, 122.6, 122.3, 55.3, 50.4, 30.6. HRMS (ESI, *m*/*z*) calcd. for C₁₄H₁₄N₂O₁Na₁[M+Na]⁺: 249.0998, found: 249.0997.

Compound (S)-7k



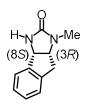
Starting from **6k** (69.6 mg, 0.20 mmol) according to the general procedure to provide **7k** as a white solid (43.9 mg, 97% yield) and with 90% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 19.4 min, t_r (minor) = 31.8 min). [α]_D²² = 97.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.91-7.74 (m, 4H), 7.57-7.38 (m, 3H), 5.35 (s, 1H), 4.89 (t, *J* = 8.1 Hz, 1H), 3.82 (t, *J* = 8.9 Hz, 1H), 3.27 (dd, *J* = 8.7, 7.2 Hz, 1H), 2.81 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.4, 139.1, 133.4, 133.3, 129.0, 128.0, 127.8, 126.6, 126.3, 125.1, 123.9, 56.0, 53.8, 30.7. HRMS (ESI, *m/z*) calcd. for C₁₄H₁₄N₂O₁Na₁[M+Na]⁺: 249.0998, found: 249.0999.

Compound (S)-7l



Starting from **61** (60.8 mg, 0.20 mmol) according to the general procedure to provide **71** as a white solid (36.1 mg, 99% yield) and with 88% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 16.0 min, t_r (minor) = 13.3 min). [α] $_{p^{22}} = 8.6^{\circ}$ (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.29 (dd, *J* = 4.8, 1.1 Hz, 1H), 7.05 (d, *J* = 2.9 Hz, 1H), 6.99 (dd, *J* = 5.0, 3.6 Hz, 1H), 5.05 (dd, *J* = 15.4, 7.1 Hz, 2H), 3.80 (t, *J* = 8.7 Hz, 1H), 3.36 (dd, *J* = 8.8, 7.1 Hz, 1H), 2.85 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.6, 145.5, 127.1, 125.4, 124.8, 56.4, 49.9, 30.6. HRMS (ESI, *m/z*) calcd. for C₈H₁₀S₁N₂O₁Na₁[M+Na]⁺: 245.0406, found: 245.0406.

Compound (3R,8S)-7m



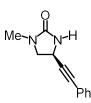
Starting from **6m** (62.0 mg, 0.20 mmol) according to the general procedure to provide **7m** as a white solid (10.9 mg, 29% yield) and with 89% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 26.2 min, t_r (minor) = 16.2 min). [α]_D²² = -131.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.19 (s, 5H), 5.03 (s, 1H), 4.94 (d, *J* = 7.8 Hz, 1H), 4.31 (ddd, *J* = 7.8, 5.1, 2.8 Hz, 1H), 3.18-3.01 (m, 2H), 2.75 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.2, 142.1, 140.5, 128.9, 127.7, 125.7, 124.5, 62.3, 58.8, 36.6, 28.8. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₂N₂O₁Na₁[M+Na]⁺: 211.0842, found: 211.0842.

Compound (4S,5S)-7n



Starting from **6n** (77.6 mg, 0.20 mmol) according to the general procedure to provide **7n** as a white solid (49.3 mg, 93% yield) and with 94% ee as determined by HPLC analysis (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 85:15, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 11.1 min, t_r (minor) = 9.7 min). [α] $_{p^{22}} = -8.2^{\circ}$ (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.30-7.14 (m, 6H), 7.14 -7.05 (m, 2H), 7.02-6.90 (m, 2H), 4.76 (s, 1H), 4.27 (d, *J* = 5.4 Hz, 1H), 3.54 (dt, *J* = 7.4, 5.3 Hz, 1H), 3.01 (dd, *J* = 13.9, 5.1 Hz, 1H), 2.90-2.68 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 161.3, 141.9, 136.6, 129.8, 128.9, 128.8, 128.1, 127.1, 126.2, 68.4, 58.0, 38.3, 29.3. HRMS (ESI, *m/z*) calcd. for C₁₇H₁₈N₂O₁Na₁ [M+Na]⁺: 289.1311, found: 289.1318.

Compound (S)-70



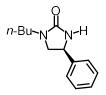
Starting from **60** (64.4 mg, 0.20 mmol) according to the general procedure to provide **70** as a white solid (35.7 mg, 89% yield) and with 87% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 10.8 min, t_r (minor) = 13.3 min). [α] $_{D}^{22}$ = 11.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.40-7.29 (m, 2H), 7.29-7.13 (m, 3H), 5.17 (s, 1H), 4.52 (dd, *J* = 8.6, 5.8 Hz, 1H), 3.61 (t, *J* = 8.6 Hz, 1H), 3.42 (dd, *J* = 8.6, 5.8 Hz, 1H), 2.74 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.6, 131.8, 128.8, 128.4, 122.2, 87.6, 83.9, 53.8, 41.5, 30.6. HRMS (ESI, *m/z*) calcd. for C₁₂H₁₃N₂O₁ [M+H]⁺: 201.1022, found: 201.1022.

Compound (S)-7p



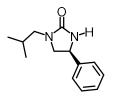
Starting from **6p** (62.4 mg, 0.20 mmol) according to the general procedure to provide **7p** as a white solid (37.9 mg, 99% yield) and with 93% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 12.1 min, t_r (minor) = 11.2 min). [α] $_{D}^{22}$ = 79.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.47-7.28 (m, 5H), 4.93-4.67 (m, 2H), 3.79 (t, *J* = 8.8 Hz, 1H), 3.47-3.14 (m, 3H), 1.11 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.8, 141.8, 129.1, 128.3, 126.2, 54.0, 53.2, 38.1, 12.8. HRMS (ESI, *m/z*) calcd. for C₁₁H₁₅N₂O₁ [M+H]⁺: 191.1179, found: 191.1178.

Compound (S)-7q



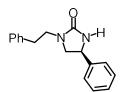
Starting from **6q** (68.0 mg, 0.20 mmol) according to the general procedure to provide **7q** as a white solid (35.7 mg, 82% yield) and with 93% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 9.7 min, t_r (minor) = 12.8 min). [α]_D²² = -61.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.41-7.28 (m, 5H), 4.93 (s, 1H), 4.81-4.66 (m, 1H), 3.78 (t, *J* = 8.9 Hz, 1H), 3.34-3.08 (m, 3H), 1.55-1.42 (m, 2H), 1.40-1.27 (m, 2H), 0.92 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 162.1, 141.9, 129.0, 128.3, 126.2, 54.0, 53.7, 43.2, 29.8, 20.1, 13.9. HRMS (ESI, *m/z*) calcd. for C₁₃H₁₉N₂O₁ [M+H]⁺: 219.1492, found: 219.1492.

Compound (S)-7r



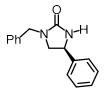
Starting from **6r** (68.0 mg, 0.20 mmol) according to the general procedure to provide **7r** as a white solid (29.7 mg, 68% yield) and with 95% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 8.3 min, t_r (minor) = 11.7 min). [**α**]**p**²² = 13.0° (*c* = 1.0, CH₂Cl₂). ¹**H NMR** (300 MHz, CDCl₃) δ 7.51-7.25 (m, 5H), 5.04 (s, 1H), 4.75 (dd, *J* = 8.6, 7.6 Hz, 1H), 3.79 (t, *J* = 8.9 Hz, 1H), 3.29-3.15 (m, 1H), 3.01 (qd, *J* = 13.7, 7.5 Hz, 2H), 1.93-1.71 (m, 1H), 0.90 (dd, *J* = 6.6, 4.0 Hz, 6H). ¹³C **NMR** (75 MHz, CDCl₃) δ 162.4, 142.0, 129.0, 128.3, 126.1, 54.4, 54.0, 51.2, 27.1, 20., 20.1. **HRMS (ESI,** *m/z*) calcd. for C₁₃H₁₉N₂O₁ [M+H]⁺: 219.1492, found: 219.1492.

Compound (S)-7s



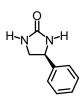
Starting from **6s** (77.6 mg, 0.20 mmol) according to the general procedure to provide **7s** as a white solid (47.8 mg, 90% yield) and with 92% ee as determined by HPLC analysis (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 40 °C, retention times: t_r (major) = 8.9 min, t_r (minor) = 10.2 min). [α]_D²² = -2.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.33-7.08 (m, 10H), 4.96 (s, 1H), 4.59 (t, *J* = 8.0 Hz, 1H), 3.59 (t, *J* = 8.8 Hz, 1H), 3.50-3.30 (m, 2H), 3.05 (dd, *J* = 8.6, 7.2 Hz, 1H), 2.75 (t, *J* = 7.4 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 161.9, 141.8, 139.0, 129.0, 128.9, 128.8, 128.6, 128.2, 126.5, 126.2, 54.1, 53.9, 44.9, 34.4. HRMS (ESI, *m/z*) calcd. for C₁₇H₁₉N₂O₁ [M+H]⁺: 267.1492, found: 267.1492.

Compound (S)-7t



Starting from **6t** (74.8 mg, 0.20 mmol) according to the general procedure to provide **7t** as a white solid (18.7 mg, 37% yield) and with 93% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*–Hexane/isopropanol = 95:5, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 33.9 min, t_r (minor) = 30.2 min). [**a**]**b**²² = -9.2° (*c* = 1.0, CH₂Cl₂). ¹**H NMR** (300 MHz, CDCl₃) δ 7.29-7.11 (m, 10H), 4.87 (s, 1H), 4.66 (t, *J* = 8.2 Hz, 1H), 4.42 (d, *J* = 14.9 Hz, 1H), 4.24 (d, *J* = 14.9 Hz, 1H), 3.59 (t, *J* = 8.9 Hz, 1H), 3.03 (dd, *J* = 8.8, 7.5 Hz, 1H). ¹³**C NMR** (75 MHz, CDCl₃) δ 161.9, 141.6, 137.0, 129.0, 128.8, 128.3, 128.2, 127.7, 126.2, 53.9, 53.3, 47.7. **HRMS (ESI,** *m***/***z***)** calcd. for C₁₆H₁₆N₂O₁Na₁ [M+Na]⁺: 275.1155, found: 275.1155.

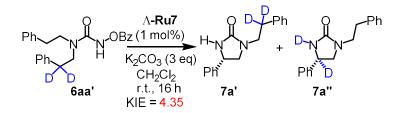
Compound (S)-7u



Starting from **6u** (56.8 mg, 0.20 mmol) according to the general procedure to provide **7u** as a white solid (29.6 mg, 91% yield) and with 91% ee as determined by HPLC analysis (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 15.2 min, t_r (minor) = 12.6 min). [α]_D²² = 23.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.46-7.27 (m, 5H), 4.95 (dd, *J* = 49.5, 41.5 Hz, 3H), 3.87 (t, *J* = 8.8 Hz, 1H), 3.35 (t, *J* = 8.1 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 141.5, 129.1, 128.4, 126.2, 56.9, 49.8. HRMS (ESI, *m*/*z*) calcd. for C₉H₁₀N₂O₁ [M+H]⁺: 163.0866, found: 163.0866.

4.4.3 Mechanistic Experiments

1) Kinetic Isotope Experiment



Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrates 1aa' (0.2 mmol), Λ -**Ru7** (0.002 mmol, 1 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (2.0 mL, 0.1 M) was added via syringe. The reaction mixture was stirred at room temperature for 16 h under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column and purified by flash chromatography on silica gel (EtOAc/*n*-*H*exane = 2:1 to EtOAc/MeOH = 95:5) to afford the mixture of products **7a'** and **7a''**. The ratio was determined by ¹H NMR analysis, resulting in KIE = 4.35 (**Figure 86**).

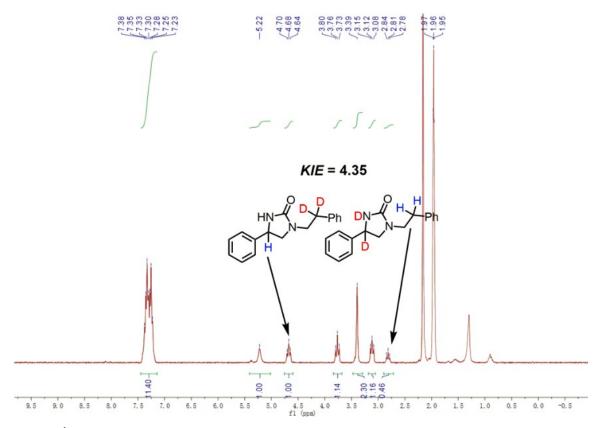
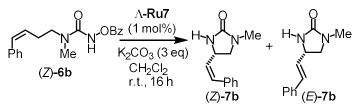


Figure 86. ¹H NMR spectrum analysis of the kinetic isotope experiment.

2) Olefin Isomerization



Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrates (*Z*)-**6b** (0.2 mmol), Λ -**Ru7** (0.002 mmol, 1 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (2.0 mL, 0.1 M) was added via syringe in sequence. The reaction mixture was stirred at room temperature for 16 h under an atmosphere of N₂. Afterwards, the crude reaction solution was filtered through a thin layer of celite and washed with 5 mL EtOAc. Filtrate was collected and organic solvent was removed under vacuo. The ratios of (*Z*)-**7b** and (*E*)-**7b** were analyzed by ¹H NMR of crude reaction solutions (see attached spectrums). The ratios of (*Z*)-**7b** is (*E*)-**7b** are 4.4:1 (at room temperature), 5.1:1 (at 4 °C) and 2.9:1 (at 40 °C).

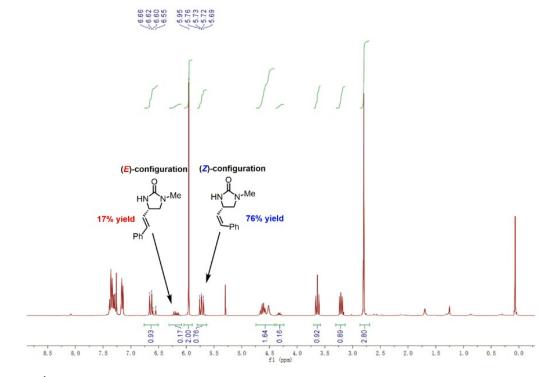


Figure 87. ¹H NMR spectrum analysis of crude reaction solution (at room temperature).

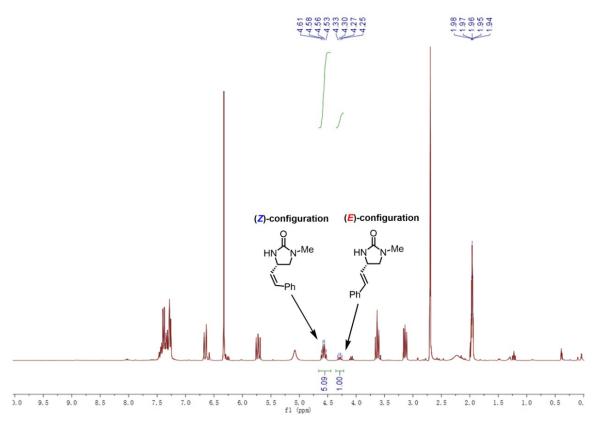


Figure 88. ¹H NMR spectrum analysis of crude reaction solution (at 4 °C).

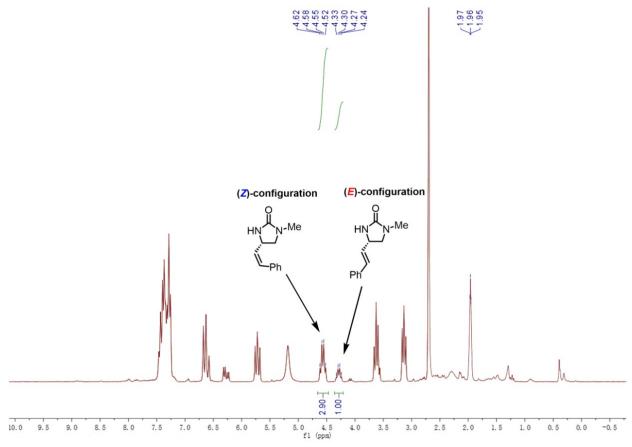
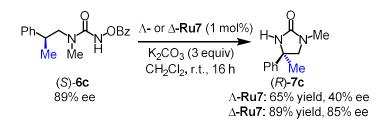


Figure 89. ¹H NMR spectrum analysis of crude reaction solution (at 40 °C).

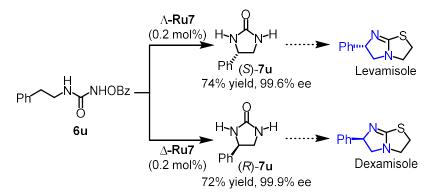
3) Stereochemistry



Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 10 mL Schlenk tube was charged with substrates (*S*)-**6c** (0.2 mmol), Λ -**Ru7** or Δ -**Ru7** (0.002 mmol, 1 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (2.0 mL, 0.1 M) was added via syringe in sequence. The reaction mixture was stirred at room temperature for 16 h under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc/*n*-Hexane = 2:1 to EtOAc/MeOH = 95:5) to afford the analytical pure product (*R*)-**7c**.

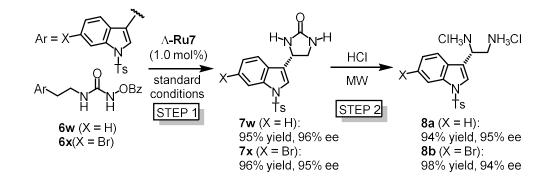
4.4.4 Synthetic applications

1) Synthetic Application to Drugs



Procedure: A pre-dried (using heating gun to dry for 3 times per tube) 25 mL Schlenk tube was charged with substrates **6u** (0.5 mmol), Λ -**Ru7** or Δ -**Ru7** (0.001 mmol, 0.2 mol%) and K₂CO₃ (207.0 mg, 1.5 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (5.0 mL, 0.1 M) was added via syringe in sequence. The reaction mixture was stirred at room temperature for 40 h under an atmosphere of N₂. Afterwards, the mixture was filtered and washed with 5 mL dichloromethane for 3 times. The filtrate was collected. Organic solvent was removed under *vacuo*. The resulting residue was dissolved in a minimum of EtOAc at 80 °C. After that, *n*-Hexane was added dropwisely until some precipitate formed. Subsequently, a minimun amount of EtOAc was added dropwisely until the solution became clear again. The resulting solution was cooled to 0 °C in freezer for 30 min. After that,

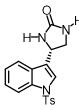
the precipitate was collected to provide enantiomeric pure (S)-7**u** with 74% yield, 99.6% ee and (R)-7**u** with 72% yield, 99.9% ee. The analytical data of compound 7**u** was already shown on page 189.



2) Synthetic Application to Natural Products

Step 1: Enantioselective C–H amination. A pre-dried (using heating gun to dry 3 times) 10 mL Schlenk tube was charged with substrates 6w or 6x (0.2 mmol), chiral ruthenium catalyst (0.002 mmol, 1 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Fresh distilled dichloromethane (2.0 mL) was added via syringe. The reaction mixture was stirred at the indicated temperature for the indicated time under an atmosphere of N₂. Afterwards, the mixture was directly transferred to a column (the Schlenk tube was rinsed with a minimal amount of CH₂Cl₂ to transfer the reaction solution completely) and purified by flash chromatography on silica gel (EtOAc/*n*–*H*exane = 2:1 to EtOAc/MeOH 95:5) to afford the analytical pure products 7w and 7x.

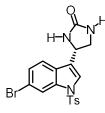
Compound (S)-7w



Starting from **6w** (95.4 mg, 0.20 mmol) according to the general procedure to provide **7w** as a white solid (67.5 mg, 95% yield) and with 96% ee as determined by HPLC analysis (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 16.9 min, t_r (minor) = 21.1 min). [α]_D²² = 17.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.02 (d, *J* = 8.3 Hz, 1H), 7.81 (d, *J* = 8.3 Hz, 2H), 7.61 (d, *J* = 6.9 Hz, 2H), 7.37 (t, *J* = 7.7 Hz, 1H), 7.31-7.22 (m, 3H), 5.25 (s,

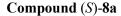
1H), 5.19-5.08 (m, 2H), 3.93 (t, J = 8.9 Hz, 1H), 3.52 (t, J = 7.9 Hz, 1H), 2.37 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 163.5, 145.3, 135.9, 135.3, 130.1, 128.2, 127.1, 125.4, 123.5, 122.6, 119.9, 114.1, 49.7, 47.5, 21.7. HRMS (ESI, *m/z*) calcd. for C₁₈H₁₇S₁N₃O₃Na₁[M+Na]⁺: 378.0833, found: 378.0885.

Compound (S)-7x



Starting from **6x** (111.0 mg, 0.20 mmol) according to the general procedure to provide **7x** as a white solid (83.2 mg, 96% yield) and with 95% ee as determined by HPLC analysis (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 50:50, flow rate: 1.0 mL/min, column temperature: 30 °C, retention times: t_r (major) = 9.5 min, t_r (minor) = 11.5 min). [α]_D²² = 28.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.10 (d, *J* = 1.2 Hz, 1H), 7.71 (d, *J* = 8.3 Hz, 2H), 7.48 (s, 1H), 7.38 (d, *J* = 8.4 Hz, 1H), 7.29 (dd, *J* = 8.5, 1.5 Hz, 1H), 7.20 (d, *J* = 8.0 Hz, 3H), 5.23 (s, 1H), 5.13-4.89 (m, 2H), 3.83 (t, *J* = 8.9 Hz, 1H), 3.39 (t, *J* = 7.9 Hz, 1H), 2.30 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 163.5, 145.7, 136.5, 135.0, 130.4, 130.1, 128.5, 127.1, 127.0, 124.0, 122.4, 121.1, 119.2, 117.2, 49.6, 47.5, 21.8. HRMS (ESI, *m/z*) calcd. for C₁₈H₁₆Br₁S₁N₃O₃Na₁[M+Na]⁺: 455.9988, 457.9969, found: 455.9989, 457.9969.

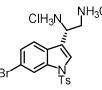
Step 2: Hydrolysis. 7w or **7x** (10.0 mg) dissolved in 0.5 mL concentrated HCl together with 0.5 mL acetic acid was placed in a sealed tube (CEM designed 20 mL pressure-rated reaction vial) with a stirring bar, and the reaction mixture was exposed to microwave irradiation for 10 min at 85 °C. After that, the resulting mixture was evaporated to dryness, redissolved with water (2 mL) and evaporated to dryness to remove traces of HCl. Redissolving with water and evaporation to dryness was repeated once. Water (2 mL) was added and the solution washed with CH_2Cl_2 (3 x 2 mL) and dried under *vacuo* to provide analytical pure products **8a** with 94% yield, 95% ee and **8b** with 98% yield, 94% ee.





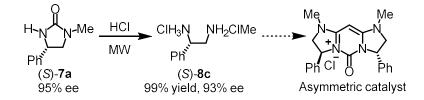
(*S*)-**8a:** 94% yield, 95% ee. light brown solid. ¹H NMR (500 MHz, DMSO) δ 9.18 (s, 3H), 8.59 (s, 3H), 8.28 (s, 1H), 7.99-7.93 (m, 4H), 7.44 (dd, J = 10.7, 4.5 Hz, 3H), 7.41-7.35 (m, 1H), 5.06 (d, J = 4.5 Hz, 1H), 3.62-3.53 (m, 1H), 3.51-3.41 (m, 1H), 2.37 (s, 3H). ¹³C NMR (126 MHz, DMSO) δ 145.8, 133.9, 133.9, 130.4, 128.3, 128.1, 127.2, 126.7, 125.5, 125.4, 123.5, 120.3, 115.4, 113.1, 108.6, 44.5, 41.2, 21.1. HRMS (ESI, *m/z*) calcd. for C₁₇H₁₇N₂O₂S₁ [M-NH₃Cl₂]⁺: 313.1005, found: 313.1007.

Compound (S)-8b



(*S*)-8b: 98% yield, 94% ee. light brown solid. ¹H NMR (500 MHz, DMSO) δ 9.16 (s, 3H), 8.53 (s, 3H), 8.33 (s, 1H), 8.06 (s, 1H), 8.01 (d, *J* = 7.9 Hz, 2H), 7.93 (d, *J* = 8.4 Hz, 1H), 7.58 (d, *J* = 8.2 Hz, 1H), 7.48 (d, *J* = 8.0 Hz, 2H), 5.05 (s, 1H), 3.53 (s, 2H), 2.39 (s, 3H). ¹³C NMR (126 MHz, DMSO) δ 142.5, 130.8, 130.0, 126.9, 123.9, 123.6, 122.9, 118.6, 114.4, 111.7, 111.7, 40.6, 37.4, 17.5. HRMS (ESI, *m/z*) calcd. for C₁₇H₁₉Br₁N₃O₂S₁ [M–HCl₂]⁺: 408.0376, 410.0355 found: 408.0379, 410.0357.

3) Synthetic Application to Asymmetric Catalyst



Hydrolysis. 7a (10.0 mg) dissolved in 1.0 mL concentrated HCl was placed in a sealed tube (CEM designed 20 mL pressure-rated reaction vial) with a stirring bar, and the reaction mixture was exposed to microwave irradiation for 15 min at 120 °C. After that, the resulting mixture was evaporated to dryness, redissolved with water (2 mL) and evaporated to dryness to remove traces of HCl to provide analytical pure products **8c** with 99% yield, 93% ee.

(*S*)-8c: 99% yield, 93% ee. white solid. ¹H NMR (300 MHz, CD₃OD_SPE) δ 7.66-7.38 (m, 5H), 3.74-3.54 (m, 2H), 3.25 (s, 1H), 2.70 (s, 3H). ¹³C NMR (75 MHz, CD₃OD_SPE) δ 133.9, 132.0, 131.3, 129.3, 53.3, 52.0, 34.7. HRMS (ESI, *m/z*) calcd. for C₉H₁₅N₂ [M–HCl₂]⁺: 151.1230 found: 151.1234.

Note: The vicinal diamines were transformed back to the corresponding cyclic ureas for the determination of the ee values using a reported method starting from the bis–Hydrochloride salt.¹



Additional informations about the set up of microwave irradiation reaction:

Figure 90. Reaction set up in the microwave reactor.

4.4.5 Single Crystal X-Ray Diffraction Studies

Crystalography of compound (*S*)-**7h:** Single crystals of (*S*)-**7h** were obtained by slow diffusion from the solution in CH_2Cl_2 layered with Et_2O at room temperature.

A suitable crystal of $C_{10}H_{11}CIN_2O$ was selected under inert oil and mounted using a MiTeGen loop. Intensity data of the crystal were recorded with a STADIVARI diffractometer (Stoe & Cie). The diffractometer was operated with Cu-K α radiation (1.54186 Å, microfocus source) and equipped with a Dectris PILATUS 300K detector. Evaluation, integration and reduction of the diffraction data was carried out using the X-Area software suite.² Multi-scan and numerical absorption corrections were applied with the X-Red32 and LANA modules of the X-Area software suite.^{3,4} The structure was solved using dual-space methods (SHELXT-2014/5) and refined against F^2 (SHELXL-2018/3 using ShelXle interface).⁵⁻⁷ All non–Hydrogen atoms were refined with anisotropic displacement parameters. The hydrogen atoms bonded to carbon atoms were refined using the "riding model" approach with isotropic displacement parameters 1.2 times (for CH₃ groups 1.5 times) of that of the preceding carbon atom. The hydrogen atom bonded to the nitrogen atom was refined isotropic without restraints. CCDC 1972573 contains the supplementary crystallographic data for this compound.

Table 8. Selected crystallographic data and details of the structure determination for (S)-7h.

Identification code	(<i>S</i>)-7h
Empirical formula	$C_{10}H_{11}ClN_2O$
Molar mass / $g \cdot mol^{-1}$	210.66
Space group (No.)	$P2_{1}2_{1}2_{1}(19)$
<i>a</i> / Å	5.66680(10)
b / Å	7.92950(10)
<i>c</i> / Å	21.8819(5)
$V/\text{\AA}^3$	983.26(3)
Ζ	4
$ ho_{calc.}$ / g·cm ⁻³	1.423
μ / mm^{-1}	3.172
Color	colorless
Crystal habitus	needle
Crystal size / mm ³	0.510 x 0.190 x 0.104
Т / К	100
λ/Å	1.54186 (Cu-K _α)
heta range / °	4.041 to 75.585
Range of Miller indices	$-6 \le h \le 2$
	$-8 \le k \le 9$
	$-27 \le l \le 27$
Absorption correction	multi-scan and numerical
T_{\min}, T_{\max}	0.1354, 0.3599
$R_{ m int}, R_{\sigma}$	0.0179, 0.0090
Completeness of the data set	0.995
No. of measured reflections	10354
No. of independent reflections	1981
No. of parameters	133
No. of restrains	0
S (all data)	1.044
$R(F)$ ($I \ge 2\sigma(I)$, all data)	0.0236, 0.0237
$wR(F^2)$ ($I \ge 2\sigma(I)$, all data)	0.0644, 0.0645
Extinction coefficient	0.0032(7)
Flack parameter <i>x</i>	0.002(6)
$\Delta ho_{ m max}, \Delta ho_{ m min}$ / e·Å ⁻³	0.269, -0.148

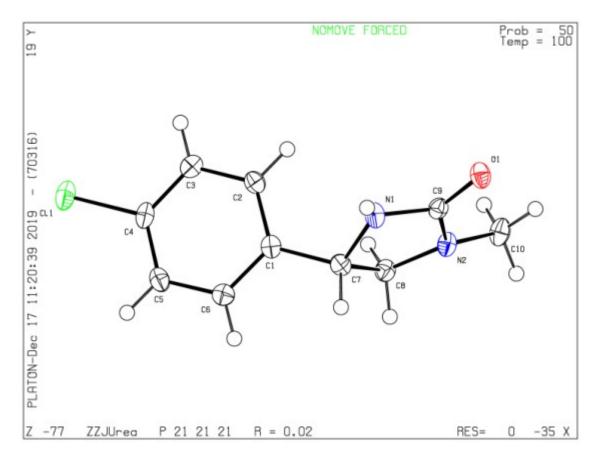
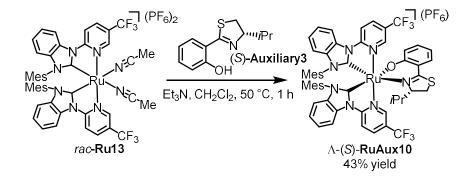


Figure 91. X-ray structure analysis of compound (S)-7h.

Reference

- 1. Y. Liu, K. Ding, J. Am. Chem. Soc. 2005, 127, 10488-10489.
- 2. X-Area, STOE & Cie GmbH, Darmstadt, Germany, 2018.
- 3. X-RED32, STOE & Cie GmbH, Darmstadt, Germany, 2018.
- 4. LANA Laue Analyzer, STOE & Cie GmbH, Darmstadt, Germany, 2019.
- 5. G. M. Sheldrick, Acta Cryst. A 2015, 71, 3.
- 6. G. M. Sheldrick, Acta Cryst. C 2015, 71, 3.
- 7. C. B. Hübschle, G. M. Sheldrick, B. Dittrich, J. Appl. Cryst. 2011, 44, 1281.

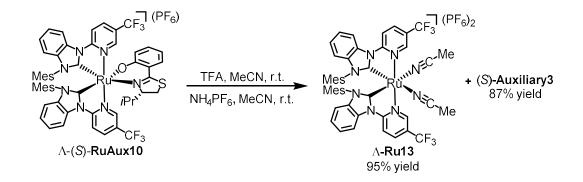
4.5 Enantioselective Synthesis of Cyclic Carbamates by Intramolecular C-H Amidation



4.5.1 Synthesis of the Ruthenium Catalyst

A mixture of racemic ruthenium complex *rac*-**Ru13** (1 equiv, 494.5 mg, 0.4 mmol), chiral auxiliary (*S*)-**5** (1 equiv, 88.5 mg, 0.4 mmol) and triethylamine (5 equiv, 278 μ L, 2.0 mmol) in CH₂Cl₂ (4 mL) was heated at 50 °C for 1 h (*Note*: longer time will decrease the yield!). The reaction mixture was cooled to room temperature and concentrated to dryness. The residue was subjected to a flash silica gel chromatography (CH₃CN:CH₂Cl₂ = 1:50 to 1:20) to separate the first colorful eluent which was assigned as Λ -(*S*)-**RuAux10**.

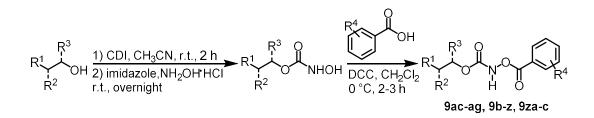
A-(*S*)-**RuAux10**: red solid (236.4 mg, 43% yield). ¹**H NMR** (300 MHz, CD₂Cl₂) δ 9.26 (s, 1H), 8.26 (s, 1H), 8.16 (t, J = 5.8 Hz, 3H), 8.04 (d, J = 8.3 Hz, 1H), 7.88 (dt, J = 8.9, 5.4 Hz, 2H), 7.63-7.47 (m, 2H), 7.32 (ddd, J = 10.9, 9.8, 4.6 Hz, 3H), 6.93-6.75 (m, 3H), 6.66 (dd, J = 12.4, 5.0 Hz, 3H), 6.51 (s, 1H), 6.36-6.21 (m, 2H), 3.91 (dd, J = 8.4, 2.3 Hz, 1H), 3.21 (dd, J = 11.9, 9.0 Hz, 1H), 3.05 (d, J = 11.9 Hz, 1H), 2.28 (s, 3H), 2.18 (s, 3H), 2.13 (s, 3H), 2.03 (s, 3H), 1.19 (s, 3H), 0.93 (s, 3H), 0.38 (d, J = 7.1 Hz, 6H), -0.44-0.64 (m, 1H). ¹³**C NMR** (75 MHz, CD₂Cl₂) δ 207.7, 205.4, 174.4, 169.9, 157.4, 156.7, 149.1, 147.6, 141.0, 137.7, 137.3, 137.1, 135.2, 134.8, 134.7, 134.5, 134.0, 133.0, 131.4, 131.2, 131.10, 131.07, 130.5, 130.2, 130.1, 126.3, 125.5, 125.4, 124.1, 123.6, 123.6, 123.0, 122.5, 119.0, 114.5, 111.8, 111.6, 111.4, 111.0, 110.6, 85.3, 32.2, 28.5, 20.9, 20.8, 19.7, 17.9, 17.5, 17.3, 16.9, 15.2. ¹⁹**F NMR** (283 MHz, CD₂Cl₂) δ -62.15, -62.79, -71.40, -73.92. **HRMS** (ESI, *m/z*) calcd for C₅₀H₅₀F₆N₇O₁Ru₁S₁ [M-PF₆]⁺: 1084.2754, found: 1084.2791.



To a mixture of single ruthenium auxiliary complex (236.4 mg) in CH₃CN (5 mL) was added CF₃COOH (50 μ L). The reaction mixture was stirred at room temperature for 30 min, then concentrated under reduced pressure. The residue was subjected to flash silica gel chromatography (CH₃CN:CH₂Cl₂ = 1:100 to 1:10) and added 500 mg NH₄PF₆ above the seasand band (to change the counter anion on the column) to afford chiral ruthenium catalyst as pale yellow solid. (*S*)-Auxiliary3 was eluted firstly and recovered in 87% yield. A-Ru13 was isolated in 202 mg as 95% yield (see following picture of the flash column).

 Λ -Ru13: pale yellow solid (202 mg, 95% yield). All other spectroscopic data of enantiopure ruthenium catalyst Λ -Ru13 were in agreement with the racemic catalyst *rac*-Ru13.

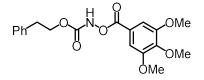
4.5.2 Synthesis of the Substrates



General procedure: 1,1'-carbonyldiimidazole (CDI, 1.5 equiv) was added to a solution of alcohol (1.0 equiv) in acetonitrile (0.2 M) and stirred at room temperature for 2 hours. After that, imidazole (2.0 equiv) and hydroxylamine hydrochloride (3.0 equiv) were added, and stirred at room temperature overnight. The reaction was quenched using a 1 N aqueous solution of HCl (3.0 equiv). The layers were separated and the aqueous layer was extracted using EtOAc (3 x 20 mL). The organic phase was washed with brine, dried over Na₂SO₄, filtered, and concentrated in *vacuo* to afford the crude *N*–Hydroxycarbamate, which was purified by flash column chromatography (*n*–hexane/EtOAc = 3:1 to 1:1). To a solution of the *N*–hydroxycarbamate (1.0 equiv) and benzoic acid or it's derivatives (1.0 equiv) in CH₂Cl₂ (0.1 M), DCC (1.0 equiv) in CH₂Cl₂ (0.33 M) was added dropwise at 0 °C. After

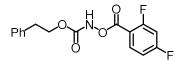
stirring for 2-3 hours (monitored by TLC), the reaction mixture was diluted by Et_2O (10 mL/mmol). The byproduct precipitated out and was removed by filtration. The solvent was removed in *vacuo* and the residue was purified by column chromatography on silica gel (*n*-hexane/CH₂Cl₂ = 1:1 to pure CH₂Cl₂).

Compound 9ae



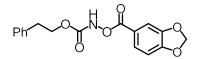
9ae: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.27 (s, 1H), 7.50-7.13 (m, 7H), 4.47 (t, *J* = 7.0 Hz, 2H), 4.06-3.83 (m, 9H), 3.03 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.8, 153.3, 143.6, 137.3, 129.1, 128.7, 126.9, 121.5, 107.5, 67.4, 61.2, 56.5, 35.3. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₁N₁O₇Na [M+Na]⁺: 398.1210, found: 398.1210.

Compound 9af



9af: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.33 (s, 1H), 8.06 (td, *J* = 8.3, 6.5 Hz, 1H), 7.29 (ddd, *J* = 18.5, 10.2, 4.5 Hz, 5H), 6.99 (tdd, *J* = 12.7, 8.4, 2.3 Hz, 2H), 4.47 (t, *J* = 7.0 Hz, 2H), 3.02 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.6, 168.5, 165.2, 165.2, 165.1, 162.9, 162.8, 161.7, 161.5, 156.5, 137.3, 134.4, 134.4, 134.2, 134.2, 129.1, 128.7, 126.9, 112.5, 112.4, 112.2, 112.2, 112.0, 112.0, 106.1, 105.8, 105.5, 67.5, 35.3. **HRMS** (ESI, *m/z*) calcd for C₁₆H₁₃F₂N₁O₄Na [M+Na]⁺: 344.0705, found: 344.0705.

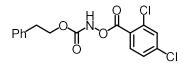
Compound 9ag



9ag: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.32 (s, 1H), 7.74 (dd, *J* = 8.2, 1.6 Hz, 1H), 7.51 (d, *J* = 1.6 Hz, 1H), 7.36-7.19 (m, 5H), 6.90 (d, *J* = 8.2 Hz, 1H), 6.10 (s, 2H), 4.45 (t, *J* = 7.0 Hz, 2H), 3.01 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.4, 156.7, 152.9, 148.2, 137.4, 129.1, 128.7,

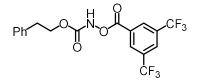
126.9, 126.5, 120.5, 109.8, 108.5, 102.3, 67.3, 35.3. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₅N₁O₆Na [M+Na]⁺: 352.0792, found: 352.0792.

Compound 9ah



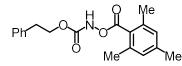
9ah: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.29 (s, 1H), 7.91 (d, *J* = 8.5 Hz, 1H), 7.57 (d, *J* = 1.8 Hz, 1H), 7.42-7.20 (m, 7H), 4.49 (t, *J* = 7.0 Hz, 2H), 3.03 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 164.0, 156.5, 140.1, 137.2, 136.0, 133.0, 131.5, 129.1, 128.8, 127.5, 126.9, 124.9, 67.5, 35.3. **HRMS** (ESI, *m/z*) calcd for C₁₆H₁₃Cl₂N₁O₄Na [M+Na]⁺: 376.0114, found: 376.0114.

Compound 9ai



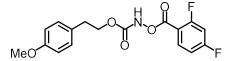
9ai: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.54 (s, 2H), 8.35 (s, 1H), 8.18 (s, 1H), 7.42-7.06 (m, 5H), 4.50 (t, J = 6.9 Hz, 2H), 3.03 (t, J = 6.9 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 163.5, 156.3, 137.1, 133.6, 133.1, 132.7, 132.2, 130.2, 129.2, 129.0, 128.7, 127.8, 127.8, 127.7, 127.0, 124.6, 121.0, 117.4, 67.7, 35.2. HRMS (ESI, m/z) calcd for C₁₈H₁₃F₆N₁O₄Na [M+Na]⁺: 444.0641, found: 444.0641.

Compound 9aj



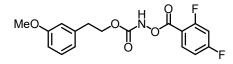
9aj: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.30 (s, 1H), 7.39-7.21 (m, 5H), 6.92 (s, 2H), 4.49 (t, *J* = 7.1 Hz, 2H), 3.05 (t, *J* = 7.1 Hz, 2H), 2.36 (d, *J* = 15.2 Hz, 9H). ¹³**C NMR** (75 MHz, CDCl₃) δ 169.1, 156.6, 141.1, 137.3, 136.9, 129.1, 128.8, 128.8, 126.9, 126.7, 67.3, 35.4, 21.4, 20.1. **HRMS** (ESI, *m/z*) calcd for C₁₈₉H₂₁N₁O₄Na [M+Na]⁺: 350.1363, found: 350.1363.

Compound 9b



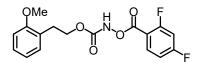
9b: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.27 (s, 1H), 8.02 (td, *J* = 8.3, 6.4 Hz, 1H), 7.18-7.06 (m, 2H), 7.05-6.90 (m, 2H), 6.86-6.73 (m, 2H), 4.40 (t, *J* = 7.0 Hz, 2H), 3.78 (s, 3H), 2.93 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.5, 165.2, 165.0, 162.9, 162.8, 161.5, 158.6, 156.5, 134.4, 134.4, 134.2, 134.2, 130.0, 129.3, 114.2, 112.5, 112.4, 112.2, 112.2, 106.1, 105.8, 105.5, 67.7, 55.4, 34.4. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₅F₂NO₅Na [M+Na]⁺: 374.0811, found: 374.0811.

Compound 9c



9c: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.03 (dd, *J* = 14.8, 8.3 Hz, 1H), 7.24-7.15 (m, 1H), 7.06-6.90 (m, 2H), 6.78 (dd, *J* = 9.7, 7.3 Hz, 3H), 4.44 (t, *J* = 7.0 Hz, 2H), 3.79 (s, 3H), 2.97 (t, *J* = 7.0 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.5, 162.9, 160.0, 156.5, 138.8, 134.4, 134.2, 129.7, 121.4, 114.9, 112.5, 112.3, 106.2, 105.8, 105.5, 67.4, 55.4, 35.4. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₅F₂NO₅Na [M+Na]⁺: 374.0811, found: 374.0811.

Compound 9d

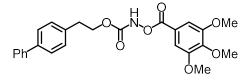


9d: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.03 (dd, J = 14.8, 8.2 Hz, 1H), 7.25-7.17 (m, 1H), 7.14 (d, J = 7.7 Hz, 1H), 7.04-6.89 (m, 2H), 6.85 (dd, J = 7.6, 4.1 Hz, 2H), 4.43 (t, J = 7.0 Hz, 2H), 3.82 (s, 3H), 3.01 (t, J = 7.0 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 157.8, 156.6, 134.4, 134.2, 131.0, 128.3, 125.5, 120.6, 112.5, 112.4, 112.2, 112.1, 110.5, 106.1, 105.8, 105.5, 66.5, 55.4, 30.3. HRMS (ESI, *m/z*) calcd for C₁₇H₁₅F₂NO₅Na [M+Na]⁺: 374.0811, found: 374.0812.

Compound 9e

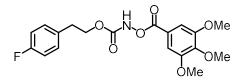
9e: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.29 (s, 1H), 8.04 (td, *J* = 8.3, 6.5 Hz, 1H), 7.21-7.08 (m, 4H), 7.06-6.88 (m, 2H), 4.41 (t, *J* = 7.4 Hz, 2H), 3.01 (t, *J* = 7.3 Hz, 2H), 2.34 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.6, 168.5, 165.2, 165.1, 162.9, 162.8, 161.7, 156.5, 136.6, 135.2, 134.4, 134.4, 134.2, 134.2, 130.6, 129.7, 127.1, 126.3, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 66.6, 32.6, 19.5. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₅F₂NO₄Na [M+Na]⁺: 358.0861, found: 358.0861.

Compound 9f



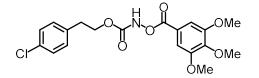
9f: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.22 (s, 1H), 7.59-7.39 (m, 6H), 7.32 (q, *J* = 7.8 Hz, 5H), 4.48 (t, *J* = 6.9 Hz, 2H), 3.91 (d, *J* = 9.0 Hz, 9H), 3.04 (t, *J* = 6.9 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.8, 153.4, 143.7, 141.0, 139.9, 136.4, 129.5, 129.0, 127.5, 127.4, 127.2, 121.5, 107.5, 67.4, 61.2, 56.5, 35.0. **HRMS** (ESI, *m/z*) calcd for C₂₅H₂₅NO₇Na [M+Na]⁺: 474.1523, found: 474.1523.

Compound 9g



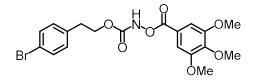
9g: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.21 (d, *J* = 5.3 Hz, 1H), 7.32 (s, 2H), 7.16 (dd, *J* = 8.3, 5.5 Hz, 2H), 6.94 (t, *J* = 8.6 Hz, 2H), 4.41 (t, *J* = 6.9 Hz, 2H), 3.92 (d, *J* = 7.6 Hz, 9H), 2.96 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 163.6, 160.3, 156.7, 153.3, 143.7, 133.0, 133.0, 130.6, 130.5, 121.4, 115.7, 115.4, 107.5, 67.2, 61.2, 56.5, 34.5. ¹⁹**F NMR** (235 MHz, CDCl₃) δ -116.30. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₀NO₇Na [M+Na]⁺: 416.1116, found: 416.1116.

Compound 9h



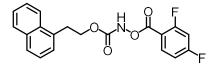
9h: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.18 (s, 1H), 7.32 (s, 2H), 7.22 (d, J = 8.4 Hz, 2H),
7.13 (d, J = 8.4 Hz, 2H), 4.41 (t, J = 6.8 Hz, 2H), 3.93 (d, J = 7.8 Hz, 9H), 2.96 (t, J = 6.8 Hz, 2H). ¹³C
NMR (75 MHz, CDCl₃) δ 165.8, 156.7, 153.4, 143.7, 135.8, 132.8, 130.4, 128.9, 121.4, 107.5, 67.0,
61.2, 56.6, 34.7. HRMS (ESI, m/z) calcd for C₁₉H₂₀ClNO₇Na [M+Na]⁺: 432.0821, found: 432.0820.

Compound 9i

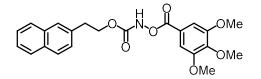


9i: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.20 (s, 1H), 7.37 (d, *J* = 8.3 Hz, 2H), 7.31 (s, 2H), 7.07 (d, *J* = 8.3 Hz, 2H), 4.41 (t, *J* = 6.8 Hz, 2H), 3.93 (d, *J* = 8.0 Hz, 10H), 2.94 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.7, 153.4, 143.7, 136.4, 131.8, 130.8, 121.4, 120.8, 107.5, 66.9, 61.2, 56.6, 34.7. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₀BrNO₇Na [M+Na]⁺: 476.0315, 478.0298, found: 476.0316, 478.0296.

Compound 9j

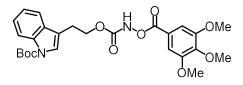


9j: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.26 (s, 1H), 8.11-7.97 (m, 2H), 7.92-7.82 (m, 1H), 7.76 (dd, *J* = 7.0, 2.3 Hz, 1H), 7.61-7.44 (m, 2H), 7.43-7.34 (m, 2H), 7.04-6.89 (m, 2H), 4.57 (t, *J* = 7.4 Hz, 2H), 3.48 (t, *J* = 7.4 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.7, 168.5, 165.1, 162., 161.6, 156.6, 134.4, 134.2, 134.1, 133.1, 132.2, 129.1, 127.8, 127.3, 126.5, 125.9, 125.7, 123.6, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 100.2, 67.0, 32.4. **HRMS** (ESI, *m/z*) calcd for C₂₀H₁₅F₂NO₄Na [M+Na]⁺: 394.0861, found: 394.0861. **Compound 9k**



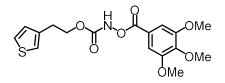
9k: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.21 (s, 1H), 7.77 (dt, *J* = 11.7, 4.4 Hz, 3H), 7.65 (s, 1H), 7.49-7.39 (m, 2H), 7.39-7.27 (m, 3H), 4.54 (t, *J* = 6.9 Hz, 2H), 3.91 (d, *J* = 16.5 Hz, 10H), 3.16 (t, *J* = 6.9 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.8, 153.3, 143.6, 134.8, 133.7, 132.5, 128.4, 127.8, 127.7, 127.6, 127.4, 126.3, 125.8, 121.4, 107.5, 67.3, 61.2, 56.5, 35.5. **HRMS** (ESI, *m/z*) calcd for C₂₃H₂₃NO₇Na [M+Na]⁺: 448.1367, found: 448.1366.

Compound 91



91: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.12 (d, J = 8.1 Hz, 1H), 7.55 (d, J = 7.8 Hz, 1H), 7.46 (s, 1H), 7.36-7.28 (m, 3H), 7.22 (d, J = 7.4 Hz, 1H), 4.51 (t, J = 7.1 Hz, 2H), 3.92 (d, J = 7.8 Hz, 9H), 3.10 (t, J = 7.1 Hz, 2H), 1.67 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 165.8, 156.8, 153.4, 143.7, 130.5, 124.7, 123.7, 122.7, 121.4, 119.0, 116.1, 115.5, 107.5, 83.8, 66.2, 61.2, 56.5, 28.4, 24.9. HRMS (ESI, *m/z*) calcd for C₂₆H₃₀N₂O₉Na [M+Na]⁺: 537.1844, found: 537.1843.

Compound 9m

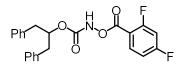


9m: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.23 (s, 1H), 7.33 (s, 2H), 7.26-7.19 (m, 1H), 7.04 (s, 1H), 6.96 (d, *J* = 4.8 Hz, 1H), 4.44 (t, *J* = 6.8 Hz, 2H), 3.92 (d, *J* = 6.0 Hz, 9H), 3.03 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.7, 153.4, 143.7, 137.5, 128.3, 125.9, 122.0, 121.4, 107.5, 66.8, 61.2, 56.5, 29.8. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₉S₁N₁O₇Na [M+Na]⁺: 404.0774, found: 404.0775.

Compound 9n

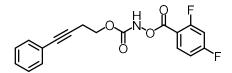
9n: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.22 (s, 1H), 8.12-7.93 (m, 1H), 7.36-7.16 (m, 4H), 7.12-6.85 (m, 2H), 5.74-5.53 (m, 1H), 3.38 (dd, J = 17.2, 6.2 Hz, 2H), 3.14 (dd, J = 17.1, 2.7 Hz, 2H).
¹³C NMR (75 MHz, CDCl₃) δ 168.6, 168.4, 165.2, 165.0, 163.0, 162.9, 161.5, 156.4, 140.1, 134.3, 134.2, 127.1, 124.9, 112.5, 112.4, 112.2, 112.1, 106.1, 105.8, 105.5, 78.6, 39.7. HRMS (ESI, *m/z*) calcd for C₁₇H₁₃F₂N₁O₄Na [M+Na]⁺: 356.0705, found: 356.0707.

Compound 9o



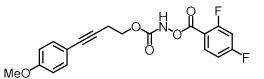
90: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.30 (s, 1H), 8.13-7.97 (m, 1H), 7.41-7.15 (m, 11H), 7.12-6.89 (m, 2H), 5.45-5.23 (m, 1H), 3.11-2.82 (m, 4H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.6, 168.4, 165.2, 165.0, 162.7, 162.7, 161.5, 156.1, 136.9, 134.4, 134.3, 134.2, 134.2, 129.6, 128.6, 126.9, 112.5, 112.4, 112.2, 112.1, 112.0, 106.1, 105.8, 105.5, 78.7, 39.7. **HRMS** (ESI, *m/z*) calcd for C₂₃H₁₉F₂N₁O₄Na [M+Na]⁺: 434.1174, found: 434.1175.

Compound 9p



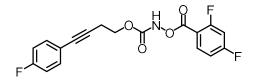
9p: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.32 (s, 1H), 8.17-7.98 (m, 1H), 7.41 (dd, *J* = 6.7, 3.0 Hz, 2H), 7.34-7.25 (m, 4H), 7.07-6.86 (m, 2H), 4.44 (t, *J* = 6.8 Hz, 2H), 2.85 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.5, 162.8, 156.3, 134.4, 134.2, 131.8, 128.4, 128.2, 123.4, 112.5, 112.2, 106.2, 105.8, 105.5, 84.8, 82.5, 64.8, 20.3. **HRMS** (ESI, *m/z*) calcd for C₁₈H₁₃F₂N₁O₄Na [M+Na]⁺: 368.0705, found: 368.0721.

Compound 9q



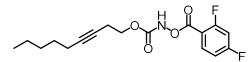
9q: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.33 (s, 1H), 8.17-7.96 (m, 1H), 7.43-7.22 (m, 3H), 7.08-6.91 (m, 2H), 6.83 (d, *J* = 8.8 Hz, 2H), 4.43 (t, *J* = 6.8 Hz, 2H), 3.83 (s, 3H), 2.82 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 173.0, 168.6, 165.2, 165.0, 162.8, 159.6, 156.3, 134.4, 134.2, 133.2, 115.5, 114.0, 112.5, 112.2, 106.2, 105.8, 105.5, 83.3, 82.3, 65.0, 55.5, 20.3. **HRMS** (ESI, *m/z*) calcd for C₁₉H₁₅F₂N₁O₅Na [M+Na]⁺: 398.0811, found: 398.0826.

Compound 9r



9r: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.29 (s, 1H), 8.12-7.97 (m, 1H), 7.40-7.30 (m, 2H), 7.05-6.82 (m, 4H), 4.40 (t, *J* = 6.8 Hz, 2H), 2.80 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.1, 164.2, 162.9, 162.8, 160.9, 134.4, 134.2, 133.7, 133.6, 119.4, 115.8, 115.5, 112.5, 112.2, 106.2, 105.8, 105.5, 84.6, 81.5, 64.8, 20.2. ¹⁹**F NMR** (235 MHz, CDCl₃) δ -98.58, -98.64, -101.39, -101.45, -111.47. **HRMS** (ESI, *m/z*) calcd for C₁₈H₁₂F₃N₁O₄Na [M+Na]⁺: 386.0626, found: 386.0626.

Compound 9s

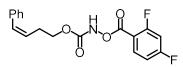


9s: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.32 (s, 1H), 8.06 (td, *J* = 8.3, 6.5 Hz, 1H), 7.09-6.84 (m, 2H), 4.39-4.20 (m, 2H), 2.54 (tt, *J* = 7.0, 2.3 Hz, 2H), 2.11 (tt, *J* = 7.1, 2.2 Hz, 2H), 1.59-1.40 (m, 2H), 1.39-1.22 (m, 4H), 0.89 (t, *J* = 7.0 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.7, 165.2, 165.1, 162.8, 162.8, 161.7, 156.3, 134.4, 134.2, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 82.8, 74.9, 65.4, 31.2, 28.7, 22.4, 19.6, 18.8, 14.2. **HRMS** (ESI, *m/z*) calcd for C₁₇H₁₉F₂N₁O₄Na [M+Na]⁺: 362.1174, found: 362.1190.

Compound (E)-9t

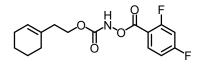
(*E*)-9t: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.28 (s, 1H), 8.13-7.97 (m, 1H), 7.38-7.14 (m, 5H),
7.04-6.86 (m, 2H), 6.49 (d, *J* = 15.9 Hz, 1H), 6.17 (dt, *J* = 15.8, 7.0 Hz, 1H), 4.38 (t, *J* = 6.6 Hz, 2H),
2.63 (q, *J* = 6.7 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 168.6, 168.4, 165.2, 165.0, 163.0, 162.9, 161.7,
161.5, 156.6, 137.3, 134.3, 134.2, 133.1, 128.7, 128.5, 127.5, 126.3, 125.0, 112.5, 112.4, 112.2, 112.1,
106.1, 105.8, 105.4, 66.3, 32.6. HRMS (ESI, *m/z*) calcd for C₁₈H₁₅F₂N₁O₄Na [M+Na]⁺: 370.0861,
found: 370.0875.

Compound (Z)-9t



(Z)-9t: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.29 (s, 1H), 8.14-7.98 (m, 1H), 7.42-7.16 (m, 5H),
7.06-6.91 (m, 2H), 6.59 (d, J = 11.7 Hz, 1H), 5.68 (dt, J = 11.7, 7.2 Hz, 1H), 4.35 (t, J = 6.7 Hz, 2H),
2.75 (qd, J = 6.9, 1.6 Hz, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 168.5, 165.2, 162.8, 156.5, 137.1, 134.4,
134.2, 132.1, 128.8, 128.6, 128.5, 127.1, 126.7, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 66.5,
28.3. HRMS (ESI, *m/z*) calcd for C₁₈H₁₅F₂N₁O₄Na [M+Na]⁺: 370.0861, found: 370.0875.

Compound 9u

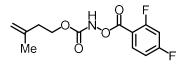


9u: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.06 (td, *J* = 8.3, 6.5 Hz, 1H), 7.06-6.84 (m, 2H), 5.46 (s, 1H), 4.29 (t, *J* = 6.9 Hz, 2H), 2.30 (t, *J* = 6.8 Hz, 2H), 1.94 (d, *J* = 3.9 Hz, 4H), 1.78 -1.33 (m, 5H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.5, 165.2, 165.1, 162.9, 162.9, 161.6, 156.7, 134.3, 134.2, 133.2, 124.2, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 77.6, 77.2, 76.8, 65.7, 37.2, 28.5, 25.4, 23.0, 22.4. **HRMS** (ESI, *m/z*) calcd for C₁₆H₁₇F₂N₁O₄Na [M+Na]⁺: 348.1018, found: 348.1033.

Compound 9v

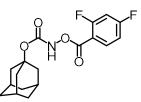
9v: colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.06 (dd, *J* = 14.8, 8.3 Hz, 1H), 6.97 (ddd, *J* = 18.8, 10.8, 2.3 Hz, 2H), 5.30 (s, 1H), 4.33-4.14 (m, 2H), 2.47-1.99 (m, 7H), 1.26 (s, 3H), 1.13 (d, *J* = 8.6 Hz, 1H), 0.81 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.6, 165.2, 165.0, 162.9, 162.9, 161.7, 156.6, 143.6, 134.4, 134.2, 119.5, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 65.4, 45.8, 40.9, 38.2, 36.1, 31.8, 31.5, 26.4, 21.3. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₁F₂N₁O₄Na [M+Na]⁺: 388.1331, found: 388.1341.

Compound 9w



9w: white solid. ¹H NMR (300 MHz, CDCl₃) δ 8.23 (s, 1H), 8.11-8.01 (m, 1H), 7.08-6.87 (m, 2H), 4.78 (d, *J* = 18.1 Hz, 2H), 4.34 (t, *J* = 6.8 Hz, 2H), 2.39 (t, *J* = 6.8 Hz, 2H), 1.75 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 168.7, 165.3, 165.1, 162.9, 156.6, 141.1, 134.4, 134.2, 112.9, 112.5, 112.2, 106.2, 105.8, 105.5, 65.3, 36.9, 22.6. HRMS (ESI, *m/z*) calcd for C₁₃H₁₂F₂N₁O₄Na [M+Na]⁺: 308.0705, found: 308.0720.

Compound 9x

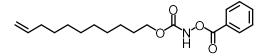


9x: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.11 (s, 1H), 8.06 (td, *J* = 8.3, 6.6 Hz, 1H), 7.04 – 6.86 (m, 2H), 2.17 (d, *J* = 13.1 Hz, 9H), 1.66 (s, 6H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.4, 165.0, 163.2, 163.1, 155.1, 134.3, 134.2, 112.5, 112.4, 112.2, 112.1, 106.1, 105.8, 105.4, 83.7, 41.5, 36.2, 31.2. **HRMS** (ESI, *m/z*) calcd for C₁₈H₁₉F₂N₁O₄Na [M+Na]⁺: 374.1174, found: 374.1192.

Compound 9y

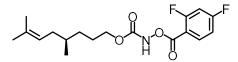
9y: colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 8.28 (s, 1H), 8.10 (d, *J* = 7.5 Hz, 2H), 7.63 (d, *J* = 7.6 Hz, 1H), 7.49 (t, *J* = 7.8 Hz, 2H), 4.22 (t, *J* = 6.7 Hz, 2H), 1.74-1.61 (m, 2H), 1.41-1.22 (m, 7H), 0.88 (t, *J* = 6.7 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.1, 156.9, 134.4, 130.2, 128.9, 127.0, 67.3, 31.5, 28.8, 25.5, 22.7, 14.1. **HRMS** (ESI, *m/z*) calcd for C₁₄H₁₉N₁O₄Na [M+Na]⁺: 288.1206, found: 288.1213.

Compound 9z



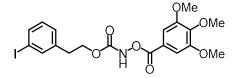
9z: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.3 (s, 1H), 8.16-8.06 (m, 2H), 7.64 (dd, *J* = 10.6, 4.3 Hz, 1H), 7.49 (t, *J* = 7.7 Hz, 2H), 5.81 (ddt, *J* = 16.9, 10.2, 6.7 Hz, 1H), 5.11-4.89 (m, 2H), 2.04 (q, *J* = 6.8 Hz, 2H), 1.74-1.61 (m, 2H), 1.32 (dd, *J* = 20.6, 9.0 Hz, 13H). ¹³**C NMR** (75 MHz, CDCl₃) δ 166.1, 156.9, 139.4, 134.4, 130.2, 128.9, 127.0, 114.3, 67.3, 34.0, 29.6, 29.5, 29.3, 29.3, 29.1, 28.9, 25.8. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₇N₁O₄Na [M+Na]⁺: 356.1832, found: 356.1849.

Compound 9za



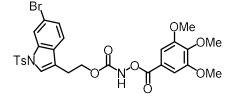
9za: colorless oil. ¹**H NMR** (300 MHz, CDCl₃) δ 8.27 (s, 1H), 8.16-7.92 (m, 1H), 7.10-6.78 (m, 2H), 5.07 (t, *J* = 7.0 Hz, 1H), 4.34-4.16 (m, 2H), 1.95 (dq, *J* = 15.4, 7.6 Hz, 2H), 1.72 (dd, *J* = 12.6, 4.9 Hz, 1H), 1.67 (s, 3H), 1.59 (s, 3H), 1.50 (ddd, *J* = 20.7, 12.5, 6.7 Hz, 2H), 1.40-1.26 (m, 1H), 1.26-1.10 (m, 1H), 0.91 (d, *J* = 6.3 Hz, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 168.6, 168.5, 165.2, 165.0, 163.0, 162.9, 161.7, 161.5, 156.7, 154.0, 134.4, 134.3, 134.2, 134.2, 131.6, 124.6, 112.5, 112.5, 112.2, 112.2, 106.2, 105.8, 105.5, 65.9, 37.1, 35.7, 29.5, 25.9, 25.6, 19.5, 17.8. **HRMS** (ESI, *m/z*) calcd for C₁₈H₂₃F₂N₁O₄Na [M+Na]⁺: 378.1487, found: 378.1502.

Compound 9zb



9zb: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.32 (s, 1H), 7.65-7.49 (m, 2H), 7.31 (t, *J* = 1.9 Hz, 2H), 7.16 (d, *J* = 7.7 Hz, 1H), 6.98 (td, *J* = 7.7, 2.0 Hz, 1H), 4.40 (ddd, *J* = 6.8, 4.4, 1.7 Hz, 2H), 3.91 (dt, *J* = 4.3, 1.8 Hz, 9H), 2.92 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.7, 156.6, 153.3, 143.7, 139.8, 138.0, 136.0, 130.4, 128.4, 121.4, 121.4, 107.5, 94.7, 66.8, 61.2, 56.6, 34.8. **HRMS** (ESI, *m/z*) calcd for C₁₉H₂₀I₁N₁O₇Na [M+Na]⁺: 524.0177, found: 524.0177.

Compound 9zc



9zc: white solid. ¹**H NMR** (300 MHz, CDCl₃) δ 8.16 (d, *J* = 11.5 Hz, 2H), 7.74 (d, *J* = 8.3 Hz, 2H), 7.42-7.30 (m, 5H), 7.23 (s, 1H), 4.45 (t, *J* = 6.8 Hz, 2H), 3.92 (d, *J* = 9.5 Hz, 10H), 3.03 (t, *J* = 6.7 Hz, 2H), 2.35 (s, 3H). ¹³**C NMR** (75 MHz, CDCl₃) δ 165.8, 156.7, 153.4, 145.4, 136.0, 135.2, 130.3, 129.6, 127.0, 126.8, 124.5, 121.3, 120.6, 118.8, 118.1, 117.0, 107.5, 65.7, 61.2, 56.5, 24.8, 21.8. **HRMS** (ESI, *m/z*) calcd for C₂₈H₂₇Br₁N₂O₉S₁Na [M+Na]⁺: 669.0513 & 671.0496, found: 669.0536 & 671.0514.

4.5.3 Ruthenium Catalyzed Asymmetric Intramolecular C-H Amidation

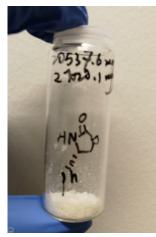
General procedure: A pre-dried Schlenk tube (10 mL) was charged with substrates (0.2 mmol), chiral ruthenium catalyst Λ -**Ru13** (0.004 mmol, 2.0 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Solvent 1,2-dichlorobezene (4.0 mL) was added via syringe. The reaction mixture was stirred at 30 °C for 20 h under an atmosphere of N₂. Afterwards, the mixture was transferred to a column and purified by flash chromatography on silica gel (EtOAc/*n*–*H*exane = 1:3 to 1:1) to afford the analytical pure product.

Additional experimental information:

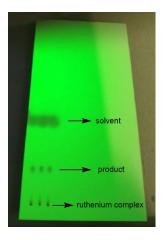
a) Execution of the reactions. All catalytic reactions were carried out in 10 mL Schlenk tubes from "Synthware" under N₂ atmosphere (see image below).



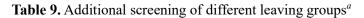
b) Isolation of the final product. The C-H amination products can be isolated by flash chromatography without any trace contaminations from the ruthenium catalyst as evident by the white color of the products (see image below).



c) TLC monitor (*n*-Hexane:EtOAc = 2:1), See image below.



9ab-akstandard conditions(S)-10a R CF_3 Λ -Ru6: R = CF3 Λ -Ru13(previous design)(new modification)
$\begin{array}{cccccccccccccccccccccccccccccccccccc$
$\begin{array}{c} \begin{array}{c} & & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ & \\ $
Entry Substrate Catalyst Conditions ^b NMR yield $(\%)^d$ ee $(\%)^d$
1 9aa Λ -Ru6 CH ₂ Cl ₂ at 25 °C without base 0 -
2 9ab A- Ru6 CH_2Cl_2 at 25 °C 69 68
3 9ac Λ - Ru6 CH ₂ Cl ₂ at 25 °C 94 62
4 9ad A- Ru6 CH_2Cl_2 at 25 °C 97 78
5 9ad Λ- Ru6 standard 98 82
6 9ad Λ-Ru13 standard 95 86
7 9ae Λ-Ru13 40 °C instead 97 90
8 9af A-Ru13 standard quant. $(99)^f$ 90
9 9af Λ -Ru13 no base < 5 n.d. ^g
10 9af Λ-Ru13 under air 92 90
11 9ag Λ-Ru13 standard 98 84
12 9ah Λ- Ru13 standard 96 83
13 9ai A-Ru13 standard 18 $n.d.^g$
14 9aj Λ-Ru13 standard 52 75



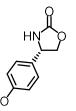
[a] Standard conditions: **9a** (0.2 mmol), K_2CO_3 (0.6 mmol), Ru catalyst (0.002 mmol) in 1,2-dichlorobenzene (4 mL) stirred at the 30 °C for 20 h under N₂ unless noted otherwise. [b] Deviations from standard conditions are shown. [c] Conversion. [d] Determined by ¹H NMR of the crude products using Cl₂CHCHCl₂ as internal standard. [e] Enantiomeric excess determined by HPLC analysis of the crude main product on a chiral stationary phase. [f] Isolated yield in brackets. [g] Not determined.

Compound 10a



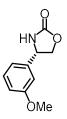
Starting from **9af** (62.2 mg, 0.20 mmol) according to the general procedure to provide **10a** as a white solid (32.3 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 15.0 min, t_r (minor) = 13.9 min). [α] $_{p^{22}}$ = +25.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.53-7.31 (m, 5H), 5.61 (s, 1H), 5.05 – 4.89 (m, 1H), 4.74 (t, *J* = 8.6 Hz, 1H), 4.20 (dd, *J* = 8.5, 6.9 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 159.6, 139.6, 129.5, 129.1, 126.2, 72.7, 56.6. HRMS (ESI, *m/z*) calcd for C₉H₉NO₂Na [M+Na]⁺: 186.0525, found: 186.0531.

Compound 10b



Starting from **9b** (70.2 mg, 0.20 mmol) according to the general procedure to provide **10b** as a white solid (38.2 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 17.3 min, t_r (minor) = 16.1 min). [α] $_{0}^{22}$ = +20.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.34-7.09 (m, 2H), 6.95-6.76 (m, 2H), 5.53 (s, 1H), 4.91-4.76 (m, 1H), 4.63 (t, *J* = 8.6 Hz, 1H), 4.09 (dd, *J* = 8.5, 7.0 Hz, 1H), 3.74 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 160.2, 159.6, 131.5, 127.6, 114.8, 72.9, 56.2, 55.6. HRMS (ESI, *m/z*) calcd for C₁₀H₁₁NO₃Na [M+Na]⁺: 216.0631, found: 216.0636.

Compound 10c



Starting from **9c** (70.2 mg, 0.20 mmol) according to the general procedure to provide **10c** as a white solid (38.3 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 92% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 20.0 min, t_r (minor) = 22.2 min). $[\alpha]_D^{22} = +18.8^\circ$ (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.30 (dd, *J* = 8.7, 7.6 Hz, 1H), 7.02-6.80 (m, 3H), 5.98 (s, 1H), 4.98-4.84 (m, 1H), 4.71 (t, *J* = 8.7 Hz, 1H), 4.17 (dd, *J* = 8.5, 7.0 Hz, 1H), 3.81 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 160.5, 159.8, 141.3, 130.5, 118.3, 114.4, 111.7, 72.6, 56.5, 55.5. HRMS (ESI, *m/z*) calcd for C₁₀H₁₁NO₃Na [M+Na]⁺: 216.0631, found: 216.0636.

Compound 10d



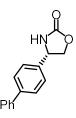
Starting from **9d** (70.2 mg, 0.20 mmol) according to the general procedure to provide **10d** as a white solid (38.1 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 93% ee (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: tr (major) = 15.7 min, tr (minor) = 13.4 min). [α] $_{D}^{22}$ = +83.1° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.32 (dd, *J* = 9.7, 7.7 Hz, 2H), 7.00 (t, *J* = 7.5 Hz, 1H), 6.90 (d, *J* = 8.1 Hz, 1H), 5.83 (s, 1H), 5.24 (dd, *J* = 8.5, 6.7 Hz, 1H), 4.80 (t, *J* = 8.7 Hz, 1H), 4.17 (dd, *J* = 8.4, 6.4 Hz, 1H), 3.86 (d, *J* = 12.0 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 160.2, 156.7, 129.6, 128.0, 125.7, 121.1, 110.7, 71.7, 55.6, 51.5. HRMS (ESI, *m/z*) calcd for C₁₀H₁₁NO₃Na [M+Na]⁺: 216.0631, found: 216.0636.

Compound 10e



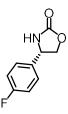
Starting from **9e** (67.0 mg, 0.20 mmol) according to the general procedure to provide **10e** as a white solid (35.1 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 92% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 24.1 min, t_r (minor) = 13.3 min). [α] $_{D}^{22}$ = +179.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.47-7.31 (m, 1H), 7.27-7.06 (m, 3H), 5.85 (s, 1H), 5.22-5.09 (m, 1H), 4.72 (t, *J* = 8.6 Hz, 1H), 4.03 (dd, *J* = 8.3, 7.0 Hz, 1H), 2.23 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 160.1, 137.7, 134.8, 131.1, 128.5, 127.1, 124.9, 71.8, 53.3, 19.2. HRMS (ESI, *m/z*) calcd for C₁₀H₁₁NO₂Na [M+Na]⁺: 200.0682, found: 200.0687.

Compound 10f



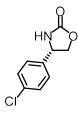
Starting from **9f** (90.2 mg, 0.20 mmol) according to the general procedure to provide **10f** as a white solid (47.3 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 98% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 254$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 26.5 min, t_r (minor) = 24.4 min). [α]_D²² = +3.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.71-7.55 (m, 4H), 7.41 (dtd, *J* = 9.6, 7.2, 3.2 Hz, 5H), 5.58 (s, 1H), 5.07-4.93 (m, 1H), 4.77 (t, *J* = 8.7 Hz, 1H), 4.24 (dd, *J* = 8.6, 6.9 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 159.5, 142.2, 140.4, 138.5, 129.1, 128.2, 127.9, 127.3, 126.7, 72.7, 56.3. HRMS (ESI, *m/z*) calcd for C₁₅H₁₃NO₂H [M+H]⁺: 240.1019, found: 240.1025.

Compound 10g



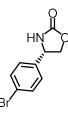
Starting from **9g** (78.6 mg, 0.20 mmol) according to the general procedure to provide **10g** as a white solid (35.7 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 23.6 min, t_r (minor) = 26.8 min). [α]_D²² = +26.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.38-7.28 (m, 2H), 7.08 (t, *J* = 8.6 Hz, 2H), 6.44 (s, 1H), 5.09-4.82 (m, 1H), 4.71 (t, *J* = 8.7 Hz, 1H), 4.13 (dd, *J* = 8.6, 6.9 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 164.7, 161.4, 160.0, 135.5, 135.5, 128.0, 127.9, 116.5, 116.2, 72.7, 72.7, 56.0. ¹⁹F NMR (235 MHz, CDCl₃) δ -112.89. HRMS (ESI, *m/z*) calcd for C₉H₈FNO₂Na [M+Na]⁺: 204.0431, found: 204.0437.

Compound 10h



Starting from **9h** (81.8 mg, 0.20 mmol) according to the general procedure to provide **10h** as a white solid (36.2 mg, 92% yield). Enantiomeric excess was established by HPLC analysis as 88% ee (column: Daicel Chiralpak IC 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 11.5 min, t_r (minor) = 18.1 min). [α] $_{0}^{22}$ = +16.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.37 (d, *J* = 8.5 Hz, 2H), 7.27 (d, *J* = 8.3 Hz, 2H), 6.29 (s, 1H), 5.03-4.85 (m, 1H), 4.13 (dd, *J* = 8.6, 6.8 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 159.9, 138.2, 134.9, 129.6, 127.6, 72.5, 56.0. HRMS (ESI, *m/z*) calcd for C₉H₈CINO₂Na [M+Na]⁺: 220.0136, found: 220.0141.

Compound 10i



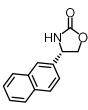
Starting from **9i** (90.6 mg, 0.20 mmol) according to the general procedure to provide **10i** as a white solid (47.5 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 90% ee (column: Daicel Chiralpak IG 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 16.9 min, t_r (minor) = 11.7 min). [α] $_{0}^{22}$ = +13.7° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.54 (d, *J* = 8.4 Hz, 2H), 7.34-7.10 (m, 2H), 6.02 (s, 1H), 5.09-4.82 (m, 1H), 4.73 (t, *J* = 8.7 Hz, 1H), 4.14 (dd, *J* = 8.4, 7.0 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 160.2, 138.8, 138.7, 132.6, 132.5, 127.9, 122.9, 72.4, 56.0. HRMS (ESI, *m/z*) calcd for C₉H₈BrNO₂Na [M+Na]⁺: 263.9631, found: 263.9638.

Compound 10j



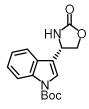
Starting from **9j** (74.2 mg, 0.20 mmol) according to the general procedure to provide **10j** as a white solid (42.2 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 93% ee (column: Daicel Chiralpak IA 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 15.8 min, t_r (minor) = 14.1 min). [α]_D²² = +186.4° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.99-7.90 (m, 1H), 7.86 (d, *J* = 8.2 Hz, 1H), 7.77 (dd, *J* = 5.8, 3.3 Hz, 1H), 7.66 (d, *J* = 7.1 Hz, 1H), 7.62-7.46 (m, 3H), 6.21 (s, 1H), 5.83-5.64 (m, 1H), 5.00 (t, *J* = 8.7 Hz, 1H), 4.22 (dd, *J* = 8.5, 6.5 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 160.0, 135.2, 134.2, 130.2, 129.5, 129.2, 127.1, 126.4, 125.8, 122.4, 121.9, 72.0, 53.3. HRMS (ESI, *m/z*) calcd for C₁₃H₁₁NO₂Na [M+Na]⁺: 236.0682, found: 236.0688.

Compound 10k



Starting from **9k** (85.0 mg, 0.20 mmol) according to the general procedure to provide **10k** as a white solid (41.3 mg, 97% yield). Enantiomeric excess was established by HPLC analysis as 92% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 60:40, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 14.1 min, t_r (minor) = 17.1 min). [α] $_{p}^{22}$ = +11.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.03-7.72 (m, 4H), 7.50 (ddt, *J* = 10.2, 8.5, 3.0 Hz, 3H), 5.59 (s, 1H), 5.20-5.04 (m, 1H), 4.81 (t, *J* = 8.7 Hz, 1H), 4.29 (dd, *J* = 8.6, 6.8 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 159.53, 136.80, 133.63, 133.38, 129.73, 128.10, 128.02, 127.05, 126.89, 125.65, 123.36, 72.57, 56.70. HRMS (ESI, *m/z*) calcd for C₁₃H₁₁NO₂Na [M+Na]⁺: 236.0682, found: 236.0688.

Compound 10l



Starting from **91** (102.8 mg, 0.20 mmol) according to the general procedure to provide **101** as a white solid (39.8 mg, 66% yield). Enantiomeric excess was established by HPLC analysis as 85% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 10.4 min, t_r (minor) = 12.1 min). [α]_D²² = +8.5° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.18 (d, *J* = 8.3 Hz, 1H), 7.61 (s, 1H), 7.54 (d, *J* = 7.7 Hz, 1H), 7.41-7.32 (m, 1H), 7.31-7.20 (m, 1H), 6.00 (s, 1H), 5.20 (dd, *J* = 8.7, 6.7 Hz, 1H), 4.76 (t, *J* = 8.7 Hz, 1H), 4.41 (dd, *J* = 8.6, 6.6 Hz, 1H), 1.67 (s, 10H). ¹³C NMR (75 MHz, CDCl₃) δ 159.6, 149.5, 136.4, 127.4, 125.4, 123.9, 123.3, 119.0, 118.8, 115.9, 84.5, 70.6, 49.8, 28.3. HRMS (ESI, *m/z*) calcd for C₁₆H₁₈N₂O₄Na [M+Na]⁺: 325.1159, found: 325.1166.

Compound 10m



Starting from **9m** (76.2 mg, 0.20 mmol) according to the general procedure to provide **10m** as a white solid (26.0 mg, 77% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak ADH column, ee = 93% (HPLC: 210 nm, *n*–Hexane/isopropanol = 85:15, flow rate 0.8 mL/min, 40 °C, t_r (major) = 11.2 min, t_r (minor) = 10.3 min). $[\alpha]_D^{22} = +9.0^\circ$ (*c* = 1.0, CH₂Cl₂). ¹H **NMR** (300 MHz, CDCl₃) δ 7.39 (dd, *J* = 5.0, 3.0 Hz, 1H), 7.08 (dd, *J* = 5.0, 1.2 Hz, 1H), 6.03 (s, 1H), 5.11-4.99 (m, 1H), 4.69 (t, *J* = 8.6 Hz, 1H), 4.23 (dd, *J* = 8.5, 6.6 Hz, 1H). ¹³C **NMR** (75 MHz, CDCl₃) δ 159.6, 140.9, 128.0, 125.2, 122.6, 72.0, 52.4. **HRMS** (ESI, *m/z*) calcd for C₇H₇N₁O₂S₁Na [M+Na]⁺: 192.0090, found: 192.0092.

Compound 10n



Starting from **9n** (66.6 mg, 0.20 mmol) according to the general procedure to provide **10n** as a white solid (34.8 mg, 99% yield). Enantiomeric excess was established by HPLC analysis as 95% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*–Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 21.0 min, t_r (minor) = 24.9 min). [α]_p²² = -58.1° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.39-7.14 (m, 4H), 6.70 (s, 1H), 5.41 (ddd, *J* = 8.1, 6.0, 2.3 Hz, 1H), 5.17 (d, *J* = 7.3 Hz, 1H), 3.50-3.24 (m, 2H). ¹³C NMR (75 MHz, CDCl₃) δ 159.9, 140.5, 139.9, 129.5, 128.1, 125.7, 125.0, 80.8, 61.4, 39.0. HRMS (ESI, *m/z*) calcd for C₁₀H₉NO₂Na [M+Na]⁺: 198.0525, found: 198.0530.

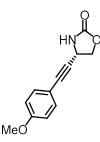
Compound 10o

Starting from **90** (82.2 mg, 0.20 mmol) according to the general procedure to provide **100** as a white solid (42.0 mg, 83% yield). Diasteriomeric ratio and enantiomeric excess was established by HPLC analysis as 1.8:1 d.r., 92%/97% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*–Hexane/isopropanol = 80:20, flow rate: 0.8 mL/min, column temperature: 30 °C, retention times: for major diastereomer, t_r (major) = 15.6 min, t_r (minor) = 14.1 min), for minor diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 17.3 min). [α] $_{D}^{22}$ = +2.4° (*c* = 1.0, CH₂Cl₂). For major diastereomer, t_r (major) = 18.5 min, t_r (minor) = 10.5 min, t_r

Compound 10p

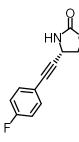
Starting from **9p** (69.0 mg, 0.20 mmol) according to the general procedure to provide **10p** as a white solid (34.0 mg, 91% yield). Enantiomeric excess was established by HPLC analysis as 91% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: tr (major) = 18.2 min, tr (minor) = 24.6 min). [α] $_{D}^{22}$ = -40.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.53-7.30 (m, 5H), 5.53 (s, 1H), 4.83 (dd, *J* = 8.4, 5.7 Hz, 1H), 4.65 (t, *J* = 8.4 Hz, 1H), 4.46 (dd, *J* = 8.3, 5.6 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 158.8, 132.0, 129.3, 128.6, 121.7, 85.9, 85.3, 70.5, 44.4. HRMS (ESI, *m/z*) calcd for C₁₁H₉NO₂Na [M+Na]⁺: 210.0525, found: 210.0530.

Compound 10q



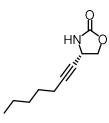
Starting from **9q** (75.0 mg, 0.20 mmol) according to the general procedure to provide **10q** as a white solid (39.5 mg, 91% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak ODH column, ee = 94% (HPLC: 220 nm, *n*–Hexane/isopropanol = 80:20, flow rate 1.0 mL/min, 25 °C, t_r (major) = 15.0 min, t_r (minor) = 16.6 min). $[\alpha]_D^{22}$ = -79.8° (*c* = 1.0, CH₂Cl₂). ¹H **NMR** (300 MHz, CDCl₃) δ 7.41-7.29 (m, 2H), 6.92-6.76 (m, 2H), 5.80 (s, 1H), 4.81 (dd, *J* = 8.4, 5.7 Hz, 1H), 4.62 (t, *J* = 8.3 Hz, 1H), 4.43 (dd, *J* = 8.3, 5.7 Hz, 1H), 3.81 (s, 3H). ¹³C **NMR** (75 MHz, CDCl₃) δ 160.4, 159.0, 133.5, 114.3, 113.7, 85.8, 84.0, 70.6, 55.5, 44.5. **HRMS** (ESI, *m/z*) calcd for C₁₂H₁₁N₁O₃Na [M+Na]⁺: 240.0631, found: 240.0631.

Compound 10r



Starting from **9r** (72.6 mg, 0.20 mmol) according to the general procedure to provide **10r** as a white solid (34.9 mg, 85% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak ODH column, ee = 90% (HPLC: 220 nm, *n*–Hexane/isopropanol = 80:20, flow rate 1.0 mL/min, 25 °C, t_r (major) = 8.3 min, t_r (minor) = 10.8 min). $[\alpha]_{D}^{22}$ = -52.4° (*c* = 1.0, CH₂Cl₂). ¹H **NMR** (300 MHz, CDCl₃) δ 7.56-7.33 (m, 2H), 7.10-6.95 (m, 2H), 5.77 (s, 1H), 4.82 (dd, *J* = 8.4, 5.6 Hz, 1H), 4.64 (t, *J* = 8.4 Hz, 1H), 4.44 (dd, *J* = 8.3, 5.6 Hz, 1H). ¹³C **NMR** (75 MHz, CDCl₃) δ 164.8, 161.5, 159.0, 134.0, 133.9, 117.8, 117.7, 116.1, 115.8, 85.1, 84.8, 70.4, 44.4. ¹⁹F **NMR** (235 MHz, CDCl₃) δ -109.44. **HRMS** (ESI, *m/z*) calcd for C₁₁H₈F₁N₁O₂Na [M+Na]⁺: 228.04319, found: 228.0435.

Compound 10s



Starting from **9s** (67.8 mg, 0.20 mmol) according to the general procedure to provide **10s** as a white solid (30.4mg, 84% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak ODH column, ee = 90% (HPLC: 205 nm, *n*–Hexane/isopropanol = 80:20, flow rate 1.0 mL/min, 25 °C, t_r (major) = 5.2 min, t_r (minor) = 6.3 min). $[\alpha]_D^{22}$ = +3.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.86 (s, 1H), 4.64-4.46 (m, 2H), 4.27 (dd, *J* = 7.3, 5.0 Hz, 1H), 2.17 (td, *J* = 7.1, 1.7 Hz, 2H), 1.58-1.42 (m, 2H), 1.38-1.22 (m, 4H), 0.89 (dd, *J* = 8.2, 5.8 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 159.2, 867.0, 76.7, 70.8, 44.1, 31.2, 28.2, 22.3, 18.7, 14.1. HRMS (ESI, *m/z*) calcd for C₁₀H₁₅N₁O₂Na [M+Na]⁺: 204.0995, found: 204.1000.

Compound (E)-10t

Рĥ

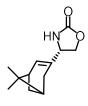
Starting from (*E*)-**9t** (69.4 mg, 0.20 mmol) according to the general procedure to provide **10t** as a white solid (33.3 mg, 88% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak IC column, ee = 63% (HPLC: 210 nm, *n*-Hexane/isopropanol = 30:70, flow rate 1.0 mL/min, 25 °C, t_r (major) = 7.7 min, t_r (minor) = 15.2 min). $[\alpha]_{D}^{22}$ = +91.6° (*c* = 1.0, CH₂Cl₂). ¹H **NMR** (300 MHz, CDCl₃) δ 7.43-7.27 (m, 5H), 6.61 (d, *J* = 15.8 Hz, 1H), 6.13 (dd, *J* = 15.8, 7.6 Hz, 1H), 5.75 (s, 1H), 4.67-4.49 (m, 2H), 4.22-4.05 (m, 1H). ¹³C **NMR** (75 MHz, CDCl₃) δ 159.6, 135.6, 134.1, 128.9, 128.7, 126.9, 126.6, 70.4, 55.3. **HRMS** (ESI, *m/z*) calcd for C₁₁H₁₁NO₂Na [M+Na]⁺: 212.0682, found: 212.0687.

Compound 10u



Starting from **9u** (65.0 mg, 0.20 mmol) according to the general procedure to provide **10u** as a white solid (32.0 mg, 96% yield). Enantiomeric excess was established by HPLC analysis as 92% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 220$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 6.0 min, t_r (minor) = 7.0 min). [α] $_{0}^{22}$ = +14.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.77 (s, 1H), 5.71 (s, 1H), 4.47 (t, *J* = 8.6 Hz, 1H), 4.35-4.24 (m, 1H), 4.08 (dd, *J* = 8.4, 6.1 Hz, 1H), 2.11-1.82 (m, 4H), 1.76-1.48 (m, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 160.1, 135.2, 125.8, 69.5, 58.3, 25.1, 23.3, 22.4, 22.4. HRMS (ESI, *m/z*) calcd for C₉H₁₃N₁O₂Na [M+Na]⁺: 190.0838, found: 190.0844.

Compound 10v



Starting from **9v** (73.0 mg, 0.20 mmol) according to the general procedure to provide **10v** as a white solid (35.9 mg, 87% yield). Enantiomeric excess was established by HPLC analysis as 96:4 d.r. (HPLC: column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 8.7 min, t_r (minor) = 10.1 min). [α] $_{D}^{22}$ = -48.8° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.54 (s, 2H), 4.39 (dt, *J* = 14.4, 8.8 Hz, 2H), 3.95 (dd, *J* = 7.8, 5.6 Hz, 1H), 2.53-2.00 (m, 5H), 1.31 (s, 3H), 1.15 (d, *J* = 8.7 Hz, 1H), 0.80 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 159.9, 145.0, 122.1, 68.3, 56.8, 41.2, 40.9, 38.1, 31.8, 31.3, 26.2, 21.4. HRMS (ESI, *m/z*) calcd for C₁₂H₁₇N₁O₂Na [M+Na]⁺: 230.1152, found: 230.1161.

Compound 10w



Starting from **9w** (57.0 mg, 0.20 mmol) according to the general procedure to provide **10w** as a white solid (20.1 mg, 79% yield). Enantiomeric excess was established by HPLC analysis as 61% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 90:10, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 10.8 min, t_r (minor) = 13.2 min). [α] $_{p}^{22}$ = +38.0° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.99 (s, 1H), 4.98 (d, *J* = 22.4 Hz, 2H), 4.52 (t, *J* = 8.6 Hz, 1H), 4.37 (dd, *J* = 8.8, 6.1 Hz, 1H), 4.08 (dd, *J* = 8.4, 6.0 Hz, 1H), 1.75 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 160.0, 142.5, 113.8, 77.6, 77.2, 76.8, 69.4, 57.8, 17.3. HRMS (ESI, *m/z*) calcd for C₆H₉N₁O₂Na [M+Na]⁺: 150.0525, found: 150.0527.

Compound 10x

Starting from **9x** (70.2 mg, 0.20 mmol) according to the general procedure to provide **10x** as a white solid (38.2 mg, 99% yield). Enantiomeric excess was established by HPLC analysis using a Bz-protected product for the testament as 99% ee (HPLC: column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 8.1 min, t_r (minor) = 9.9 min). [α]_D²² = +14.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.30 (d, *J* = 3.5 Hz, 1H), 3.66 (d, *J* = 1.6 Hz, 1H), 2.28 (s, 2H), 2.22-1.95 (m, 4H), 1.93-1.54 (m, 7H). ¹³C NMR (75 MHz, CDCl₃) δ 161.1, 80.7, 64.2, 40.2, 37.3, 36.5, 36.4, 31.4, 31.2, 29.4, 29.2. HRMS (ESI, *m/z*) calcd for C₁₁H₁₅NO₂Na [M+Na]⁺: 216.0995, found: 216.1000.

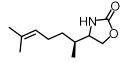
Compound 10y

Starting from **9y** (53.0 mg, 0.20 mmol) according to the general procedure to provide **10y** as a colorless oil (15.0 mg, 53% yield). Enantiomeric excess was established by HPLC analysis using a Bz-protected product for the testament as 80% ee (HPLC: column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 11.5 min, t_r (minor) = 18.7 min). [α]_D²² = -9.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 5.83 (s, 1H), 4.48 (t, *J* = 8.4 Hz, 1H), 4.02 (dd, *J* = 8.4, 6.2 Hz, 1H), 3.93-3.73 (m, 1H), 1.58 (dt, *J* = 12.4, 6.5 Hz, 2H), 1.34 (tdd, *J* = 13.9, 9.4, 4.3 Hz, 4H), 0.91 (t, *J* = 6.9 Hz, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 159.9, 70.5, 52.8, 35.2, 27.5, 22.6, 14.0. All other datas were in agreement with literature report.⁴

Compound 10z

Starting from **9z** (53.0 mg, 0.20 mmol) according to the general procedure to provide **10z** as a colorless oil (17.3 mg, 41% yield). Enantiomeric excess was established by HPLC analysis using a Bz-protected product for the testament as 79% ee (column: Daicel Chiralpak ODH 250 x 4.6 mm, absorption: $\lambda = 210$ nm, mobile phase: *n*-Hexane/isopropanol = 80:20, flow rate: 1.0 mL/min, column temperature: 25 °C, retention times: t_r (major) = 14.3 min, t_r (minor) = 19.9 min). [α]_D²² = -11.2° (*c* = 1.0, CH₂Cl₂). ¹**H** NMR (300 MHz, CDCl₃) δ 5.92-5.62 (m, 2H), 5.07-4.85 (m, 2H), 4.48 (t, *J* = 8.4 Hz, 1H), 4.01 (dd, *J* = 8.5, 6.2 Hz, 1H), 3.90-3.77 (m, 1H), 2.04 (q, *J* = 6.8 Hz, 2H), 1.59 (d, *J* = 14.7 Hz, 2H), 1.44-1.17 (m, 12H). ¹³**C** NMR (75 MHz, CDCl₃) δ 159.9, 139.2, 114.5, 70.5, 52.8, 35.5, 33.9, 29.5, 29.4, 29.1, 29.0, 25.4. **HRMS** (ESI, *m/z*) calcd for C₁₂H₂₁N₁O₂Na [M+Na]⁺: 234.1465, found: 234.1471.

Compound 10za



Starting from **9za** (71.0 mg, 0.20 mmol) according to the general procedure to provide **10za** as a colorless oil (7.1 mg, 18% yield). Diastereomeric ratio was deteremined by H NMR spectrum as > 20:1 d.r. ¹H NMR (300 MHz, CDCl₃) δ 5.55 (s, 1H), 5.18-4.96 (m, 1H), 4.43 (t, *J* = 8.7 Hz, 1H), 4.11 (dd, *J* = 8.7, 6.3 Hz, 1H), 3.70 (dd, *J* = 14.9, 7.1 Hz, 1H), 2.19-1.85 (m, 2H), 1.69 (s, 3H), 1.60 (s, 3H), 1.46-1.35 (m, 1H), 1.16 (dd, *J* = 8.9, 4.7 Hz, 1H), 0.90 (d, *J* = 6.8 Hz, 4H). ¹³C NMR (75 MHz, CDCl₃) δ 159.87, 132.58, 123.74, 68.31, 57.31, 37.00, 32.62, 25.88, 25.14, 17.89, 14.28. All other datas were in agreement with literature report.

4.5.4 Mechanistic Experiments

1) Proposed mechanism

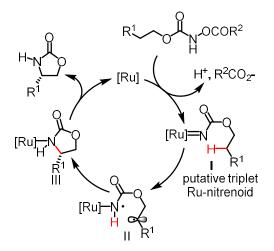
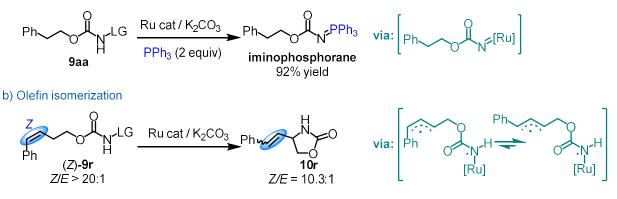


Figure 92. Proposed reaction pathway.

Proposed mechanism: Upon release of benzoic acid from the *N*-benzoyloxycarbamate, the ruthenium catalyst forms a ruthenium nitrenoid intermediate (I). The ruthenium nitrenoid from its triplet state subsequently performs a 1,5-hydrogen atom transfer (HAT) to provide the radical intermediate II. This is followed by C-N bond formation through radical-radical recombination to provide the ruthenium-coordinated product (III), which is released to regenerate the active catalyst for a new catalytic cycle.

2) Mechanistic experiments

a) Nitrene trapping with PPh3



Nitrene trapping with PPh₃: A dry Schlenk tube (10 mL) was charged with substrates (64.2 mg, 0.2 mmol), chiral ruthenium catalyst Λ -Ru13 (0.004 mmol, 2.0 mol%), PPh₃ (105 mg, 0.4 mmol) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Solvent 1,2-dichlorobezene (4.0 mL) was added via syringe. The reaction mixture was stirred at 30 °C for 20 h under an atmosphere of N₂. Afterwards, the mixture was transferred to a column and purified by flash chromatography on silica gel (EtOAc/*n*-Hexane = 1:3 to 1:1) to afford the analytical pure product **iminophosphorane** in 92% yield.

Iminophosphorane: colorless oil. ¹**H NMR** (300 MHz, CD₃CN) δ 7.75-7.59 (m, 9H), 7.56-7.46 (m, 6H), 7.30-7.15 (m, 5H), 4.15 (t, *J* = 6.8 Hz, 2H), 2.84 (t, *J* = 6.8 Hz, 2H). ¹³**C NMR** (75 MHz, CD₃CN) δ 162.5, 140.2, 133.9, 133.7, 133.6, 133.5, 130.1, 129.89, 129.87, 129.7, 129.3, 128.7, 127.1, 66.6, 66.6, 36.4. **HRMS** (ESI, *m/z*) calcd for C₂₇H₂₄N₁O₂P₁H₁ [M+H]⁺: 426.1617, found: 426.1628.

Olefin isomerization: A dry Schlenk tube (10 mL) was charged with substrates (*Z*)-**9b** (0.2 mmol), Λ -**Ru13** (0.002 mmol, 1 mol%) and K₂CO₃ (82.8 mg, 0.6 mmol) under an atmosphere of N₂. Solvent 1,2-dichlorobezene (4.0 mL) was added via syringe. The reaction mixture was stirred at 30 °C for 20 h under an atmosphere of N₂. Afterwards, the crude reaction solution was filtered through a thin layer of celite and washed with 5 mL EtOAc. Filtrate was collected and organic solvent was removed under vacuo. The ratio of (*Z*)-**10r** and (*E*)-**10r** was analyzed by ¹H NMR of crude reaction solutions (see attached spectrum). The ratio of (*Z*)-**10r**:(*E*)-**10r** are 10.3:1.

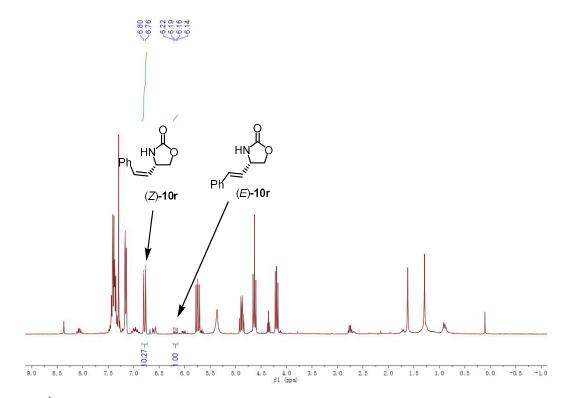


Figure 93. ¹H NMR of crude reaction solutions for analysis of Z/E ratio.

4.5.5 Synthetic Applications

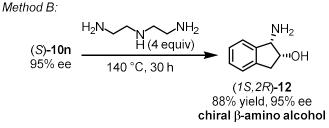
1) Synthesis of chiral β-amino alcohols

Method A:

(S)-10a
> 99% ee
$$\frac{CH_2CI_2, r.t., 16 h}{2}$$
 BocHN OH
2) Cs₂CO₃, MeOH, r.t., 2 h Ph
(S)-11
94% yield, > 99% ee
chiral β-amino alcohol

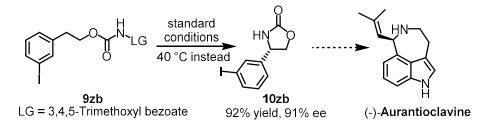
Procedure: To a solution of (*S*)-**10a** (101 mg, 0.62 mmol) and Boc₂O (214 mg, 0.98 mmol) in 16 mL THF was added Et₃N (120 μ L, 0.88 mmol). After stirring at room temperature for 10 min, DMAP (7.6 mg, 10 mol%) was added in one portion. The resulting solution was stirred at room temperature for 16 h. Solvent was removed in *vacuo*. The resulting solid dissolved in 6 mL MeOH was added Cs₂CO₃ (20 mol%). The resulting solution was stirred at room temperature for 2 h. After that, citric acid was added until pH = 7. Solvent was removed in *vacuo*. The residue was subjected to a flash silica gel chromatography (*n*-Hexane/EtOAc= 5:1 to 2:1) to provide analytical pure (*S*)-**11** in 94% yield. Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak IG column, > 99% ee (HPLC: 210 nm, *n*-Hexane/isopropanol = 90:10, flow rate 1.0 mL/min, 25 °C, t_r (major) = 12.1 min). ¹**H NMR** (300 MHz, CDCl₃) δ 7.42-7.26 (m, 5H), 5.22 (d, *J* = 6.5 Hz, 1H), 4.77 (s, 1H),

3.96-3.74 (m, 2H), 2.29 (s, 1H), 1.43 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 156.3, 139.7, 129.0, 128.0, 126.8, 80.2, 67.2, 57.2, 28.5.



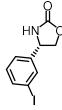
Procedure: (*3S*,*8R*)-**10n** (35 mg) in diethylenetriamine (86.4 µL, 4 equiv) was heated at 140 °C for 30 h. Afterwards, it was directly subjected to a flash silica gel chromatography (CH₂Cl₂/MeOH/Et₃N= 100:10:1) to provide analytical pure (*IS*,*2R*)-**13** as light pink solid in 88% yield. Enantiomeric excess was determined by transforming back to corresponding carbamate as 95% ee. ¹H NMR (300 MHz, CDCl₃) δ 7.42-7.18 (m, 4H), 4.52-4.23 (m, 2H), 3.12 (dd, *J* = 16.4, 5.4 Hz, 1H), 2.97 (dd, *J* = 16.4, 2.7 Hz, 1H), 2.81 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 144.1, 141.1, 128.2, 127.1, 125.6, 124.1, 72.9, 58.7, 39.6.

2) Synthesis of Natural Products



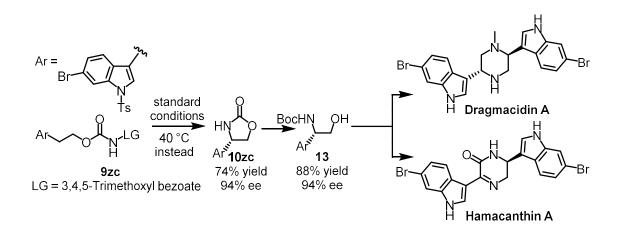
(S)-10zb was synthesized according to the general produce shown before.

Compound 10zb



Starting from **9zb** (100.2 mg, 0.20 mmol) according to the general procedure to provide **10zb** as a white solid (53.0 mg, 92% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak ODH column, 91% ee (HPLC: 220 nm, *n*–Hexane/isopropanol = 60:40, flow rate 1.0

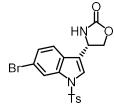
mL/min, 25 °C, t_r (major) = 13.7 min, t_r (minor) = 9.1 min). $[\alpha]_D^{22}$ = +39.2° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 7.74-7.65 (m, 2H), 7.31 (d, *J* = 7.8 Hz, 1H), 7.14 (t, *J* = 8.0 Hz, 1H), 6.15 (s, 1H), 4.99-4.80 (m, 1H), 4.72 (t, *J* = 8.7 Hz, 1H), 4.15 (dd, *J* = 8.6, 6.7 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 159.6, 142.0, 138.2, 135.3, 131.1, 125.4, 95.1, 72.4, 55.8. HRMS (ESI, *m/z*) calcd for C₉H₈I₁N₁O₂Na [M+Na]⁺: 311.9492, found: 311.9503.



(S)-10zc was synthesized according to the general produre shown before.

(S)-13 was synthesized according to the general produce shown before (Method A).

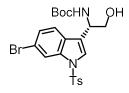
Compound 10zc



Starting from **9zc** (64.6 mg, 0.10 mmol) according to the general procedure to provide **10zc** as a white solid (32.1 mg, 74% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak IA column, 94% ee (HPLC: 254 nm, *n*–Hexane/isopropanol = 85:15, flow rate 1.0 mL/min, 25 °C, t_r (major) = 17.4 min, t_r (minor) = 20.8 min). $[\alpha]_D^{22}$ = +18.6° (*c* = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.18 (s, 1H), 7.79 (d, *J* = 8.3 Hz, 2H), 7.55 (s, 1H), 7.38 (s, 2H), 7.29 (d, *J* = 8.2 Hz, 2H), 5.76 (s, 1H), 5.15 (dd, *J* = 8.6, 6.6 Hz, 1H), 4.74 (t, *J* = 8.8 Hz, 1H), 4.31 (dd, *J* = 8.6, 6.4 Hz, 1H), 2.38 (s, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 159.1, 146.0, 136.5, 134.9, 130.5, 127.3, 127.2, 126.4, 124.5, 120.7, 120.4, 119.6, 117.3, 70.4, 49.5, 21.8. HRMS (ESI, *m/z*) calcd for C₁₈H₁₆Br₁N₂O₄S₁H₁

[M+H]⁺: 435.0009 & 436.9989, found: 435.0017 & 436.9997.

Compound 13



Starting from **10zc** (30.4 mg, 0.07 mmol) according to the general procedure to provide **13** as a white solid (31.2 mg, 88% yield). Enantiomeric excess was established by HPLC analysis by using a Daicel Chiralpak IA column, 94% ee (HPLC: 210 nm, *n*–Hexane/isopropanol = 85:15, flow rate 1.0 mL/min, 25 °C, t_r (major) = 9.7 min, t_r (minor) = 13.7 min). $[\alpha]_{D}^{22} = +20.9^{\circ}$ (c = 1.0, CH₂Cl₂). ¹H NMR (300 MHz, CDCl₃) δ 8.15 (d, J = 1.1 Hz, 1H), 7.75 (d, J = 8.3 Hz, 2H), 7.54 (s, 1H), 7.43 (d, J = 8.5 Hz, 1H), 7.35 (dd, J = 8.5, 1.6 Hz, 1H), 7.26 (t, J = 4.0 Hz, 3H), 5.07 (t, J = 14.2 Hz, 2H), 3.95 (s, 2H), 2.36 (s, 3H), 2.20 (s, 1H), 1.44 (s, 9H). ¹³C NMR (75 MHz, CDCl₃) δ 155.9, 145.6, 136.1, 135.1, 130.3, 128.2, 127.1, 127.0, 124.3, 121.2, 119.0, 117.0, 80.45, 65.1, 28.5, 21.8. HRMS (ESI, *m/z*) calcd for C₂₂H₂₅Br₁N₂O₅S₁Na₁ [M+Na]⁺: 531.0560 & 533.0541, found: 531.0571 & 533.0550.

4.5.6 Single Crystal X-Ray Diffraction Studies

Crystallography of compound (*S*)-10i: Single crystals of (*S*)-10i were obtained by slow diffusion from the solution in EtOAc layered with *n*-Hexane at room temperature. A suitable crystal of $C_9H_8BrNO_2$ was selected under inert oil and mounted using a MiTeGen loop. Intensity data of the crystal were recorded with a D8 Quest diffractometer (Bruker AXS). The instrument was operated with Mo-K α radiation (0.71073 Å, microfocus source) and equipped with a PHOTON 100 detector. Evaluation, integration and reduction of the diffraction data was carried out using the Bruker APEX 3 software suite.¹ Multi-scan and numerical absorption corrections were applied using the SADABS program.^{2,3} The structure was solved using dual-space methods (SHELXT-2014/5) and refined against F^2 (SHELXL-2018/3 using ShelXle interface).⁴⁻⁶ All non–Hydrogen atoms were refined with anisotropic displacement parameters. The hydrogen atoms attached to carbon atoms were refined using the "riding model" approach with isotropic displacement parameters 1.2 times of that of the preceding carbon atom. The hydrogen atom at the nitrogen atom was refined using a restraint on the bond length to prevent too strong underestimation of the atom distance. CCDC 2012187 contains the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/structures.

	, , ,
Identification code	(<i>S</i>)-10i
Empirical formula	C ₉ H ₈ BrNO ₂
Molar mass / $g \cdot mol^{-1}$	242.07
Space group (No.)	$P2_{1}2_{1}2_{1}$ (19)
<i>a</i> / Å	5.6166(3)
b / Å	7.7164(4)
<i>c</i> / Å	21.6106(11)
$V/\text{\AA}^3$	936.60(8)
Ζ	4
$ ho_{calc.}$ / g·cm ⁻³	1.717
μ / mm^{-1}	4.353
Color	colorless
Crystal habitus	block
Crystal size / mm ³	0.265 x 0.239 x 0.168
Т / К	100
$\lambda / \text{\AA}$	0.71073 (Mo-K _α)
heta range / °	2.803 to 31.638
Range of Miller indices	$-8 \le h \le 8$
	$-11 \le k \le 11$
	$-31 \le l \le 31$
Absorption correction	multi-scan and numerical
T_{\min}, T_{\max}	0.4622, 0.5826
$R_{ m int}, R_{\sigma}$	0.0210, 0.0160
Completeness of the data set	0.999
No. of measured reflections	17155
No. of independent reflections	3139
No. of parameters	123
No. of restrains	1
S (all data)	1.091
$R(F)$ ($I \ge 2\sigma(I)$, all data)	0.0171, 0.0188
$wR(F^2)$ ($I \ge 2\sigma(I)$, all data)	0.0405, 0.0412
Extinction coefficient	0.0127(10)
Flack parameter <i>x</i>	-0.006(3)
$\Delta ho_{ m max}, \Delta ho_{ m min}$ / e·Å ⁻³	0.336, -0.238

Table 10. Selected crystallographic data and details of the structure determination for C₉H₈BrNO₂.

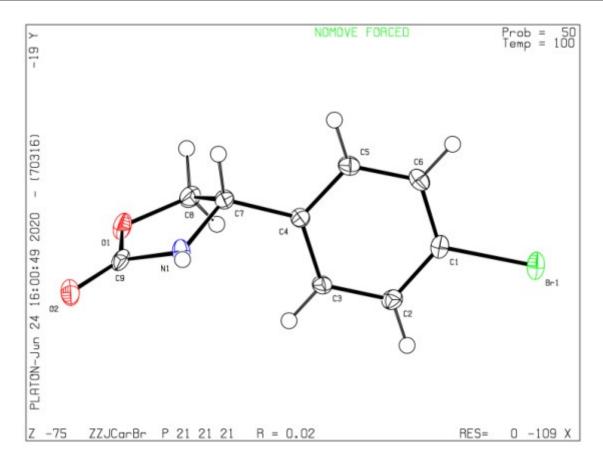


Figure 94. Crystal structure of compound (S)-10i.

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Chapter 5: Appendices

5.1 List of Abbreviations

¹ H NMR	proton nuclear magnetic resonance spectroscopy
¹³ C NMR	carbon nuclear magnetic resonance spectroscopy
⁹ F NMR	fluorine nuclear magnetic resonance spectroscopy
δ	chemical shift
J	coupling constant
br	broad
S	singlet
d	doublet
t	triplet
q	quartet
m	multiplet
ppm	parts per million
AcOH	acetic acid
aq	aqueous
Ar	Aryl-group
bpy	2,2'-bipyridine
CD	circular dichroism
CH ₂ Cl ₂ /DCM	dichloromethane
CD_2Cl_2	dideuteromethylenechloride
CHCl ₃	chloroform
CDCl ₃	deuterochloroform
CH ₃ CN/ MeCN	acetonitrile
conc	concentrated
DMAP	4-dimethylaminopyridine
DMF	dimethylformamide
DMSO	dimethyl sulfoxide
dr	diastereomeric ratio

ee	enantiomeric excesses
et al.	et alii (lat.: and others)
EtOH	ethanol
Et ₂ O	diethyl ether
Et ₃ N	triethyl amine
EtOAc	ethyl acetate
EDG	electron donating group
EWG	electron withdrawing group
HAT	hydrogen atom transfer
h	hour(s)
HPLC	high performance liquid chromatography
HRMS	high resolution mass spectrometry
IR spectra	infrared spectra
Ir	iridium
Rh	rhodium
Ru	ruthenium
Fe	iron
	••
Ag	silver
Ag Mn	silver manganese
-	
Mn	manganese
Mn L	manganese liter(s)
Mn L M	manganese liter(s) mol/liter
Mn L M m	manganese liter(s) mol/liter meta-
Mn L M m min	manganese liter(s) mol/liter meta- minute(s)
Mn L M <i>m</i> min mL	manganese liter(s) mol/liter meta- minute(s) milliliter(s)
Mn L M m min mL mmol	manganese liter(s) mol/liter meta- minute(s) milliliter(s) millimole
Mn L M m min mL mmol MS	manganese liter(s) mol/liter meta- minute(s) milliliter(s) millimole mass spectroscopy
Mn L M m min mL mmol MS N ₂	manganese liter(s) mol/liter meta- minute(s) milliliter(s) millimole mass spectroscopy nitrogen

rac	racemate
rt	room temperature
TFA	trifluoroacetic acid
THF	tetrahydrofuran
TLC	thin layer chromatography

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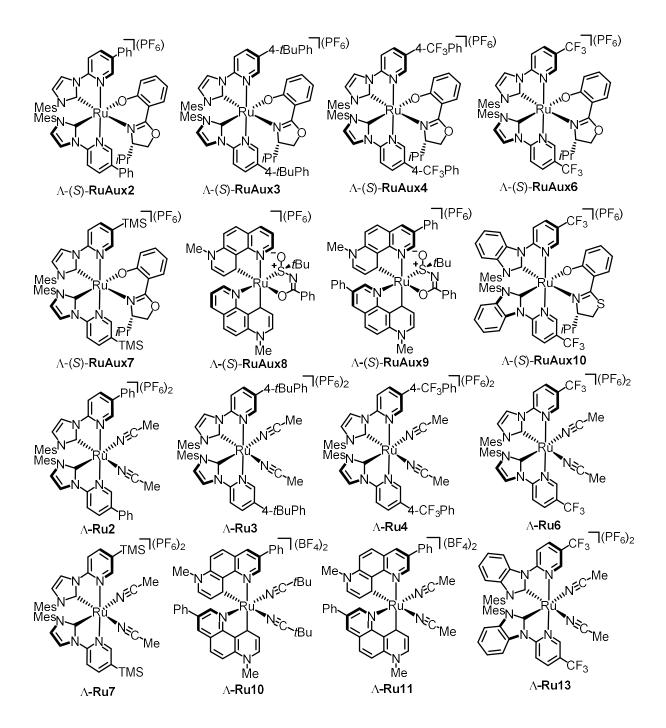
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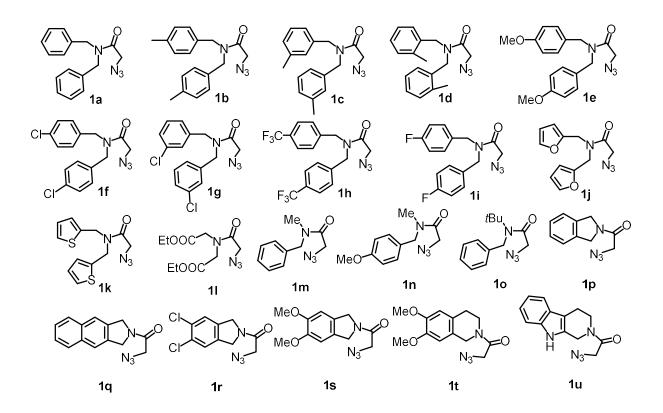
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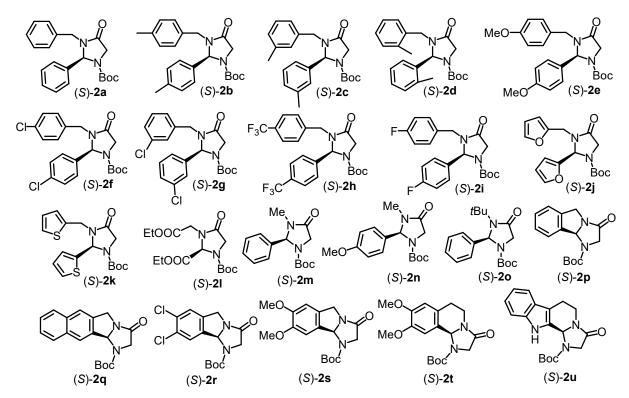
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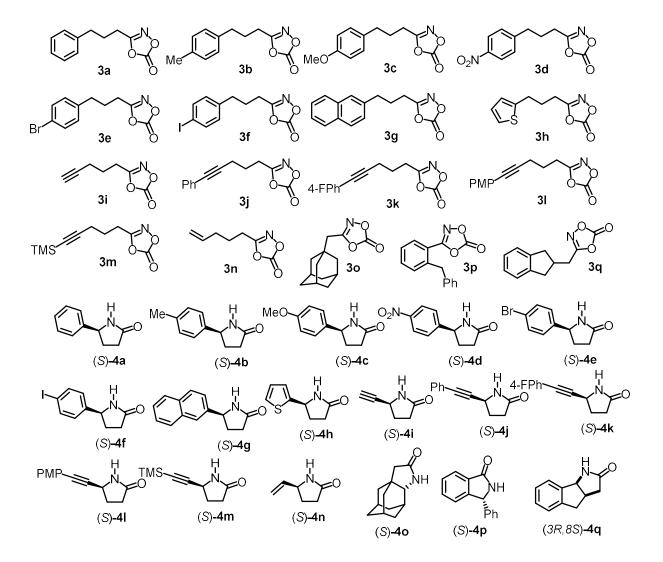


2) New Organic Compounds

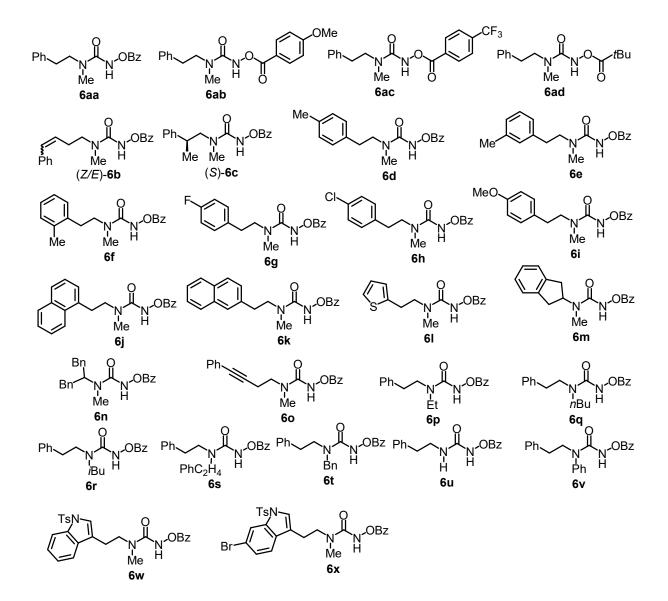
Chapter 2.1 and its Experimental Part



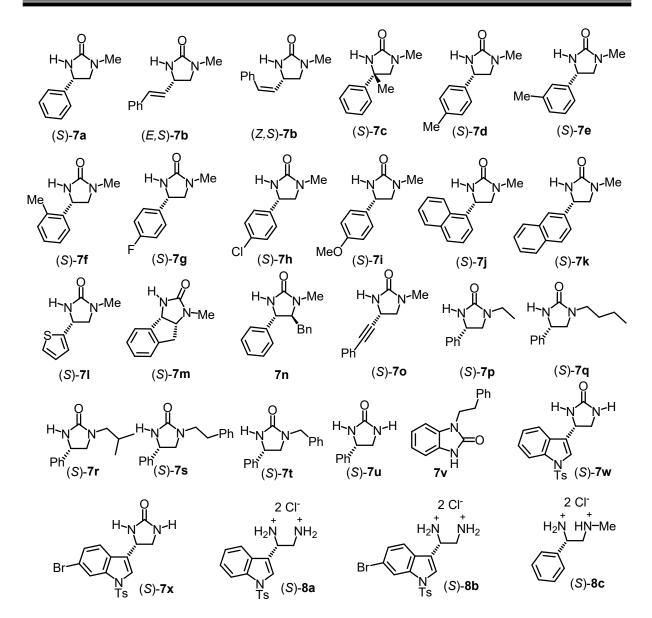


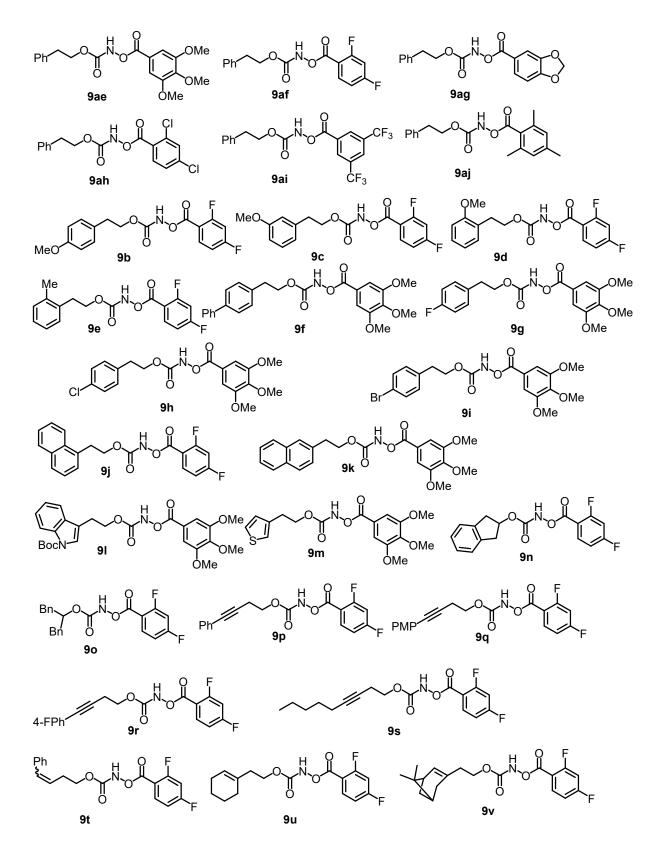


Chapter 2.2 and its Experimental Part

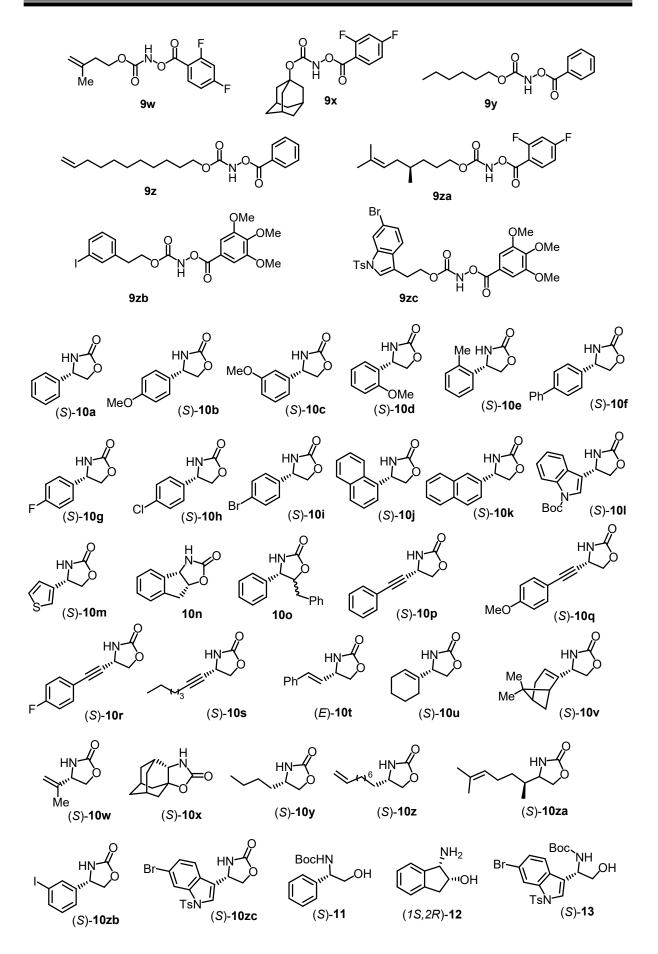


Chapter 2.3 and its Experimental Part





Chapter 2.4 and its Experimental Part



Statement

gemäß § 10, Abs. 1 der Promotionsordnung der mathematisch-naturwissenschaftlichen Fachbereiche und des Medizinischen Fachbereichs für seine mathematischnaturwissenschaftlichen Fächer der Philipps-Universität Marburg vom 15.Oct.2020

Ich erkläre, dass eine Promotion noch an keiner anderen Hochschule als der Philipps-Universität Marburg, Fachbereich Chemie, versucht wurde und versichere, dass ich meine vorgelegte Dissertation

Asymmetric Intramolecular C-H Aminations with Chiral-at-Ruthenium

Complexes

selbst und ohne fremde Hilfe verfasst, nicht andere als die in ihr angegebenen Quellen oder Hilfsmittl benutz, alle vollständig oder sinngemäß übernommenen Zitate als solche gekennzeichnet sowie die Dissertation in der vorliegenden oder ähnlichen Form noch bei keiner anderen in- oder ausländischen Hochschule anlässlich eines Promotionsgesuchs oder zu anderen Prüfungszwecken eingereicht habe.

Zijun Zhou Marburg, den 15.Oct.2020

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- Z. Zhou, Y. Tan, T. Yamahira, S. Ivlev, X. Xie, R. Riedel, M. Hemming, M. Kimura, E. Meggers, Chem 2020, 6, 2024.
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